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Author(s): Katie Hancock , Anne M. Zajac , Francois Elvinger , and David S. Lindsay

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- , D. J. FORRESTER, AND E. C. GREINER. 1987. Pathogenicity of *Haemoproteus meleagridis* (Haemosporina: Haemoproteidae) in experimentally infected domestic turkeys. *Journal of Parasitology* **74**: 228–239.
- DIGGS, C. L., AND A. G. OSLER. 1969. Humoral immunity in rodent malaria. Estimation of parasitemia by electronic particle counting. *Journal of Immunology* **102**: 292–297.
- DI RUBERTO, C., A. DEMPSTER, S. KHAN, AND B. JARRA. 2002. Analysis of infected blood cell images using morphological operators. *Image and Vision Computing* **20**: 133–146.
- FEDYNICH, A. M., D. B. PENCE, AND R. D. GODFREY JR. 1995. Hematozoa in thin blood smears. *Journal of Wildlife Diseases* **31**: 436–438.
- GODFREY, R. D. JR., A. M. FEDYNICH, AND D. B. PENCE. 1987. Quantification of hematozoa in blood smears. *Journal of Wildlife Diseases* **23**: 558–565.
- JARVI, S. I., J. J. SCHULTZ, AND C. T. ATKINSON. 2002. PCR diagnostics underestimate the prevalence of avian malaria (*Plasmodium relictum*) in experimentally-infected passerines. *Journal of Parasitology* **88**: 153–158.
- MCKENZIE, F. E., AND W. H. BOSSERT. 1997. The dynamics of *Plasmodium falciparum* blood-stage infection. *Journal of Theoretical Biology* **188**: 127–140.
- MERINO, S., J. MORENO, J. J. SANZ, AND E. ARRIERO. 2000. Are avian blood parasites pathogenic in the wild? A medication experiment in blue tits (*Parus caeruleus*). *Proceedings of the Royal Society of London B* **267**: 2507–2510.
- RASBAND, W. 2002. ImageJ [PC program], version 1.25s. National Institutes of Health, Bethesda, Maryland.
- RIDLER, T., AND S. CALVARD. 1978. Picture thresholding using an iterative selection method. *IEEE Transactions on Systems, Man and Cybernetics* **8**: 630–632.

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## Prevalence of Agglutinating Antibodies to *Sarcocystis neurona* in Raccoons (*Procyon lotor*) From an Urban Area of Virginia

Katie Hancock, Anne M. Zajac, Francois Elvinger\*, and David S. Lindsay†, Center for Molecular Medicine and Infectious Diseases, Department of Biomedical Sciences and Pathobiology, Virginia–Maryland Regional College of Veterinary Medicine, Virginia Tech, 1410 Prices Fork Road, Blacksburg, Virginia 24061-0342; \*Department of Large Animal Clinical Sciences, Virginia–Maryland Regional College of Veterinary Medicine, Virginia Tech, 1410 Prices Fork Road, Blacksburg, Virginia 24061-0342; †To whom correspondence should be addressed. e-mail: [lindsayd@vt.edu](mailto:lindsayd@vt.edu)

**ABSTRACT:** Equine protozoal myeloencephalitis is the most important protozoan disease of horses in North America and is usually caused by *Sarcocystis neurona*. Natural and experimentally induced cases of encephalitis caused by *S. neurona* have been reported in raccoons (*Procyon lotor*) and raccoons are an intermediate host for this parasite. A 3-yr-long serological survey was conducted to determine the prevalence of agglutinating antibodies to *S. neurona* in raccoons collected from Fairfax County, Virginia, a suburban–urban area outside Washington, D.C. Samples from 469 raccoons were examined, and agglutinating antibodies ( $\geq 1:50$  dilution) were found in 433 (92.3%) of the raccoons. This study indicates that exposure to *S. neurona* is high in this metropolitan area.

Equine protozoal myeloencephalitis is a neurologic disease in horses from the Americas and is usually caused by infection with the apicomplexan parasite *Sarcocystis neurona* (Dubey et al., 1991). It is the most important protozoan disease of horses in North America (reviewed by Dubey, Lindsay, Saville et al., 2001). The Virginia opossum *Didelphis virginiana* is the only known definitive host in North America (Dubey and Lindsay, 1998), whereas *Didelphis albiventris* is a host in South America (Dubey, Lindsay, Kerber et al., 2001). Nine-banded armadillos (*Dasypus novemcinctus*), raccoons (*Procyon lotor*), and sea otters (*Enhydra lutris*) are natural intermediate hosts (Cheadle, Tanhauser et al., 2001; Dubey, Rosypal et al., 2001; Dubey, Saville et al., 2001; Tanhauser et al., 2001). Domestic cats (*Felis domesticus*), striped skunks (*Mephitis mephitis*), and raccoons are known experimental intermediate hosts (Dubey et al., 2000; Cheadle, Yowell et al., 2001; Dubey, Saville et al., 2001).

Dubey, Saville et al. (2001) reported clinical encephalitis in experimentally infected raccoons. Stanek et al. (2002) described the life cycle of *S. neurona* in experimentally infected raccoons.

This study was conducted to determine the serological prevalence of antibodies to *S. neurona* in a common intermediate host, the raccoon. The direct *S. neurona* agglutination test (SAT) described by Lindsay and Dubey (2001) was used.

Raccoons originated in various locations in Fairfax County, Virginia, a suburban–urban area outside Washington, D.C. Raccoons used for this study were livetrapped as part of a larger study on rabies in Fairfax County. Blood samples were collected from all trapped raccoons. Rac-

coons were released immediately after sampling was completed. The serum was collected, placed in a tube, and frozen at  $-70$  C. Frozen sera were sent to the Center for Molecular Medicine and Infectious Diseases, Department of Biomedical Sciences and Pathobiology, Virginia–Maryland Regional College of Veterinary Medicine, Virginia Tech, Blacksburg, Virginia for agglutination testing. Samples from 137, 120, and 212 raccoons from 2000, 2001, and 2002, respectively, were examined (total = 469 for 3 yr). The SAT was used to test sera at dilutions of 1:50. The SAT has previously been validated using sera from experimentally infected raccoons (Dubey, Saville et al., 2001; Lindsay et al., 2001; Stanek et al., 2002).

The prevalence of antibodies in 2000 was 128 (93.4%) of the 137 samples, in 2001 it was 109 (90.8%) of 120 samples, and in 2002 it was 196 (92.5%) of 212 samples. The total prevalence of antibodies in the 469 raccoons was 93.3% (N = 433 positives).

This study is the largest conducted to date on the prevalence of *S. neurona* antibodies in raccoons. Mitchell et al. (2002) examined sera from 12 raccoons from Connecticut and found that all 12 were positive in the SAT. Lindsay et al. (2001) demonstrated that sera from 33% of raccoons (n = 24) from Florida, 72% of raccoons (n = 25) from New Jersey, 52% of raccoons (n = 25) from Pennsylvania, and 96% of raccoons (n = 25) from Massachusetts were positive in the SAT.

Clinical *Sarcocystis* sp.-associated encephalitis and myocarditis are common in raccoons (Stoffregen and Dubey, 1991; Thulin et al., 1992; Dubey, Hamir et al., 1990; Hamir and Dubey, 2001). These animals are often coinfecting with canine distemper (Stoffregen and Dubey, 1992; Thulin et al., 1992). Experimental infection of raccoons with *S. neurona* is also potentially pathogenic for raccoons (Dubey, Saville et al., 2001; Stanek et al., 2002). *Sarcocystis kirkpatricki* is the only other named species of *Sarcocystis* infecting the muscles of raccoons (Snyder et al., 1990). Kirkpatrick et al. (1987) found sarcocysts in 26 (50%) of 52 raccoons from Ohio, Pennsylvania, Florida, and Maryland. Snyder et al. (1990) found *S. kirkpatricki* sarcocysts in 66 of 100 raccoons examined from Illinois. Demonstration of sarcocysts in tissues is not as accurate as acid–pepsin digestion of tissues or serological methods, and this method usually underestimates prevalence. The life cycle of *S. kirkpatricki* is not known. Molecular studies are needed to determine the taxonomic relationship between *S. neurona* and *S. kirkpatricki*.

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#### LITERATURE CITED

- CHEADLE, M. A., S. M. TANHAUSER, J. B. DAME, D. C. SELTON, D. HINES, P. E. GINN, R. J. MACKAY, AND E. C. GREINER. 2001. The nine-banded armadillo (*Dasypus novemcinctus*) is an intermediate host for *Sarcocystis neurona*. *International Journal for Parasitology* **31**: 330–335.
- , C. A. YOWELL, D. C. SELTON, M. HINES, P. E. GINN, A. E. MARSH, J. B. DAME, AND E. C. GREINER. 2001. The striped skunk (*Mephitis mephitis*) is an intermediate host for *Sarcocystis neurona*. *International Journal for Parasitology* **31**: 843–849.
- DUBEY, J. P., AND D. S. LINDSAY. 1998. Isolation of *Sarcocystis neurona* from opossum (*Didelphis virginiana*) faeces in immunodeficient mice and its differentiation from *Sarcocystis falcatula*. *International Journal for Parasitology* **28**: 1823–1828.
- , S. W. DAVIS, C. A. SPEER, D. D. BOWMAN, A. DE LAHUNTA, D. E. GRANSTROM, M. J. TOPPER, A. N. HAMIR, AND M. M. SUTER. 1991. *Sarcocystis neurona* n. sp. (Protozoa: Apicomplexa), the etiologic agent of equine protozoal myeloencephalitis. *Journal of Parasitology* **77**: 212–218.
- , A. N. HAMIR, C. A. HANLON, M. J. TOPPER, AND C. E. RUPPRECHT. 1990. Fatal necrotizing encephalitis in a raccoon associated with a *Sarcocystis*-like protozoon. *Journal of Veterinary Diagnostic Investigation* **2**: 345–347.
- , D. S. LINDSAY, C. E. KERBER, N. KASAI, H. F. T. PENA, O. C. H. KWOK, S. K. SHEN, AND B. M. ROSENTHAL. 2001. First isolation of *Sarcocystis neurona* from the South American opossum, *Didelphis albiventris*, from Brazil. *Veterinary Parasitology* **95**: 295–304.
- , D. S. LINDSAY, W. J. A. SAVILLE, S. M. REED, D. E. GRANSTROM, AND C. A. SPEER. 2001. A review of *Sarcocystis neurona* and equine protozoal myeloencephalitis (EPM). *Veterinary Parasitology* **95**: 89–132.
- , A. C. ROSYPAL, B. M. ROSENTHAL, N. J. THOMAS, D. S. LINDSAY, J. F. STANEK, W. J. A. SAVILLE, AND S. M. REED. 2001. *Sarcocystis neurona* infections in Sea otters (*Enhydra lutris nereis*): Evidence for natural infections with sarcocysts and transmission of infection to opossums (*Didelphis virginiana*). *Journal of Parasitology* **87**: 1387–1393.
- , W. J. A. SAVILLE, D. S. LINDSAY, R. W. STICH, J. F. STANEK, C. A. SPEER, B. M. ROSENTHAL, C. J. NJOKU, S. K. SHEN, AND S. M. REED. 2000. Completion of the life cycle of *Sarcocystis neurona*. *Journal of Parasitology* **86**: 1276–1280.
- , W. J. A. SAVILLE, J. F. STANEK, D. S. LINDSAY, B. M. ROSENTHAL, M. OLGLESBEE, A. C. ROSYPAL, C. J. NJOKU, R. W. STICH, O. C. H. KWOK, S. K. SHEN, A. N. HAMIR, AND S. M. REED. 2001. *Sarcocystis neurona* infections in raccoons (*Procyon lotor*): Evidence for natural infection with sarcocysts, transmission of infection to opossums (*Didelphis virginiana*), and experimental induction of neurologic disease in raccoons. *Veterinary Parasitology* **100**: 117–129.
- HAMIR, A. N., AND J. P. DUBEY. 2001. Myocarditis and encephalitis associated with *Sarcocystis neurona* infection in raccoons (*Procyon lotor*). *Veterinary Parasitology* **95**: 335–340.
- KIRKPATRICK, C. E., A. N. HAMIR, J. P. DUBEY, AND C. E. RUPPRECHT. 1987. *Sarcocystis* in muscles of raccoons (*Procyon lotor* L.). *Journal of Protozoology* **34**: 445–447.
- LINDSAY, D. S., AND J. P. DUBEY. 2001. Direct agglutination test for the detection of antibodies to *Sarcocystis neurona* in experimentally infected animals. *Veterinary Parasitology* **95**: 179–186.
- , A. C. ROSYPAL, J. A. SPENCER, M. A. CHEADLE, A. M. ZAJAC, C. RUPPRECHT, J. P. DUBEY, AND B. L. BLAGBURN. 2001. Prevalence of agglutinating antibodies to *Sarcocystis neurona* in raccoons, *Procyon lotor*, from the United States. *Veterinary Parasitology* **100**: 131–134.
- MITCHELL, S. M., D. J. RICHARDSON, M. A. CHEADLE, A. M. ZAJAC, AND D. S. LINDSAY. 2002. Prevalence of agglutinating antibodies to *Sarcocystis neurona* in skunks (*Mephitis mephitis*), raccoons, (*Procyon lotor*), and opossums (*Didelphis virginiana*) from Connecticut. *Journal of Parasitology* **88**: 1027–1029.
- SNYDER, D. E., G. C. SANDERSON, M. TOIVIO-KINNUCAN, AND B. L. BLAGBURN. 1990. *Sarcocystis kirkpatricki* n. sp. (Apicomplexa: Sarcocystidae) in muscles of raccoons (*Procyon lotor*) from Illinois. *Journal of Parasitology* **76**: 495–500.
- STANEK, J. F., J. P. DUBEY, M. J. OLGLESBEE, S. M. REED, D. S. LINDSAY, L. A. CAPITINI, C. J. NJOKU, K. L. VITITOW, AND W. J. A. SAVILLE. 2002. Life cycle of *Sarcocystis neurona* in its natural intermediate host, the raccoon, *Procyon lotor*. *Journal of Parasitology* **88**: 1151–1158.
- STOFFREGEN, D. A., AND J. P. DUBEY. 1991. A *Sarcocystis* sp.-like protozoan and concurrent canine distemper virus infection associated with encephalitis in a raccoon (*Procyon lotor*). *Journal of Wildlife Disease* **27**: 688–692.
- TANHAUSER, S. M., M. A. CHEADLE, E. T. MASSEY, B. A. MAYER, D. E. SCHROEDTER, J. B. DAME, E. C. GREINER, AND R. J. MACKAY. 2001. The nine-banded armadillo (*Dasypus novemcinctus*) is naturally infected with *Sarcocystis neurona*. *International Journal for Parasitology* **31**: 325–329.
- THULIN, J. D., D. E. GRANSTROM, H. B. GELBERG, D. G. MORTON, R. A. FRENCH, AND R. C. GILES. 1992. Concurrent protozoal encephalitis and canine distemper virus infection in a raccoon (*Procyon lotor*). *Veterinary Record* **130**: 162–164.

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## *Parvicapsula minibicornis* in Anadromous Sockeye (*Oncorhynchus nerka*) and Coho (*Oncorhynchus kisutch*) Salmon From Tributaries of the Columbia River

Simon Jones, Gina Proserpi-Porta, Sheila Dawe, Kimberley Taylor, and Benjamin Goh, Department of Fisheries and Oceans, Pacific Biological Station, 3190 Hammond Bay Road, Nanaimo, British Columbia, V9T 6N7 Canada. e-mail: jones@pac.dfo-mpo.gc.ca

**ABSTRACT:** The myxosporean parasite *Parvicapsula minibicornis* is described from adult sockeye and coho salmon during spawning migrations in tributaries of the Columbia River in Canada and the United States. These observations extend the known distribution of this parasite from the Fraser River drainage basin. The parasite was identified in Columbia River salmonids using polymerase chain reaction (PCR) and by in situ hybridization, but unlike in Fraser River salmon, it was not observed in conventional histological preparations of the kidney. Prevalence of the parasite determined by PCR was higher in spawning sockeye from the Fraser River than in those from the Okanagan River. Our ability to explain the relatively low prevalence and absence of clinical

*P. minibicornis* infections in Columbia River salmon is hampered by our poor understanding of the life cycle of this parasite.

The myxosporean parasite *Parvicapsula minibicornis* was first described from the kidney of adult sockeye salmon (*Oncorhynchus nerka*) spawning at Weaver Creek, a tributary of the Fraser River in British Columbia, Canada (Kent et al., 1997). Pathological changes of the kidney were associated with *P. minibicornis* (Raverty et al., 2000; St-Hilaire et al., 2002), suggesting that severe infections may contribute to prespawn mortality observed among prematurely migrating sockeye