

LIST OF REFERENCES

1. M.B. Sporn, A.B. Roberts, (1992) "Autocrine Secretion-10 Years Later". *Ann. Int. Med.* **117**, 408-414.
2. B. Cercek, M.C. Fishbein, J.S. Forrester, R.H. Helfant, J.A. Fagin, (1990) "Induction of Insulin-Like Growth Factor I in Messenger RNA in Rat Aorta After Balloon Denudation". *Circ. Res.* **66**, 1757-1760.
3. S. Buisguina, J.A. Chowen, J. Argente, I. Torres-Aleman, (1996) "Specific Alterations of the Insulin-Like Growth Factor I System in the Cerebellum of Diabetic Rats". *Endocrinology* **137**, 4980-4987.
4. D. LeRoith, H. Werner, D. Beitner-Johnson, C.T. Roberts, Jr., (1995) "Molecular and Cellular Aspects of the Insulin-Like Growth Factor I Receptor". *Endo. Rev.* **16**, 143-163.
5. L. Stryer, (1988) *Biochemistry*, 3rd ed., W.H. Freeman and Company, New York, 5-6, 15-16, 177-200, 288-297
6. H. Lodish, D. Baltimore, A. Berk, S.L. Zipursky, P. Matsudaira, J. Darnell, (1995) *Molecular Cell Biology*, 3rd ed., Scientific American Books, Inc., New York, 853-924, 1136-1142, 1253-1261
7. M.B. Sporn, A.B. Roberts, (1990) "TGF- β : Problems and Prospects". *Cell Regul.* **1**, 875-882.
8. V.A. Blakesley, A. Scrimgeour, D. Esposito, D. LeRoith, (1996) "Signaling Via the Insulin-Like Growth Factor-I Receptor: Does It Differ From Insulin Receptor Signaling". *Cyto. Grow. Fact. Rev.* **7**, 153-159.
9. I.J. Mason, (1994) "The Ins and Outs of Fibroblast Growth Factors". *Cell* **78**, 547-552.
10. J.R. Singleton, V.M. Dixit, E.L. Feldman, (1996) "Type I Insulin-Like Growth Factor Receptor Activation Regulates Apoptotic Proteins". *J. Biol. Chem.* **271**, 31791-31794.
11. D.A. Lauffenberger, L. Chu, A. French, G. Oehrtman, C. Reddy, A. Wells, S. Niyogi, H.S. Wiley, (1996) "Engineering Dynamics of Growth Factors and Other Therapeutic Ligands". *Biotech. Bioeng.* **52**, 61-80.

12. R. Flaumenhaft, D.B. Rifkin, (1992) "The Extracellular Regulation of Growth Factor Action". *Mol. Biol. Cell* **3**, 1057-1065.
13. M.A. Cascieri, M.L. Bayne, (1988) "Serum Half-life and Biological Activity of Mutants of Human Insulin-Like Growth Factor I Which Do Not Bind to Serum Binding Proteins". *Endocrinology* **123**, 373-381.
14. A.C. Rapraeger, (1993) "The Coordinated Regulation of Heparan Sulfate, Syndecans and Cell Behavior". *Curr. Opin. Cell Biol.* **5**, 844-853.
15. E. Ruoslahti, Y. Yamaguchi, (1991) "Proteoglycans as Modulators of Growth Factor Activities". *Cell* **64**, 867-869.
16. M.A. Nugent, E.R. Edelman, (1992) "Kinetics of Basic Fibroblast Growth Factor Binding to Its Receptor and Heparan Sulfate Proteoglycan: A Mechanism for Cooperativity". *Am. Chem. Soc.* **31**, 8876-8883.
17. S.C. Thornton, S.N. Mueller, E.M. Levine, (1983). "Human Endothelial Cells: Use of Purified Heparin in Cloning and Long-term Serial Cultivation". *Science* **222**, 623-625.
18. A.B. Schreiber, J. Kenney, W.J. Kowalski, R. Friesel, T. Mehlman, T. Maciag, (1985) "Interaction of Endothelial Cell Growth Factor with Heparin: Characterization By Receptor and Antibody Recognition". *Proc. Natl. Acad. Sci. USA* **82**, 6138-6142.
19. D. Gospodarowicz, J. Cheng, (1986) "Heparin Protects Basic and Acidic FGF from Inactivation". *J. Cell. Physiol.* **128**, 475-484.
20. J. Schlessinger, I. Lax, M. Lemmon, (1995) "Regulation of Growth Factor Activation by Proteoglycans: What Is the Role of the Low Affinity Receptors?". *Cell* **83**, 357-360.
21. A.C. Rapraeger, A. Krufka, B.B. Olwin, (1991) "Requirement of Heparan Sulfate for bFGF-mediated Fibroblast Growth and Myoblast Differentiation". *Science* **252**, 1705-1708.
22. A. Yayon, M. Klagsbrun, J.D. Esko, P. Leder, D.M. Ornitz, (1991) "Cell Surface, Heparin-like Molecules Are Required for Binding of Basic Fibroblast Growth Factor to Its High Affinity Receptor". *Cell* **64**, 841-848.
23. M.Fannon, M.A. Nugent, (1996) "Basic Fibroblast Growth Factor Binds Its Receptors, Is Internalized, and Stimulates DNA Synthesis in Balb/c3T3 Cells in

- the Absence of Heparan Sulfate". *J. Biol. Chem.* **271**, 17949-17956.
24. L.M. Wakefield, D.M. Smith, K.C. Flanders, M.B. Sporn, (1988) "Latent Transforming Growth Factor- from Human Platelets
 25. E. Ruoslahti, (1988) "Structure and Biology of Proteoglycans". *Ann. Rev. Cell Biol.* **4**, 229-250.
 26. R.L. Jackson, S.J. Busch, A.D. Cardin, (1991) "Glycosaminoglycans: Molecular Properties, Protein Interactions, and Role in Physiological Processes". *Phys. Rev.* **71**, 481-539.
 27. R.D. Sanderson, J.E. Turnbull, J.T. Gallagher, A.D. Lander, (1994) "Fine Structure of Heparan Sulfate Regulates Syndecan-1 Function and Cell Behavior". *J. Biol. Chem.* **269**, 13100-13106.
 28. R.S. Bar, B.L. Dake, S. Stueck, (1987) "Stimulation of Proteoglycans by IGF-I and II in Microvessel and Large Vessel Endothelial Cells". *Am. J. Physiol.* **253**, E21-E27.
 29. P. Vijayagopal, H.P. Ciolino, G.S. Berenson, (1992) "Endothelial Cell-conditioned Medium Modulates the Synthesis and Structure of Proteoglycans in Vascular Smooth Muscle Cells". *Biochem. Biophys. Acta.* **1135**, 129-140.
 30. A. Bassols, J. Massague, (1988) "Transforming Growth Factor Regulates the Expression and Structure of Extracellular Matrix Chondroitin/Dernatan Sulfate Proteoglycans". *J. Biol. Chem.* **263**, 3039-3045.
 31. W.E. Benitz, R.T. Kelley, C.M. Anderson, D.E. Lorant, M. Bernfield, (1990) "Endothelial Heparan Sulfate Proteoglycan. I. Inhibitory Effects on Smooth Muscle Cell Proliferation". *Am. J. Respir. Cell Mol. Biol.* **2**, 13-24.
 32. M.A. Nugent, M.J. Karnovsky, E.R. Edelman, (1993) "Vascular Cell-Derived Heparan Sulfate Shows Coupled Inhibition of Basic Fibroblast Growth Factor Binding and Mitogenesis in Vascular Smooth Muscle Cells". *Circ. Res.* **73**, 1051-1060.
 33. K.E. Forsten, N.A. Courant, M.A. Nugent, (1997) "Endothelial Proteoglycans Inhibit bFGF Binding and Mitogenesis". *J. Cell. Phys.*, accepted.
 34. E. Rinderknecht, R.H. Humbel, (1978) "The Amino Acid Sequence of Human Insulin-Like Growth Factor I and Its Structural Homology with Proinsulin". *J. Biol. Chem.* **253**, 2769-2776.

35. R.S. Bar, K. Siddle, S. Dolash, M. Boes, B. Dake, (1988) "Actions of Insulin and Insulin-Like Growth Factors I and II in Cultured Microvessel Endothelial Cells From Bovine Adipose Tissue". *Metabolism* **37**, 714-720.
36. M. Grant, J. Jerdan, T.J. Merimee, (1987) "Insulin-Like Growth Factor-I Modulates Endothelial Cell Chemotaxis". *J. Clin. Endo. Metabol.* **65**, 370-371.
37. C.M. Gadjusek, Z. Luo, M.R. Mayberg, (1993) "Sequestration and Secretion of Insulin-Like Growth Factor-I by Bovine Aortic Endothelial Cells". *J. Cell. Phys.* **154**, 192-198.
38. Z. Dai, A.D. Stiles, B. Moats-Staats, J.J. Van Wyk, A. J. D'Ercole, (1992) "Interaction of Secreted Insulin-Like Growth Factor-I (IGF-I) with Cell Surface Receptors Is the Dominant Mechanism of IGF-I's Autocrine Actions". *J. Biol. Chem.* **267**, 19565-19571.
39. J.I. Jones, D.R. Clemmons, (1995) "Insulin-Like Growth Factors and Their Binding Proteins: Biological Actions". *Endocrine Rev.* **16**, 3-34.
40. R.S. Bar, M. Boes, M. Yorek, (1986) "Processing of Insulin-Like Growth Factors I and II by Capillary and Large Vessel Endothelial Cells". *Endocrinology* **118**, 1072-1080.
41. M.C. Kiefer, R.S. Ioh, D.M. Bauer, J. Zapf, (1991) "Molecular Cloning of a New Human Insulin-Like Growth Factor Binding Protein". *Bioch. Biophys. Res. Comm.* **176**, 219-225.
42. R.G. Elgin, W.H. Busby, D.R. Clemmons, (1987) "An Insulin-Like Growth Factor (IGF) Binding Protein Enhances the Biologic Response to IGF-I". *Proc. Natl. Acad. Sci. USA* **84**, 3254-3258.
43. R. Gopinath, P.E. Walton, T.D. Etherton, (1989) "An Acid-Stable Insulin-Like Growth Factor (IGF)-Binding Protein From Pig Serum Inhibits Binding of IGF-I and IGF-II to Vacular Endothelial Cells". *J. Endocrinol.* **120**, 231-236.
44. R.S. Bar, B.A. Booth, M. Boes, B.L. Dake, (1989) "Insulin-Like Growth Factor-Binding Proteins from Vacular Endothelial Cells: Purification, Characterization, and Intrinsic Biological Activities". *Endocrinology* **125**, 1910-1919.
45. W.F. Blum, E.W. Jenne, F. Reppin, K. Kitzmann, M.B. Ranke, J.R. Bierich, (1989) "Insulin-Like Growth Factor I (IGF-I)- Binding Protein Complex Is a Better Mitogen than Free IGF-I". *Endocrinology* **125**, 766-772.

46. R.H. McCusker, C. Camacho-Hubner, M.L. Bayne, M.A. Cascieri, D.R. Clemmons, (1990) "Insulin-Like Growth Factor (IGF) Binding to Human Fibroblast and Glioblastoma Cells: The Modulating Effect of Cell Released IGF Binding Proteins (IGFBPs)". *J. Cell. Physiol.* **144**, 244-253.
47. R.H. McCusker, W.H. Busby, M.H. Dehoff, C. Camacho-Hubner, D.R. Clemmons, (1991) "Insulin-Like Growth Factor (IGF) Binding to Cell Monolayers Is Directly Modulated by the Addition of IGF-Binding Proteins". *Endocrinology* **129**, 939-949.
48. E.P. Smith, P.T. Cheung, A. Ferguson, S.D. Chernauek, (1992) "Mechanisms of Sertoli Cell Insulin-Like Growth Factor (IGF)-Binding Protein-3 Regulation by IGF-I and Adenosine 3',5'-Monophosphate". *Endocrinology* **131**, 2733-2741.
49. Y. Oh, H.L. Muller, G. Lamson, R.G. Rosenfeld, (1993) "Insulin-Like Growth Factor (IGF)-Independent Action of IGF Binding Protein-3 in Hs578T Human Breast Cancer Cells". *J. Biol. Chem.* **268**, 14964-14971.
50. E.P. Smith, L.Lu, S.D Chernauek, D.J. Klein, (1994) "Insulin-Like Growth Factor-Binding Protein-3 (IGFBP-3) Concentration in Rat Sertoli Cell-Conditioned Medium Is Regulated by a Pathway Involving Association of IGFBP-3 With Cell Surface Proteoglycans". *Endocrinology* **135**, 359-364.
51. D.R. Clemmons, L.E. Underwood, P.G. Chatelain, J.J. Van Wyk, (1983) "Liberation of Immunoreactive Somatomedin-C From Its Binding Proteins by Proteolytic Enzymes and Heparin". *J. Clin. Endocrinol. Metab.* **56**, 384-389.
52. R.C. Baxter, (1990) "Glycosaminoglycans Inhibit Formation of the 140kDa Insulin-Like Growth Factor-Binding Protein Complex". *Biochem. J.* **271**, 773-777.
53. T. Arai, A. Parker, W. Busby, Jr, D.R. Clemmons, (1994) "Heparin, Heparan Sulfate, and Dermatan Sulfate Regulate Formation of the Insulin-Like Growth Factor-I and Insulin-Like Growth Factor-Binding Protein Complexes". *J.Biol. Chem.* **269**, 20388-20393.
54. B.A. Booth, M. Boes, D.L. Andress, B.L. Dake, C. Maack, R.J. Linhardt, K. Bar, E.E.O. Caldwell, J. Weiler, R.S. Bar, (1995) "IGFBP-3 and IGFBP-3 Association from Endothelial Cells: Role of C-terminal Heparin Binding Domain". *Growth Regul.* **5**, 1-17.
55. J.L. Martin, M. Ballesteros, R.C. Baxter, (1992) "Insulin-Like Growth Factor (IGF-I) and Transforming Growth Factor-1 Release IGF-Binding Protein-3 from Human Fibroblasts by Different Mechanisms". *Endocrinology* **131**, 1703-1710.

56. C.A. Conover, D.R. Powell, (1991) "Insulin-Like Growth Factor (IGF)-Binding Protein-3 Blocks IGF-I Induced Receptor Down-Regulation and Cell Desensitization in Cultured Bovine Fibroblasts". *Endocrinology* **129**, 710-716.
57. M. Yanagishita, V.C. Hascall, (1992) "Cell Surface Heparan Sulfate Proteoglycans". *J. Biol. Chem.* **267**, 9451-9454.
58. M.A. Gimbrone, (1976) "Culture of Vascular Endothelium". *Prog. Hemost. Thromb.* **3**, 1-28.
59. E.E. Van Obberrghen-Schilling, M.M. Rechler, J.A. Romanus, A.B. Knight, Sp.P Nissley, R.E. Humbel, (1981) "Receptors for Insulin-Like Growth Factor I are Defective in Fibroblasts Cultured from a Patient with Leprechaunism". *J. Clin. Invest.* **82**, 1359-1365.
60. R.W. Farndale, D.J. Buttle, A.J. Barrett, (1986) "Improved Quantitation and Discrimination of Sulphated Glycosaminoglycan by Use of Dimethylmethylene Blue". *Biochem. Biophys. Acta* **883**, 173-177.
61. H. Ohno, J. Blackwell, A.M Jamieson, D.A. Carrino, A. Caplan, (1986) "Calibration of the Relative Molecular Mass of Proteoglycan Subunit by Column Chromatography on Sepharose CL-2B". *Biochem. J.* **235**, 553-557.
62. B.A. Booth, R.S. Bar, M. Boes, B.L. Dake, M. Bayne, M. Cascieri, (1990) "Intrinsic Bioactivity of Insulin-Like Growth Factor-Binding Proteins from Vascular Endothelial Cells". *Endocrinology* **127**, 2630-2637.
63. J.L. Martin, R.C. Baxter, (1986) "Insulin-Like Growth Factor-Binding Protein from Human Plasma". *J. Biol. Chem.* **261**, 8754-8760.
64. A. Baird, D. Schubert, N. Ling, R. Guillemin, (1988) "Receptor- and Heparin-Binding Domains of Basic Fibroblast Growth Factor". *Proc. Natl. Acad. Sci. USA* **85**, 2324-2328.
65. J.L. Van den Brande, (1992) "Structure of the Human Insulin-Like Growth Factors: Relationship to Function" in *Insulin -Like Growth Factors Structure and Biological Functions* P.N. Schofield, Eds. Oxford University Press, Oxford, 12-44.
66. K.E. Forsten, N.A. Courant, M.A. Nugent, (1995) "Quantitative Analysis of basic Fibroblast Growth Factor Binding to Endothelial Cell-Derived Heparan Sulfate Proteoglycan". American Institute of Chemical Engineers National Meeting, Miami Florida.

67. D.A. Lauffenberger, J.J. Lindeman, (1993) *Receptors: Models for Binding, Trafficking, and Signaling*. Oxford University Press, New York
68. D. Romagnolo, R.M. Akers, J.C. Byatt, E.A. Wong, J.D. Turner, (1994) "IGF-I-Induced IGFBP-3 Potentiates the Mitogenic Actions of IGF-I in Mammary Epithelial MD-IGF-I Cells". *Mol. Cell Endocrinol.* **102**, 131-139.
69. D.A. Lauffenberger, C. Cozens, (1989) "Regulation of Mammalian Cell Growth by Autocrine Growth Factors: Analysis of Consequences for Inoculum Cell Density Effects". *Biotechnol. Bioeng.* **33**, 1365-1378.
70. K.E. Forsten, D.A. Lauffenberger, (1992) "Autocrine Ligand Binding to Cell Receptors". *Biophys. J.* **61**, 518-529.
71. K.E. Forsten, D.A. Lauffenberger, (1994) "The Role of Low-Affinity Interleukin-2 Receptors in Autocrine Ligand Binding: Alternative Mechanisms for Enhanced Binding Effect". *Mol. Immunol.* **31**, 739-751.
72. H.C. Berg, (1983) *Random Walks in Biology*. Princeton University Press, Princeton
73. M.V. Smoluchowski, (1917) "Versuch einer Mathematischen Theorie Der Koagulationskinetik Kolloider Losungen". *Z. Phys. Chem.* **92**, 129-168.
74. W. Hu, D.I.C. Wang, (1987) "A Mechanistic Analysis of Inoculum Requirement for the Cultivation of Mammalian Cells on Microcarriers.". *Biotechnol. Bioeng.* **30**, 548-557.
75. C.J. Geankoplis, (1993) *Transport Processes and Unit Operations*. 3rd ed., Prentice Hall, New Jersey
76. O.T. Hanna, O.C. Sandall, (1995) *Computational Methods in Chemical Engineering*. Prentice Hall, New Jersey
77. L.S. Matthews, R.E. Hammer, R.R. Behringer, A.J. D'Ercole, G.I. Bell, R.L. Brinster, R.D. Palmiter, (1988) "Growth Enhancement of Transgenic Mice Expressing Human Insulin Growth Factor I". *Endocrinology* **123**, 2827-2833.
78. P. Wilton, (1992) "Treatment with Recombinant Human Insulin-Like Growth Factor I in the Prevention of Vincristine Neuropathy in Mice". *Ann. NY Acad. Sci.* **692**, 243-245.
79. M. Barinaga, (1994) "Neurotrophic Factors Enter the Clinic". *Science* **264**, 772-774.

APPENDIX A. BINDING AFFINITY (K_D) VALUES: LEAST SQUARES ANALYSIS

Least squares analysis is a distributed error approximation method. This involves the decrease of some global error measure with respect to the whole approximation interval as the order of approximation increases. It is valuable in fitting equations to discrete data points and in analyzing measurement errors.

Based on an form of the equation:

$$y_p(x_i) = aC_1 + C_2, \quad (1)$$

the least squares procedure forms the sum of the squares of the differences between observed values (y_i) and the predicted values $y_p(x_i)$. The equation is minimized with respect to the unknown parameters C_1 and C_2 . Therefore, a normalized error quantity predicts the goodness of fit of the data to the model.

$$\text{Error} = \frac{1}{N} \sum_{i=1}^N [y_i - (aC_1 + C_2)]^2 / (N-p), \quad (2)$$

where N = number of data points; p = number of parameters to be considered.

Error is minimized with respect to C_1 and C_2 by using:

$$0 = \text{Error} / C_1 = \frac{1}{N} \sum_{i=1}^N [y_i - (aC_1 + C_2)], \quad (3)$$

$$0 = \text{Error} / C_2 = \frac{1}{N} \sum_{i=1}^N [y_i - (aC_1 + C_2)]x_i^k. \quad k= 0,1... \quad (4)$$

Expansion of the sums generates two linear algebraic equations for the coefficients C_1 and C_2 . Once the parameters are known, error can be determined⁷⁵.

Time derivatives describing the bound complex of IGF-I (I) and IGFBP-3 (BP) or p9 HS (PG) were established. The time derivative for the IGF-1/IGFBP-3 complex (B) is

as follows:

$$dB/dt = k_{on}^s[I][BP] - k_{off}^s[B], \quad (5)$$

where $I = I_o - B$, $BP = BP_o - B$. At steady state the rate of change of the IGF-I/IGFBP-3 complex (B) is zero and Equation 5 becomes:

$$I_o = K_D^s[B]/[BP_o - B] + [B], \quad (6)$$

where $K_D^s = k_{off}^s/k_{on}^s$. Equation 6 becomes y “predicted” (Equation 1) with $I_o = y_p$, $K_D = a$, $B/[BP_o - B] = C_1$ and $[B] = C_2$. Least squares analysis from experimental assay data yielded K_D values of 6.2×10^{-9} M for the biodot assay and 3×10^{-9} M for the charcoal assay. As discussed in Chapter III.A and B, the charcoal assay was the preferable assay for quantitating binding affinity.

Likewise, a time derivative describing the rate of change of the IGF-I/p9 HS complexes (P) is:

$$dP/dt = k_{on}^p[I][PG] - k_{off}^p[P], \quad (7)$$

where $I = I_o - P$, $PG = PG_o - P$. At steady state the rate of change of IGF-I/p9 HS complex (P) is zero and Equation 7 becomes:

$$I_o = K_D^p[P]/[PG_o - P] + [P], \quad (8)$$

where $K_D^p = k_{off}^p/k_{on}^p$. Equation 8 becomes y “predicted” (Equation 1) with $I_o = y_p$, $K_D = a$, $P/[PG_o - P] = C_1$ and $[P] = C_2$. Least squares analysis from experimental assay data yielded K_D values of 2.2×10^{-7} M for the biodot assay and 1.5×10^{-8} M for the charcoal assay.

APPENDIX B. BAE CELL SYSTEM MODEL FORTRAN CODES

Basic BAE Cell System Model:

```
C
C***** ABSTRACT
C*****
C
C THIS PROGRAM APPLIES THE LIBRARY ROUTINE LSODE TO THE
C INTEGRATION ON A NON-STIFF SET OF ODE'S. NOMENCLATURE FOR THIS
C PROGRAM CAN BE FOUND IN FILE LSDP1.
C
C*****
C
c Basic BAE Cell- Y(1) = u (dimensionless unbound receptors, R/Ro)
c - Y(2) = v (dimensionless bound receptors, C/Ro)
c - Y(3) = y (dimensionless ligand, Lo/Kd)
c initial conditions: Y(1) = 1
c Y(2) = 0
c Y(3) = Lo/Kd
C integrate from zero time (t=0) to 3 hours (t=10800 secs)
C
C IMPLICIT REAL*8(A-H,O-Z)
C NONSTIFF PROBLEMS (MF = 10)
C RWORK(20+16*NEQ), IWORK(20)
C DIMENSION Y(3),ATOL(3),RWORK(52),IWORK(20)
C
C STIFF PROBLEMS (MF=21(WITH JACOBIAN),22(WITHOUT JACOBIAN)
C RWORK(22+9*NEQ+NEQ**2),IWORK(20+NEQ)
C DIMENSION Y(3), ATOL(3),RWORK(58),IWORK(22)
C
C NONSTIFF PROBLEMS/STIFF (MF=22)
C EXTERNAL FEX
C STIFF PROBLEMS (MF=21)
C EXTERNAL FEX,JAC
C
C common/class/b,rho
c Parameter values
C Ro = 2d4
c Ro = # of receptors/cell
```

```

xkd = .6d-9
c Kd= koff/kon = affinity of ligand to receptor = M
a = 5.0d-4
c a = radius of a single cell = cm
D = 5.0d-7
c D = diffusivity of ligand through binding buffer = cm2/s
xkon = 4.0d7
c kon = association rate constant for ligand and receptor = M-1s-1
xLo = 5.7E-10
c Lo = initial ligand concentration=M
V=3.33d-5
c V = binding buffer volume/avg. # of cells (15,000) = cm3
xN=6.023d23
c N = Avogadro's number = # sites/mol
C
C Dimensionless Groups
b = xkon*Ro*1000/(4.0d0*3.1416d0*a*D*xN)
c [(L/mol*s)*(#/cell)*(L/1000 cm3)]/[(cm/cell)*(cm2/s)*(#/mol)]
rho = (Ro*1000)/(V*xkd*xN)
c [(#/cell)*(1000cm3/L)]/[(cm3)*(mol/L)*(#/mol)]
write(*,*) b,rho
C INITIAL CONDITIONS
Y(1)=1.0d0
Y(2)=0.0d0
Y(3)=xlo/xkd
Tin=0.0d0
Tout =10.8d3
Tin=tin*xkd*xkon
Tout=Tout*xkd*xkon
N = 100
deltax =(tout-tin)/float(N)
C DGEAR PARAMETER SPECIFICATIONS
NEQ=3
RTOL=1.E-6
ITOL=1
ATOL(1)=1.E-6
ITASK=1
ISTATE=1
IOPT=0
C nonstiff integrator
C MF=10
C LRW = 20*16*NEQ

```

```

C LIW = 20
c stiff integrator with jacobian
  MF = 21
  LRW = 22+9*NEQ+NEQ**2
  LIW = 20+NEQ
C INTEGRATE IN EQUALLY SPACED SEGMENTS USING A DO-LOOP
  DO 10 K=1,N
  Tout=Tin+deltax
  CALL LSODE(FEX,NEQ,Y,Tin,Tout,ITOL,RTOL,ATOL,ITASK,ISTATE,IOPT,
  1RWORK,LRW,IWORK,LIW,JAC,MF)
C PRINT OUT RESULTS
  WRITE(6,12) Tout/(xkd*xkon),Y(1),Y(2),Y(3)
  OPEN(Unit=95,FILE='BAE.DAT',STATUS='OLD')
  WRITE(95,22) Tout/(xkd*xkon),Y(1),Y(2),Y(3)
  tin=tout
10 continue
12 FORMAT( 14X,2HX=,G8.3,2X,5HY(1)=,D11.6,2X,5HY(2)=,D11.5,
          2X,5HY(3)=,E14.2)
22 FORMAT( E14.6,E14.6,E14.6,E14.6)

      STOP
      END
C
C***** SUBROUTINE FCN
*****
C
C THIS SUBROUTINE CALCULATES THE DERIVATIVE OF EACH DEPENDENT
C VARIABLE WITH RESPECT TO X, YPRIME(I).
C
C*****
C
SUBROUTINE FEX(N,X,Y,YPRIME)
  IMPLICIT REAL*8(A-H,O-Z)
  INTEGER N
  DIMENSION Y(3),YPRIME(3)
  common/class/b,rho
  YPRIME(1)=(y(2)-y(3)*y(1))/(1.0d0+b*y(1))
  YPRIME(2)=(y(3)*y(1)-y(2))/(1.0d0+b*y(1))
  YPRIME(3)=((-y(3)*rho*y(1) + rho*y(2))/(1.0d0+b*y(1)))
  RETURN
  END
C

```

```

C***** SUBROUTINE FCNJ
C*****
C
C THIS IS A DUMMY SUBROUTINE IF MF IS EQUAL TO 10/22 AND THE
C JACOBIAN IS ESTIMATED NUMERICALLY.
C
C*****
C
SUBROUTINE JAC(N,X,Y,ML,MU,PD,NRPD)
IMPLICIT REAL*8(A-H,O-Z)
REAL*8 Y(3),PD(NRPD,3),X
common/class/b,rho
C
pd(1,1) = ((-b*(y(2)-y(3)*y(1)))/((1.0d0+b*y(1))*(1.0d0+b*y(1))))-
          (y(3)/(1.0d0+b*y(1)))
pd(1,2) = 1.0d0/(1.0d0+b*y(1))
pd(1,3) = -y(1)/(1.0d0+b*y(1))
pd(2,1) = ((b*(y(2)-y(3)*y(1)))/
          ((1.0d0+b*y(1))*(1.0d0+b*y(1))))+(y(3)/(1.0d0+b*y(1)))
pd(2,2) = -1.0d0/(1.0d0+b*y(1))
pd(2,3) = y(1)/(1.0d0+b*y(1))
pd(3,1) = ((rho*b*(y(3)*y(1)-y(2)))/
          ((1.0d0+b*y(1))*(1.0d0+b*y(1))))-(y(3)*rho/
          (1.0d0+b*y(1)))
pd(3,2) = rho/(1.0d0+b*y(1))
pd(3,3) = (-rho*y(1)/(1.0d0+b*y(1)))
RETURN
END

```

IGF-I/IGFBP-3 BAE Cell System Model:

```
C
C***** ABSTRACT
C*****
C
C THIS PROGRAM APPLIES THE LIBRARY ROUTINE LSODE TO THE
C INTEGRATION ON A NON-STIFF SET OF ODE'S. NOMENCLATURE FOR THIS
C PROGRAM CAN BE FOUND IN FILE LSDP1.
C
C*****
C IGF-I/IGFBP-3 BAE Cell System Model
c      - Y(1) = u (dimensionless unbound receptors, R/Ro)
c      - Y(2) = v (dimensionless bound receptors, C/Ro)
c      - Y(3) = y (dimensionless ligand, Lo/Kd)
c      - Y(4) = w (dimensionless unbound IGFBP-3, S/So)
c      - Y(5) = z (dimensionless bound IGFBP-3, X/So)
c      IGFBP-3      pick Kons = 4 x 10+7 M-1s-1
c                   KDs = 3 x 10-9 M
c initial conditions: Y(1) = 1
c                   Y(2) = 0
c                   Y(3) = Lo/Kd
c                   Y(4) = 1
c                   Y(5) = 0
C integrate from zero time (t=0) to 3 hours (t=10800 secs)
C
C IMPLICIT REAL*8(A-H,O-Z)
C NONSTIFF PROBLEMS (MF = 10)
C RWORK(20+16*NEQ), IWORK(20)
C DIMENSION Y(5),ATOL(5),RWORK(100),IWORK(20)
C
C STIFF PROBLEMS (MF=21(WITH JACOBIAN),22(WITHOUT JACOBIAN)
C RWORK(22+9*NEQ+NEQ**2),IWORK(20+NEQ)
C DIMENSION Y(5), ATOL(5),RWORK(92),IWORK(22)
C
C NONSTIFF PROBLEMS/STIFF (MF=22)
C EXTERNAL FEX
C STIFF PROBLEMS (MF=21)
C EXTERNAL FEX,JAC
C
common/class/b,rho,alpha,beta,gamma
```

```

c Parameter values
  Ro = 2.0d4
c   Ro=# receptors/cell
  xkd = .6d-9
c   Kd = koff/kon = affinity of ligand to receptor =M
  a = 5.0d-4
c   a = radius of a single cell
  D = 5.0d-7
c   D = diffusivity of ligand through binding buffer = cm2/s
  xkon = 4.0d7
c   kon = association rate constant for ligand and receptor = M-1s-1
  xLo = 5.7d-10
c   Lo = initial ligand concentration =M
  V=3.33d-5
c   V = binding buffer volume/avg. # of cells (15,000) = cm3
  So=2.8d-9
c   So = exogenous IGFBP-3 added = M
  xkons=4.0d7
c   kons = association rate constant for ligand and IGFBP-3 = M-1s-1
  xkds=3d-9
c   Kds = affinity of ligand to IGFBP-3 = M
  xN = 6.023d23
c   N = Avogadro's # = #/mol
C   dimensionless groups
  b = xkon*Ro*1000/(4.0d0*3.1416d0*a*D*xN)
c   [(L/mol*s)*(#/cell)*(L/1000cm3)]/[(cm/cell)*(cm2/s)*(#/mol)]
  rho = (Ro*1000)/(V*xkd*xN)
c   [(#/cell)*(1000cm3/L)]/[(cm3*(mol/L)*(#/mol)]
  alpha = xkds/xkd
c   [M/M]
  beta = xkons/xkon
c   [(M-1s-1)/(M-1s-1)]
  gamma = So/xkd
c   [M/M]
  write(*,*) b,rho,alpha,beta,gamma
C INITIAL CONDITIONS
  Y(1)=1.0d0
  Y(2)=0.0d0
  Y(3)=xlo/xkd
  Y(4)=1.0d0
  Y(5)=0.0d0
  Tin=0.0d0

```

```

    Tout =10.8d3
    Tin=tin*xkd*xkon
    Tout=Tout*xkd*xkon
    N = 100
    deltax =(tout-tin)/float(N)
C DGEAR PARAMETER SPECIFICATIONS
    NEQ=5
    RTOL=1.E-6
    ITOL=1
    ATOL(1)=1.E-6
    ITASK=1
    ISTATE=1
    IOPT=0
C
c nonstiff integrator
C MF=10
C LRW = 20*16*NEQ
C LIW = 20
c stiff integrator with jacobian
MF = 21
LRW = 22+9*NEQ+NEQ**2
LIW = 20+NEQ
C INTEGRATE IN EQUALLY SPACED SEGMENTS USING A DO-LOOP
DO 10 K=1,N
    Tout=Tin+deltax
    CALL LSODE(FEX,NEQ,Y,Tin,Tout,ITOL,RTOL,ATOL,ITASK,ISTATE,IOPT,
              RWORK,LRW,IWORK,LIW,JAC,MF)
C PRINT OUT RESULTS
WRITE(6,12) Tout/(xkd*xkon),Y(1),Y(2),Y(3),Y(4),Y(5)
OPEN(Unit=95,FILE='BP.DAT',STATUS='OLD')
WRITE(95,22) Tout/(xkd*xkon),Y(1),Y(2),Y(3),Y(4),Y(5)
tin=tout
10 continue
12 FORMAT( 14X,2HX=,G8.3,2X,5HY(1)=,D11.6,2X,5HY(2)=,D11.6,
          2X,5HY(3)=,E14.2,2X,5HY(4)=,D11.6,2X,5HY(5)=,D11.6)
22 FORMAT( E14.6,E14.6,E14.6,E14.6,E14.6,E14.6)

    STOP
END
C
C***** SUBROUTINE FCN
*****

```



```

C
C   THIS SUBROUTINE CALCULATES THE DERIVATIVE OF EACH DEPENDENT
C   VARIABLE WITH RESPECT TO X, YPRIME(I).
C
C*****
C
SUBROUTINE FEX(N,X,Y,YPRIME)
IMPLICIT REAL*8(A-H,O-Z)
INTEGER N
DIMENSION Y(5),YPRIME(5)
common/class/b,rho,alpha,beta,gamma
YPRIME(1)=(y(2)-y(3)*y(1))/(1.0d0+b*y(1))
YPRIME(2)=(y(3)*y(1)-y(2))/(1.0d0+b*y(1))
YPRIME(3)=((-y(3)*rho*y(1) + rho*y(2))/(1.0d0+b*y(1)))-
           (beta*Y(3)*gamma*Y(4))+(alpha*beta*Y(5)*gamma)
YPRIME(4)=(-beta*Y(3)*Y(4))+(alpha*beta*Y(5))
YPRIME(5)=(beta*Y(3)*Y(4))-(alpha*beta*Y(5))
RETURN
END

C
C***** SUBROUTINE FCNJ
C*****
C
C   THIS IS A DUMMY SUBROUTINE IF MF IS EQUAL TO 10/22 AND THE
C   JACOBIAN IS ESTIMATED NUMERICALLY.
C
C*****
C
SUBROUTINE JAC(N,X,Y,ML,MU,PD,NRPD)
IMPLICIT REAL*8(A-H,O-Z)
REAL*8 Y(5),PD(NRPD,5),X
common/class/b,rho,alpha,beta,gamma
c
pd(1,1) = ((-b*(y(2)-y(3)*y(1)))/((1.0d0+b*y(1))*
           (1.0d0+b*y(1))))-(y(3)/(1.0d0+b*y(1)))
pd(1,2) = 1.0d0/(1.0d0+b*y(1))
pd(1,3) = -y(1)/(1.0d0+b*y(1))
pd(1,4) = 0
pd(1,5) = 0
pd(2,1) = ((b*(y(2)-y(3)*y(1)))/((1.0d0+b*y(1))*
           (1.0d0+b*y(1))))+(y(3)/(1.0d0+b*y(1)))
pd(2,2) = -1.0d0/(1.0d0+b*y(1))

```

```

pd(2,3) = y(1)/(1.0d0+b*y(1))
pd(2,4) = 0
pd(2,5) = 0
pd(3,1) = ((rho*b*(y(3)*y(1)-y(2)))/((1.0d0+b*y(1))*
          (1.0d0+b*y(1))))-(y(3)*rho/(1.0d0+b*y(1)))
pd(3,2) = rho/(1.0d0+b*y(1))
pd(3,3) = (-rho*y(1)/(1.0d0+b*y(1)))
pd(3,4) = -beta*Y(3)*gamma
pd(3,5) = alpha*beta*gamma
pd(4,1) = 0
pd(4,2) = 0
pd(4,3) = -beta*Y(4)
pd(4,4) = -beta*Y(3)
pd(4,5) = alpha*beta
pd(5,1) = 0
pd(5,2) = 0
pd(5,3) = beta*Y(4)
pd(5,4) = beta*Y(3)
pd(5,5) = -alpha*beta
RETURN
END

```

IGF-I/p9 HS BAE Cell System Model:

```
C
C***** ABSTRACT
C*****
C
C THIS PROGRAM APPLIES THE LIBRARY ROUTINE LSODE TO THE
C INTEGRATION ON A NON-STIFF SET OF ODE'S. NOMENCLATURE FOR THIS
C PROGRAM CAN BE FOUND IN FILE LSDP1.
C
C*****
C IGF-I/p9 HS BAE Cell System Model
c          - Y(1) = u (dimensionless unbound receptors, R/Ro)
c          - Y(2) = v (dimensionless bound receptors, C/Ro)
c          - Y(3) = y (dimensionless ligand, Lo/Kd)
c          - Y(4) = w (dimensionless unbound p9 HS, S/So)
c          - Y(5) = z (dimensionless bound p9 HS, X/So)
c          p9 HS      pick Kons = 4 x 10+7 M-1s-1
c                      KDS = 1.5 x 10-8 M
c  initial conditions:  Y(1) = 1
c                      Y(2) = 0
c                      Y(3) = Lo/Kd
c                      Y(4) = 1
c                      Y(5) = 0
C  integrate from zero time (t=0) to 3 hours (t=10800 secs)
C
C IMPLICIT REAL*8(A-H,O-Z)
C NONSTIFF PROBLEMS (MF = 10)
C RWORK(20+16*NEQ), IWORK(20)
C DIMENSION Y(5),ATOL(5),RWORK(100),IWORK(20)
C
C STIFF PROBLEMS (MF=21(WITH JACOBIAN),22(WITHOUT JACOBIAN)
C RWORK(22+9*NEQ+NEQ**2),IWORK(20+NEQ)
C DIMENSION Y(5), ATOL(5),RWORK(92),IWORK(22)
C
C NONSTIFF PROBLEMS/STIFF (MF=22)
C EXTERNAL FEX
C STIFF PROBLEMS (MF=21)
C EXTERNAL FEX,JAC
C
C common/class/b,rho,alpha,beta,gamma
```

c Parameter values
 $R_o = 2.0d4$
c $R_o = \# \text{ receptors/cell}$
 $xkd = .6d-9$
c $Kd = koff/kon = \text{affinity of ligand to receptor} = M$
 $a = 5.0d-4$
c $a = \text{radius of a single cell}$
 $D = 5.0d-7$
c $D = \text{diffusivity of ligand through binding buffer} = \text{cm}^2/\text{s}$
 $xkon = 4.0d7$
c $kon = \text{association rate constant for ligand and receptor} = M^{-1}s^{-1}$
 $xLo = 5.7d-10$
c $Lo = \text{initial ligand concentration} = M$
 $V = 3.33d-5$
c $V = \text{binding buffer volume/avg. \# of cells (15,000)} = \text{cm}^3$
 $So = 1.4d-10$
c $So = \text{exogenous p9 HS added} = M$
 $xkons = 4.0d7$
c $kons = \text{association rate constant for ligand and p9 HS} = M^{-1}s^{-1}$
 $xkds = 1.5d-8$
c $Kds = \text{affinity of ligand to p9 HS} = M$
 $xN = 6.023d23$
c $N = \text{Avogadro's \#} = \#/\text{mol}$
C dimensionless groups
 $b = xkon * R_o * 1000 / (4.0d0 * 3.1416d0 * a * D * xN)$
c $[(L/\text{mol} * s) * (\#/\text{cell}) * (L/1000\text{cm}^3)] / [(cm/\text{cell}) * (cm^2/s) * (\#/\text{mol})]$
 $\rho = (R_o * 1000) / (V * xkd * xN)$
c $[(\#/\text{cell}) * (1000\text{cm}^3/L)] / [(cm^3 * (\text{mol}/L) * (\#/\text{mol})]$
 $\alpha = xkds / xkd$
c $[M/M]$
 $\beta = xkons / xkon$
c $[(M^{-1}s^{-1}) / (M^{-1}s^{-1})]$
 $\gamma = So / xkd$
c $[M/M]$
write(*,*) b,rho,alpha,beta,gamma
C INITIAL CONDITIONS
 $Y(1) = 1.0d0$
 $Y(2) = 0.0d0$
 $Y(3) = xlo / xkd$
 $Y(4) = 1.0d0$
 $Y(5) = 0.0d0$
 $Tin = 0.0d0$

```

    Tout =10.8d3
    Tin=tin*xkd*xkon
    Tout=Tout*xkd*xkon
    N = 100
    deltax =(tout-tin)/float(N)
C DGEAR PARAMETER SPECIFICATIONS
    NEQ=5
    RTOL=1.E-6
    ITOL=1
    ATOL(1)=1.E-6
    ITASK=1
    ISTATE=1
    IOPT=0
C
c nonstiff integrator
C MF=10
C LRW = 20*16*NEQ
C LIW = 20
c stiff integrator with jacobian
    MF = 21
    LRW = 22+9*NEQ+NEQ**2
    LIW = 20+NEQ
C INTEGRATE IN EQUALLY SPACED SEGMENTS USING A DO-LOOP
    DO 10 K=1,N
        Tout=Tin+deltax
        CALL LSODE(FEX,NEQ,Y,Tin,Tout,ITOL,RTOL,ATOL,ITASK,ISTATE,IOPT,
                RWORK,LRW,IWORK,LIW,JAC,MF)
C PRINT OUT RESULTS
    WRITE(6,12) Tout/(xkd*xkon),Y(1),Y(2),Y(3),Y(4),Y(5)
    OPEN(Unit=95,FILE='p9.DAT',STATUS='OLD')
    WRITE(95,22) Tout/(xkd*xkon),Y(1),Y(2),Y(3),Y(4),Y(5)
    tin=tout
10 continue
12 FORMAT( 14X,2HX=,G8.3,2X,5HY(1)=,D11.6,2X,5HY(2)=,D11.6,
        2X,5HY(3)=,E14.2,2X,5HY(4)=,D11.6,2X,5HY(5)=,D11.6)
22 FORMAT( E14.6,E14.6,E14.6,E14.6,E14.6,E14.6)

    STOP
END
C
C***** SUBROUTINE FCN
*****

```

```

C
C   THIS SUBROUTINE CALCULATES THE DERIVATIVE OF EACH DEPENDENT
C   VARIABLE WITH RESPECT TO X, YPRIME(I).
C
C*****
C
SUBROUTINE FEX(N,X,Y,YPRIME)
IMPLICIT REAL*8(A-H,O-Z)
INTEGER N
DIMENSION Y(5),YPRIME(5)
common/class/b,rho,alpha,beta,gamma
YPRIME(1)=(y(2)-y(3)*y(1))/(1.0d0+b*y(1))
YPRIME(2)=(y(3)*y(1)-y(2))/(1.0d0+b*y(1))
YPRIME(3)=((-y(3)*rho*y(1) + rho*y(2))/(1.0d0+b*y(1)))-
           (beta*Y(3)*gamma*Y(4))+(alpha*beta*Y(5)*gamma)
YPRIME(4)=(-beta*Y(3)*Y(4))+(alpha*beta*Y(5))
YPRIME(5)=(beta*Y(3)*Y(4))-(alpha*beta*Y(5))
RETURN
END

C
C***** SUBROUTINE FCNJ
C*****
C
C   THIS IS A DUMMY SUBROUTINE IF MF IS EQUAL TO 10/22 AND THE
C   JACOBIAN IS ESTIMATED NUMERICALLY.
C
C*****
C
SUBROUTINE JAC(N,X,Y,ML,MU,PD,NRPD)
IMPLICIT REAL*8(A-H,O-Z)
REAL*8 Y(5),PD(NRPD,5),X
common/class/b,rho,alpha,beta,gamma
c
pd(1,1) = ((-b*(y(2)-y(3)*y(1)))/((1.0d0+b*y(1))*
           (1.0d0+b*y(1))))-(y(3)/(1.0d0+b*y(1)))
pd(1,2) = 1.0d0/(1.0d0+b*y(1))
pd(1,3) = -y(1)/(1.0d0+b*y(1))
pd(1,4) = 0
pd(1,5) = 0
pd(2,1) = ((b*(y(2)-y(3)*y(1)))/((1.0d0+b*y(1))*
           (1.0d0+b*y(1))))+(y(3)/(1.0d0+b*y(1)))
pd(2,2) = -1.0d0/(1.0d0+b*y(1))

```

```
pd(2,3) = y(1)/(1.0d0+b*y(1))
pd(2,4) = 0
pd(2,5) = 0
pd(3,1) = ((rho*b*(y(3)*y(1)-y(2)))/((1.0d0+b*y(1))*
          (1.0d0+b*y(1))))-(y(3)*rho/(1.0d0+b*y(1)))
pd(3,2) = rho/(1.0d0+b*y(1))
pd(3,3) = (-rho*y(1)/(1.0d0+b*y(1)))
pd(3,4) = -beta*Y(3)*gamma
pd(3,5) = alpha*beta*gamma
pd(4,1) = 0
pd(4,2) = 0
pd(4,3) = -beta*Y(4)
pd(4,4) = -beta*Y(3)
pd(4,5) = alpha*beta
pd(5,1) = 0
pd(5,2) = 0
pd(5,3) = beta*Y(4)
pd(5,4) = beta*Y(3)
pd(5,5) = -alpha*beta
RETURN
END
```

Complex BAE Cell System Model:

```
C
C***** ABSTRACT
C*****
C
C THIS PROGRAM APPLIES THE LIBRARY ROUTINE LSODE TO THE
C INTEGRATION ON A NON-STIFF SET OF ODE'S. NOMENCLATURE FOR THIS
C PROGRAM CAN BE FOUND IN FILE LSDP1.
C
C*****
C IGFBP and p9 HS Complex BAE Cell
c          - Y(1) = u (dimensionless unbound receptors, R/Ro)
c          - Y(2) = v (dimensionless bound receptors, C/Ro)
c          - Y(3) = y (dimensionless ligand, Lo/Kd)
c          - Y(4) = w (dimensionless unbound IGFBP-3, S/So)
c          - Y(5) = z (dimensionless bound IGFBP-3, X/So)
c          - Y(6) = p (dimensionless unbound p9 HS, P/Po)
c          - Y(7) = q (dimensionless bound p9 HS, Q/Po)
c          IGFBP-3          pick Kons = 4 x 10+7 M-1s-1
c          p9 HS           pick konp = 4 x 10+7 M-1s-1
c                          KDs = 3 x 10-9 M
c                          KDp = 1.5 x 10-8 M
c initial conditions:      Y(1) = 1
c                          Y(2) = 0
c                          Y(3) = Lo/Kd
c                          Y(4) = 1
c                          Y(5) = 0
c                          Y(6) = 1
c                          Y(7) = 0
C integrate from zero time (t=0) to 3 hours (t=10800 secs)
C
C IMPLICIT REAL*8(A-H,O-Z)
C NONSTIFF PROBLEMS (MF = 10)
C RWORK(20+16*NEQ), IWORK(20)
C DIMENSION Y(7),ATOL(7),RWORK(132),IWORK(27)
C
C STIFF PROBLEMS (MF=21(WITH JACOBIAN),22(WITHOUT JACOBIAN)
C RWORK(22+9*NEQ+NEQ**2),IWORK(20+NEQ)
C DIMENSION Y(7), ATOL(7),RWORK(135),IWORK(27)
```


C
 C NONSTIFF PROBLEMS/STIFF (MF=22)
 C EXTERNAL FEX
 C STIFF PROBLEMS (MF=21)
 C EXTERNAL FEX,JAC
 C
 common/class/b,rho,alpha,beta,gamma,sigma,xkappa,phi
 c Parameter values
 Ro = 2.0d4
 c Ro=# receptors/cell
 xkd = .6d-9
 c Kd = koff/kon = affinity of ligand to receptor =M
 a = 5.0d-4
 c a = radius of a single cell
 D = 5.0d-7
 c D = diffusivity of ligand through binding buffer = cm2/s
 xkon =4d7
 c kon = association rate constant for ligand and receptor = M-1s-1
 xLo = 5.7d-10
 c Lo = initial ligand concentration =M
 V=3.33d-5
 c V = binding buffer volume/avg. # of cells (15,000) = cm3
 So=2.8d-9
 c So = exogenous IGFBP-3 added = M
 Po=1.4d-10
 c Po = exogenous p9 HS added = M
 xkons=4.0d7
 c kons = association rate constant for ligand and IGFBP-3 = M-1s-1
 xkonp=4.0d7
 c konp = association rate constant for ligand and p9 HS = M-1s-1
 xkds=3.0d-9
 c Kds = affinity of ligand to IGFBP-3 = M
 xkdp=1.5d-8
 c Kdp = affinity of ligand to p9 HS = M
 xN = 6.023d23
 c N = Avogadro's # = #/mol
 C dimensionless groups
 b = xkon*Ro*1000/(4.0d0*3.1416d0*a*D*xN)
 c [(L/mol*s)*(#/cell)*(L/1000cm3)]/[(cm/cell)*(cm2/s)*(#/mol)]
 rho = (Ro*1000)/(V*xkd*xN)
 c [(#/cell)*(1000cm3/L)]/[(cm3*(mol/L)*(#/mol)]
 alpha =xkds/xkd

```

c  [M/M]
sigma = xkdp/xkd
c  [M/M]
beta = xkons/xkon
c  [(M-1s-1)/(M-1s-1)]
xkappa = xkonp/xkon
c  [(M-1s-1)/(M-1s-1)]
gamma = So/xkd
c  [M/M]
phi = Po/xkd
c  [M/M]
write(*,*) b,rho,alpha,beta,gamma,sigma,xkappa,phi
C INITIAL CONDITIONS
Y(1)=1.0d0
Y(2)=0.0d0
Y(3)=xlo/xkd
Y(4)=1.0d0
Y(5)=0.0d0
Y(6) = 1.0d0
Y(7) = 0.0d0
Tin=0.0d0
Tout =5d0
Tin=tin*xkd*xkon
Tout=Tout*xkd*xkon
N = 100
deltax =(tout-tin)/float(N)
C DGEAR PARAMETER SPECIFICATIONS
NEQ=7
RTOL=1.E-6
ITOL=1
ATOL(1)=1.E-6
ITASK=1
ISTATE=1
IOPT=0

C
c  nonstiff integrator
C  MF=10
C  LRW = 20*16*NEQ
C  LIW = 20
c  stiff integrator with jacobian
MF = 21
LRW = 22+9*NEQ+NEQ**2

```

```

    LIW = 20+NEQ
C INTEGRATE IN EQUALLY SPACED SEGMENTS USING A DO-LOOP
    DO 10 K=1,N
        Tout=Tin+deltax
        CALL LSODE(FEX,NEQ,Y,Tin,Tout,ITOL,RTOL,ATOL,ITASK,ISTATE,IOPT,
            RWORK,LRW,IWORK,LIW,JAC,MF)
C PRINT OUT RESULTS
        WRITE(6,12) Tout/(xkd*xkon),Y(1),Y(2),Y(3),Y(4),Y(5),Y(6),Y(7)
        OPEN(Unit=95,FILE='COMBO.DAT',STATUS='OLD')
        WRITE(95,22) Tout/(xkd*xkon),Y(1),Y(2),Y(3),Y(4),Y(5),Y(6),Y(7)
        tin=tout
10 continue
12 FORMAT( 14X,2HX=,G8.3,2X,5HY(1)=,D11.6,2X,5HY(2)=,D11.6,
        2X,5HY(3)=,E14.2,2X,5HY(4)=,D11.6,2X,5HY(5)=,D11.6,
        2X,5HY(6)=,D11.6,2X,5HY(7)=,D11.6)
22 FORMAT( E14.6,E14.6,E14.6,E14.6,E14.6,E14.6,E14.6)

    STOP
    END
C
C***** SUBROUTINE FCN
C*****
C
C THIS SUBROUTINE CALCULATES THE DERIVATIVE OF EACH DEPENDENT
C VARIABLE WITH RESPECT TO X, YPRIME(I).
C
C*****
C
SUBROUTINE FEX(N,X,Y,YPRIME)
    IMPLICIT REAL*8(A-H,O-Z)
    INTEGER N
    DIMENSION Y(7),YPRIME(7)
    common/class/b,rho,alpha,beta,gamma,sigma,xkappa,phi
    YPRIME(1)=(y(2)-y(3)*y(1))/(1.0d0+b*y(1))
    YPRIME(2)=(y(3)*y(1)-y(2))/(1.0d0+b*y(1))
    YPRIME(3)=((-y(3)*rho*y(1) + rho*y(2))/(1.0d0+b*y(1)))-
        (beta*Y(3)*gamma*Y(4))+(alpha*beta*Y(5)*gamma)
        -(xkappa*Y(3)*phi*Y(6))+(sigma*xkappa*Y(7)*phi)
    YPRIME(4)=(-beta*Y(3)*Y(4))+(alpha*beta*Y(5))
    YPRIME(5)=(beta*Y(3)*Y(4))-(alpha*beta*Y(5))
    YPRIME(6)=(-xkappa*Y(3)*Y(6))+(sigma*xkappa*Y(7))
    YPRIME(7)=(xkappa*Y(3)*Y(6))-(sigma*xkappa*Y(7))

```

RETURN
END

C

C***** SUBROUTINE FCNJ

C*****

C

C THIS IS A DUMMY SUBROUTINE IF MF IS EQUAL TO 10/22 AND THE
C JACOBIAN IS ESTIMATED NUMERICALLY.

C

C*****

C

SUBROUTINE JAC(N,X,Y,ML,MU,PD,NRPD)

IMPLICIT REAL*8(A-H,O-Z)

REAL*8 Y(7),PD(NRPD,7),X

common/class/b,rho,alpha,beta,gamma,sigma,xkappa,phi

C

pd(1,1) = ((-b*(y(2)-y(3)*y(1)))/((1.0d0+b*y(1))*
(1.0d0+b*y(1))))-(y(3)/(1.0d0+b*y(1)))

pd(1,2) = 1.0d0/(1.0d0+b*y(1))

pd(1,3) = -y(1)/(1.0d0+b*y(1))

pd(1,4) = 0

pd(1,5) = 0

pd(1,6) = 0

pd(1,7) = 0

pd(2,1) = ((b*(y(2)-y(3)*y(1)))/((1.0d0+b*y(1))*
(1.0d0+b*y(1))))+(y(3)/(1.0d0+b*y(1)))

pd(2,2) = -1.0d0/(1.0d0+b*y(1))

pd(2,3) = y(1)/(1.0d0+b*y(1))

pd(2,4) = 0

pd(2,5) = 0

pd(2,6) = 0

pd(2,7) = 0

pd(3,1) = ((rho*b*(y(3)*y(1)-y(2)))/((1.0d0+b*y(1))*
(1.0d0+b*y(1))))-(y(3)*rho/(1.0d0+b*y(1)))

pd(3,2) = rho/(1.0d0+b*y(1))

pd(3,3) = (-rho*y(1)/(1.0d0+b*y(1)))

pd(3,4) = -beta*Y(3)*gamma

pd(3,5) = alpha*beta*gamma

pd(3,6) = -xkappa*Y(3)*phi

pd(3,7) = sigma*xkappa*phi

```
pd(4,1) = 0
pd(4,2) = 0
pd(4,3) = -beta*Y(4)
pd(4,4) = -beta*Y(3)
pd(4,5) = alpha*beta
pd(4,6) = 0
pd(4,7) = 0
pd(5,1) = 0
pd(5,2) = 0
pd(5,3) = beta*Y(4)
pd(5,4) = beta*Y(3)
pd(5,5) = -alpha*beta
pd(5,6) = 0
pd(5,7) = 0
pd(6,1) = 0
pd(6,2) = 0
pd(6,3) = -xkappa*Y(6)
pd(6,4) = 0
pd(6,5) = 0
pd(6,6) = -xkappa*Y(3)
pd(6,7) = sigma*xkappa
pd(7,1) = 0
pd(7,2) = 0
pd(7,3) = xkappa*Y(4)
pd(7,4) = 0
pd(7,5) = 0
pd(7,6) = xkappa*Y(3)
pd(7,7) = -sigma*xkappa
RETURN
END
```

CURRICULUM VITAE

- EDUCATION:** Virginia Polytechnic Institute and State University
Master of Science, Chemical Engineering, May 1997
- University Of Mississippi
Bachelor of Science, Chemical Engineering, May 1995
- RESEARCH EXPERIENCE:** Dr. Kim Forsten's Lab, VA Tech
January 1996 - May 1997
- TEACHING EXPERIENCE:** Unit Operations Lab, VA Tech
May 1996 - June 1996
- Mass and Energy Balances, VA Tech
Teaching Assistant, Fall 1995, 1996
- Process Modeling, VA Tech
Teaching Assistant, Spring 1996
- Heat Transfer, VA Tech
Teaching Assistant, Spring 1997
- HONORS / ACTIVITIES:** Graduate Student Advisory Delegate, VA Tech
College of Engineering Graduate Student Committee, VA Tech
National Merit Scholar
Chancellor's and Dean's Honor Roll
Who's Who Among National College Students
ASB Cabinet - Director of Student Development
Sorority Treasurer
ASB Committee(s) Chairman: *Health Advisory,*
Student Wellness,
Environmental Awareness
- Student Alumni Council
American Institute of Chemical Engineers - Treasurer
Society of Women Engineers - Secretary / Treasurer
Phi Eta Sigma - Academic Honorary
Alpha Lambda Delta - Academic Honorary

Student Programming Board

Order of Omega

Intramural Sports - softball, football, volleyball

Ole Miss Sexual Harassment Hearing Panel

Three Time Miss Majorette of Virginia

Four Time Virginia State Solo Twirling Champion

Three Time Virginia State Strutting Champion

Second Place, World Solo Twirling Championships

Second Place, World Strutting Championships

Fifth Place, Miss Majorette of America