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CHARACTERIZATION OF TEMPERATURE-SENSITIVE STRAINS OF NEOSPA
CANUM IN MICE

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ABSTRACT: Temperature-sensitive (ts) strains of the Neospora canum tachyzoites were selected by chemical mutagenesis and selection for growth at 32°C. Three ts strains and the parental, N. canum wild-type strain, NC-1, were examined in the present study for their ability to cause disease in inbred BALB/c mice, outbred ICR mice, and chemically immunosuppressed ICR mice. In BALB/c mice, all strains failed to induce clinical disease, whereas infection with the NC-1 strain caused central nervous system disease and death in some mice. No disease was observed in ICR mice inoculated with the 3 ts strains or the NC-1 strain. All immunosuppressed ICR mice inoculated with the NC-1 strain died, whereas no immunosuppressed mice inoculated with the NCts-4 strain and only 1 of 5 mice inoculated with the NCts-8 and NCts-12 strains died. The NCts-4 and NCts-12 strains reverted to a wild-type phenotype when grown at 37°C. Vaccination of BALB/c mice with live, but not frozen NCts-8 strain tachyzoites induced significant (P < 0.05) protection following NC-1 strain challenge.

Neospora canum is a major cause of abortion in dairy cattle worldwide (see Dubey and Lindsay, 1996). Cattle that abort do not become immune and can reabort during several consecutive pregnancies. Little is known about the immune response to N. canum in cattle, and no vaccines are available against N. canum in any animal species.

Modified live vaccines have been developed for the vacci
nation of animals against Toxoplasma gondii, a protozoan structurally and biologically related to N. canum. The temperature-sensitive (ts) mutant ts-4 (Pfefferkorn and Pfefferkorn, 1976) has been demonstrated to be safe and afford protection in several mammalian hosts, including mice, hamsters, monkeys, and pigs (Waldeland and Frenkel, 1983; Waldeland et al., 1983; Elwell and Frenkel, 1984a, 1984b; Escayladillo and Frenkel, 1991; Lindsay et al., 1993; Pinckney et al., 1994, 1995). The S48 mutant has been evaluated against abortion in sheep and is safe and effective (O’Connel et al., 1988; Buxton et al., 1991; Buxton and Innes, 1995; Wastling et al., 1994, 1995). The objective of the present study was to develop and characterize ts strains of N. canum and to evaluate their safety and efficacy in an N. canum encephalitic mouse model (Lindsay et al., 1995; Long et al., 1998).

MATERIALS AND METHODS

Generation of cloned parasite strains

The NC-1–2C line (Lindsay et al., 1996) of the NC-1 strain of N. canum was used to generate ts mutants. This line was exposed to 0.5 μmol N-methyl-N-nitro-N-nitrosoguanidine for 24 hr. It was then grown at 32.5°C for 3 mo after which it was cloned by limiting dilution. Twelve clones were isolated and 3 selected for further study. The NCs-4, NCs-8, and NCts-12 clones had been maintained at 32.5°C for >8 mo before use in the present studies. These strains were examined for reversion to wild-type phenotype by continuous propagation at 37°C for 88 days (25 cell culture passages) prior to use (NCts-4–37, NCts-8–37, NCts-12–37) in experiment 3.

Mice

Mice used were outbred ICR and inbred BALB/c strains obtained from the same commercial supplier (Harlan Sprague Dawley, Indianapolis, Indiana). Mice were 18–22-g females, housed in groups of 4 or 5 in plastic box cages, and provided rodent chow and water ad libitum.

Sero logical testing

The indirect immunofluorescent antibody (IFA) test was used to examine sera for antibodies to N. canum from mice (Cole et al., 1995). Sera were obtained from mice immediately prior to challenge inoculation in vaccination studies (experiment 4) and obtained from all mice that survived pathogenicity experiments. Sera were examined for IgG N. canum–specific antibodies at doubling dilutions beginning at 1:50 and end-point titrated. Air-dried tachyzoites were used as antigen. Positive samples exhibited a complete tachyzoite surface fluorescence. Negative samples exhibited no fluorescence or only anterior end fluorescence.

Cell culture assay for identification for N. canum tissue cysts in mouse brain

The brain from each mouse was removed at necropsy, and half was fixed in 10% neutral buffered formalin solution and used for histopathologic examination (see lesion scores below). The remaining half was used for acid-pepsin digestion and inoculation of cell cultures to detect tissue cysts of N. canum. The portion of brain was placed in 3 ml of 0.25% trypsin and incubated at 37°C for 15 min. After incubation, the brain was washed with media and incubated in maintenance medium. The brains were washed for 28 days after which they were considered negative if no stages of N. canum were isolated.

Microscopic lesion scores

The following criteria were used for histologic lesion scoring (Lindsay et al., 1995). Number of inflammatory or necrotic foci: no foci = 1; 1–5 foci = 2; 6–10 foci = 3; >10 foci = 4. Average size of foci: none = 1; 100–200 μm = 2; 200–500 μm = 3; >500 μm = 4. Severity of lesions: no lesions = 1; slight = 2; mild = 3; moderate = 4; marked = 5. A mean lesion score was obtained from these 3 values. A normal noninfected mouse brain would have a mean lesion score of 3.0. Mean lesion scores were evaluated using a Kruskal-Wallis nonparametric test and distribution free multiple comparisons methods (Zar, 1984). The numbers of mice in each group with lesions was examined using Fisher’s exact test. Significance was established at a cutoff of P < 0.05 prior to conduct of analysis of data.
**Experiment 1: Pathogenicity of NC-1 and NCts strains of *N. caninum* in BALB/c mice**

Forty-four BALB/c mice were used in experiment 1. Mice were inoculated subcutaneously (s.c.) with HBSS or 500,000 tachyzoites of the NC-1, NCts-4, NCts-8, or NCts-12 strains of *N. caninum*, and all surviving mice were killed and examined at necropsy 42 or 56 days post-inoculation (PI) (Table I). Serum was collected from surviving mice at necropsy. Brains were obtained and used for microscopic lesions scores and parasite isolation.

**Experiment 2: Pathogenicity of NC-1 and NCts strains of *N. caninum* in ICR mice**

Twenty-five ICR mice were used in experiment 2. Mice were inoculated s.c. with HBSS or 500,000 tachyzoites of the NC-1, NCts-4, NCts-8, or NCts-12 strains of *N. caninum*. All surviving mice were killed 56 days PI. Sera were collected for IFA testing. Brains were collected for microscopic lesions scores and parasite isolation.

**Experiment 3: Pathogenicity of NC-1, NCts strains, and NCts revertant strains of *N. caninum* in immunosuppressed ICR mice**

Forty ICR mice were used in experiment 3. All mice were immunosuppressed by intramuscular administration of 2 mg of methylprednisolone acetate on days 7, 0, and 7 PI (Lindsay and Dubey, 1989). Mice were inoculated s.c. with HBSS or 200,000 tachyzoites of the NC-1, NCts-4, NCts-8, or NCts-12 strains or reversion controls NCts-4–37, NCts-8–37, and NCts-12–37 on day 0. Surviving mice were killed and examined at necropsy 56 days PI. Sera were collected for IFA testing. Brains were collected for microscopic lesions scores and parasite isolation.

**Experiment 4: Vaccination of BALB/c mice against *N. caninum* induced encephalitis**

Thirty-five BALB/c mice were used in experiment 4. Mice were vaccinated s.c. with HBSS, 500,000 freshly isolated tachyzoites of the NCts-8 strain, or 2 × 10⁷ killed (frozen) tachyzoites by s.c. inoculation (Table VI). Mice were boosted 21 days PI with identical preparations of HBSS or parasites. Mice were challenged s.c. with 1 × 10⁶ NC-1 strain tachyzoites or HBSS 14 days after the second vaccination. Surviving mice were killed and examined at necropsy 56 days PI. Sera were collected for IFA testing. Brains were collected for microscopic lesion scores and parasite isolation. The viability of the frozen vaccine was examined by inoculation of HS68 cell cultures immediately following primary and secondary immunizations.

**RESULTS**

**Experiment 1**

Clinical neosporosis and mortalities occurred only in BALB/c mice infected with the NC-1 strain (Table I). Three of 10 BALB/c mice challenged with NC-1 survived through the 8-wk study period. None of the sham-infected or NCts-mutant-infected BALB/c mice died, and significant differences in mean lesion scores between NCts-mutant groups and the NC-1-infected group were found (Table II). No tissue cysts were observed in histological sections of mouse brain, and no parasites were reisolated from cell culture from any group.

No antibodies to *N. caninum* were detected in sham-infected mice examined 42 days PI, or 56 days PI. Mice inoculated with the NCts-4 strain had *N. caninum* antibody titers ranging from 1:800 (1 mouse) to 1:1,600 (4 mice) 42 days PI, and all mice had a 1:1,600 titer 56 days PI. Mice inoculated with the NCts-8 strain showed antibody titers ranging from 1:1,600 (3 mice) to 1:3,200 (2 mice) at both 42 and 56 days PI. Similar antibody titers from NCts-12 strain inoculated mice were also detected. Mice inoculated with the NC-1 strain and examined 56 days PI had titers of 1:1,600 (3 mice).

**Experiment 2**

None of the parasite strains caused mortality in ICR mice, and no significant differences were observed in numbers of mice with lesions or in the mean lesions scores (Table III). No tissue cysts were observed in histological sections of mouse brain, and parasites were not isolated from brain tissue inoculated cell cultures.

No antibodies to *N. caninum* were detected in 5 control mice examined 56 days PI. Mice inoculated with the NCts-4 strain and examined 56 days PI had titers ranging from 1:400 (1 mouse), to 1:800 (3 mice), with a fifth mouse at 1:1,600. Mice inoculated with the NCts-8 strain and examined 56 days PI had titers of 1:50 (1 mouse), 1:800 (2 mice), 1:1,600 (1 mouse), and 1:3,200 (1 mouse). Mice inoculated with the NCts-12 strain and examined 56 days PI had titers of 1:50 (1 mouse), 1:800 (3 mice), and 1:1,600 (1 mouse). Mice inoculated with the NC-

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**Table I. Protocol and clinical outcome of BALB/c mice inoculated subcutaneously with Hanks’ balanced salt solution (HBSS) or 500,000 tachyzoites of NC-1, NCts-4, NCts-8, or NCts-12 strains of Neospora caninum.**

<table>
<thead>
<tr>
<th>Group/treatment</th>
<th>No. of mice</th>
<th>Strain</th>
<th>Clinical outcome</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>4</td>
<td>HBSS</td>
<td>Two mice killed 42 and 56 days.</td>
</tr>
<tr>
<td>2</td>
<td>5</td>
<td>NCts-4</td>
<td>All mice killed 42 days PI. No mortality.</td>
</tr>
<tr>
<td>3</td>
<td>5</td>
<td>NCts-8</td>
<td>All mice killed 56 days PI. No mortality.</td>
</tr>
<tr>
<td>4</td>
<td>5</td>
<td>NCts-8</td>
<td>All mice killed 42 days PI. No mortality.</td>
</tr>
<tr>
<td>5</td>
<td>5</td>
<td>NCts-8</td>
<td>All mice killed 56 days PI. No mortality.</td>
</tr>
<tr>
<td>6</td>
<td>5</td>
<td>NCts-12</td>
<td>All mice killed 42 days PI. No mortality.</td>
</tr>
<tr>
<td>7</td>
<td>5</td>
<td>NCts-12</td>
<td>All mice killed 56 days PI. No mortality.</td>
</tr>
<tr>
<td>8</td>
<td>5</td>
<td>NC-1</td>
<td>Mice developed clinical neosporosis at around 16 days PI. They were less active than groups 1–7 and had rough hair coats. Two mice were killed because of neosporosis 34 days PI, and an additional mouse died 39 days PI. The remaining 2 mice were killed 56 days PI.</td>
</tr>
<tr>
<td>9</td>
<td>5</td>
<td>NC-1</td>
<td>Clinical signs same as group 8. Mice were found dead on days 26, 30, 32, and 41 after inoculation. The remaining mouse was killed 56 days PI.</td>
</tr>
</tbody>
</table>

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**Table II. Mean lesion scores of BALB/c mice inoculated subcutaneously with Hanks’ balanced salt solution (HBSS) or 500,000 tachyzoites of NC-1, NCts-4, NCts-8, or NCts-12 strains of Neospora caninum in experiment 1.**

<table>
<thead>
<tr>
<th>Group/treatment</th>
<th>No. of mice with lesions/no.</th>
<th>Score</th>
</tr>
</thead>
<tbody>
<tr>
<td>Group 1/HBSS</td>
<td>0/44</td>
<td>3.00</td>
</tr>
<tr>
<td>Groups 2 + 3/NCts-4</td>
<td>4/10/10*</td>
<td>4.80</td>
</tr>
<tr>
<td>Groups 4 + 5/NCts-8</td>
<td>6/10/10*</td>
<td>5.50</td>
</tr>
<tr>
<td>Groups 6 + 7/NCts-12</td>
<td>5/10/10*</td>
<td>4.90</td>
</tr>
<tr>
<td>Groups 8 + 9/NC-1</td>
<td>8/3/3†</td>
<td>9.38†</td>
</tr>
</tbody>
</table>
TABLE III. Results of lesion scores of ICR mice inoculated subcutaneously with Hanks’ balanced salt solution (HBSS) or 500,000 tachyzoites of NC-1, NCts-4, NCts-8, and NCts-12 strains of Neospora caninum in experiment 2.

<table>
<thead>
<tr>
<th>Group/treatment</th>
<th>No. of mice with lesions/no. examined/ Mean lesion score*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Group 10/HBSS</td>
<td>0/5/5/3.0</td>
</tr>
<tr>
<td>Group 11/NC-1</td>
<td>3/5/5/6.0</td>
</tr>
<tr>
<td>Group 12/NCts-4</td>
<td>0/5/5/3.0</td>
</tr>
<tr>
<td>Group 13/NCts-8</td>
<td>1/5/5/4.0</td>
</tr>
<tr>
<td>Group 14/NCts-12</td>
<td>0/5/5/3.0</td>
</tr>
</tbody>
</table>

* No significant differences (P > 0.05) were present.

1 strain and examined 56 days PI had titers ranging from 1:1,600 (2 mice) to 1:3,200 (3 mice).

Experiment 3

The NC-1, NCts-4–37, and NCts-12–37 strains caused 100% mortality in immunosuppressed ICR mice (Table IV). The NCts-4, NCts-8, NCts-12, and NCts-8–37 strains were less pathogenic for immunosuppressed mice and caused 0–20% mortality. Microscopic lesion scores are presented in Table V. No tissue cysts were observed in histological sections of mouse brain, and parasites were not isolated from brain tissue–inoculated cell cultures.

All surviving mice were tested for antibody titers at 56 days PI. Mice inoculated with the NCts-4 strain had titers of 1:100 (1 mouse), 1:800 (1 mouse), 1:1,600 (1 mouse), 1:3,200 (1 mouse), and 1:6,400 (1 mouse). Mice inoculated with the NCts-8 strain displayed titers of 1:1,600 (2 mice), 1:3,200 (1 mouse), and 1:6,400 (1 mouse). Mice inoculated with the NCts-12 strain possessed titers of 1:1,600 (1 mouse), 1:3,200 (2 mice), and 1:6,400 (1 mouse). Finally, mice inoculated with the NCts-8–37 strain had titers of 1:3,200 (1 mouse), 1:6,400 (2 mice), and 1:12,800 (1 mouse).

Experiment 4

None of the live, NCts-8–immunized mice died prior to NC-1 challenge. Mortality and mean lesion scores following challenge are shown in Table VI. Two of 8 sham-immunized (HBSS) mice and 3 of 10 mice inoculated with frozen NCts-8 tachyzoites died following NC-1 challenge, and attempts at parasite reisolation in cell culture were unsuccessful.

HBSS-immunized, challenged mice had antibody titers ranging from 1:1,600 (1/10) to 1:6,400 (3/10). Postchallenge titers in NCts-8–immunized mice ranged from 1:400 to 1:6,400 (9/20) with titers ranging from 1:800 to 1:3,200 by end of study. Postchallenge titers in NCts-8–immunized mice ranged from 1:800 (1/10) through 1:1,600 (8/10) to 1:3,200 (1 mouse) and 1:6,400 (1 mouse). Finally, mice inoculated with frozen NCts-8–37 strain had titers of 1:3,200 (1 mouse), 1:6,400 (2 mice), and 1:12,800 (1 mouse). No parasites were isolated in cultures inoculated with frozen tachyzoites.

DISCUSSION

The present study demonstrates that the NCts-4, NCts-8, and NCts-12 strains are less pathogenic than the parental wild-type NC-1 strain in BALB/c mice (experiment 1) and immunosuppressed ICR mice (experiment 3). Although lesions were present in brains of some mice inoculated with these ts strains, they were not statistically significant compared with controls (Tables I, V). A significant difference in lesion scores and numbers of...
mice with lesions were found when the mice inoculated with the NC-1 strain were compared with mice inoculated with HBSS or NCts strains of N. caninum.

Two of the 3 ts strains (NCts-4-37 and NCts-12-37) were 100% lethal for immunosuppressed ICR mice after 25 continuous passages at 37°C (Table V). This suggests a reversion to a wild-type phenotype and were not further evaluated. In contrast, the NCts-8–37 strain caused mortality in only 20% of immunosuppressed mice examined, indicating a much lower incidence of reversion to virulence.

Experiment 4 demonstrated that vaccination of BALB/c mice with 2 doses of the NCts-8 strain provided significant protection against challenge with the NC-1 strain (Table VI). Mice immunized with live NCts-8 strain tachyzoites failed to develop clinical disease or die. NCts-8–immunized, challenged mice had almost identical lesion scores as NCts–8–immunized, nonchallenged mice, providing further evidence for protection using a live, experimental vaccine. However, immunization with nonviable, frozen NCts-8 strain tachyzoites (experiment 4) offered little protection from neosporosis. This lack of protection with killed NCts-8 tachyzoites is similar to what has been observed in sheep given killed T. gondii vaccines (Beverly et al., 1971; Wilkins et al., 1987; Buxton et al., 1989).

**LITERATURE CITED**


