

## Review



**Cite this article:** Marr LC, Tang JW, Van Mullekom J, Lakdawala SS. 2019 Mechanistic insights into the effect of humidity on airborne influenza virus survival, transmission and incidence. *J. R. Soc. Interface* **16**: 20180298. <http://dx.doi.org/10.1098/rsif.2018.0298>

Received: 2 May 2018

Accepted: 20 December 2018

### Subject Category:

Life Sciences—Engineering interface

### Subject Areas:

biometeorology

### Keywords:

humidity, temperature, environment, transmission, flu, aerosol

### Author for correspondence:

Linsey C. Marr

e-mail: [lmarr@vt.edu](mailto:lmarr@vt.edu)

Electronic supplementary material is available online at <https://dx.doi.org/10.6084/m9.figshare.c.4351247>.

# Mechanistic insights into the effect of humidity on airborne influenza virus survival, transmission and incidence

Linsey C. Marr<sup>1</sup>, Julian W. Tang<sup>2,3</sup>, Jennifer Van Mullekom<sup>4</sup>  
and Seema S. Lakdawala<sup>5</sup>

<sup>1</sup>Civil and Environmental Engineering, Virginia Tech, Blacksburg, VA 24061, USA

<sup>2</sup>Clinical Microbiology, University Hospitals Leicester NHS Trust, Leicester, UK

<sup>3</sup>Infection, Immunity and Inflammation, University of Leicester, Leicester, UK

<sup>4</sup>Statistics, Virginia Tech, Blacksburg, VA 24061, USA

<sup>5</sup>Microbiology and Molecular Genetics, University of Pittsburgh School of Medicine, Pittsburgh, PA 15219, USA

LCM, 0000-0003-3628-6891

Influenza incidence and seasonality, along with virus survival and transmission, appear to depend at least partly on humidity, and recent studies have suggested that absolute humidity (AH) is more important than relative humidity (RH) in modulating observed patterns. In this perspective article, we re-evaluate studies of influenza virus survival in aerosols, transmission in animal models and influenza incidence to show that the combination of temperature and RH is equally valid as AH as a predictor. Collinearity must be considered, as higher levels of AH are only possible at higher temperatures, where it is well established that virus decay is more rapid. In studies of incidence that employ meteorological data, outdoor AH may be serving as a proxy for indoor RH in temperate regions during the wintertime heating season. Finally, we present a mechanistic explanation based on droplet evaporation and its impact on droplet physics and chemistry for why RH is more likely than AH to modulate virus survival and transmission.

## 1. Introduction

There has been a surge of interest in understanding transmission and seasonality of influenza and other infectious diseases. Humidity, among other factors, appears to play an important role in influenza transmission, and a confluence of recent studies has suggested that absolute humidity (AH) rather than relative humidity (RH) modulates influenza virus survival, transmission and seasonality [1–4]. While we have an intuitive understanding of humidity in general, the relationship between RH and AH is less obvious. AH is a mass concentration that describes the amount of water vapour per volume of air. Air contains water vapour at concentrations ranging from near-zero grams of water vapour per cubic metre of air in the driest deserts to approximately  $40 \text{ g m}^{-3}$  in hot and humid environments. The amount of water vapour in air can also be expressed as a partial pressure in units such as atmospheres or millibars. A related concept is specific humidity, which is the mass of water vapour per mass of air, such that absolute and specific humidity are related by the density of air.

RH is defined as the ratio of the actual water vapour pressure (or concentration) to the saturation vapour pressure (or concentration), which is defined as the equilibrium partial pressure of water vapour above a flat surface. The saturation vapour pressure increases with temperature, so the concentration of water vapour can be higher in warmer air. The relationship between AH and RH depends on temperature. At  $25^\circ\text{C}$  and 100% RH, the corresponding AH is  $23 \text{ g m}^{-3}$ , while at  $0^\circ\text{C}$  and 100% RH, the corresponding AH is  $4.8 \text{ g m}^{-3}$ . On the other hand, an AH of  $4 \text{ g m}^{-3}$  corresponds to an RH of

83% at 0°C and 17% at 25°C. Greater detail on different measures of atmospheric moisture and their relationship to human health can be found in Davis *et al.* [5] and Wolkoff [6].

Although recent studies suggest that AH modulates influenza virus survival, transmission and seasonality [1–4], we contend that the combination of RH and temperature provides a more consistent, mechanistically sound explanation for the observations. To support our contention, we review and reanalyse previously published data while adding mechanistic insights to the findings in this perspective article. In evaluating studies of influenza virus survival in aerosols and droplets, transmission in animal models and influenza incidence, we make two critical observations. First, there is strong evidence that temperature modulates influenza virus survival independent of humidity [7]; survival is reduced at higher temperatures. Higher levels of AH are physically possible only at higher temperatures, and because of this relationship, directly comparing the effect of AH versus RH alone on virus survival paints an incomplete picture. Second, most influenza transmission between humans probably takes place indoors within a narrow range of temperatures, where AH and RH move in tandem. When the outdoor temperature is low and the indoor environment is heated, indoor RH is closely correlated with outdoor AH but is only weakly correlated with outdoor RH. Thus, we expect outdoor AH, rather than outdoor RH, to be a better predictor of influenza incidence. Finally, we propose a mechanistic explanation for the relationship between RH and transmission via droplets and aerosols, where RH affects their physics and chemistry.

## 2. Material and methods

### 2.1. Statistical analyses

To investigate the relationships between the dependent variable of virus viability (quantified as log % viability) and independent variables of temperature ( $T$ ), AH or RH, we fitted statistical models in JMP<sup>®</sup> Pro, v. 13 using ordinary least-squares regression with normal error structure. We used techniques consistent with best statistical practices in regression modelling, including considering a variety of metrics of fit and prediction, using plots to inform modelling and removing collinear terms. We evaluated models on the basis of  $F$  and  $t$ -tests for statistical significance,  $R^2$ , mean squared error, residual plots, variance inflation factors and predicted versus actual plots.

In selecting among predictive statistical models for virus viability, we must account for the relationship between AH and RH. Based on an improved form of the Magnus equation for the saturation vapour pressure of water [8], the ideal gas law and the definitions of AH and RH, we derived the following empirical equation for AH as a function of RH and temperature ( $T$ ):

$$\text{AH} = \frac{1322.7 \exp(17.625 T / (T + 243.04)) \times (\text{RH}/100)}{T + 273.15}, \quad (2.1)$$

where AH is in  $\text{g m}^{-3}$ ,  $T$  is in °C and RH is in per cent. This equation is valid over the temperature range of  $-40$  to  $50^\circ\text{C}$ . Further details of the derivation appear in the electronic supplementary material.

### 2.2. Equilibrium droplet size

We followed a parametrization for the relationship between RH and droplet size presented in a study of the interaction of aerosols composed of salt and protein with water vapour (eqn 28

in [9]). The equation assumes that the solutes in the droplet do not interact with each other and that the volume of the droplet is the sum of the volume of the dry solutes and of the water. We represented salt by NaCl at a concentration of  $9 \text{ mg ml}^{-1}$  protein as having a molecular weight of  $640\,000 \text{ g mol}^{-1}$  and a concentration of either  $3 \text{ mg ml}^{-1}$  as found in nasal airway surface fluid [10] or  $76 \text{ mg ml}^{-1}$  [11] as estimated for aerosols of respiratory breath condensate, and surfactant by dipalmitoylphosphatidylcholine (DPPC) with a molecular weight of  $734 \text{ g mol}^{-1}$  and a concentration of  $0.5 \text{ g l}^{-1}$  [12]. We applied the simplified ion-interaction parametrization of the osmotic coefficient ( $\Phi$ ) for the strong electrolyte NaCl (eqn 13 in [9]) and the osmotic pressure parametrization of  $\Phi$  for protein and surfactant, assumed to behave like rigid spheres. We assumed that the surface tension of the solution equalled that of water; the predicted diameter was not sensitive to this value (i.e. if it were a factor of three lower, as in the lungs) for the size of droplets under consideration.

## 3. Results and discussion

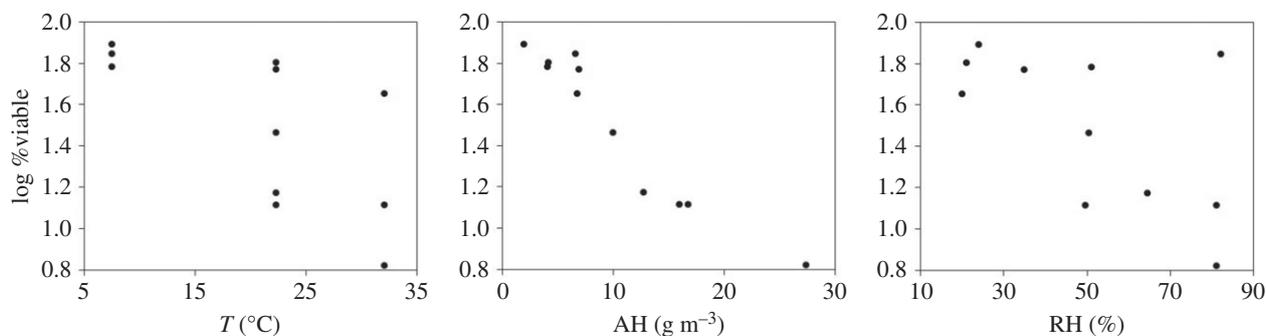
### 3.1. Prior study of airborne virus survival

To our knowledge, only one study has examined airborne influenza virus survival as a function of both temperature and humidity. Harper [13] used a Collison nebulizer to aerosolize virus from a suspension in ‘casein McIlvaine’s buffer’ into a 75-l rotating drum. He measured viability in terms of infectious dose in egg membranes, corrected for physical loss, at three temperatures ( $7.0$ – $8.0^\circ\text{C}$ ,  $20.5$ – $24.0^\circ\text{C}$  and  $32.0^\circ\text{C}$ ) and five RHs ( $20$ – $25\%$ ,  $34$ – $36\%$ ,  $49$ – $51\%$ ,  $64$ – $65\%$  and  $81$ – $82\%$ ). His results showed that the virus survived better at the lowest temperature and the lowest RH tested. Shaman & Kohn [3] reanalysed Harper’s results and showed that AH, in terms of the vapour pressure of water, provided a much stronger fit to virus viability than did RH.

### 3.2. Effect of temperature and humidity on virus survival

A systematic review of influenza virus persistence concluded that temperature, over an environmentally relevant range, is a significant predictor of persistence across all environmental matrices considered, including air, water and soil [7]; higher temperature is associated with a shorter half-life. The effect of temperature on influenza viruses has also been studied in *in vitro* culture systems in the context of live virus vaccine attenuation. A serial passage of viruses at restricted temperatures produced viruses with temperature sensitivity (*ts*), such as influenza viruses with restricted growth at  $39^\circ\text{C}$ , or a cold-adaptive (*ca*) phenotype, whereby virus growth was enhanced at  $33^\circ\text{C}$  compared with  $37^\circ\text{C}$  [14,15]. However, these studies on live vaccine platforms focused on the growth restriction and not the survival of virus strains at various temperatures.

Researchers have proposed mechanisms for the thermal degradation of influenza virus, and we expect these to apply across a wide variety of systems, such as aerosols, droplets, surfaces or bulk culture medium. Woese [16] identified thermal denaturation of proteins and nucleic acids as mechanisms of virus inactivation and invoked the Eyring equation, used to describe the rate of chemical reactions and functionally similar to the Arrhenius equation, to explain temperature dependence. Polozov *et al.* [17] suggested that increased ordering of lipids of



**Figure 1.** Scatterplots showing influenza virus viability in aerosols [13], quantified as the log of the per cent viable after 1 h, versus temperature ( $T$ ), AH and RH.

the viral envelope may contribute to improved stability at lower temperatures. There are several lines of evidence suggesting a relationship between the pH of haemagglutinin activation and stability of influenza viruses at higher temperatures; viruses with a pH of fusion of greater than 5.6 are more stable at 50°C compared with those with a pH of fusion less than 5.4 [18–23]. These studies demonstrate that temperature can impact influenza viruses at a molecular level.

We have previously reviewed the effect of humidity on survival of a broad range of viruses, including influenza virus [24]. Studies of influenza virus survival in droplets and aerosols as a function of RH have shown either a monotonic decrease in viability with increasing RH or a U-shaped relationship with reduced viability at mid-range RHs [25,26]. Disparities in results may be due to differences in the suspension media, whether it included protein or not [24].

When weighing the effect of humidity and limiting the variables under consideration to AH and RH only, and not temperature, one drawback is that high values of AH are only achievable at high temperature. Zhao *et al.* [27] recognized this in a study of survival of aerosolized Gumboro virus, concluding, ‘A large part of the AH effect on viral survival actually was attributable to the effect of temperature’. They found that temperature and RH together were the best predictors of virus survival and that survival was not significantly affected by AH. Because Gumboro virus is non-enveloped, its relationship between viability and RH is likely to differ from that of influenza virus [26], but the point about the relationship among temperature, AH and RH is universally applicable. It is well established that influenza virus is less stable at higher temperatures. Higher values of AH are only achievable at higher temperature, at which virus stability is compromised. Under this constraint, we might expect the correlation of viability with AH alone, which includes temperature by proxy, to be stronger than the correlation of viability with RH alone.

### 3.3. Reanalysis of virus survival data

Ideal linear regression models (i.e. linear in the coefficients) arise from uncorrelated independent variables. The functional form for AH in eqn (2.1) indicates that AH is dependent on  $T$ . Linear regression models with correlated independent variables often have inflated variances of the estimated coefficients, coefficients with signs in the opposite direction of known relationships and inaccurate predictions for dependent variable combinations inconsistent with the regressor correlation. Given this information, only two possible general linear models are statistically advised in this circumstance: either viability must be modelled as a function

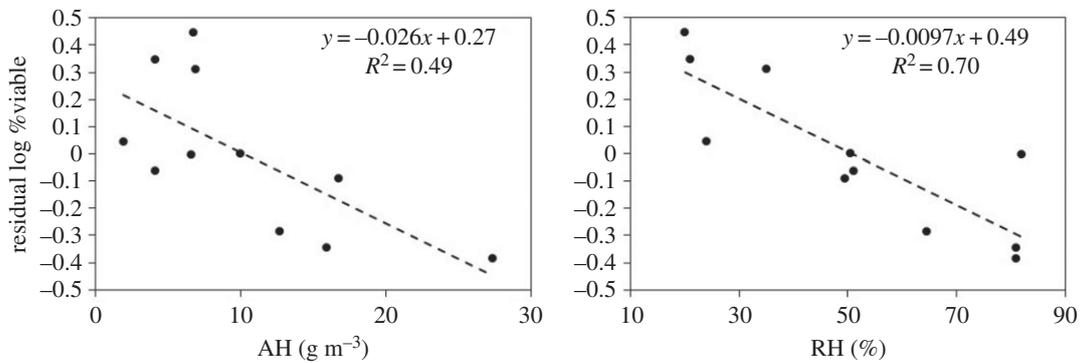
**Table 1.** Measures of goodness of fit comparing models of airborne virus viability [13] as a function of RH and  $T$  and as a function of AH only.

measure of fit	$T$ , RH, $T \times$ RH	AH, AH $\times$ AH
$R^2$	94.8%	94.3%
mean square error	0.10	0.10

of RH and  $T$ , or it must be modelled as a function of AH. In general, when a known physical relationship indicates the presence of collinearity among regressors, there are two choices to fit the best-performing model: (i) fit a model with the less correlated or uncorrelated regressors or (ii) fit a model that accounts for collinear regressors (e.g. ridge regression, principal components regression and LASSO). The first, simpler approach is possible with this dataset.

Figure 1 shows scatterplots of Harper’s original data [13] on influenza virus survival, where viability is quantified as the log of the percentage of infectious virus remaining after 1 h. Figure 1 indicates linear relationships of virus viability with the individual variables of RH and  $T$  and a nonlinear relationship with AH. Figure 1a shows a poor fit at higher temperatures, which can be explained by RH, as shown in a modified version of the figure in the electronic supplementary material, where viability is coloured by RH. At first glance, the relationship with AH shown in figure 1b appears to be linear; however, there is a slight curvature in the centre of the plot which is reflected in the predictive models described below. As shown in figure 1c, RH alone is a poor predictor, although it is evident that at low RH, viability is high. Over the range of values in the experiment, AH and  $T$  have a correlation coefficient of 0.651 ( $p < 0.0001$ ).

A model containing  $T$ , RH and their interaction as predictors is comparable, in terms of  $R^2$ , mean square error and predicted versus actual values, to a quadratic model containing only AH, as shown in table 1. Fitting the additional AH quadratic term results in a 15% reduction in the model root mean square error in comparison with the linear AH model, indicating a better fit to the data. Because AH is a function of both RH and  $T$ , any model that is a function of AH is effectively a function of RH and  $T$ . The electronic supplementary material contains parameter estimates for the models. While it may appear to be more appropriate to compare a model containing  $T$ , RH,  $T \times$  RH with a model containing  $T$ , AH,  $T \times$  AH, this is not the best approach for the Harper data. The correlation between  $T$  and AH ( $\rho = 0.651$ ) in this specific dataset results in statistical models indicating no relationship between viability and  $T$ , contrary



**Figure 2.** Residuals of influenza virus viability, quantified as the log of the per cent viable after 1 h, versus AH and RH after removing the temperature effect.

to scientific knowledge. In addition, the root mean square error increases by 20% in the  $T$ , AH,  $T \times \text{AH}$  model. This is a classic manifestation of multicollinearity (correlated regressors) in a regression model and is the reason this model is not presented for discussion. We fitted multiple additional models to the data, but do not include them in the discussion because they either contain correlated regressors or do not have the same predictive capability as the chosen models.

Another way of showing the influence of both temperature and humidity on virus viability is to first remove the temperature dependence and then examine the relationship between humidity and the residuals. Figure 2 shows the residuals of the linear regression of virus viability against temperature as a function of AH and RH. In both cases, the slope is negative, but the goodness of fit is better with RH than with AH ( $R^2$  of 0.70 versus 0.49).

In summary, the  $T$  and RH linear model with interactions and the quadratic AH model perform similarly in their ability to explain and predict influenza virus viability for this experimental dataset. The statistical models agree with the fundamental physical laws and equations that govern the relationships among the three quantities of  $T$ , AH and RH. For statistical purposes, we can interchange the two because a model based on AH is a function of  $T$  and RH and vice versa. First- and second-order models approximate relationships well in local areas. However, as the ranges of the independent variables change, the values of the coefficients, the order of the model and/or the significant terms are subject to change in order to obtain the best approximation [28].

At first glance, this result appears to conflict with previous studies concluding that AH modulates influenza virus survival, transmission and seasonality [1–4]. However, closer inspection shows that our result is consistent with these studies. While Shaman & Kohn's [3] analysis of Harper's data used simple linear regression with only one variable at a time and did not account for an increase in the variance of the response, we fit a more complex model that was appropriate given multicollinearity in the dataset. Had we employed Shaman & Kohn's [3] approach, we would have obtained the same results as they did. Using an epidemiological model, Shaman *et al.* [4] subsequently showed that AH alone could explain influenza seasonality in the USA. They demonstrated a statistically stronger association of AH than RH with the onset of wintertime influenza on the basis of comparing  $p$ -values of less than 0.00002 for AH and 0.00166 or lower for RH (table 1 in [4]), but small differences in  $p$ -values should not be used to rule out the influence of one variable versus another. McDevitt *et al.* [2] studied the inactivation of influenza virus on stainless steel surfaces and

found that 'The multiple linear regression model including independent variables for RH, temperature, and time... provides a reasonable means of predicting virus survival, but it does so at the expense of adding an additional parameter to the model', compared with the model including only AH and time. There was very little practical difference in the adjusted  $R^2$  values of the two models (0.90 versus 0.93, respectively), as in our results (table 1). Although the authors selected the model with AH and time because it was simpler, this choice does not invalidate the strength of the model containing RH, temperature and time. In an analysis of global observational data, Deyle *et al.* [1] stated that the effect of RH on influenza incidence was weaker than that of AH or temperature, but it was not absent. The magnitude of the weakness was not indicated, and the authors did not consider a model with the interaction term  $T \times \text{RH}$ , which might have produced different results.

We recently conducted a study that was explicitly designed to discriminate between AH and RH in the survival of the bacteriophage phi6, which has been suggested as a surrogate for influenza virus [29], in aerosols and droplets. The application of linear and log-linear multiple regression models to the data produced modest fits and varying results about the significance of AH and RH, but we achieved the best fit to the results using a random forest model, which indicated that RH and temperature were stronger predictors of virus survival than was AH [30].

We caution against using these results to draw conclusions about virus survival under real-world conditions. Influenza virus morphology can range from spherical, as in the laboratory-adapted strain A/Puerto Rico/8/34 (PR8), to filamentous, as maintained in nature [31], and the effect of morphology on virus survival is not known. Numerous studies have shown that the relationship between virus survival and humidity depends on the chemical composition of the suspension media. Both salt and protein concentrations affect virus viability [25]. We have recently shown that in a physiologically realistic suspension medium, specifically extracellular matrix from human bronchial epithelial cells, the strain A/California/07/2009 (H1N1pdm09) survives well in both droplets and aerosols independent of RH [32]. More studies are needed to confirm the relationship between strain, morphological phenotype, temperature, humidity and influenza virus survival in real respiratory fluid.

### 3.4. Influenza transmission in animal models

Limited animal studies have been performed to elucidate the role of humidity on the transmission of influenza viruses.

Studies in guinea pigs [33] and ferrets [34] have shown that dry conditions favour transmission. Reanalysis of the guinea pig results suggested that AH, rather than RH, modulates transmission [3], although the authors of the guinea pig study [35] subsequently noted, 'several of our transmission results are not explained by this relationship'. Transmission was completely blocked at 30°C and at two different fixed values of AH, transmission efficiency spanned the full range from 0 to 100% of contacts infected. Analysis of the transport of the virus between cages inside the chamber suggested that viral growth kinetics and background airflow in the chamber could explain the observations without invoking virus survival [36]. In addition, given the rapid forced airflow rates within experimental animal transmission systems, an estimated 10 s would have elapsed between release of the virus from the infected subjects and exposure to the test subjects [36]. This amount of time is likely to be insufficient for environmental conditions to affect the viability of viruses in released aerosols.

A carefully designed study is needed to test the hypothesis that humidity affects transmission by modulating virus survival. To isolate the effect, researchers would need to collect viral aerosols from infected animals and keep them suspended to allow sufficient time for ageing and inactivation at different humidities before exposing uninfected animals, ideally using a method that isolated transmission by the aerosol mode. Simultaneous measurements of virus loading in air, both in terms of total genome copies and infectious virus, are advisable to control for varying amounts of virus released by infected animals.

If the results from animal model influenza transmission studies are to be extrapolated to humans, greater understanding is needed of how the anatomical and symptomatic differences between animal models and humans contribute to the observed transmission characteristics [37,38]. Ferrets have been considered the 'gold standard' for influenza transmission studies, but there are several key physical differences between ferrets and humans that may impact how any transmission patterns (at different temperatures and humidities) seen in ferrets can be extrapolated to humans due to various allometric scaling issues. Some of these differences include body posture and the direction in which nostrils point; diameter, length and morphology of the airways; airflow velocity; respiratory rate and body temperature. All of these variables may affect the dynamics and deposition of inhaled particles of different sizes at different temperatures and humidities.

### 3.5. Epidemiology

In studies of influenza incidence versus meteorological conditions, some have found a stronger relationship with AH than RH, while others have reached the opposite conclusion. Davis *et al.* [39] reported an association between cold, dry conditions and influenza mortality in New York City. Using causality analysis on a global flu data, Deyle *et al.* [1] showed that AH, and to a lesser extent temperature, drives influenza incidence. Tang *et al.* [40] documented that the incidence of influenza A in Hong Kong increased with higher RH, while the incidence of influenza B decreased with a higher temperature and the incidence of neither was related to AH. In a study of paediatric hospital admissions in Singapore, Loh *et al.* [41] found that upper respiratory tract infections, which were correlated with positive test

results for influenza A, increased with lower RH and were not related to AH. Some of the apparent disagreement in results may be due to differences in climate. On average, Hong Kong and Singapore are warmer and more humid than New York City, and air conditioning and heating practices, which can dramatically influence indoor humidity, are likely to differ among locations. In some cases, the association with one measure of humidity versus another is weaker but not absent.

Because people spend over 90% of their time indoors [42], it is likely that influenza transmission among humans occurs mainly indoors rather than outdoors. If so, then indoor, rather than outdoor, environmental conditions should be more closely related to transmission, but indoor data are not widely available, especially for humidity. Nearly all of the epidemiological studies investigating links between temperature, humidity and influenza incidence have relied on outdoor measures of temperature and humidity. One exception is a study in a low-resource tropical setting with a limited sample size of 34 households [43]; the study did not detect differences in transmission efficiency as a function of temperature or humidity. To aid interpretation of epidemiological studies that use outdoor environmental data, we must understand the relationship between outdoor and indoor conditions.

Despite its seeming simplicity, or perhaps because of it, there have been few studies of the relationship between indoor and outdoor temperature and humidity. Indoor and outdoor temperatures were associated during the warm season but not during the cool season in New York City [44]. Indoor temperature measured in eight locations ranging from near the equator to the Arctic showed little variation across location and season [45]. A study in Boston, Massachusetts, showed that indoor and outdoor temperatures were strongly correlated when outdoor temperatures were greater than 13°C but only weakly correlated when outdoor temperatures were lower [46]. In all three studies, outdoor RH was only weakly correlated with indoor RH, while outdoor AH was strongly correlated with both indoor AH and indoor RH [44–46]. Unless water vapour is removed by an air conditioning unit or dehumidifier, or added by a humidifier, we expect indoor AH to be similar to that outdoor AH because of air exchange in a building, although sources such as humans, vegetation and water use could contribute water vapour to indoor air while absorptive materials could remove it. Indoor AH and RH are expected to be strongly correlated because indoor temperature is usually controlled to within a fairly narrow range [47], especially during winter unless the heating system employs humidification. Because AH and RH are proportional and move in tandem at a fixed temperature, it would be difficult to identify one versus the other as having a more significant effect indoors. We agree with Nguyen & Dockery [45] that outdoor AH serves a reliable proxy for indoor AH and RH in temperate regions, while outdoor RH does not.

In temperate regions during the cold season, indoor air is usually heated, and indoor and outdoor environmental parameters differ significantly. The cold season typically coincides with the peaks of influenza incidence in these regions. Although some studies have speculated that influenza seasonality may be related to indoor crowding effects [48,49], a causal relationship has not been shown [50]. There is now a need to study the relationship between

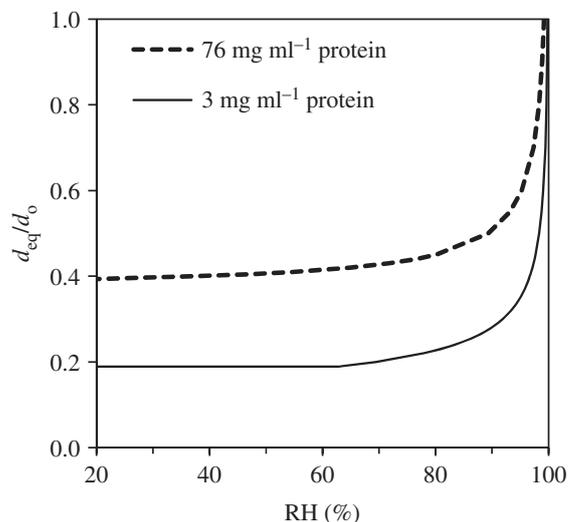
these seasonal winter influenza incidence peaks and indoor, rather than outdoor, environmental parameters. In studies showing that influenza seasonality is linked to outdoor AH in temperate regions, it is likely that outdoor AH is serving as a proxy for indoor AH or RH.

In tropical regions, the relationship between indoor humidity and outdoor conditions is not as easily predicted. Air conditioning removes water vapour from air, but the resulting indoor humidity will depend on the temperature differential between air entering and leaving the unit and the fraction of air that is recirculated. The prevalence of air conditioning varies with income and wealth [51], and thus, there may be differences in the indoor–outdoor humidity relationship as a function of these factors. One study found lower values of temperature and RH in businesses that needed to appear ‘high-status’, such as banks, dining facilities, retail shops and offices, compared with other buildings in tropical countries [52]; the researchers suggested that such conditions were more favourable for aerosol transmission of influenza. In a small study of influenza transmission in tropical settings that found no differences in transmission efficiency with indoor temperature or humidity, Tamerius *et al.* [43] concluded, ‘observational studies investigating the relationship between environment and influenza activity should use caution using outdoor environmental measurements since they can be imprecise estimates of the conditions that mediate transmission indoors’.

### 3.6. Mechanisms

When influenza viruses are emitted into air from an infected host by coughing, sneezing, talking or breathing [53–55], they are just one component of respiratory droplets that are a complex mixture of inorganic and organic ions, protein and surfactant [56]. The respiratory droplets can range in size from submicrometre to thousands of micrometres [57–64], although those larger than 100  $\mu\text{m}$  remain suspended for less than 5 s before settling to the ground from a height of 1.5 m. We expect temperature to affect the stability of viruses in the environment, including those that are airborne. In a review of the effect of environmental parameters on the survival of airborne pathogens, Tang [26] concluded that temperature and RH always interact to affect the survival of aerosolized viruses. One mystery is how RH could affect viruses in a respiratory droplet because unless they are present at the surface of the droplet, they are not exposed to ambient air and so would not directly interact with water vapour in the air.

Upon contact with ambient air, respiratory droplets are subject to evaporation until they reach equilibrium. Evaporation may be partial, in which case some water remains in liquid or semi-solid form, or complete, in which case only solutes and the virus remain, and likely some trapped water molecules. Air expired from the respiratory tract is saturated [65], whereas ambient air usually is not, so there is a driving force in the form of a vapour pressure gradient between the surface of a droplet and ambient air. The extent of evaporation and final, equilibrium size of the droplet is determined by ambient RH, not AH. This equilibrium size, which is attained within a few seconds at most [3,36,66], has important effects on the droplet’s physics and chemistry, although inactivation of any virus contained in the droplet appears to proceed more slowly [13].



**Figure 3.** Ratio of equilibrium diameter to initial diameter ( $d_{\text{eq}}/d_0$ ) as a function of RH for a droplet consisting of 9 mg ml<sup>-1</sup> NaCl, 3 or 76 mg ml<sup>-1</sup> protein and 0.5 mg l<sup>-1</sup> surfactant.

Combining the effects of curvature at the air–liquid interface (i.e. the Kelvin effect) and of solutes on the saturation vapour pressure of a droplet produces an equation that can be used to predict the equilibrium diameter of a droplet as a function of RH [67]. Figure 3 shows how the equilibrium size of a droplet of model respiratory fluid depends on RH in terms of the ratio of the equilibrium diameter to the initial diameter. Initially, the model fluid contains 9 mg ml<sup>-1</sup> salt as NaCl, 3 or 76 mg ml<sup>-1</sup> protein (range of values found in nasal surface airway fluid [10] and aerosols in exhaled breath condensate [11], respectively) and 0.5 mg ml<sup>-1</sup> surfactant as DPPC [12]. For reference, two other studies report total protein contents of respiratory fluid that are closer to the lower end of the range: 4 mg ml<sup>-1</sup> in unstimulated saliva [68] and 7 mg ml<sup>-1</sup> in nasal mucus [69].

The ratio shown in figure 3 applies for droplets of initial diameter as small as approximately 0.5  $\mu\text{m}$ , at which point the Kelvin effect becomes important. Figure 3 shows that at 100% RH, there is no evaporation, and the ratio is 1. At 90% RH, the droplets are dramatically smaller at equilibrium, 0.28 or 0.51 of their initial size for low and high-protein contents (3 mg ml<sup>-1</sup> and 76 mg ml<sup>-1</sup>), respectively. At 50% RH, these values are 0.19 and 0.41, respectively. For comparison, Liu *et al.* [70] predicted that dried droplet nuclei would be 0.32 of their original diameter. In fact, at RH below 64% for the low-protein droplets and below 42% for the high-protein droplets, the equilibrium diameter is unchanged because all bulk liquid water has been lost and only the solutes, or what some have called the droplet nucleus, remain.

The equilibrium droplet size is critical in determining its fate (e.g. [71–74]). Size determines how long the droplet remains suspended in air before it is removed by gravitational settling and where it deposits in the respiratory system if inhaled. The transformation of a droplet subject to evaporation in ambient air has a dramatic impact on its lifetime. We calculated the settling time for a droplet initially 10  $\mu\text{m}$  in diameter containing 9 mg ml<sup>-1</sup> salt as NaCl and 3 mg ml<sup>-1</sup> protein at RHs of 100%, 90% and 64% and lower. Table 2 shows that the droplet shrinks to 2.8  $\mu\text{m}$  at 90% RH and to 1.9  $\mu\text{m}$  at less than 64% RH. These diameters correspond to vastly different settling times; the 10  $\mu\text{m}$

**Table 2.** Parameters for a droplet initially 10  $\mu\text{m}$  in size containing 9  $\text{mg ml}^{-1}$  NaCl, 3  $\text{mg ml}^{-1}$  protein and 0.5  $\text{mg ml}^{-1}$  surfactant.

	100% RH	90% RH	<64% RH
equilibrium size	10 $\mu\text{m}$	2.8 $\mu\text{m}$	1.9 $\mu\text{m}$
time to settle 1.5 m	8 min	102 min	216 min
deposition efficiency in head airways	81%	73%	57%
deposition efficiency in tracheobronchial region	2%	6%	6%
deposition efficiency in alveolar region	2%	10%	12%
concentration factor	1	65	>7000

droplet at 100% RH remains suspended for only 8 min, whereas the 1.9  $\mu\text{m}$  droplet at less than 64% RH remains suspended for more than 3 h, posing a much longer opportunity for airborne transmission. Xie *et al.* [74] and Parienta *et al.* [72] present a much more complete analysis of droplet transformation and transport distance as a function of initial size and RH.

Furthermore, we can calculate the deposition efficiency in different regions of the respiratory system to contrast where the droplets might deposit if inhaled [75]. Table 2 shows that the 10  $\mu\text{m}$  droplet has a high chance of being deposited in the head airways (81%) and only a small chance of being deposited in the alveolar region (2%), whereas a 1.9  $\mu\text{m}$  droplet has a lower chance of being deposited in the head airways (57%) and a relatively higher chance of being deposited in the alveolar region (12%) than does the 10  $\mu\text{m}$  droplet. This calculation does not account for hygroscopic growth of droplets upon entering the saturated airway, although we have shown that model respiratory droplets containing surfactant do not reabsorb water even after extended exposure to saturated conditions [56]. A review of aerosol transmission of influenza suggests that infection initiated in the lower respiratory tract requires a lower dose and produces more severe symptoms compared to intranasal inoculation [76]. In summary, at lower RH, droplets stay suspended longer and are more likely to deposit in the lower respiratory tract. As mentioned earlier, the dynamics and deposition efficiencies of aerosols in animal models are likely to be quite different, for example in ferrets due to their more horizontal posture and much smaller airway diameters compared to those of humans [37,38].

Besides its effects on physics of a droplet, evaporation also affects its chemistry, which could affect the stability of a virus contained within the droplet. The mass of solutes in a respiratory fluid droplet containing one virus is expected to be at least five orders of magnitude larger than the mass of the virus [56], so these components should not be ignored when considering virus viability in droplets and aerosols. Table 2 shows the concentration factor, calculated as the ratio of the initial mass of water in the droplet to that remaining at equilibrium. This is the factor by which solutes that remain in the aqueous phase become concentrated due to loss of water if they do not precipitate out of solution. The solubility of NaCl is 360  $\text{mg ml}^{-1}$ , or 40 times the initial

concentration of 9  $\text{mg ml}^{-1}$ . Thus, we expect it to crystallize at an ambient RH of approximately 90% (concentration factor of 65) and lower, and indeed, we have observed crystallization of salt in model respiratory fluid droplets exposed to varying RH [25,56]. Under these concentrated conditions in droplets, protein may form aggregates [56], and there may be spatial gradients in pH [77]. In studies of atmospheric aerosols, researchers have observed phase separation, crystallization and changes in pH under conditions of changing RH [78–81]. While such changes in droplet chemistry could very well cause virus decay, the exact mechanism of inactivation is not known. Various possibilities have been proposed, such as osmotic bursting [82] or pH-sensitive changes in protein structure [24], but more studies are needed.

## 4. Conclusion

The debate over the importance of AH versus RH for influenza virus survival and transmission dates to the mid-twentieth century. Writing in *Nature* in 1960, Hemmes *et al.* [83] claimed, ‘it became evident that the relative humidity and not the absolute humidity is the determining factor’, for virus survival in aerosols. Analyses of the early twenty-first century have pointed to AH as the modulating factor. While it is appealing to try to isolate a single controlling environmental factor that modulates influenza virus survival, transmission and incidence, our analysis suggests that the combination of temperature and RH provides a consistent, mechanistically sound explanation of the observations. Temperature can be considered an intrinsic factor in virus stability because rates of inactivation of proteins and nucleic acid are expected to increase with temperature [16]. RH can be considered an extrinsic factor in virus stability because it controls evaporation, which affects a droplet’s size and physical fate and its chemical microenvironment. By contrast, there is no mechanism by which AH is expected to affect droplet diameter except through its relationship with RH.

The story is not yet complete, but we know that RH determines the extent of evaporation of an airborne droplet and thus the resulting chemical microenvironment to which the virus is exposed. If virus survival in the environment is a dominant factor in influenza transmission, then RH is expected to influence incidence and seasonality, too.

**Data accessibility.** All data used in this work have been previously published and can be found in the cited references.

**Authors’ contributions.** L.C.M. conceived of the study, analysed and interpreted data, and drafted the manuscript. J.W.T. conceived of the study and helped draft the manuscript. J.V.M. designed, carried out and interpreted the statistical analysis and helped draft the manuscript. S.S.L. provided information about influenza viruses and helped draft the manuscript.

**Competing interests.** We have no competing interests.

**Funding.** This work was supported by the National Institutes of Health (NIH) through the NIH Director’s New Innovator Award Program (1-DP2-A1112243) and the National Science Foundation (CBET-1438103 and ECCS-1542100).

**Acknowledgements.** The idea for this work originated at a workshop, ‘Viral Infectious Disease Transmission in the Built Environment’, at Yale-NUS in Singapore on 12–13 January 2017 which was sponsored by the Alfred P. Sloan Foundation. L.C.M. received support from a Fulbright scholarship during the preparation of this work.

## References

- Deyle ER, Maher MC, Hernandez RD, Basu S, Sugihara G. 2016 Global environmental drivers of influenza. *Proc. Natl Acad. Sci. USA* **113**, 13 081–13 086. (doi:10.1073/pnas.1607747113)
- McDevitt J, Rudnick S, First M, Spengler J. 2010 Role of absolute humidity in the inactivation of influenza viruses on stainless steel surfaces at elevated temperatures. *Appl. Environ. Microbiol.* **76**, 3943–3947. (doi:10.1128/aem.02674-09)
- Shaman J, Kohn M. 2009 Absolute humidity modulates influenza survival, transmission, and seasonality. *Proc. Natl Acad. Sci. USA* **106**, 3243–3248. (doi:10.1073/pnas.0806852106)
- Shaman J, Pitzer VE, Viboud C, Grenfell BT, Lipsitch M. 2010 Absolute humidity and the seasonal onset of influenza in the continental United States. *PLoS Biol.* **8**, e1000316. (doi:10.1371/journal.pbio.1000316)
- Davis RE, McGregor GR, Enfield KB. 2016 Humidity: a review and primer on atmospheric moisture and human health. *Environ. Res.* **144**, 106–116. (doi:10.1016/j.envres.2015.10.014)
- Wolkoff P. 2018 Indoor air humidity, air quality, and health — an overview. *Int. J. Hyg. Environ. Health* **221**, 376–390. (doi:10.1016/j.ijheh.2018.01.015)
- Irwin CK, Yoon KJ, Wang C, Hoff SJ, Zimmerman JJ, Denagamage T, O'Connor AM. 2011 Using the systematic review methodology to evaluate factors that influence the persistence of influenza virus in environmental matrices. *Appl. Environ. Microbiol.* **77**, 1049–1060. (doi:10.1128/aem.02733-09)
- Alduchov OA, Eskridge RE. 1996 Improved Magnus form approximation of saturation vapor pressure. *J. Appl. Meteorol.* **35**, 601–609. (doi:10.1175/1520-0450(1996)035<0601:IMFAOS>2.0.CO;2)
- Mikhailov E, Vlasenko S, Niessner R, Poschl U. 2004 Interaction of aerosol particles composed of protein and salts with water vapor: hygroscopic growth and microstructural rearrangement. *Atmos. Chem. Phys.* **4**, 323–350. (doi:10.5194/acp-4-323-2004)
- Gould JM, Weiser JN. 2001 Expression of C-reactive protein in the human respiratory tract. *Infect. Immun.* **69**, 1747–1754. (doi:10.1128/IAI.69.3.1747-1754.2001)
- Effros RM, Hoagland KW, Bosbous M, Castillo D, Foss B, Dunning M, Gare M, Lin W, Sun F. 2002 Dilution of respiratory solutes in exhaled condensates. *Resp. Crit. Care Med.* **165**, 663–669. (doi:10.1164/ajrccm.165.5.2101018)
- Veldhuizen EJ, Haagsman HP. 2000 Role of pulmonary surfactant components in surface film formation and dynamics. *Biochim. Biophys. Acta* **1467**, 255–270. (doi:10.1016/S0005-2736(00)00256-X)
- Harper GJ. 1961 Airborne micro-organisms: survival tests with four viruses. *J. Hyg. Cambridge* **59**, 479–486. (doi:10.1017/S0022172400039176)
- Spring SB, Maassab HF, Kendal AP, Murphy BR, Chanock RM. 1977 Cold-adapted variants of influenza virus A: I. Comparison of the genetic properties of ts mutants and five cold-adapted variants of influenza virus A. *Virology* **77**, 337–343. (doi:10.1016/0042-6822(77)90430-5)
- Spring SB, Maassab HF, Kendal AP, Murphy BR, Chanock RM. 1977 Cold adapted variants of influenza A. *Arch. Virol.* **55**, 233–246. (doi:10.1007/bf01319909)
- Woese C. 1960 Thermal inactivation of animal viruses. *Ann. NY Acad. Sci.* **83**, 741–751. (doi:10.1111/j.1749-6632.1960.tb40943.x)
- Polozov IV, Bezrukov L, Gawrisch K, Zimmerberg J. 2008 Progressive ordering with decreasing temperature of the phospholipids of influenza virus. *Nat. Chem. Biol.* **4**, 248–255. (doi:10.1038/nchembio.77)
- DuBois RM, Zaraket H, Reddivari M, Heath RJ, White SW, Russell CJ. 2011 Acid stability of the hemagglutinin protein regulates H5N1 influenza virus pathogenicity. *PLoS Pathog.* **7**, e1002398. (doi:10.1371/journal.ppat.1002398)
- Galloway SE, Reed ML, Russell CJ, Steinhauer DA. 2013 Influenza HA subtypes demonstrate divergent phenotypes for cleavage activation and pH of fusion: implications for host range and adaptation. *PLoS Pathog.* **9**, e1003151. (doi:10.1371/journal.ppat.1003151)
- Haywood AM, Boyer BP. 1986 Time and temperature dependence of influenza virus membrane fusion at neutral pH. *J. Gen. Virol.* **67**, 2813–2817. (doi:10.1099/0022-1317-67-12-2813)
- Imai M *et al.* 2012 Experimental adaptation of an influenza H5 HA confers respiratory droplet transmission to a reassortant H5 HA/H1N1 virus in ferrets. *Nature* **486**, 420. (doi:10.1038/nature10831)
- O'Donnell CD, Vogel L, Matsuoka Y, Jin H, Subbarao K. 2014 The matrix gene segment destabilizes the acid and thermal stability of the hemagglutinin of pandemic live attenuated influenza virus vaccines. *J. Virol.* **88**, 12 374–12 384. (doi:10.1128/jvi.01107-14)
- Scholtissek C. 1985 Stability of infectious influenza A viruses at low pH and at elevated temperature. *Vaccine* **3**, 215–218. (doi:10.1016/0264-410X(85)90109-4)
- Yang W, Marr LC. 2012 Mechanisms by which ambient humidity may affect viruses in aerosols. *Appl. Environ. Microbiol.* **78**, 6781. (doi:10.1128/AEM.01658-12)
- Yang W, Elankumaran S, Marr LC. 2012 Relationship between humidity and influenza A viability in droplets and implications for influenza's seasonality. *PLoS ONE* **7**, e46789. (doi:10.1371/journal.pone.0046789)
- Tang JW. 2009 The effect of environmental parameters on the survival of airborne infectious agents. *J. R. Soc. Interface* **6**, S737–S746. (doi:10.1098/rsif.2009.0227.focus)
- Zhao Y, Aarnink AJA, Dijkman R, Fabri T, de Jong MCM, Groot Koerkamp PWG. 2012 Effects of temperature, relative humidity, absolute humidity, and evaporation potential on survival of airborne Gumboro vaccine virus. *Appl. Environ. Microbiol.* **78**, 1048–1054. (doi:10.1128/aem.06477-11)
- Myers RH, Montgomery DC, Anderson-Cook CM. 2016 *Response surface methodology: process and product optimization using designed experiments*, 4th edn. Hoboken, NJ: John Wiley & Sons, Inc.
- Turgeon N, Toulouse M-J, Martel B, Moineau S, Duchaine C. 2014 Comparison of five bacteriophages as models for viral aerosol studies. *Appl. Environ. Microbiol.* **80**, 4242–4250. (doi:10.1128/aem.00767-14)
- Prussin II AJ, Schwake DO, Lin K, Gallagher DL, Buttlng L, Marr LC. 2018 Survival of the enveloped virus Phi6 in droplets as a function of relative humidity, absolute humidity, and temperature. *Appl. Environ. Microbiol.* **84**, e00551-18. (doi:10.1128/AEM.00551-18)
- Seladi-Schulman J, Steel J, Lowen AC. 2013 Spherical influenza viruses have a fitness advantage in embryonated eggs, while filament-producing strains are selected *in vivo*. *J. Virol.* **87**, 13 343–13 353. (doi:10.1128/jvi.02004-13)
- Kormuth KA, Lin K, Prussin II AJ, Vejerano EP, Tiwari AJ, Cox SC, Myerburg MM, Lakdawala SS, Marr LC. 2018 Influenza virus infectivity is retained in aerosols and droplets independent of relative humidity. *J. Infect. Dis.* **218**, 739–747. (doi:10.1093/infdis/jiy221)
- Lowen AC, Mubareka S, Steel J, Palese P. 2007 Influenza virus transmission is dependent on relative humidity and temperature. *PLoS Pathog.* **3**, 1470–1476. (doi:10.1371/journal.ppat.0030151)
- Gustin KM, Belsler JA, Veguilla V, Zeng H, Katz JM, Tumpey TM, Maines TR. 2015 Environmental conditions affect exhalation of H3N2 seasonal and variant influenza viruses and respiratory droplet transmission in ferrets. *PLoS ONE* **10**, e0125874. (doi:10.1371/journal.pone.0125874)
- Lowen AC, Steel J. 2014 Roles of humidity and temperature in shaping influenza seasonality. *J. Virol.* **88**, 7692–7695. (doi:10.1128/jvi.03544-13)
- Halloran SK, Wexler AS, Ristenpart WD. 2012 A comprehensive breath plume model for disease transmission via expiratory aerosols. *PLoS ONE* **7**, e7088. (doi:10.1371/journal.pone.0037088)
- Belsler JA, Katz JM, Tumpey TM. 2011 The ferret as a model organism to study influenza A virus infection. *Dis. Model. Mech.* **4**, 575–579. (doi:10.1242/dmm.007823)
- Buhnerkempe MG, Gostic K, Park M, Ahsan P, Belsler JA, Lloyd-Smith JO. 2015 Mapping influenza transmission in the ferret model to transmission in humans. *eLife* **4**, e07969. (doi:10.7554/eLife.07969)
- Davis RE, Rossier CE, Enfield KB. 2012 The impact of weather on influenza and pneumonia mortality in New York City, 1975–2002: a retrospective study. *PLoS ONE* **7**, e34091. (doi:10.1371/journal.pone.0034091)
- Tang JW, Lai FY, Wong F, Hon KL. 2010 Incidence of common respiratory viral infections related to climate factors in hospitalized children in Hong

- Kong. *Epidemiol. Infect.* **138**, 226–235. (doi:10.1017/s0950268809990410)
41. Loh TP, Lai FY, Tan ES, Thoon KC, Tee NW, Cutter J, Tang JW. 2011 Correlations between clinical illness, respiratory virus infections and climate factors in a tropical paediatric population. *Epidemiol. Infect.* **139**, 1884–1894. (doi:10.1017/s0950268810002955)
  42. Klepeis NE, Nelson WC, Ott WR, Robinson JP, Tsang AM, Switzer P, Behar JV, Hern SC, Engelmann WH. 2001 The National Human Activity Pattern Survey (NHAPS): a resource for assessing exposure to environmental pollutants. *J. Expo. Sci. Environ. Epidemiol.* **11**, 231–252. (doi:10.1038/sj.jea.7500165)
  43. Tamerius J, Ojeda S, Uejio CK, Shaman J, Lopez B, Sanchez N, Gordon A. 2017 Influenza transmission during extreme indoor conditions in a low-resource tropical setting. *Int. J. Biometeorol.* **61**, 613–622. (doi:10.1007/s00484-016-1238-4)
  44. Tamerius J, Perzanowski M, Acosta L, Jacobson J, Goldstein I, Quinn J, Rundle A, Shaman J. 2013 Socioeconomic and outdoor meteorological determinants of indoor temperature and humidity in New York City dwellings. *Weather Clim. Soc.* **5**, 168–179. (doi:10.1175/wcas-d-12-00030.1)
  45. Nguyen JL, Dockery DW. 2016 Daily indoor-to-outdoor temperature and humidity relationships: a sample across seasons and diverse climatic regions. *Int. J. Biometeorol.* **60**, 221–229. (doi:10.1007/s00484-015-1019-5)
  46. Nguyen JL, Schwartz J, Dockery DW. 2014 The relationship between indoor and outdoor temperature, apparent temperature, relative humidity, and absolute humidity. *Indoor Air* **24**, 103–112. (doi:10.1111/ina.12052)
  47. Hoppe P, Martinac I. 1998 Indoor climate and air quality. Review of current and future topics in the field of ISB study group 10. *Int. J. Biometeorol.* **42**, 1–7. (doi:10.1007/s004840050075)
  48. Dowell SF. 2012 Seasonality — still confusing. *Epidemiol. Infect.* **140**, 87–90. (doi:10.1017/s0950268811001695)
  49. Murray EL *et al.* 2012 Rainfall, household crowding, and acute respiratory infections in the tropics. *Epidemiol. Infect.* **140**, 78–86. (doi:10.1017/s0950268811000252)
  50. Lofgren E, Fefferman NH, Naumov YN, Gorski J, Naumova EN. 2007 Influenza seasonality: underlying causes and modeling theories. *J. Virol.* **81**, 5429–5436. (doi:10.1128/jvi.01680-06)
  51. International Energy Agency. 2018 The future of cooling: opportunities for energy-efficient air conditioning. See <https://www.iea.org/futureofcooling/>.
  52. Hanley BP, Borup B. 2010 Aerosol influenza transmission risk contours: a study of humid tropics versus winter temperate zone. *Virol. J.* **7**, 98. (doi:10.1186/1743-422x-7-98)
  53. Lindsley WG *et al.* 2010 Measurements of airborne influenza virus in aerosol particles from human coughs. *PLoS ONE* **5**, e15100. (doi:10.1371/journal.pone.0015100)
  54. Milton DK, Fabian MP, Cowling BJ, Grantham ML, McDevitt JJ. 2013 Influenza virus aerosols in human exhaled breath: particle size, culturability, and effect of surgical masks. *PLoS Pathog.* **9**, e1003205. (doi:10.1371/journal.ppat.1003205)
  55. Yan J, Grantham M, Pantelic J, Bueno de Mesquita PJ, Albert B, Liu F, Ehrman S, Milton DK. 2018 Infectious virus in exhaled breath of symptomatic seasonal influenza cases from a college community. *Proc. Natl Acad. Sci. USA* **115**, 1081–1086. (doi:10.1073/pnas.1716561115)
  56. Vejerano EP, Marr LC. 2018 Physico-chemical characteristics of evaporating respiratory fluid droplets. *J. R. Soc. Interface* **15**, 20170939. (doi:10.1098/rsif.2017.0939)
  57. Chao CYH *et al.* 2009 Characterization of expiration air jets and droplet size distributions immediately at the mouth opening. *J. Aerosol Sci.* **40**, 122–133. (doi:10.1016/j.jaerosci.2008.10.003)
  58. Duguid JP. 1946 The size and the duration of air-carriage of respiratory droplets and droplet-nuclei. *J. Hyg. Cambridge* **44**, 471–479. (doi:10.1017/S0022172400019288)
  59. Johnson GR *et al.* 2011 Modality of human expired aerosol size distributions. *J. Aerosol Sci.* **42**, 839–851. (doi:10.1016/j.jaerosci.2011.07.009)
  60. Lindsley WG *et al.* 2012 Quantity and size distribution of cough-generated aerosol particles produced by influenza patients during and after illness. *J. Occup. Environ. Hyg.* **9**, 443–449. (doi:10.1080/15459624.2012.684582)
  61. Loudon RG, Roberts RM. 1967 Droplet expulsion from the respiratory tract. *Am. Rev. Respir. Dis.* **95**, 435–442. (doi:10.1164/arrd.1967.95.3.435)
  62. Morawska L, Johnson GR, Ristovski ZD, Hargreaves M, Mengersen K, Corbett S, Chao CYH, Katoshevski D. 2009 Size distribution and sites of origin of droplets expelled from the human respiratory tract during expiratory activities. *J. Aerosol Sci.* **40**, 257–269. (doi:10.1016/j.jaerosci.2008.11.002)
  63. Papineni RS, Rosenthal FS. 1997 The size distribution of droplets in the exhaled breath of healthy human subjects. *J. Aerosol Med.* **10**, 105–116. (doi:10.1089/jam.1997.10.105)
  64. Xie X, Li Y, Sun H, Liu L. 2009 Exhaled droplets due to talking and coughing. *J. R. Soc. Interface* **6**, S703–S714. (doi:10.1098/rsif.2009.0388)
  65. Walker JEC, Wells RE. 1961 Heat and water exchange in the respiratory tract. *Am. J. Med.* **30**, 259–267. (doi:10.1016/0002-9343(61)90097-3)
  66. Nicas M, Jones RM. 2009 Relative contributions of four exposure pathways to influenza infection risk. *Risk Anal.* **29**, 1292–1303. (doi:10.1111/j.1539-6924.2009.01253.x)
  67. Seinfeld JH, Pandis SN. 2006 *Atmospheric chemistry and physics — from air pollution to climate change*, 2nd edn. Hoboken, NJ: John Wiley & Sons, Inc.
  68. Sanchez GA, Miozza V, Delgado A, Busch L. 2011 Determination of salivary levels of mucin and amylase in chronic periodontitis patients. *J. Periodontol Res.* **46**, 221–227. (doi:10.1111/j.1600-0765.2010.01332.x)
  69. Ruocco L, Fattori B, Romanelli A, Martelloni M, Casani A, Samolewska M, Rezzonico R. 1998 A new collection method for the evaluation of nasal mucus proteins. *Clin. Exp. Allergy* **28**, 881–888. (doi:10.1046/j.1365-2222.1998.00312.x)
  70. Liu L, Wei J, Li Y, Ooi A. 2017 Evaporation and dispersion of respiratory droplets from coughing. *Indoor Air* **27**, 179–190. (doi:10.1111/ina.12297)
  71. Knight V. 1980 Viruses as agents of airborne contagion. *Ann. NY Acad. Sci.* **353**, 147–156. (doi:10.1111/j.1749-6632.1980.tb18917.x)
  72. Parienta D *et al.* 2011 Theoretical analysis of the motion and evaporation of exhaled respiratory droplets of mixed composition. *J. Aerosol Sci.* **42**, 1–10. (doi:10.1016/j.jaerosci.2010.10.005)
  73. Wells WF. 1934 On air-borne infection. Study II. Droplets and droplet nuclei. *Am. J. Epidemiol.* **20**, 611–618. (doi:10.1093/oxfordjournals.aje.a118097)
  74. Xie X, Li Y, Chwang ATY, Ho PL, Seto WH. 2007 How far droplets can move in indoor environments — revisiting the Wells evaporation-falling curve. *Indoor Air* **17**, 211–225. (doi:10.1111/j.1600-0668.2007.00469.x)
  75. ICRP. 1994 Human respiratory tract model for radiological protection. ICRP publication 66. *Ann. ICRP* **24**, 1–482.
  76. Tellier R. 2006 Review of aerosol transmission of influenza A virus. *Emerg. Infect. Dis.* **12**, 1657–1662. (doi:10.3201/eid1211.060426)
  77. Wei H, Vejerano EP, Leng W, Huang Q, Willner MR, Marr LC, Vikesland PJ. 2018 Aerosol droplets exhibit a stable pH gradient. *Proc. Natl Acad. Sci. USA* **115**, 7272–7277. (doi:10.1073/pnas.1720488115)
  78. Bertram AK *et al.* 2011 Predicting the relative humidities of liquid–liquid phase separation, efflorescence, and deliquescence of mixed particles of ammonium sulfate, organic material, and water using the organic-to-sulfate mass ratio of the particle and the oxygen-to-carbon elemental ratio of the organic component. *Atmos. Chem. Phys.* **11**, 10 995–11 006. (doi:10.5194/acp-11-10995-2011)
  79. Laskina O, Morris HS, Grandquist JR, Qin Z, Stone EA, Tivanski AV, Grassian VH. 2015 Size matters in the water uptake and hygroscopic growth of atmospherically relevant multicomponent aerosol particles. *J. Phys. Chem. A* **119**, 4489–4497. (doi:10.1021/jp510268p)
  80. Rindelaub JD, Craig RL, Nandy L, Bondy AL, Dutcher CS, Shepson PB, Ault AP. 2016 Direct measurement of pH in individual particles via Raman microspectroscopy and variation in acidity with relative humidity. *J. Phys. Chem. A* **120**, 911–917. (doi:10.1021/acs.jpca.5b12699)
  81. You Y *et al.* 2012 Images reveal that atmospheric particles can undergo liquid–liquid phase separations. *Proc. Natl Acad. Sci. USA* **109**, 13 188–13 192. (doi:10.1073/pnas.1206414109)
  82. Ud-Dean SMM. 2010 Structural explanation for the effect of humidity on persistence of airborne virus: seasonality of influenza. *J. Theor. Biol.* **264**, 822–829. (doi:10.1016/j.jtbi.2010.03.013)
  83. Hemmes JH, Winkler KC, Kool SM. 1960 Virus survival as a seasonal factor in influenza and poliomyelitis. *Nature* **188**, 430–431. (doi:10.1038/188430a0)