

**EFFECTS OF DIETARY TYROSINE AND TRYPTOPHAN SUPPLEMENTATION
ON IMMUNITY AND BRAIN NEUROTRANSMITTER LEVELS
AFTER SRBC INJECTION IN CHICKENS**

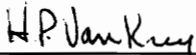
by
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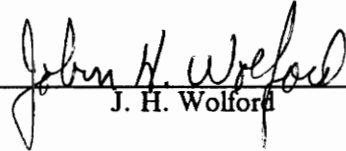
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(ABSTRACT)

This study investigated the effects of dietary tyrosine or tryptophan supplementation on immunity and brain neurotransmitter levels after antigen challenge. Chickens were given 0.1%, 0.5% or 1% supplemental dietary L-tyrosine or L-tryptophan prior to the injection of sheep red blood cells (SRBC). The 0.1% tyrosine supplementation increased primary IgM and secondary IgG titers at some time periods in Leghorns and decreased secondary IgM titers at Day 5 in broilers, while 0.1% tryptophan addition decreased Leghorn secondary IgM titers and increased secondary IgG titers at Day 9 and broiler secondary IgM titers at Day 9. The phytohemagglutinin (PHA) wattle response in Leghorns and broilers and resistance of Leghorns to *E. coli* challenge were not affected with the 0.1% supplemental level. With higher levels, 0.5% tyrosine supplementation increased Leghorn primary IgM titers at Day 11. In broilers, 0.5 and 1% tryptophan supplementation decreased secondary total antibody titers at Day 2, while the secondary IgM titers at Day 6 with the 0.5% tyrosine supplementation were higher than those with 1% tyrosine or tryptophan supplementation. The 0.5% or 1% tryptophan supplementation also lowered stressed broiler primary IgM titers at Day 3. After pooling the titer results within two dietary supplemental levels (0.5% and 1%) of a given amino acid, the tyrosine treatment appeared to suppress antibody response in unstressed broilers, but not stressed ones, while tryptophan displayed a suppressive trend in broilers under both situations. Supplementation with 0.5% or 1% dietary tyrosine did not alter brain catecholamine or serotonin (5-HT) levels in Leghorns. In contrast, 0.5% or 1% dietary tryptophan supplementation generally increased 5-HT, and its metabolite 5-HIAA, and 5-HIAA/5-HT ratios in the diencephalon, telecephalon, and brain stem in Leghorns. Tryptophan

supplementation caused a dose-response increase in 5-HT and 5-HIAA levels of the brain stem. The results demonstrated that 0.5-1% dietary tryptophan supplementation suppressed broiler, but not Leghorn, antibody response, and the mechanism was probably via enhancing 5-HT synthesis and release. The results also suggested that lower levels (0.1-0.5%) of dietary tyrosine supplementation may enhance immune response in Leghorns and stressed broilers, but ingesting large quantity of tyrosine, occurred in broilers, suppresses the response.

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Chapter 1

LITERATURE REVIEW

Effects of Brain Catecholamines and Serotonin on Neuroimmunomodulation

It has been understood for some time that psychological factors, such as stressors, decrease immune function and increase the incidence of disease, thus, leading to the conclusion that the brain is able to influence the activity of the immune system (Dunn, 1989). Not all stressors, however, cause suppression of the immune response; some even increase it. For example, continued auditory stimulation enhanced the lymphocyte response to mitogens (Monjan and Collector, 1977). Accordingly, the effect of stress on the immune system may depend on how the brain reacts to the stressor. The disparate effects of stressors may involve different neurohormonal responses.

There are two available routes by which the brain can signal the immune system. These include the neuroendocrine route and the autonomic nerves (Felen *et al.*, 1988). The neuroendocrine route involves an activation of the hypothalamo-hypophyseal-adrenocortical (HPA) axis resulting in the secretion of adrenal glucocorticoids and neuropeptides (Bateman *et al.*, 1989). The autonomic nerves, on the other hand, directly innervate immune organs (Felen *et al.*, 1988). Much attention has been focused on the effect of the neuroendocrine route on the immunological response.

The immediate products resulting from activation of the HPA axis: *i.e.* adrenocorticotrophic hormone (ACTH), β -endorphin, and a number of other peptides may participate in neuroimmunomodulation (reviewed by Bateman *et al.*, 1989). β -Endorphin enhanced the proliferative response of splenic lymphocytes to T-cell mitogens (Gilman *et al.*, 1982) and natural killer cell activity (Carr *et al.*, 1986), whereas ACTH inhibited spleen cell antibody response against both T-cell dependent and T-cell independent antigens (Johnson *et al.*, 1982). Glucocorticoids, whose release is stimulated by ACTH, inhibited the production of IL-2 (Kelso *et al.*, 1984), which is an important T-cell growth factor. Furthermore, glucocorticoids decreased cell

growth and number in LPS-stimulated lymphocytes (Roess *et al.*, 1982), though considerable species differences existed in the glucocorticoid sensitivity of B lymphocytes (Bateman *et al.*, 1989). Catecholamines and serotonin appeared able to influence the activity of the HPA axis, as will be discussed below.

Corticotrophic-releasing factor (CRF) is elaborated primarily by the paraventricular nucleus (PVN) and is transported to the pituitary gland via the hypothalamo-hypophysial portal system (Bateman *et al.*, 1989). There, it stimulates ACTH release. The endorphins and other proopiomelanocortin (POMC)-derived peptides may also be secreted in response to CRF stimulation (Bateman *et al.*, 1989).

The injection of norepinephrine (NE) and epinephrine into the paraventricular nucleus (PVN) can stimulate CRF release, and catecholaminergic inputs in this area originate in the brain stem (Plotsky *et al.*, 1989). However, the secretion of ACTH is not controlled only by CRF. Vasopressin (Whitnall, 1988) and oxytocin (Gibbs, 1986) are also important ACTH secretagogues. Clarke *et al.* (1989) proposed that the inputs from the brain stem provide information about the physiological state, while the descending inputs from the limbic system provide information predominantly about the emotional state. The inputs from the limbic system are oxytocinergic and vasopressinergic, with some axons extending to the pituitary (Swanson *et al.*, 1983). Accordingly, vasopressin and oxytocin in the limbic system may play a more important role in release of ACTH in stress than catecholamines do in the PVN.

Moreover, brain catecholamines have also been shown to participate in inhibition of ACTH release. Ganong *et al.* (1977) found that a variety of catecholamine-releasing drugs, including amphetamine, methamphetamine and dihydroxyphenylalanine (L-dopa), caused similar inhibition. Furthermore, the inhibitory effect of L-dopa could be blocked by the decarboxylase inhibitor benserazide. He also reported that clonidine inhibited ACTH secretion, and the α -adrenergic blocker phenoxybenzamine prevented the effect of clonidine when administered into the third cerebroventricle. On the other hand, the β -adrenoceptor antagonist prazosin and the tyrosine hydroxylase inhibitor α -methyl-p-tyrosine increased the hypothalamic CRF content and the plasma ACTH concentration, and exaggerated the HPA axis response to stress (Buckingham

et al., 1987). Injection of 6-hydroxydopamine (6-OHDA) into the third cerebroventricle also caused a significant rise of the plasma cortisol level during stress (Domanski *et al.*, 1985). Therefore, it appears that the catecholaminergic neurons located outside the brain stem may play an inhibitory role in the HPA axis by decreasing CRF release.

Dopamine (DA) also participates in the regulation of the HPA axis. Sapun-Malcolm *et al.* (1986) reported that dopamine had an inhibitory effect in the control of pituitary released β -endorphin-like immunoreactivity. This result was confirmed by Loeffler (1988) who reported that dopamine inhibited POMC gene expression in the intermediate lobe of the pituitary. The pituitary secretion of these hormones was increased during periods of physical and psychological stress, and, therefore, contributed to stress-induced immunosuppression (reviewed by Cannon *et al.*, 1986).

Further evidence shows that catecholaminergic neurons participate in immune regulation. Cross *et al.* (1986) demonstrated that injection of 6-OHDA into the cisterna magna resulted in marked inhibition of the antibody response to SRBC, and inhibition was not related to changes in circulating corticosterone. Immunizing rats with SRBC caused decreased turnover of norepinephrine (NE) in the hypothalamus (Besedovsky *et al.*, 1983). Conversely, Carlson *et al.* (1987) reported an increased turnover of NE in the PVN, but not other sites of the hypothalamus, following inoculation with SRBC. Probably the strongest evidence, which was reported by Saphier (1989), was that an increase in antibody titers was accompanied by a rise in NE levels in the preoptic area/anterior hypothalamus (PO/AH), and an increase in plasma corticosterone concentration occurred following a large decrease of NE levels in the PO/AH during the primary response to SRBC. In addition, in an extensive series of experiments, Devoino *et al.* (1987) found that administration of L-dopa, the precursor of dopamine, enhanced the immune response, an effect believed to be mediated by the central nervous system.

In contrast to the catecholamines, indoleamines appear to stimulate the HPA axis. Serotonin was reported to possess a stimulatory role in the control of CRF secretion (Holmes *et al.*, 1982). Furthermore, it has been shown to exert a stimulatory effect on the pituitary release of β -endorphin-like immunoreactivity, with dopamine and serotonin neurons appearing to exert

reciprocal and independent control of pituitary β -endorphin secretion *in vivo* (Sapum-Malcom *et al.*, 1986).

Serotonin plays an inhibitory role in the immune system. Devoino and Ilyutchenok (1968) reported that increasing brain serotonin (5-HT) levels by giving daily injections of 5-hydroxytryptophan (5-HPT), the precursor of serotonin, decreased delayed-type hypersensitivity in a dose-dependent manner. Likewise, increasing the brain serotonin level, using either monoamine oxidase (MAO) inhibitors or 5-HPT, resulted in suppression of humoral immunity (Devoino *et al.*, 1973; 1987) and transplantation immunity (Draskoci *et al.*, 1964; Pierpaoli *et al.*, 1978). The immunological deficits seen after increasing brain 5-HT were a result of decreased IgM antibody titers in the primary response, and a lack of a secondary IgG response (Devoino *et al.*, 1973).

Conversely, decreasing brain serotonin appeared to increase the immune response. Lesioning the midbrain raphe nucleus, the major location of serotonergic cell bodies, resulted in a two-fold increase in antibody titers to bovine serum albumin (reviewed by Devoino *et al.*, 1987). Injections of the tryptophan hydroxylase inhibitor p-chlorophenylalanine also resulted in an enhanced immune response (Devoino *et al.*, 1987).

The catecholamines and serotonin can also influence the immune response peripherally. In chickens, injection of serotonin 30 min. prior to antigen injection decreased the IgM and IgG plaque-forming cell response against SRBC (Gray *et al.*, 1986). However, the peripheral role of catecholamines appeared less clear. Besedovesky *et al.* (1979) showed that norepinephrine suppressed the *in vitro* anti-SRBC response of murine spleen cells, whereas dopamine enhanced the response. Conversely, Kouassi *et al.* (1988) found that the catecholamines dopamine and norepinephrine exerted stimulatory and inhibitory effects, respectively, on murine polyclonal B-cell activation.

Effect of Tryptophan Availability on Serotonin Synthesis and Physiological Activities

Hydroxylation of tryptophan to 5-hydroxytryptophan, catalyzed by the enzyme tryptophan hydroxylase (TpOH), is the rate limiting step in the synthesis of serotonin. The factors regulating the activity of TpOH are not fully understood, but availability of tryptophan can influence the reaction (Sved, 1983). The enzyme was reported to be unsaturated (Kaufman, 1974) since brain tryptophan levels are about 20 μM , well below the 50 μM K_m of TpOH. End product inhibition does not appear to operate, at least at physiological levels of serotonin (Cooper *et al.*, 1978). Increasing brain tryptophan levels elevated brain serotonin concentration (Ashcroft *et al.*, 1964; Weber *et al.*, 1965), and its major metabolite 5-hydroxyindoleacetic acid (5-HIAA) (Curzon *et al.*, 1974; Green *et al.*, 1975). On the other hand, decreasing tryptophan availability has an opposite effect (Fernstrom *et al.*, 1975). Studies using HPLC with electrochemical detection to measure serotonin and 5-HIAA confirmed the above conclusion (Marsden *et al.*, 1980; Reinhard *et al.*, 1980). Using the rate at which brain serotonin accumulates after inhibition of monoamine oxidase (MAO), the enzyme that breaks down the indoleamine, also provided evidence that the rate of serotonin synthesis varied directly with the brain tryptophan level (Knott *et al.*, 1974). This effect occurred in regions containing primarily serotonergic cell bodies as well as in regions containing terminals of serotonergic neurons.

Administration of tryptophan can also influence physiological activities. Tryptophan administration consistently reduced sleep latency, and in some studies, also increased total sleep time (Griffiths *et al.*, 1972; Hartmann *et al.*, 1977). Sved *et al.* (1982) found that tryptophan injection lowered blood pressure in spontaneously hypertensive rats. The action of tryptophan could be blocked by the serotonin receptor antagonist metergoline, or by the inhibitor of serotonin synthesis, p-chlorophenylalanine. Regarding anterior pituitary function, Sapun *et al.* (1981) provided evidence that β -endorphin release from the pituitary was increased by the administration of tryptophan, as well as by drugs that enhanced serotonergic neurotransmission.

Data on the effect of tryptophan administration on blood ACTH and corticosterone levels is conflicting, with some laboratories reporting a decrease (Woolf *et al.*, 1977) and others reporting an increase (Modlinger *et al.*, 1979). Kennett and Joseph (1981) examined the effect of valine injection on the stress-induced elevation of serum corticosterone levels. Valine injection suppressed the stress-induced rise in serum corticosterone and in brain 5-HIAA. They argued that valine prevented the increase in brain serotonin synthesis, since valine and tryptophan use the same transport system to enter the brain.

Effect of Tyrosine Supplement on Catecholamine Synthesis and Stress

The initial reaction in the biosynthesis of DA, NE, and epinephrine is hydroxylation of tyrosine to form L-dopa. This is the rate-limiting step in catecholamine synthesis. The availability of tyrosine have been proposed as one of the regulators of the rate at which tyrosine is hydroxylated, and, thus, catecholamines synthesized (Sved, 1983). Tyrosine hydroxylase (TOH) is mostly, but not fully, saturated with tyrosine *in vivo* (Kaufman, 1975), which may limit the regulatory effect of tyrosine availability on catecholamine synthesis. Studies on the effect of increasing brain tyrosine showed that DA and NE levels were not changed (Dairman, 1979; Sved, 1983). However, Acworth *et al.* (1988) recently demonstrated that tyrosine administration caused a short-lived dose-related increase in extracellular DA levels in the nigrostriatum, as well as a non dose-related increase in 3, 4-dihydroxyphenylacetic acid (DOPAC) and 4-hydroxy-3-methoxyphenylacetic acid (HVA), the major metabolites of dopamine. On the other hand, high frequency firing markedly affected the responsiveness of the neurons to added tyrosine (Weiner *et al.*, 1978). Tyrosine increased DA synthesis within neurons with high spontaneous firing frequencies and bursting activities (Bannon *et al.*, 1983).

Although the relationship between tyrosine loading and catecholamine synthesis is not significant under normal physiological conditions, administration of tyrosine reduced behavioral and neurochemical deficits in stressful situations. Banderet *et al.* (1989) reported that tyrosine

significantly decreased symptoms, adverse moods, and performance impairments in subjects who exhibited average or greater responses to environmental conditions. There are other experiments showing that tyrosine, given acutely or in the diet, protected rodents from both the neurochemical and behavioral effects of acute stressors such as tail shock or cold exposure (Brady *et al.*, 1980; Lehnert *et al.*, 1984).

In addition, stress can affect the metabolism of catecholamines. TOH activity was decreased in the septal area of tree shrews exposed to a chronic social stress (the visual presence of a dominant animal that subjugated them). In contrast, animals that were able to avoid the visual presence of the victor survived much longer, and those animals showed increased activity of TOH in the limbic septum and adrenal glands (Raab *et al.*, 1980). Stress also caused an increase in NE turnover and decreased NE in the locus ceruleus, hypothalamus, and hippocampus (Lehnert *et al.*, 1984), and even resulted in depletion of NE, and perhaps dopamine, in catecholaminergic neurons (Stone, 1975). During stressful situations, highly active catecholaminergic neurons may require additional precursor so that catecholamine synthesis can keep pace with the increased turnover of neurotransmitters (Milner *et al.*, 1985). Further evidence supporting the concept that dietary tyrosine can enhance the immune response during stress was done by Reinstein *et al.* (1985). They found that tyrosine supplementation suppressed the rise in plasma corticosterone following acute stress in rats.

Objectives

Dietary supplementation of amino acids, particularly tryptophan, can increase respective neurotransmitters. Catecholamines appear to enhance the immune response, whereas 5-HT appears to inhibit the response. In addition, feed-grade amino acids have recently become available making it practical to provide these in commercial diets. Therefore, the objective of this study was to determine what effect dietary manipulation of endogenous serotonin and catecholamines has on immunity.

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Chapter 2

EFFECTS OF DIETARY TRYOSINE AND TRYPTOPHAN SUPPLEMENTATION ON IMMUNITY AND BRAIN NEUROTRANSMITTER LEVELS AFTER SRBC INJECTION

Introduction

Serotonin and catecholamines function as neurotransmitters and are synthesized from the amino acids tryptophan and tyrosine, respectively. Since the precursors of the neurotransmitters are supplied to the body via the diet, diet-induced alterations in plasma levels of the amino acids may affect brain serotonin and catecholaminic synthesis. It has become apparent that ingesting increased quantities of these amino acids, under certain circumstances, increases respective brain neurotransmitter levels (Sved, 1983; Milner *et al.*, 1987). Furthermore, administration of the precursors can alter physiological activities and behavior of animals including sleep, blood pressure, hormone release from the pituitary, and the stress response (Sved, 1983; Reinstein *et al.*, 1985; Milner *et al.*, 1985; 1987).

Recently, studies on neuroimmunomodulation indicated that the brain participates in the regulation of the immune response (Roszman, *et al.*, 1988; Saphier, 1988; Bateman *et al.*, 1989; Rabin *et al.* 1989). Alterations in brain indoleamines and catecholamines influenced the ability of an animal to respond to an antigen challenge. Devoino *et al.* (1987) found that the dopamine precursor L-Dopa enhanced the antibody response to SRBC, an effect believed to be mediated by the central nervous system. On the other hand, lesioning catecholaminergic neurons by injecting 6-OHDA into the cisterna magna resulted in marked inhibition of the antibody response to SRBC (Cross *et al.*, 1986). In contrast, increasing brain serotonin levels by giving daily injections of 5-hydroxytryptophan (5-HPT), the precursor of serotonin, or by using monoamine oxidase inhibitors, decreased delayed-type hypersensitivity in a dose-dependent manner (Devoino and Ilyutchenok, 1968), suppressed humoral immunity (Devoino *et al.*, 1973; 1987) and transplantation

immunity (Draskoci *et al.*, 1964; Pierpaoli *et al.*, 1978). On the other hand, decreasing brain serotonin levels by lesioning the raphe nucleus increased antibody titers (reviewed by Devoino *et al.*, 1987).

Stress was found to alter the metabolism of catecholamines and indoleamines (reviewed by Stone, 1975; Anisman, 1978; Dunn *et al.*, 1984; Glavin, 1985). It increased NE turnover and decreased, or even depleted, NE content if sufficiently severe (Stone, 1975; Glavin, 1985), while the content of 5-HT was generally unchanged although the 5-HT turnover increased (Morgan *et al.*, 1975; Joseph *et al.*, 1981; Kennett *et al.*, 1981). Stressors are well-known immune suppressors (Dunn, 1989). Alteration in brain neurotransmitter levels, especially the depletion during stress, probably results in changes in immunoregulation. Thus, highly active catecholaminergic neurons may require additional precursor so that catecholamine synthesis can keep pace with increased turnover (Milner *et al.*, 1987). Reinstein *et al.* (1985) successfully suppressed the increase in plasma corticosterone of stressed rats by supplying tyrosine in the diet.

Given the fact that diet has an effect on brain neurotransmitters and that brain neurotransmitters can influence the immune response, it appears possible that diet may be able to influence disease resistance. To date, little if any research has focussed on the possible role of diet in enhancing animal production through its possible effect on the immune response. The purpose of this study was to investigate whether such a dietary supplementation could affect an immune response.

Materials and Methods

Leghorns and broilers were obtained on the day of hatch from a commercial hatchery, weighed, wing-banded, and randomly placed in heated batteries with 10 birds per pen until three weeks of age. The chickens were then moved to wire-floor batteries with 5 birds per pen. The birds were provided a starter diet (Appendix B) for the first three weeks, after which they were fed a grower diet (Appendix B). The starter diet was calculated to contain 3134 Kcal ME/Kg, 21.3% crude protein, 0.84% tyrosine and 0.24% tryptophan, and the grower diet was 3160 Kcal ME/Kg,

19.7% crude protein, 0.77% tyrosine and 0.22% tryptophan. Feed and water were provided *ad libitum* and lighting was continuous. This study consisted of five experiments.

Experiment 1

This experiment was to investigate the effect of 0.1% dietary tyrosine and tryptophan supplementation on humoral and cell-mediated immunity, as well as disease resistance. At three weeks of age, twenty one pens of male Leghorns were randomly assigned to one of 3 dietary treatments as follows: 1) birds fed a control grower diet, 2) birds fed a grower diet with 0.1% tyrosine supplementation, and 3) birds fed a grower diet with 0.1% tryptophan supplementation. Two pens in each group were injected with 1 ml of a 5% SRBC suspension, and two pens with 0.5 ml of a 0.5% SRBC suspension. To investigate the secondary immune response, the birds were inoculated again with the same dose 14 days after the first injection. Blood samples (0.3-0.4 ml) were taken via the brachial vein every two days starting 3 days after injection. The blood was allowed to clot, and the serum used to measure antibody titers as briefly described below.

Total antibody titers and 2-mercaptoethanol resistant (MER) titers to SRBC were measured using the microtiter procedure (Wegman *et al.*, 1966). The MER titers were determined as a measure of immunoglobulin G (IgG) levels, and 2-mercaptoethanol sensitive (MES) titers were associated with immunoglobulin M (IgM) levels, obtained by subtracting MER titers from total titers. Titers were expressed as \log_2 of the reciprocal of the highest dilution in which there was hemagglutination. MES and MER titers will be referred to as IgG and IgM titers, respectively.

The same birds were used for the phytohemagglutinin (PHA) wattle response (McCorkle *et al.*, 1979). One wattle was injected with 0.1 ml of 100 $\mu\text{g}/0.1$ ml PHA (pH 7.5), while the other wattle injected with 0.1 ml of saline, served as a control. The thickness of each wattle was measured by a micrometer before and 16 hours after the injection. The fold differences were calculated by dividing the wattle thickness post injection with the thickness before injection.

At 5 weeks of age, the other three pens of birds in each group were injected with 0.1 ml of a 10^{-4} dilution of *E. coli* via the posterior thoracic air sac. The *E. coli* was from a 24-hour incubated serotype of KI in tryptose broth. Body weights were recorded at 24, 48, 72 and 96 hr after the challenge. Following the final weighing, the birds were killed via cervical dislocation and

scored for air sac and heart lesions as described by Collins and Siegel (1987). Relative weight loss was calculated by dividing the weight loss by body weight before each weight loss, and the scores of lesion were transformed using the formula $(\text{score} + 1)^{1/2}$.

Experiment 2

This experiment was similar to Experiment 1 with the exception that male broilers were used instead of Leghorns. Twelve pens of broilers (four pens per treatment) were given the same dietary treatments and injected with 1 ml of a 5% or 0.5 ml of a 0.5% SRBC suspension. The birds injected with the 5% SRBC suspension were inoculated again 14 days after the first injection. Antibody titers were measured using the procedure described above. The PHA wattle response was also conducted on these birds using the same procedure as that in the Experiment 1.

Experiment 3

This experiment was designed to examine the effect of higher tyrosine and tryptophan supplementation levels on humoral immunity using a lower injection dose of SRBC. At 4 weeks of age, 7 pens of Leghorns were randomly assigned to each of 5 dietary treatments including: 1) grower diet with 0.5% supplemental tyrosine, 2) grower diet with 1% supplemental tyrosine, 3) grower diet with 0.5% supplemental tryptophan, 4) grower diet with 1% supplemental tryptophan, and 5) grower diet only. These diets were fed for one week after which the birds returned to control grower diet. Feed intake and body weight gain were measured while the birds were fed the amino acid supplemented diets.

The Leghorns were inoculated intravenously with 0.1 ml of a 0.5% SRBC suspension one day after starting dietary treatments. The same birds were inoculated again 14 days after the primary inoculation. Blood samples were collected from the brachial vein of 10 birds per group every two days starting 3 days and 2 days after injection of the primary and secondary inoculation, respectively, for measuring antibody titers. The remaining birds were used for measuring brain neurotransmitter levels. Five chickens in each group were randomly sacrificed by cervical dislocation at days 2, 4, 6, 8, and 10 after the first SRBC injection. The brains were quickly removed, placed on dry ice, and sectioned into four brain parts including telencephalon,

diencephalon, brain stem, and cerebellum. The brain parts were frozen at -70°C for further analysis.

Brain catecholamines, serotonin and their metabolite concentrations were measured by high performance liquid chromatography with electrochemical detection (HPLC-EC) using a modified procedure of Myers *et al.* (1986). Briefly, brain parts were homogenized in 0.05 M HClO_4 with 3,4 dihydroxybenzylamine (DHBA) added as an internal standard. Using a Valco injector, 100 μl samples were loaded into the HPLC-EC system consisting of a pump (Model SM-909; Anspec, Ann Arbor, Michigan), C-18 column (Bioanalytical Systems, Inc., 250 X 4.6 mm, 5 μm Biophase), amperometric detector (Model 400, Princeton Applied Res. Corp., Princeton, NJ) and Chromatochart, a computerized data collection system (Interactive Microware, Inc., State College, PA). The mobile phase contained 0.12 M citric acid, 350 mg/l NaOH, 150 mg/l Na-EDTA and 200 mg disodium octyl sulfate, 86 ml acetonitrile and 1.8 ml of triethanolamine adjusted to a pH of 3.1. The mobile phase was run through an inline degasser (Model ERC-3001; ERMA Inc., Tokyo, Japan).

Experiment 4

This experiment was designed to examine the effect of a higher level dietary amino acid supplementation on antibody response in broilers. Fifty male broilers were assigned to each of the 5 dietary treatments, and the experimental procedure was the same as that of Experiment 3. Briefly, ten birds (two pens) per group were inoculated with the SRBC suspension, and serum samples were collected for antibody titration. Feed intakes and body weight gain during the supplemental period were also measured.

Experiment 5

Since stress is known to affect the immune response, this experiment was designed to investigate the interaction of dietary amino acid supplementation and stress on humoral immunity. Immobilization stress, in addition to dietary treatment, was used in the experiment. Ten (two pens) male broilers 4 weeks of age were randomly assigned to one of each of following treatments: 1) stressed and fed 0.5% supplemental tyrosine, 2) stressed and fed 1% supplemental tyrosine, 3) stressed and fed 0.5% supplemental tryptophan, 4) stressed and fed 1% supplemental tryptophan,

5) stressed and fed grower diet, and 6) fed grower diet without stress. The birds were given the supplemental diets for 10 days, then returned to the control grower diet. The immobilization stress was conducted by restraining both wings and legs for three hours just prior to the first SRBC injection. The SRBC inoculation and the collection of blood samples for measuring antibody titers was done as described for Exp. 1. Feed intake and body weight gain were measured during the dietary supplemental period.

The antibody titers, brain neurotransmitter levels, relative body weight loss, transformed lesion scores, fold differences of wattle thickness, feed intakes, and body weight gain were analyzed by analysis of variance using the Statistical Analysis System (SAS Inst. Inc., 1985) for a completely randomized design. The model was:

$$Y_{ij} = \mu + T_j + e_{ij}$$

Where μ is the mean of Y_{ij} , T equals the effect of the treatments and e_{ij} is the random variation unique to each group. If significant treatment effects were observed, then the means were separated using Duncan's new multiple range test (Duncan, 1955). Significance implies $p \leq 0.05$.

Results

The dietary addition of 0.1% tyrosine increased the primary IgM titers at Day 9 (Table 1) and secondary IgG titers at Days 3 and 9 in Leghorns injected with 0.5% SRBC (Table 2). No differences in titers were observed in Leghorns injected with 5% SRBC. The addition of 0.1% supplemental tryptophan increased secondary IgG titers and decreased secondary IgM titers at Day 9 in Leghorns injected with 5% SRBC suspension (Table 2). In broilers, 0.1% tyrosine supplementation decreased secondary IgM titers at Day 5 with 5% SRBC injection, and 0.1% tryptophan supplementation increased the IgM titers at Day 9 (Table 4), whereas the primary antibody response was not affected by the tryptophan supplementation (Table 3).

Neither 0.1% tyrosine or tryptophan supplementation affected the PHIA wattle response in Leghorns or broilers (Table 5). The relative weight loss (Table 6) and transformed lesion score (Table 7) of Leghorns in response to the *E. coli* challenge were also not changed by the dietary supplementation.

With higher supplemental levels, 0.5% tyrosine supplementation significantly enhanced the primary IgM response at Day 11 in Leghorns (Table 8). Antibody titers at other time periods were not significantly affected by 0.5% or 1% tyrosine or tryptophan supplementation.

In broilers, the primary antibody titers were not significantly affected by 0.5% or 1% tryptophan supplementation (Table 9). However, both levels of added tryptophan significantly decreased the secondary IgM titers at Day 2 (Table 9). The secondary total antibody titers at Day 6 with 1% tyrosine or tryptophan supplementation were significantly lower than those with 0.5% tyrosine supplementation.

The stress increased the primary IgM titers at Day 3 in broilers, while 0.5% or 1% tryptophan supplementation significantly lowered the titers (Table 10). The primary IgM titers at Day 3 with the 1% tyrosine supplementation were significantly lower than those stressed as well as with 0.5% tyrosine dietary supplementation or stressed only. The secondary IgM titers at Day 2 with 0.5% tryptophan supplementation were also lower than those with 1% tryptophan supplementation.

Since there were no significant differences in antibody titers and brain neurotransmitter levels between the two levels (0.5% and 1%) of amino acid supplementation, except at a few time periods, the titers within the two dietary supplemental levels of each amino acid were pooled. The antibody titers after grouping were statistically analyzed to determine if there were effects due to the amino acid supplementation.

Tyrosine supplementation caused an increase in the Leghorn primary IgM titers at Day 11 (Fig. 2), but it decreased broiler primary total antibody titers at Day 7 (Fig. 1). The restraint stress increased primary total antibody titers at Day 3 (Fig. 7) and IgM titers at Days 2, 5, 7 in the broilers (Fig. 8). The increase in IgM titers was also found in the stressed broilers fed the tyrosine supplemented diet, but not the tryptophan added diet (Fig. 8).

The antibody titers of the Leghorns remained affected by the tryptophan supplementation (Fig. 1-6); nevertheless, the addition of tryptophan decreased the primary total titers at Days 7 and 9 (Fig. 2) and IgM titers at Day 9 (Fig. 3), secondary IgM titers at Day 2 (Fig. 5), and IgG titers at Day 8 (Fig. 6) in broilers. It also decreased primary IgM titers at Days 3, 9 (Fig. 8) and secondary total titers at Day 8 (Fig. 7), and IgG titers at Day 6 (Fig. 9) in stressed broilers.

In Leghorn chicks, dietary tryptophan supplementation increased 5-HT, 5-HIAA levels, and 5-HIAA to 5-HT ratios in the brains. The addition of 1% dietary tryptophan significantly increased 5-HT at Days 4 and 6, 5-HIAA levels at Days 2, 4, and 6, and their ratios at Day 2 post-inoculation in the brain stem (Table 11). It also increased 5-HT levels at Day 2, 5-HIAA levels at Days 2, 4, and 6, and the 5-HIAA/5-HT ratios at Days 4 and 6 in the diencephalon (Table 12). The 5-HIAA levels and the ratios were increased in the telencephalon at Days 2 and 6 (Table 13). There were no changes in 5-HIAA or 5-HT levels and their ratios in the cerebellum (Table 14). The addition of 0.5% tryptophan also increased 5-HT levels at Day 2, 5-HIAA levels and 5-HIAA/5-HT ratios at Days 4 and 6 in the brain stem (Table 11); the 5-HIAA levels and the ratios at Day 4 in the diencephalon (Table 12) and at Day 6 in the telencephalon (Table 13), and the 5-HIAA/5-HT ratios at Day 6 in the cerebellum (Table 14). There were dose-dependent increases in 5-HT and 5-HIAA levels in the brain stem (Table 11), though the higher tryptophan addition caused significant increases in more measurements.

Dietary supplementation of tryptophan also influenced brain catecholamine levels. Both tryptophan supplementation levels increased NE in the brain stem at Days 2 and 4 post inoculation (Table 15); it also increased epinephrine in the brain stem and diencephalon at day 2 (Table 15, 16), and DA in the diencephalon at Day 4 (Table 16). Brain DA concentrations were increased dose-dependently at Day 6 in the diencephalon and at Day 4 in the telencephalon.

Tyrosine supplementation had no effect on brain catecholamine levels (Table 15-18) except for a significant increase in L-dopa levels in the diencephalon at Day 4 (Table 16). Brain 5-HT were also not affected by tyrosine supplementation (Table 11-14). However, 1% tyrosine supplementation increased 5-HIAA levels in the diencephalon at Day 4 (Table 12) and the ratios of 5-HIAA to 5-HT in the brain stem (Table 11), telencephalon (Table 13), and cerebellum (Table

14) at Day 6. It also decreased the ratios in the cerebellum at Day 2 (Table 14). No significant changes occurred in 5-HT and 5-HIAA levels, or their ratios with 0.5% tyrosine supplementation (Table 11-14).

Feed intake and body weight gain were not significantly influenced by the 0.5% or 1% dietary amino acid addition during the supplementation period; however, the daily average feed consumption of broilers was twice that of Leghorns (Table 19, 20).

Discussion

The present study demonstrated that dietary tryptophan supplementation significantly increased brain 5-HT and 5-HIAA levels, as well as the ratios of 5-HIAA to 5-HT. The increase in 5-HT levels probably reflects an increase in the synthesis of 5-HT from tryptophan, since the rate limiting enzyme in this conversion, tryptophan hydroxylase, was estimated to be only 50% saturated under normal conditions (Fernstrom *et al.*, 1971; Young *et al.*, 1977). As 5-HIAA is the major metabolite of 5-HT, the 5-HIAA level can be used as a measurement of 5-HT turnover (Reinhard *et al.*, 1977). The increase of 5-HIAA concentration and its ratio to 5-HT indicates an increase in the activity of serotonergic neurons. These results are similar to those seen in rats (Fernstrom *et al.*, 1971; Sved, 1983). Other evidence shows that tryptophan loading increased extracellular serotonin (Schwartz *et al.*, 1990) and enhanced serotonin release (Schacchter *et al.*, 1989). It has been suggested that alterations in tetrahydrobiopterin (BH₄) availability after tryptophan loading may serve as a mechanism by which brain tryptophan hydroxylation, and, therefore, serotonin turnover, can be regulated (Bengtsson *et al.*, 1991).

In the brain stem, tryptophan supplementation caused dose-dependent increases in 5-HT and 5-HIAA levels. The tryptophan requirement for chickens has been reported to be no greater than 0.16% of a diet containing 3,200Kcal ME/Kg (Smith *et al.*, 1987), which is far below the supplementation levels used in this study. When supplementing the diet with 0.1 to 0.3% tryptophan, 5-HT and 5-HIAA levels were increased dose-dependently in turkeys (Hobbs, 1989). Therefore, it can be concluded that the 0.5% of tryptophan supplementation in the basal diet

(Appendix B) of poultry does not saturate tryptophan hydroxylase and allows for incremental increases in 5-HT synthesis and its turnover. Tryptophan binding to albumin may slow down the substrate saturation of tryptophan hydroxylase, because such binding lowers the free tryptophan concentration in the plasma. The percentage of serum free tryptophan ranges from 10-50% in the serum (Fernstrom *et al.*, 1976). The free tryptophan can cross the blood-brain barrier, while albumin-bound tryptophan requires a catalyst-enhanced dissociation of tryptophan from its albumin binding site within brain microcirculation before being transported into the brain (Pardridge *et al.*, 1987).

Tryptophan supplementation was also found to occasionally increase brain catecholamine levels in selected brain regions, a result similar to that observed in turkeys (Hobbs, 1989). While the mechanism is not known, there is evidence of morphological connections and interrelationships between serotonergic and catecholaminergic systems in their neurotransmitter concentration, turnover, and release (Samanin *et al.*, 1975). In addition, serotonergic systems have an inhibitory influence on catecholamine function (Andrews *et al.*, 1978; Dray *et al.*, 1978). Such evidence suggests that the increase of catecholamine levels may result from increasing the inhibitory effect of the serotonergic system on catecholaminergic release and, thus, reducing the amount of catecholamines broken down by MAO and thereby increasing the storage of catecholamines.

The failure of tyrosine addition to the diet to increase brain levels of catecholamines in the present study is also in agreement with a previous report that systemic administration of tyrosine, in general, does not increase the synthesis or brain levels of catecholamines and their major metabolites (Carlsson *et al.*, 1972). This may be primarily due to the 75% saturation of tyrosine hydroxylase with its substrate, tyrosine, under basal conditions (Gibson *et al.*, 1978). However, using brain microdialysis, it was recently found that tyrosine administration increased DA release with minor elevations in the levels of DOPAC and HVA in the striatum, but the rise in DA was short-lived (Acworth *et al.*, 1988). Unlike tyrosine supplementation, L-dopa administration was found to increase brain DA level (Sved, 1983). Accordingly, substrate saturation and end-product inhibition of tyrosine hydroxylase may play major roles in maintaining stable catecholamine levels during normal physiological condition.

Moreover, tyrosine supplementation occasionally increased the ratios of 5-HIAA/5-HT. The mechanism is unknown; however, tyrosine, unlike tryptophan, supplementation did not actually increase 5-HT or 5-HIAA levels, which may not necessarily indicate that the supplementation increased serotonin turnover.

Although the tryptophan treatment significantly suppressed broiler antibody titers at only a few time periods, there was a nearly significant ($p \leq 0.10$) decrease in the primary antibody response in stressed and unstressed broilers. This suggests that the tryptophan supplementation played a suppressive role in antibody production under both physiological and stressful conditions in broilers, but not Leghorns.

Previously, serotonin has been demonstrated to cause an inhibition of immune function via action in the central nervous system. Increases in brain serotonin caused by various chemicals including MAO inhibitors, 5-hydroxytryptophan, and reserpine, resulted in inhibition of immunogenesis (Draskoci *et al.*, 1964; Devoino *et al.*, 1968; 1973; Pierpaoli *et al.*, 1978; Boranic *et al.*, 1984). The inhibition in immunological function included the humoral antibody response, delayed type hypersensitivity, and transplantation immunity in different mammals with different antigens, including SRBC. On the other hand, reducing the level of serotonin by tryptophan hydroxylase blockade, or lesioning of the raphe nuclei was observed to increase rosette forming cells to bovine serum albumin (BSA) and SRBC (reviewed by Devoino *et al.*, 1987).

It has been reported that serotonergic terminals in the hypothalamus activated secretion of ACTH-releasing hormone (reviewed by Devoino *et al.*, 1987) and serotonin stimulated CRF secretion from the hypothalamus *in vitro* (Jones *et al.*, 1976). On the other hand, hypophysectomy or pituitary stalk destruction prevented stimulation of the immune response after lesioning of the serotonergic raphe nuclei, as well as the inhibition of the immune response after increasing serotonin levels (Devoino *et al.*, 1987). Furthermore, adrenalectomy not only prevented the inhibition of the immune response, but stimulated it after 5-hydroxytryptophan injection (Devoino *et al.*, 1987). According to the evidence above, Devoino *et al.*, (1987) suggested that the serotonergic system has an inhibitory effect on the immune response which is modulated via the HPA axis. Their work using syngenic transfer of cells from bone marrow, spleen, and thymus

demonstrated that the inhibition of serotonergic system in immune response by activating antigen-nonspecific suppression.

Although peripheral levels of serotonin were not measured in the current study, plasma levels have been found to increase following dietary supplementation of tryptophan in turkeys (Hobbs, 1989). Thus, a peripheral role for serotonin in the immune response should not be neglected. Systemic administration of serotonin, which can not pass the blood brain barrier, prior to inoculation resulted in suppression of IgM and IgG plaque-forming cells in response to SRBC (Cross *et al.*, 1986; Gray *et al.*, 1988). The finding that receptors for neurotransmitters are present on the surface membrane of lymphocytes and monocytes (Eliseeva *et al.*, 1982) also supports that serotonin is capable of influencing immune function peripherally.

In the present study, lower (0.1 and 0.5%) dietary tyrosine supplementation increased Leghorn and stressed broiler immune response. According to previous reports, dopaminergic neurons in the striatum were found to have a stimulating role on the immune response. Increasing endogenous DA levels by 3,4-dihydroxyphenylalanine administration stimulated antibody formation (reviewed by Devoino *et al.*, 1987). A similar result was observed with phenamine, a drug that attenuates the release of catecholamines from storage and inhibits their reuptake (Devoino *et al.*, 1987). Furthermore, activation of the dopaminergic system by increasing DA receptor activity or decreasing serotonin levels caused a more intense immune reaction which was mediated via the hypothalamus-hypophysis-thymus axis (Devoino *et al.*, 1987).

Tyrosine supplementation failed to alter catecholamine levels. However, both 0.5% and 1% supplemental tyrosine, instead of increasing catecholamine levels, had an effect on maintaining a stable DA level in the brain stem of Leghorns, while DA levels fluctuated significantly in other groups (Table 15). This indicates that tyrosine supplementation may have a facilitating role on the catecholaminergic system, eliciting a stimulative effect on the immune response, thus, causing increases in the primary IgM titers in Leghorns.

Since brain catecholamine levels remained unchanged following dietary tyrosine supplementation treatment in Leghorns, it is assumed that this was also true in broilers. The suppressive effect of tyrosine supplementation in broilers, but not Leghorns, may be due to the

peripheral catecholamines (not measured). Because of significant differences in feed intake behavior between Leghorns and broilers, the broiler peripheral catecholamine levels may greatly differ from that of Leghorns. Tyrosine availability has been shown to affect peripheral catecholamine synthesis. Administration of oral tyrosine to human subjects increased urinary levels of three catecholamines and their major metabolites (Agharanya *et al.*, 1981; Alonso *et al.*, 1982). The evidence suggests that the plasma levels of catecholamines may increase after the oral administration. The increasing levels of catecholamines may result in immune suppression. Johnson *et al.* (1981) reported a suppression of the proliferation response of mouse spleen cells to LPS in the presence of NE, and Crary *et al.* (1983) found decreases in mitogen responsiveness of mononuclear cells from peripheral blood after epinephrine administration in humans. There was also a negative relationship between adrenaline and the T helper cell/T suppressor cell ratio (Landmann *et al.*, 1984). In the current study, 0.1% tyrosine supplementation enhanced Leghorn antibody response, and 0.5% tyrosine supplementation resulted in higher antibody response than 1% tyrosine supplementation. Accordingly, ingesting very large quantity of dietary tyrosine due to high feed intake or high supplementation level caused immune suppression probably via the effect of high peripheral catecholamines.

The stimulating effect of restraint stress on the immune response may have resulted from the activation of catecholaminergic neurons, especially those in the striatum, which enhanced DA synthesis and release. This may have resulted in enhancing the immune response via stimulation of the hypothalamus-hypophysis-thymus axis as stated previously. The following evidence supports this hypothesis. Immobilization stress significantly enhanced L-dopa accumulation in the striatum after decarboxylase inhibition (Algeri *et al.*, 1988). Footshock and restraint stress resulted in a significant increase in DOPAC to DA ratios in the striatum (Dunn, 1988). Moreover, tyrosine administration increased DA synthesis and release in rats given stress (Gibson *et al.*, 1978; Lehnert *et al.*, 1984).

Furthermore, the immobilization stress used in the current study may not have been strong enough to cause depletion of brain catecholamines, which usually occurs in severe stress and may result in behavioral deficits (Lehnert *et al.*, 1984; Stone, 1975) and immune suppression. As stated

previously, stress activates catecholamine neurons and enhances catecholamine synthesis and release, including firing rate. An increase in firing frequency of the neurons causes allosteric changes in tyrosine hydroxylase, thereby increasing its affinity for its cofactor BH₄ and reducing its susceptibility to end-product inhibition (Lovenberg *et al.*, 1975). Thus, the immune stimulative effect due to increased activity of the catecholaminergic system by stress and tyrosine supplementation may overcome the immune suppressive effect of increasing peripheral catecholamines which probably occurred in unstressed broilers. The stress stimulating effect on the immune response was not found in broilers fed the tryptophan supplemented diets which must be prevented by the suppressive effect of the tryptophan supplementation as described above.

There were no significant differences in *E. coli* challenge and PHA wattle response between the dietary treatments. The *E. coli* challenge has been used as an index of resistance to disease (Gross and Siegal, 1983), and the wattle swelling response to mitogen stimulation shown to be an indicator of cell-mediated immunity in chickens (Klesius *et al.*, 1977). Due to the great deviation in the observations, it may be too early to conclude that cell-mediated immunity and disease resistance were not affected by the dietary treatments.

Disparate results within the same dietary treatment were found between broilers and Leghorns. Leghorns appeared to be more resistant to dietary tryptophan supplementation than broilers. Tyrosine dietary supplementation enhanced Leghorn immune response, but suppressed the broiler response. Tryptophan supplementation decreased antibody titers in broilers, but not Leghorns. The reason for these varying responses is unclear. Similar phenomena were reported in broilers and Leghorns responding to ICV injection of biogenic amines in feed intake and drinking (Denbow *et al.*, 1982; 1983). On the other hand, it is well-known that Leghorns possess a higher response to antigen challenges and much lower feed intake than broilers, which may contribute to the disparate results.

Summary

The purpose of the present study was to determine if dietary supplementation with tyrosine or tryptophan affected brain neurotransmitter levels and humoral immune response in chickens. A basal diet supplemented with 0.1%, 0.5% or 1% of tyrosine or tryptophan were given to chickens at 3-4wks of age. The chickens were injected with SRBC suspension one Day after the supplementation and 14 Days after the first injection. Brain neurotransmitter levels and antibody titers were measured using HPLC-EC and the microtiter procedure, respectively, at different time intervals following the dietary treatment and SRBC injection.

The dietary addition of 0.1% tyrosine increased the primary IgM and secondary IgG titers at some time periods, while 0.1% tryptophan increased the secondary IgG titers and decreased IgM titers at Day 9 in Leghorns. In broilers, 0.1% tyrosine supplementation decreased secondary IgM titers at Day 5 and 0.1% tryptophan dietary addition increased the IgM titers at Day 9. The PHA wattle response and resistance to *E. Coli* challenge were not different between the dietary treatments.

The addition of 0.5% tyrosine significantly enhanced the Leghorn primary IgM response at Day 11 and broiler secondary total antibody response at Day 6. In broilers, secondary total antibody titers at Day 6 with 1% tyrosine supplementation were significantly lower than those with 0.5% tyrosine supplementation. The primary IgM titers at Day 3 with 1% tyrosine supplementation were also significantly lower than those stressed as well as with 0.5% tyrosine supplementation or stressed only. Both 0.5% or 1% tryptophan addition resulted in suppression of the broiler secondary IgM titers at Day 2 and stressed broiler primary IgM titers at Day 3 but had no effect on Leghorn immune response. When the titers within the two supplemental levels of a given amino acid were pooled, tyrosine supplementation appeared to enhance the antibody production in stressed broilers and unstressed Leghorns and decreased antibody titers in unstressed broilers, while tryptophan showed a suppressive effect on stressed and unstressed broilers antibody response.

Dietary supplementation with 0.5% and 1% Tyrosine had no effect on brain catecholamine levels in Leghorns, though it significantly increased L-dopa levels. Serotonin was also not

influenced by tyrosine supplementation. However, 1% tyrosine altered the 5-HIAA levels and the 5-HIAA/5-HT ratios. Dietary tryptophan supplementation significantly increased 5-HT, 5-HIAA levels, and their ratios in all brain parts at some time periods during the dietary supplementation period except in the cerebellum. There were dose-dependent increases in 5-HT and 5-HIAA levels in the brain stem, although 1% tryptophan supplementation caused more significant increases.

These results demonstrate that 0.5-1% dietary tryptophan supplementation suppressed the antibody response in broilers, which is probably via enhancing serotonin synthesis and release. They also suggest that lower (0.1-0.5%) dietary tyrosine supplementation probably enhances the immune response in Leghorns and stressed broilers, but ingesting large quantity of tyrosine which occurred in broilers suppress the response.

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Table 1. Primary anti-SRBC antibody titers of male Leghorns fed 0.1% supplemental dietary tyrosine or tryptophan^{1,2,3}

Measurement	Dietary Treatment	Days Post Inoculation			
		3	5	7	9
5% SRBC Inoculation					
Total	Control	2.6 ± 0.8	6.0 ± 0.8	4.1 ± 1.5	3.6 ± 1.2
	0.1% Tyr	2.3 ± 0.9	5.5 ± 1.3	3.3 ± 1.3	3.1 ± 0.9
	0.1% Trp	2.1 ± 0.6	5.4 ± 0.8	3.6 ± 1.3	2.9 ± 0.9
IgG	Control	0	0.3 ± 0.5	0.7 ± 0.9	0.9 ± 0.9
	0.1% Tyr	0	0.1 ± 0.3	0.5 ± 0.8	0.6 ± 0.8
	0.1% Trp	0	0	1.2 ± 1.2	1.6 ± 0.7
IgM	Control	2.6 ± 0.8	5.7 ± 0.7	3.4 ± 1.6	2.7 ± 1.3
	0.1% Tyr	2.3 ± 0.9	5.4 ± 1.3	2.8 ± 1.5	2.5 ± 0.8
	0.1% Trp	2.1 ± 0.6	5.4 ± 0.8	2.4 ± 2.1	2.3 ± 0.9
0.5% SRBC Inoculation					
Total	Control	1.5 ± 0.8	3.6 ± 1.4	4.2 ± 1.4	3.2 ± 1.5
	0.1% Tyr	2.3 ± 1.3	4.8 ± 1.8	4.7 ± 1.2	4.4 ± 0.7
	0.1% Trp	1.5 ± 0.7	4.6 ± 1.9	4.3 ± 1.5	4.0 ± 1.5
IgG	Control	0	0.2 ± 0.4	1.3 ± 0.9	1.1 ± 0.8
	0.1% Tyr	0	0.1 ± 0.3	0.9 ± 0.8	1.1 ± 0.7
	0.1% Trp	0	0	0.8 ± 0.8	0.9 ± 0.7
IgM	Control	1.5 ± 0.8	3.4 ± 1.6	2.9 ± 1.2	2.1 ± 1.2 ^b
	0.1% Tyr	2.3 ± 1.3	4.7 ± 1.9	3.8 ± 1.3	3.3 ± 0.9 ^a
	0.1% Trp	1.5 ± 0.7	4.6 ± 1.9	3.5 ± 1.5	3.1 ± 1.2 ^{ab}

¹ The birds were given the treated diets one day before SRBC injection, and the supplementation diets were given throughout the experiment.

² The values represent means ± SE.

^{a,b} Means within a row with different superscript are significantly different ($p \leq 0.05$).

Table 2. Secondary anti-SRBC antibody titers of male Leghorns fed 0.1% dietary supplemental tyrosine or tryptophan^{1,2,3}

Measurement	Dietary Treatment	Days Post Inoculation			
		3	5	7	9
5% SRBC Inoculation					
Total	Control	5.1 ± 0.6	7.6 ± 1.1	6.4 ± 0.8	6.4 ± 0.8
	0.1% Tyr	5.6 ± 1.3	7.9 ± 1.4	7.0 ± 0.8	7.1 ± 0.7
	0.1% Trp	5.1 ± 1.0	7.8 ± 0.9	6.4 ± 0.5	6.3 ± 0.8
IgG	Control	2.4 ± 1.0	3.8 ± 1.5	3.4 ± 1.3	2.3 ± 0.7 ^b
	0.1% Tyr	2.7 ± 0.8	4.5 ± 1.5	3.8 ± 1.1	2.9 ± 1.1 ^{ab}
	0.1% Trp	2.4 ± 0.5	4.2 ± 1.2	3.6 ± 1.0	3.6 ± 0.8 ^a
IgM	Control	2.7 ± 1.1	3.8 ± 1.5	3.0 ± 1.2	4.1 ± 0.9 ^a
	0.1% Tyr	2.9 ± 1.4	3.4 ± 1.3	3.2 ± 1.4	4.2 ± 1.0 ^a
	0.1% Trp	2.7 ± 0.9	3.6 ± 1.2	2.8 ± 1.0	2.7 ± 1.3 ^b
0.5% SRBC Inoculation					
Total	Control	4.5 ± 1.1	7.6 ± 1.1	6.9 ± 1.0	6.1 ± 1.3
	0.1% Tyr	4.4 ± 1.4	7.7 ± 1.3	6.7 ± 0.8	5.8 ± 1.3
	0.1% Trp	4.7 ± 1.1	7.5 ± 1.6	6.7 ± 1.3	5.9 ± 1.5
IgG	Control	1.2 ± 0.6 ^b	3.1 ± 0.9	2.9 ± 1.2	3.0 ± 0.8 ^b
	0.1% Tyr	2.0 ± 0.8 ^a	3.9 ± 1.4	3.7 ± 1.3	4.2 ± 1.4 ^a
	0.1% Trp	1.8 ± 0.6 ^{ab}	3.3 ± 0.9	3.5 ± 0.8	3.2 ± 0.9 ^b
IgM	Control	3.3 ± 1.3	4.5 ± 1.4	4.0 ± 1.6	3.2 ± 1.8
	0.1% Tyr	2.4 ± 1.3	3.8 ± 1.4	3.0 ± 1.2	1.6 ± 1.2
	0.1% Trp	2.9 ± 1.3	4.2 ± 1.8	3.2 ± 1.6	2.7 ± 1.3

¹ The birds were given the treated diets one day before SRBC injection, and the supplementation diets were given throughout the experiment.

² The values represent means ± SE.

^{a,b} Means within a row with different superscript are significantly different (≤ 0.05).

Table 3. Primary anti-SRBC antibody titers of male broilers fed 0.1% dietary supplemental tyrosine or tryptophan^{1,2,3}

Measurement	Dietary Treatment	Days Post Inoculation				
		3	5	7	9	11
5% SRBC Inoculation						
Total	Control	2.3±1.6	5.7±1.3	5.3±1.1	3.8±0.9	2.8±1.0
	0.1% Tyr	2.3±1.4	6.2±0.8	5.5±0.7	4.3±0.8	3.2±0.6
	0.1% Trp	3.1±0.7	6.3±1.4	5.8±1.0	4.0±1.1	3.0±0.9
IgG	Control	0	0	1.3±1.2	0.6±1.0	0.4±0.8
	0.1% Tyr	0	0	1.8±1.0	0.9±1.2	0
	0.1% Trp	0	0	1.4±1.3	0.9±1.2	0.3±0.9
IgM	Control	2.3±1.6	5.7±1.3	4.0±1.1	3.2±1.0	2.4±1.3
	0.1% Tyr	2.3±1.4	6.2±0.8	3.7±0.9	3.4±1.2	3.2±0.6
	0.1% Trp	3.1±0.7	6.3±1.4	4.4±1.1	3.1±1.4	2.7±1.1
		3	5	6	7	9
0.5% SRBC Inoculation						
Total	Control	2.3±0.5	5.5±1.8	6.6±1.2	5.6±1.7	4.4±1.8
	0.1% Tyr	2.2±0.4	5.2±0.9	6.1±1.4	5.9±0.7	4.8±0.9
	0.1% Trp	2.0±0.0	5.5±0.8	5.8±1.4	4.6±1.6	3.8±1.3
IgG	Control	0.7±0.8	0.4±0.8	0.7±1.0	0.8±1.4	0.2±0.7
	0.1% Tyr	0.6±0.8	0.4±0.8	0.5±1.1	0.5±1.1	0
	0.1% Trp	0.4±0.8	0	0.4±0.8	0.7±1.2	0.2±0.6
IgM	Control	1.6±0.8	0.4±0.8	5.9±1.2	4.7±1.8	4.2±1.7
	0.1% Tyr	1.6±0.8	0.4±0.8	5.6±1.4	5.4±1.0	4.8±0.9
	0.1% Trp	1.6±0.8	0	5.4±1.4	3.9±1.5	3.6±1.1

¹ The birds were given the treated diets one day before SRBC injection, and the supplementation diets were given throughout the experiment.

² The values represent means ± SE.

³ No significant differences between the treatments ($P \leq 0.05$).

Table 4. Secondary anti-SRBC antibody titers of male broilers fed 0.1% dietary supplemental tyrosine or tryptophan^{1,2,3}

Measurement	Dietary Treatment	Days Post Inoculation			
		3	5	7	9
5% SRBC Inoculation					
Total	Control	5.6 ± 2.6	5.1 ± 2.0	4.9 ± 1.3	3.4 ± 2.2
	0.1% Tyr	4.9 ± 1.5	5.0 ± 1.6	4.4 ± 2.0	3.7 ± 2.1
	0.1% Trp	6.2 ± 2.1	5.5 ± 2.1	5.1 ± 1.7	4.5 ± 1.6
IgG	Control	1.9 ± 1.2	3.1 ± 1.3	2.6 ± 0.9	2.5 ± 1.4
	0.1% Tyr	2.1 ± 1.3	3.2 ± 1.9	1.8 ± 1.1	2.3 ± 1.3
	0.1% Trp	2.0 ± 1.6	3.1 ± 1.7	2.9 ± 1.6	2.2 ± 1.6
IgM	Control	3.7 ± 2.3	2.9 ± 1.1 ^a	2.1 ± 1.2	0.9 ± 1.0 ^b
	0.1% Tyr	2.8 ± 1.0	1.8 ± 0.4 ^b	2.6 ± 1.3	1.4 ± 1.0 ^b
	0.1% Trp	4.2 ± 1.6	2.4 ± 0.7 ^{ab}	2.2 ± 1.1	2.3 ± 0.8 ^a

¹ The birds were given the treated diet one day before SRBC injection and the supplementation diets were given throughout the experiment.

² The values represent means ± SE.

^{a,b} Means with different superscripts are significantly different ($p \leq 0.05$).

Table 5. Fold differences of wattle thicknesses as affected by phytohemagglutinin injection in Leghorns and broilers fed 0.1% tyrosine or tryptophan supplemental diets^{1,2}

Dietary Treatment	Fold Difference	
	Leghorns	Broilers
Controll	3.655	2.152
0.1% Tyrosine	3.909	2.147
0.1% Tryptophan	4.272	2.258

¹ The values represent means \pm SE.

² No significant difference between the treatments ($P \leq 0.05$).

³ Fold differces = (wattle diameter after PHA injection)/(wattle diameter before PHA injection).

Table 6. Relative body weight changes (%) in response to injection of *E. coli* into air sac of Leghorns fed 0.1% tyrosine or tryptophan supplemented diets^{1,2,3}

Dietary Treatment	Days Post <i>E. Coli</i> Injection			
	1	2	3	4
Controll	-4.10 ± 5.02	0.14 ± 3.29	0.16 ± 3.24	0.48 ± 5.07
0.1% Tyrosine	-3.72 ± 4.55	1.93 ± 2.93	2.96 ± 2.51	-0.24 ± 2.10
0.1% Tryptophan	-2.79 ± 4.60	1.21 ± 3.64	1.25 ± 3.44	-1.50 ± 3.77

¹ The values represent means ± SE.

² No significant difference between the treatments ($P \leq 0.05$).

³ Relative body weight changes were calculated by dividing body weight loss by body weight before each weight loss.

Table 7. Transformed scores of lesion after *E. coli* injection in Leghorns fed diets supplemented with 0.1% tyrosine or tryptophan^{1,2,3}

Dietary Treatment	Transformed Score of Lesion
Control	1.954 ± 0.673
0.1% Tyrosine	1.826 ± 0.611
0.1% Tryptophan	1.828 ± 0.700

¹ The values represent means ± SE.

² No significant difference between the treatments ($P \leq 0.05$).

³ The transformed scores are from the formula of $(\text{score} + 1)^{1/2}$.

Table 8. Anti-SRBC antibody titers of male Leghorns fed varying levels of added tyrosine and tryptophan in the diets from 27 to 34 days of age^{1,2}

Measurement	Days ³	Dietary Treatment				
		Control	0.5% Tyr	1% Tyr	0.5% Trp	1% Trp
Primary Antibody Response						
Total	3	1.4 ± 1.1	1.5 ± 0.5	1.4 ± 0.8	1.1 ± 1.2	0.8 ± 0.8
	5	6.9 ± 1.1	6.7 ± 0.9	7.4 ± 0.8	6.9 ± 0.6	6.7 ± 1.1
	7	7.1 ± 1.5	7.5 ± 1.1	7.6 ± 1.0	7.6 ± 0.8	6.7 ± 1.2
	9	5.3 ± 0.8	6.1 ± 0.9	6.0 ± 0.8	5.8 ± 0.9	5.7 ± 1.4
	11	3.3 ± 0.8	4.0 ± 0.9	3.9 ± 1.1	3.8 ± 1.0	3.8 ± 1.3
IgG	3	0	0	0	0	0
	5	0.9 ± 0.3	0.9 ± 0.7	0.6 ± 0.7	0.5 ± 0.5	0.9 ± 0.9
	7	1.7 ± 0.5	1.6 ± 0.5	2.1 ± 0.6	1.9 ± 0.3	2.1 ± 1.0
	9	1.5 ± 0.7	1.6 ± 0.8	1.7 ± 0.7	2.1 ± 0.6	1.4 ± 0.7
	11	1.2 ± 0.4	0.9 ± 0.7	1.3 ± 0.7	1.3 ± 0.5	1.9 ± 1.2
IgM	3	1.4 ± 1.1	1.5 ± 0.5	1.4 ± 0.8	1.1 ± 1.2	0.8 ± 0.8
	5	6.0 ± 0.9	5.8 ± 1.0	6.8 ± 1.1	6.4 ± 0.5	5.8 ± 0.6
	7	5.4 ± 1.3	5.9 ± 1.3	5.5 ± 1.4	5.7 ± 1.1	4.6 ± 1.2
	9	3.8 ± 0.8	4.5 ± 1.0	4.3 ± 0.9	3.7 ± 1.3	4.3 ± 1.2
	11	2.1 ± 0.7 ^b	3.1 ± 1.1 ^a	2.6 ± 1.1 ^{ab}	2.5 ± 0.7 ^{ab}	1.9 ± 0.7 ^b
Secondary Antibody Response						
Total	2	2.3 ± 0.7	2.4 ± 0.7	2.5 ± 0.8	2.2 ± 0.8	2.4 ± 0.5
	4	7.5 ± 1.2	7.0 ± 1.8	7.8 ± 0.9	7.0 ± 0.9	6.7 ± 0.7
	6	8.1 ± 1.0	7.6 ± 1.1	8.3 ± 1.1	7.7 ± 0.9	7.5 ± 1.0
	8	6.4 ± 0.8	6.2 ± 1.5	6.9 ± 0.7	6.2 ± 1.1	5.8 ± 1.0
IgG	2	0.9 ± 0.3	0.7 ± 0.5	0.7 ± 0.5	0.4 ± 0.5	0.8 ± 0.4
	4	2.9 ± 0.9	2.4 ± 0.7	2.6 ± 0.5	2.2 ± 0.4	2.6 ± 0.5
	6	3.1 ± 0.7	3.0 ± 0.7	3.5 ± 0.5	3.3 ± 0.5	3.2 ± 0.4
	8	2.9 ± 0.3	2.7 ± 0.7	2.8 ± 0.6	2.9 ± 0.3	2.5 ± 0.7
IgM	2	1.4 ± 0.7	1.7 ± 0.9	1.8 ± 0.8	1.8 ± 1.1	1.6 ± 0.5
	4	4.6 ± 1.1	4.6 ± 1.7	5.2 ± 0.8	4.8 ± 1.0	4.1 ± 0.9
	6	5.0 ± 1.3	4.6 ± 0.7	4.8 ± 0.8	4.3 ± 0.9	4.3 ± 1.2
	8	3.5 ± 1.0	3.6 ± 1.1	4.1 ± 0.7	3.3 ± 1.0	3.4 ± 1.2

¹ The birds were given the treated diet one day before SRBC injection(0.5%, 0.1ml), and the supplemented diets were given for 7 days.

² The values represent means ± SE.

³ The number of the days indicates days after SRBC injection.

^{a,b} Means within a row with different superscripts are significantly different (p≤0.05).

Table 9. Anti-SRBC antibody titers of male broilers fed varying levels of added tyrosine and tryptophan in the diets from 27 to 34 days of age^{1,2}

Measurement	Days ³	Dietary Treatment				
		Control	0.5% Tyr	1% Tyr	0.5% Trp	1% Trp
Primary Antibody Response						
Total	3	0.6±0.7	0.2±0.4	0.5±0.7	0.4±0.8	0.3±0.5
	5	4.4±1.1	4.0±1.5	4.5±1.2	4.4±1.3	3.7±1.7
	7	6.0±1.1	4.9±1.5	4.9±1.4	4.8±1.3	4.7±0.9
	9	4.5±1.4	3.6±1.1	3.3±1.1	3.2±1.3	3.4±1.0
IgG	3	0	0	0	0	0
	5	0	0	0	0	0.1±0.3
	7	0.4±0.7	0	0.1±0.3	0.1±0.3	0.6±0.8
	9	0.4±0.7	0.1±0.3	0	0.4±0.5	0.4±0.7
IgM	3	0.6±0.7	0.2±0.4	0.5±0.7	0.4±0.8	0.3±0.5
	5	4.4±1.1	4.0±1.5	4.5±1.2	4.4±1.3	3.6±1.5
	7	5.6±0.8	4.9±1.5	4.8±1.4	4.7±1.2	4.1±1.3
	9	4.1±1.0	3.5±1.0	3.3±1.1	2.8±1.3	3.0±0.8
Secondary Antibody Response						
Total	2	1.4±0.5 ^a	1.5±0.5 ^a	1.7±0.9 ^a	0.8±0.8 ^b	0.7±0.6 ^b
	4	6.2±1.4	6.6±1.3	5.6±1.4	5.4±1.4	5.3±1.1
	6	5.8±1.2 ^{ab}	6.4±0.8 ^a	5.4±0.7 ^b	5.6±1.1 ^{ab}	5.0±0.5 ^b
	8	4.9±1.6	4.6±0.8	4.2±1.0	4.3±1.0	3.9±0.5
IgG	2	0	0	0.1±0.3	0	0
	4	1.4±0.5	1.1±1.0	0.8±0.8	0.7±1.1	0.8±1.0
	6	1.8±0.7	1.9±1.0	1.4±0.7	1.2±1.3	1.4±0.7
	8	2.0±0.9	2.0±0.9	1.3±0.7	1.0±1.3	1.1±0.9
IgM	2	1.4±0.5 ^{ab}	1.5±0.5 ^a	1.6±1.1 ^a	0.8±0.8 ^b	0.7±0.6 ^b
	4	4.8±1.2	5.5±0.7	4.8±1.2	4.8±0.8	4.5±1.4
	6	4.0±1.0	4.5±0.7	4.0±0.9	4.3±0.7	3.6±0.8
	8	2.9±1.8	2.6±0.7	2.9±0.8	3.3±0.7	2.8±1.0

¹ The birds were given the treated diet one day before SRBC injection(0.5%, 0.1ml), and the supplemented diets were given for 7 days.

² The values represent means ± SE.

³ The number of the days indicates days after SRBC injection.

^{a,b} Means within a row with different superscripts are significantly different (p≤0.05).

Table 10. Anti-SRBC antibody titers of stressed male broilers fed varying levels of added tyrosine and tryptophan in the diets from 27 to 37 days of age^{1,2}

Measurement	Days ³	Dietary Treatment					
		Controll	Stress	Stress + 0.5% Tyr	Stress + 1% Tyr	Stress + 0.5% Trp	Stress + 1% Trp
Primary Antibody Response							
Total	3	0.7±0.5 ^{bc}	1.3±0.5 ^a	1.0±0.5 ^{ab}	0.3±0.7 ^{cd}	0.1±0.3 ^d	0 ^d
	5	5.2±1.3	5.9±1.1	5.9±0.6	5.4±0.8	4.8±0.6	5.4±0.8
	7	5.8±1.5	6.4±1.1	6.6±1.2	6.0±0.9	5.2±1.4	5.8±1.1
	9	4.7±1.6	5.1±0.9	5.3±1.1	4.7±1.1	4.2±1.8	3.9±1.5
IgG	3	0	0	0	0	0	0
	5	0.1±0.3	0.1±0.3	0	0	0	0.0
	7	0.8±1.0	0.5±0.7	0.3±0.5	0.1±0.3	0.2±0.4	0.4±0.5
	9	0.6±1.0	0.2±0.4	0.3±0.5	0.1±0.3	0.1±0.3	0.5±0.7
IgM	3	0.7±0.5 ^{bc}	1.3±0.5 ^a	1.0±0.5 ^{ab}	0.3±0.7 ^{cd}	0.1±0.3 ^d	0 ^d
	5	5.1±1.4	5.8±0.9	5.9±0.6	5.4±0.8	4.8±0.6	5.4±0.8
	7	5.0±1.3	5.9±1.0	6.3±0.9	5.9±1.0	5.0±1.3	5.4±1.0
	9	4.1±1.4	4.9±0.7	5.0±0.8	4.6±1.1	4.1±1.7	3.4±1.6
Secondary Antibody Response							
Total	2	1.0±0.5	1.2±0.4	1.2±0.4	1.3±0.5	1.0±0.7	1.4±0.7
	4	6.8±1.3	6.7±1.3	6.2±1.0	6.7±1.2	5.9±1.8	6.2±1.4
	6	7.1±1.3	6.8±1.3	6.6±0.7	7.0±1.0	6.3±1.4	6.8±1.6
	8	6.4±1.0	5.8±1.1	5.6±0.7	5.7±1.0	5.2±1.1	5.2±1.0
IgG	2	0.1±0.3	0	0	0	0.2±0.4	0
	4	1.7±0.9	1.4±1.1	1.4±1.0	1.1±0.7	1.3±1.1	1.4±0.5
	6	3.0±0.7	2.8±0.9	2.2±0.9	2.2±1.1	2.1±1.0	2.2±0.9
	8	2.3±0.7	1.9±1.0	1.9±1.1	1.8±1.0	1.6±1.3	1.7±0.7
IgM	2	0.9±0.3 ^{bc}	1.2±0.4 ^{abc}	1.2±0.4 ^{abc}	1.3±0.5 ^{ab}	0.8±0.4 ^c	1.4±0.7 ^a
	4	5.1±1.0	5.3±1.4	4.8±0.8	5.6±1.2	4.6±1.3	4.8±1.3
	6	4.1±1.4	4.0±1.1	4.4±0.8	4.8±1.0	4.2±1.2	4.6±1.6
	8	4.1±1.3	3.9±1.4	3.7±1.1	3.9±1.3	3.6±0.7	3.5±1.3

¹ The birds were given the treated diet one day before SRBC injection(0.5%, 0.1ml), and the supplemented diets were given for 10 days.

² The values represent means ± SE.

³ The number of the days indicates days after SRBC injection.

^{a,b,c,d} Means within a row with different superscripts are significantly different ($p \leq 0.05$).

Table 11. 5-HIAA, 5-HT levels (ng/g), and their ratio in the brain stem of male Leghorns fed varying levels of added dietary tyrosine and tryptophan from 27 to 34 days of age.^{1,2}

Measurement	Days ³	Dietary Treatment			
		Control	0.5% Tyr	1% Tyr	1% Trp
5-HT	2	1038.8 ± 160.8 ^b	934.6 ± 234.7 ^b	981.8 ± 101.3 ^b	1268.7 ± 68.8 ^a
	4	1119.8 ± 177.5	1363.6 ± 445.3	1164.0 ± 140.1	1366.0 ± 374.0
	6	1154.1 ± 204.6 ^{abc}	977.4 ± 59.3 ^{bc}	907.4 ± 315.6 ^c	1211.1 ± 223.4 ^{ab}
	8	1167.6 ± 178.4	1003.7 ± 212.4	932.5 ± 115.8	941.2 ± 149.5
	10	991.8 ± 154.7	1062.4 ± 252.2	1116.7 ± 180.4	1115.7 ± 112.2
5-HIAA	2	341.2 ± 62.2 ^{bc}	292.3 ± 53.5 ^c	272.3 ± 56.4 ^c	404.7 ± 61.2 ^b
	4	281.4 ± 61.2 ^c	313.8 ± 101.1 ^c	343.7 ± 38.9 ^{bc}	473.9 ± 127.0 ^{ab}
	6	273.1 ± 48.4 ^b	238.7 ± 46.1 ^b	261.6 ± 60.0 ^b	377.0 ± 49.2 ^a
	8	300.4 ± 54.7	343.3 ± 75.0	365.2 ± 35.2 ^x	342.2 ± 105.9
	10	289.2 ± 47.6	348.9 ± 57.0	333.8 ± 37.0 ^x	295.2 ± 69.6
5-HIAA/5-HT	2	0.332 ± 0.068	0.339 ± 0.147	0.276 ± 0.047 ^y	0.319 ± 0.048
	4	0.252 ± 0.045 ^{cd}	0.233 ± 0.040 ^d	0.300 ± 0.060 ^{bc}	0.352 ± 0.060 ^{ab}
	6	0.238 ± 0.033 ^c	0.244 ± 0.043 ^{bc}	0.300 ± 0.056 ^{ab}	0.315 ± 0.038 ^a
	8	0.265 ± 0.073	0.371 ± 0.190	0.397 ± 0.065 ^x	0.377 ± 0.152
	10	0.301 ± 0.092	0.336 ± 0.056	0.302 ± 0.025 ^y	0.271 ± 0.092

¹ The birds were given the treated diet one day before SRBC injection, and the dietary supplementation lasted for 7 days.

² The values represent means ± standard error.

³ The number of the days indicates days after SRBC injection.

^{a,b,c} Means within a row with different superscripts are significantly different ($P \leq 0.05$).

^{x,y,z} Means within a column of measurement with different superscripts are significantly different ($P \leq 0.05$).

Table 12. 5-HT and 5-HIAA levels (ng/g) and their ratio in the diencephalon of male Leghorns fed varying levels of added dietary tyrosine and tryptophan from 27 to 34 days of age^{1,2}

Measurement	Days ³	Dietary Treatment				
		Control	0.5% Tyr	1% Tyr	0.5% Trp	1% Trp
5-HT	2	1099.9 ± 292.5	1096.7 ± 161.8 ^x	1151.1 ± 380.7 ^x	1401.1 ± 425.2 ^x	1235.0 ± 139.0 ^y
	4	969.7 ± 185.4 ^b	983.4 ± 132.8 ^{bx}	1003.32 ± 190.5 ^{bx}	1163.5 ± 81.6 ^{abxy}	1235.0 ± 139.0 ^{ay}
	6	1257.5 ± 92.0 ^{bc}	1140.8 ± 221.6 ^{cx}	1150.3 ± 111.4 ^{cx}	1416.2 ± 105.8 ^{abx}	1587.4 ± 155.0 ^{ax}
	8	1010.9 ± 238.2	978.1 ± 149.3 ^{xy}	971.9 ± 66.3 ^{xy}	1054.6 ± 131.8 ^y	1079.8 ± 163.3 ^y
	10	819.3 ± 190.8	763.5 ± 181.0 ^y	766.1 ± 106.3 ^y	773.6 ± 156.3 ^z	836.4 ± 145.8 ^z
5-HIAA	2	322.8 ± 104.3 ^{ab}	223.0 ± 48.4 ^b	239.6 ± 89.0 ^b	315.6 ± 112.7 ^{abyz}	465.5 ± 154.0 ^{ax}
	4	261.6 ± 64.7 ^c	304.1 ± 49.6 ^{bc}	361.1 ± 46.6 ^{ab}	411.6 ± 57.0 ^{axy}	403.3 ± 59.5 ^{axy}
	6	299.7 ± 103.3 ^b	311.0 ± 80.9 ^b	355.0 ± 73.6 ^{ab}	447.8 ± 118.5 ^{abx}	477.1 ± 122.7 ^{ax}
	8	270.9 ± 63.8	317.7 ± 15.7	338.5 ± 45.2	314.1 ± 56.5 ^{yz}	312.1 ± 32.8 ^y
	10	250.4 ± 46.0	289.8 ± 99.8	286.9 ± 30.5	273.0 ± 41.8 ^z	321.0 ± 18.5 ^y
5-HIAA/5-HT	2	0.295 ± 0.073 ^{ab}	0.205 ± 0.046 ^{bz}	0.213 ± 0.048 ^{by}	0.250 ± 0.047 ^y	0.373 ± 0.100 ^a
	4	0.270 ± 0.045 ^c	0.311 ± 0.047 ^{bcxy}	0.406 ± 0.055 ^{ax}	0.353 ± 0.035 ^{abx}	0.316 ± 0.045 ^{bc}
	6	0.237 ± 0.074	0.272 ± 0.049 ^{yz}	0.309 ± 0.062 ^x	0.314 ± 0.066 ^{xy}	0.299 ± 0.063
	8	0.269 ± 0.024	0.334 ± 0.073 ^{xy}	0.347 ± 0.030 ^x	0.298 ± 0.035 ^{xy}	0.293 ± 0.047
	10	0.324 ± 0.103	0.373 ± 0.037 ^x	0.379 ± 0.060 ^x	0.360 ± 0.069 ^x	0.393 ± 0.073

¹ The birds were given the treated diet one day before SRBC injection, and the dietary supplementation lasted for 7 days.

² The values represent means ± standard error.

³ The number of the days indicates days after SRBC injection.

^{a,b,c} Means within a row with different superscripts are significantly different ($P \leq 0.05$).

^{x,y,z} Means within a column of measurement with different superscripts are significantly different ($P \leq 0.05$).

Table 13. 5-HT, 5-HIAA levels (ng/g), and their ratio in the telencephalon of male Leghorns fed varying levels of added dietary tyrosine and tryptophan from 27 to 34 days of age^{1,2}

Measurement	Days ³	Dietary Treatment			
		Control	0.5% Tyr	1% Tyr	1% Trp
5-HT	2	1339.9 ± 86.9 ^x	1285.6 ± 155.8 ^y	1243.5 ± 65.3 ^{xy}	1467.4 ± 78.3 ^x
	4	1179.5 ± 99.5 ^y	1247.4 ± 283.6 ^y	1186.3 ± 194.4 ^{xyz}	1233.3 ± 177.5 ^y
	6	1119.3 ± 74.7 ^y	1030.9 ± 47.4 ^z	1009.3 ± 93.4 ^{yz}	1136.5 ± 128.8 ^{yz}
	8	1542.2 ± 251.0 ^x	1551.2 ± 152.9 ^x	1441.5 ± 189.8 ^x	1667.0 ± 189.9 ^x
	10	1005.8 ± 146.3 ^y	914.3 ± 164.2 ^z	966.1 ± 310.8 ^z	963.8 ± 247.9 ^z
5-HIAA	2	129.4 ± 49.0 ^{bc}	121.6 ± 24.8 ^{bc}	103.4 ± 15.0 ^c	153.2 ± 9.2 ^{bx}
	4	102.6 ± 19.6	116.2 ± 8.8	143.3 ± 81.5	170.8 ± 54.9 ^x
	6	97.4 ± 8.9 ^b	95.3 ± 5.8 ^b	120.7 ± 17.8 ^{ab}	138.8 ± 22.6 ^{ax}
	8	104.9 ± 20.9	110.1 ± 10.4	115.8 ± 8.9	109.2 ± 11.2 ^{yz}
	10	98.0 ± 9.3	99.9 ± 14.1	95.6 ± 13.2	98.6 ± 16.5 ^z
5-HIAA/5-HT	2	0.097 ± 0.041 ^b	0.096 ± 0.024 ^b	0.083 ± 0.011 ^b	0.105 ± 0.006 ^{ab}
	4	0.088 ± 0.018	0.097 ± 0.022	0.131 ± 0.099	0.147 ± 0.079
	6	0.088 ± 0.013 ^b	0.092 ± 0.004 ^{ab}	0.120 ± 0.021 ^a	0.124 ± 0.027 ^a
	8	0.069 ± 0.006 ^{abc}	0.074 ± 0.005 ^{ab}	0.082 ± 0.014 ^a	0.066 ± 0.006 ^{bc}
	10	0.099 ± 0.018	0.113 ± 0.031	0.113 ± 0.063	0.112 ± 0.047

¹ The birds were given the treated diet one day before SRBC injection, and the dietary supplementation lasted for 7 days.

² The values represent means ± standard error.

³ The number of the days indicates days after SRBC injection.

^{a,b,c} Means within a row with different superscripts are significantly different ($P \leq 0.05$).

^{x,y,z} Means within a column of measurement with different superscripts are significantly different ($P \leq 0.05$).

Table 14. 5-HT, 5-HIAA levels, and their ratios of in the cerebellum of male Leghorns fed varying levels of added dietary tyrosine and tryptophan from 27 to 34 days of age^{1,2}

Measurement	Days ³	Dietary Treatment				
		Control	0.5% Tyr	1% Tyr	0.5% Trp	1% Trp
5-HT	2	221.1 ± 41.8	204.8 ± 40.8	220.2 ± 67.2	239.8 ± 70.5	304.7 ± 30.2
	4	215.8 ± 63.1	359.8 ± 258.7	316.2 ± 101.5	204.6 ± 73.8	329.5 ± 125.1
	6	427.6 ± 175.0	242.9 ± 44.1	240.3 ± 77.0	281.7 ± 78.2	358.9 ± 127.0
	8	267.3 ± 82.9	198.5 ± 76.2	199.2 ± 63.4	330.2 ± 99.7	411.7 ± 278.2
	10	282.9 ± 92.2	265.4 ± 105.9	226.9 ± 82.3	351.2 ± 114.3	297.2 ± 59.2
5-HIAA	2	66.9 ± 10.4 ^{ab}	42.2 ± 14.8 ^{bc}	33.4 ± 11.0 ^c	59.2 ± 18.6 ^{abcy}	82.7 ± 19.2 ^{axy}
	4	41.2 ± 23.8	40.7 ± 10.4	53.1 ± 16.2	31.2 ± 7.5 ^y	48.7 ± 18.7 ^y
	6	57.0 ± 21.7	47.0 ± 15.7	65.2 ± 21.9	56.8 ± 15.3 ^y	67.9 ± 20.7 ^y
	8	81.9 ± 52.4	61.6 ± 8.0	74.7 ± 38.9	100.5 ± 34.4 ^x	103.4 ± 33.9 ^x
	10	45.4 ± 18.1	44.9 ± 14.1	49.0 ± 18.2	59.5 ± 20.9 ^y	58.6 ± 13.2 ^y
5-HIAA/5-HT	2	0.264 ± 0.082 ^a	0.184 ± 0.037 ^{ab y}	0.139 ± 0.017 ^{b z}	0.248 ± 0.045 ^{axy}	0.276 ± 0.069 ^{ax}
	4	0.169 ± 0.047	0.113 ± 0.044 ^y	0.175 ± 0.061 ^{yz}	0.165 ± 0.055 ^y	0.135 ± 0.058 ^y
	6	0.137 ± 0.015 ^b	0.190 ± 0.033 ^{ab y}	0.246 ± 0.063 ^{ay}	0.207 ± 0.059 ^{axy}	0.197 ± 0.043 ^{ab xy}
	8	0.374 ± 0.373	0.327 ± 0.123 ^x	0.365 ± 0.103 ^x	0.325 ± 0.136 ^x	0.296 ± 0.089 ^x
	10	0.170 ± 0.061	0.177 ± 0.046 ^y	0.218 ± 0.051 ^{yz}	0.171 ± 0.035 ^y	0.200 ± 0.044 ^{xy}

¹ The birds were given the treated diet one day before SRBC injection, and the dietary supplementation lasted for 7 days.

² The values represent means ± standard error.

³ The number of the days indicates days after SRBC injection.

^{a,b,c} Means within a row with different superscripts are significantly different ($P \leq 0.05$).

^{x,y,z} Means within a column of measurement with different superscripts are significantly different ($P \leq 0.05$).

Table 15. L-Dopa and catecholamine levels (ng/g) in the brain stem of male Leghorns fed varying levels of added dietary tyrosine and tryptophan from 27 to 34 days of age.^{1,2}

Measurement	Days ³	Dietary Treatment				
		Control	0.5% Tyr	1% Tyr	0.5% Trp	1% Trp
L-Dopa	2	466.5 ± 67.1 ^x	447.6 ± 42.9 ^x	473.7 ± 33.8 ^x	458.7 ± 95.8 ^x	484.0 ± 42.2 ^x
	4	354.0 ± 95.8 ^{xyz}	332.4 ± 118.6 ^{xy}	398.9 ± 91.1 ^x	449.8 ± 44.3 ^x	428.8 ± 57.6 ^{xy}
	6	381.0 ± 221.0 ^{xy}	315.6 ± 119.6 ^y	267.4 ± 118.4 ^y	406.8 ± 79.1 ^x	315.2 ± 115.6 ^{yz}
	8	216.0 ± 41.2 ^{ab yz}	136.1 ± 32.3 ^{b z}	260.7 ± 18.6 ^{a y}	155.4 ± 109.9 ^y	187.8 ± 56.2 ^{ab z}
	10	172.7 ± 86.6 ^z	182.8 ± 100.0 ^z	180.3 ± 64.6 ^y	153.5 ± 98.2 ^y	206.5 ± 154.1 ^z
Dopamine	2	131.9 ± 34.4 ^{xy}	153.7 ± 27.6	165.7 ± 12.2	159.1 ± 17.2 ^x	172.4 ± 25.1 ^x
	4	134.3 ± 21.5 ^{xy}	168.5 ± 29.4	148.6 ± 9.9	142.7 ± 36.5 ^{xy}	142.6 ± 11.9 ^{xy}
	6	149.6 ± 16.2 ^x	158.3 ± 3.5	155.7 ± 25.0	157.0 ± 18.0 ^x	167.6 ± 44.4 ^x
	8	157.2 ± 17.8 ^x	146.1 ± 22.8	145.6 ± 22.2	151.4 ± 15.9 ^x	144.2 ± 20.7 ^{xy}
	10	109.6 ± 21.6 ^y	129.9 ± 26.3	139.2 ± 24.0	115.7 ± 20.7 ^y	111.7 ± 14.0 ^y
Norepinephine	2	876.9 ± 102.9 ^{c x}	772.1 ± 96.1 ^{c x}	859.5 ± 85.9 ^{c x}	1011.1 ± 105.7 ^{b x}	1161.3 ± 102.3 ^{a x}
	4	667.0 ± 222.5 ^{b xy}	698.9 ± 100.0 ^{ab x}	705.3 ± 127.2 ^{ab y}	901.3 ± 171.0 ^{a x}	909.2 ± 107.8 ^{a y}
	6	743.9 ± 223.9 ^{xy}	638.2 ± 155.0 ^{xy}	682.3 ± 120.6 ^y	836.8 ± 130.5 ^x	808.7 ± 247.4 ^y
	8	569.0 ± 167.3 ^y	668.9 ± 100.9 ^x	581.2 ± 141.8 ^y	569.2 ± 127.7 ^y	541.9 ± 69.6 ^z
	10	507.7 ± 170.7 ^y	507.6 ± 86.2 ^y	551.8 ± 47.4 ^y	563.7 ± 88.4 ^y	505.3 ± 89.3 ^z
Epinephine	2	137.5 ± 16.9 ^b	137.7 ± 8.3 ^{b y}	144.8 ± 26.1 ^b	213.1 ± 31.6 ^{a xy}	221.1 ± 47.6 ^a
	4	140.6 ± 9.8	181.5 ± 31.9 ^x	122.2 ± 35.3	309.9 ± 177.0 ^x	280.8 ± 208.2
	6	146.2 ± 28.2	139.2 ± 22.7 ^y	162.3 ± 77.2	200.2 ± 45.5 ^{xy}	284.2 ± 198.1
	8	115.5 ± 25.5	118.3 ± 29.1 ^y	94.8 ± 30.9	120.1 ± 26.1 ^y	139.7 ± 52.1
	10	135.3 ± 24.5	124.9 ± 32.4 ^y	147.4 ± 27.2	131.5 ± 40.8 ^y	144.9 ± 16.4

¹ The birds were given the treated diet one day before SRBC injection, and the dietary supplementation lasted for 7 days.

² The values represent means ± standard error.

³ The number of the days indicates that the samples were collected after the days of SRBC injection.

^{a,b,c} Means within a row with different superscripts are significantly different ($P \leq 0.05$).

^{x,y,z} Means within a column of measurement with different superscripts are significantly different ($P \leq 0.05$).

Table 16. L-Dopa and catecholamine levels (ng/g) in the diencephalon of male Leghorns fed varying levels of added dietary tyrosine and tryptophan from 27 to 34 days of age^{1,2}

Measurement	Days ³	Dietary Treatment			
		Control	0.5% Tyr	1% Tyr	1% Trp
L-Dopa	2	351.5±276.5 ^y	86.6± 71.0 ^y	219.5± 95.7 ^y	283.6±126.7 ^y
	4	74.0± 12.8 ^{c,z}	122.6± 34.0 ^{b,y}	167.7± 44.6 ^{a,y}	80.5± 21.3 ^{c,z}
	6	106.4± 69.3 ^z	119.8± 71.5 ^y	180.6± 82.3 ^y	90.6± 41.8 ^z
	8	781.3±164.2 ^x	860.9±187.2 ^x	691.2±123.8 ^x	786.1±222.6 ^w
	10	568.4±209.0 ^{xy}	747.1±197.0 ^x	560.6±122.5 ^x	476.0± 98.8 ^x
Dopamine	2	332.7±246.6	271.5± 60.2	242.5± 82.3 ^x	386.6±127.5 ^x
	4	201.2± 61.5 ^c	187.7± 28.1 ^c	176.7± 40.4 ^{c,xy}	309.9±104.7 ^{b,xy}
	6	241.9±113.1 ^b	194.5± 72.7 ^b	198.5± 24.9 ^{b,xy}	262.5± 42.6 ^{b,yz}
	8	215.1± 17.3	223.7± 71.9	248.9± 19.0 ^x	225.8± 37.7 ^{yz}
	10	189.6± 62.8	189.6± 55.5	163.0± 46.2 ^y	176.5± 44.8 ^z
Norepinephine	2	540.3±165.0 ^z	471.8± 71.0 ^y	490.1± 57.4 ^z	576.8±131.7 ^z
	4	690.4±151.7 ^{yz}	736.0±123.3 ^y	740.1±229.4 ^{yz}	691.4± 68.1 ^{yz}
	6	793.2±237.4 ^{yz}	679.2±142.0 ^y	736.4± 97.8 ^{yz}	715.9± 85.9 ^{yz}
	8	1256.4±265.9 ^x	1324.7±300.2 ^x	1212.4±422.7 ^x	1049.1±199.2 ^x
	10	921.1±158.0 ^y	1247.3±358.4 ^x	978.7±257.1 ^{xy}	897.7±244.6 ^{xy}
Epinephine	2	93.7± 18.9 ^{b,y}	105.0± 8.8 ^{b,z}	112.7± 31.0 ^b	264.0±189.3 ^a
	4	146.9± 43.1 ^y	166.0± 42.0 ^{yz}	140.4± 47.3	175.4± 22.1
	6	173.0± 53.8 ^y	113.6± 31.0 ^{yz}	147.9± 36.9	136.6± 32.5
	8	298.8±114.7 ^x	301.2±133.0 ^{xy}	373.4±344.5	218.8± 49.9
	10	191.5± 41.2 ^y	429.8±285.0 ^x	283.2±235.2	183.4± 22.9

¹ The birds were given the treated diet one day before SRBC injection, and the dietary supplementation lasted for 7 days.

² The values represent means ± standard error.

³ The number of the days indicates days after SRBC injection.

• Only one observation in the measurement.

^{a,b,c} Means within a row with different superscripts are significantly different ($P \leq 0.05$).

^{w,x,y,z} Means within a column of measurement with different superscripts are significantly different ($P \leq 0.05$).

Table 17. L-Dopa and norepinephrine levels (ng/g) in the telencephalon of male Leghorns fed varying levels of added dietary tyrosine and tryptophan from 27 to 34 days of age^{1,2,3}

Measurement	Days ⁴	Dietary Treatment			
		Control	0.5% Tyr	1% Tyr	1% Trp
Dopamine	2	479.8 ± 90.4	617.9 ± 128.0	590.6 ± 102.0	609.4 ± 85.9
	4	587.5 ± 64.1 ^b	604.2 ± 111.5 ^b	531.5 ± 107.7 ^b	627.9 ± 253.8 ^b
	6	545.8 ± 142.2	616.43 ± 169.9	558.8 ± 136.8	570.9 ± 125.8
	8	604.1 ± 151.3	474.4 ± 85.5	466.2 ± 186.8	399.7 ± 120.2
	10	558.1 ± 121.2	496.0 ± 17.8	443.6 ± 52.5	455.9 ± 83.6
Norepinephrine	2	195.2 ± 33.4 ^y	237.7 ± 51.4	216.6 ± 13.7	202.4 ± 17.3
	4	225.0 ± 24.9 ^y	233.1 ± 46.4	286.2 ± 136.0	281.3 ± 107.5
	6	293.9 ± 55.4 ^x	270.2 ± 65.7	233.6 ± 32.2	232.6 ± 46.7
	8	253.3 ± 55.2 ^{xy}	278.4 ± 101.4	219.3 ± 63.8	238.9 ± 36.9
	10	258.4 ± 38.7 ^y	257.5 ± 48.5	247.7 ± 49.0	253.1 ± 33.0

¹ The birds were given the treated diet one day before SRBC injection, and the dietary supplementation lasted for 7 days.

² The values represent means ± standard error.

³ The peaks of L-dopa and norepinephrine was not picked up by HPLC-EC.

⁴ The number of the days indicates days after SRBC injection.

^{a,b} Means within a row with different superscripts are significantly different ($P \leq 0.05$).

^{x,y,z} Means within a column of measurement with different superscripts are significantly different ($P \leq 0.05$).

• Only one observation in the measurement.

Table 18. Norepinephrine in the cerebellum of male Leghorns fed varying levels of added dietary tyrosine and tryptophan from 27 to 34 days of age^{1,2,3}

Measurement	Days ⁴	Dietary Treatment			
		Control	0.5% Tyr	1% Tyr	1% Trp
Norepinephrine	2	199.4 ± 75.9 ^y	321.2 ± 53.4	376.7 ± 139.1	304.1 ± 89.2
	4	206.2 ± 59.3 ^y	339.0 ± 78.4	255.2 ± 80.6	233.7 ± 81.0
	6	187.6 ± 65.2 ^y	310.7 ± 148.1	288.6 ± 101.8	322.3 ± 109.5
	8	312.5 ± 44.3 ^y	312.6 ± 113.9	232.8 ± 48.6	257.4 ± 73.1
	10	390.4 ± 142.8 ^x	340.3 ± 52.0	347.3 ± 11.6	312.2 ± 118.6

¹ The birds were given the treated diet one day before SRBC injection, and the dietary supplementation lasted for 7 days.

² The values represent means ± standard error.

³ Only peak of norepinephrine was picked up by HPLC-EC

⁴ The number of the days indicates days after SRBC injection.

^{a,b,c} Means within a row with different superscripts are significantly different ($P \leq 0.05$).

^{x,y,z} Means within a column of measurement with different superscripts are significantly different ($P \leq 0.05$).

Table 19. Daily feed intake and body weight gain (g) of Leghorns and broilers during the dietary supplementation period^{1,2}

Measurement	Dietary Treatment				
	Control	0.5% Tyr	1% Tyr	0.5% Trp	1% Trp
Leghorns					
Feed intake	59.6 ± 0.6	59.9 ± 4.0	59.1 ± 2.0	61.0 ± 3.2	58.6 ± 2.2
Body weight gain	40.1 ± 15.6	45.4 ± 10.2	48.6 ± 9.1	48.3 ± 5.6	47.4 ± 3.3
Broilers					
Feed intake	123.9 ± 8.0	128.8 ± 5.0	131.7 ± 1.0	128.1 ± 0.9	131.4 ± 3.2
Body weight gain	56.3 ± 4.4	57.3 ± 5.9	59.6 ± 1.9	55.6 ± 0.6	56.9 ± 0

¹ The values represent Means ± SE.

² No significant difference between the treatment at the level of $P \leq 0.05$.

Table 20. Daily feed intake and body weight gain (g) of stressed broilers during the dietary supplementation period^{1,2,3}

Measurement	Dietary treatment					
	Controll	Stress	Stress + 0.5% Tyr	Stress + 1% Tyr	Stress + 0.5% Trp	Stress + 1% Trp
Feed intake	1159.5± 13.5	128.4± 5.4	126.9± 5.4	122.4± 4.3	129.9± 4.7	121.7± 3.3
Body weight gain	58.3± 8.6	61.4± 4.5	62.0± 1.7	61.0± 2.0	63.0± 2.8	56.5± 2.1

¹ The broilers were tied both legs and wings for thress hours.

² The values represent means ± SE.

³ No significant difference between the treatments ($p \leq 0.05$).

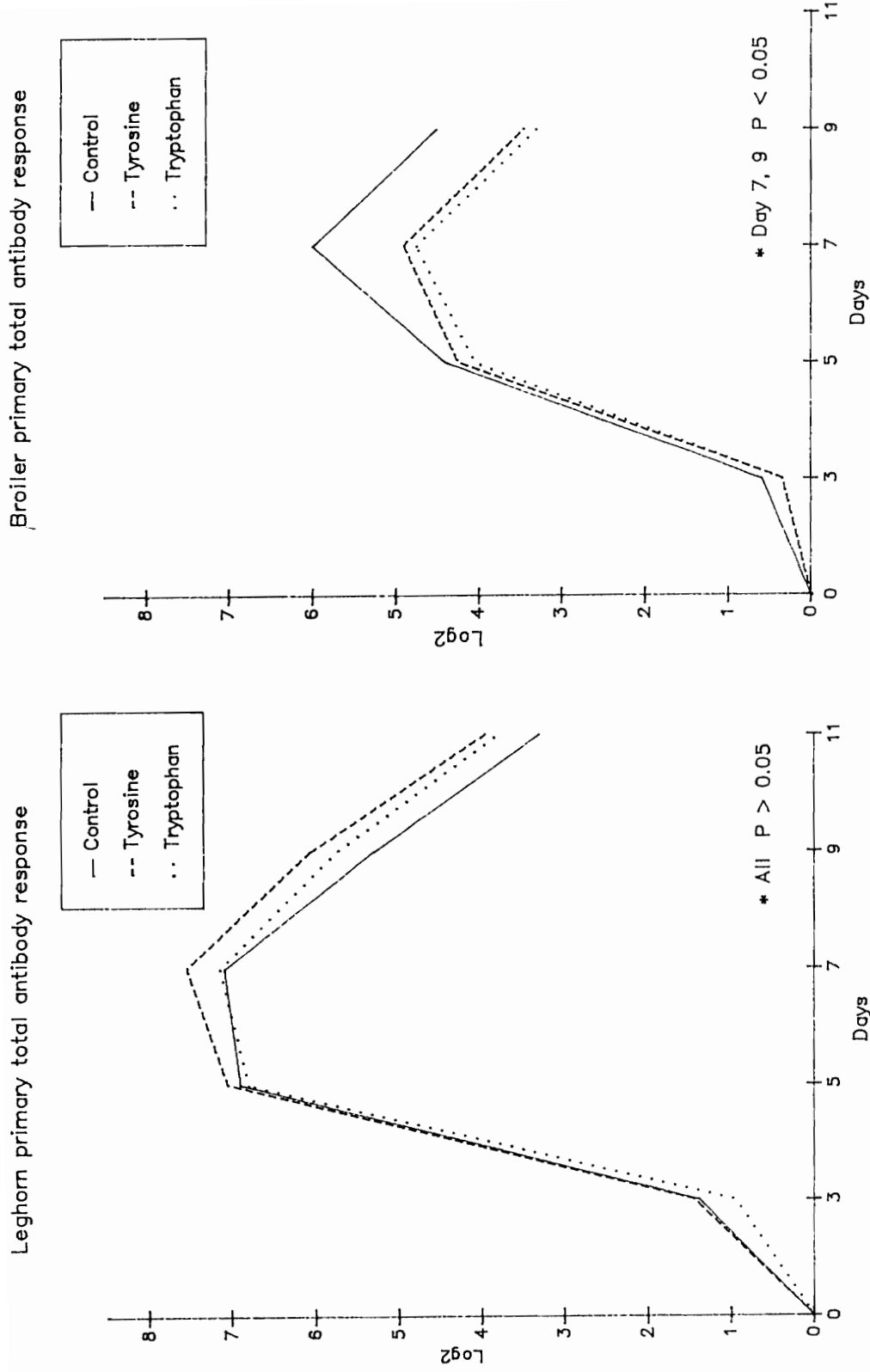


Figure 1. Primary total antibody titers in Leghorns and broilers after pooling the two dietary addition levels of a given amino acid

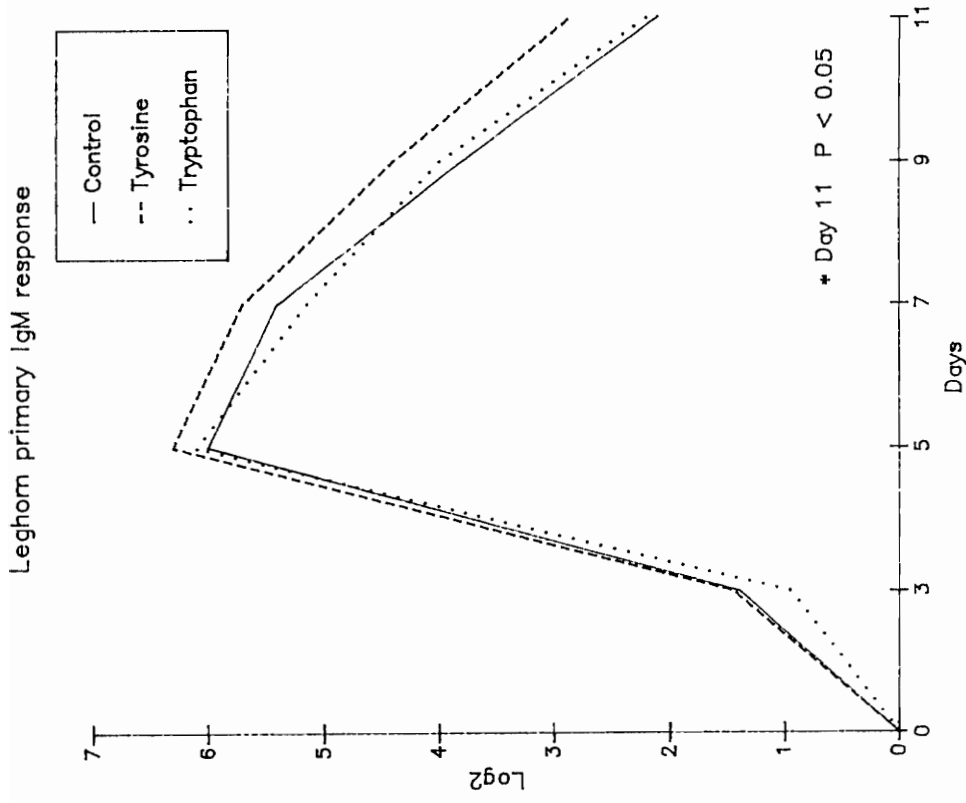
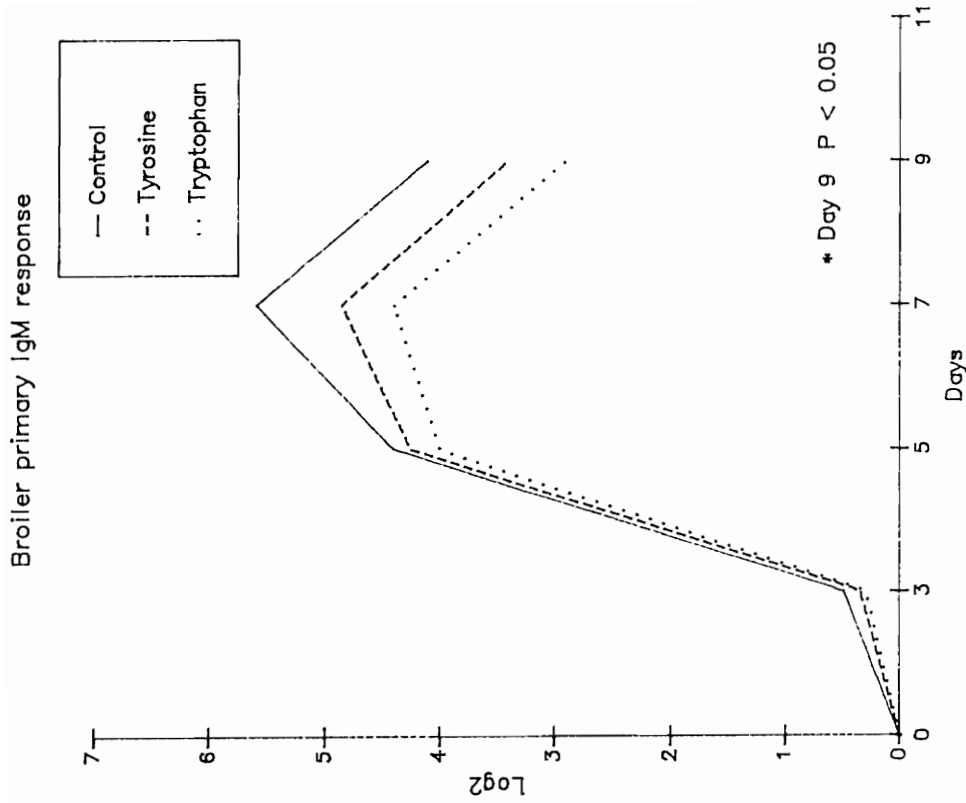


Figure 2. Primary IgM titers in Leghorns and broilers after pooling the two dietary addition levels of a given amino acid

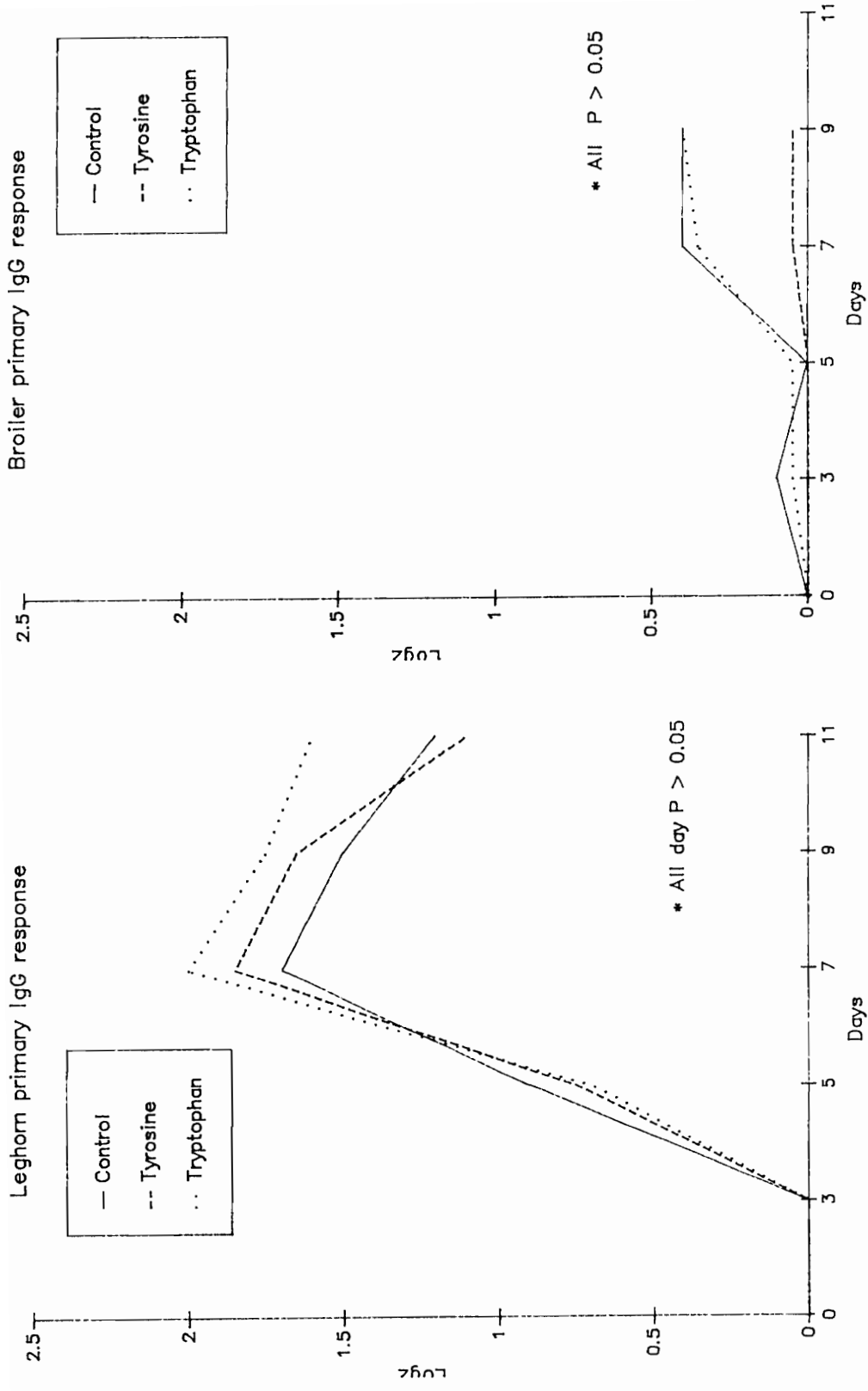
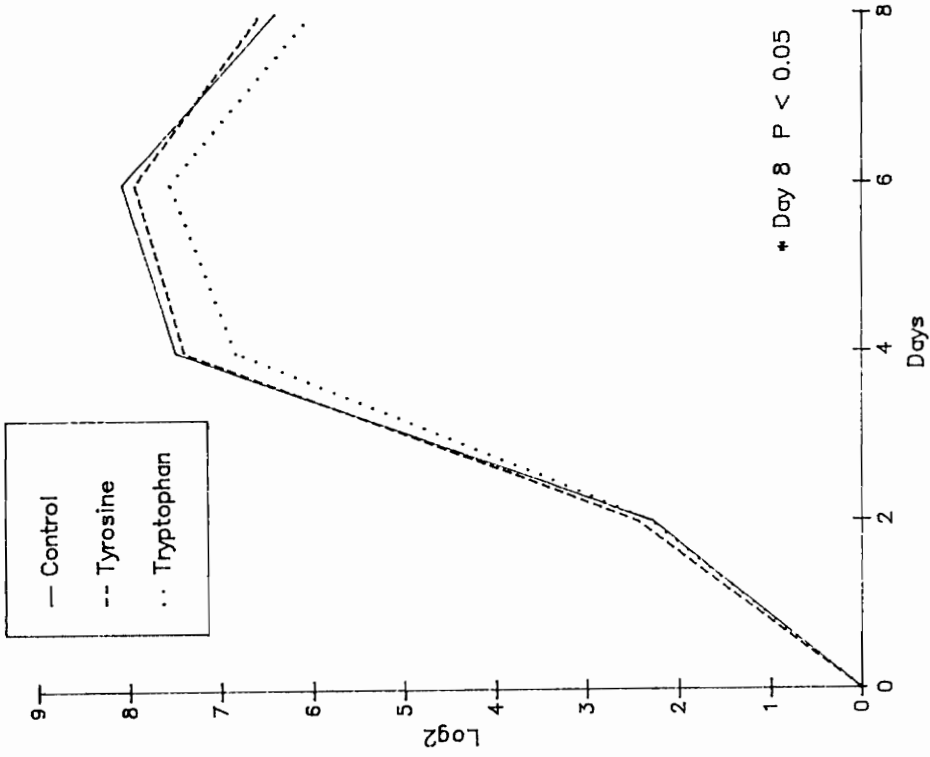


Figure 3. Primary IgG titers in Leghorns and broilers after pooling the two dietary addition levels of a given amino acid

Leghorn secondary total antibody response



Broiler secondary total antibody response

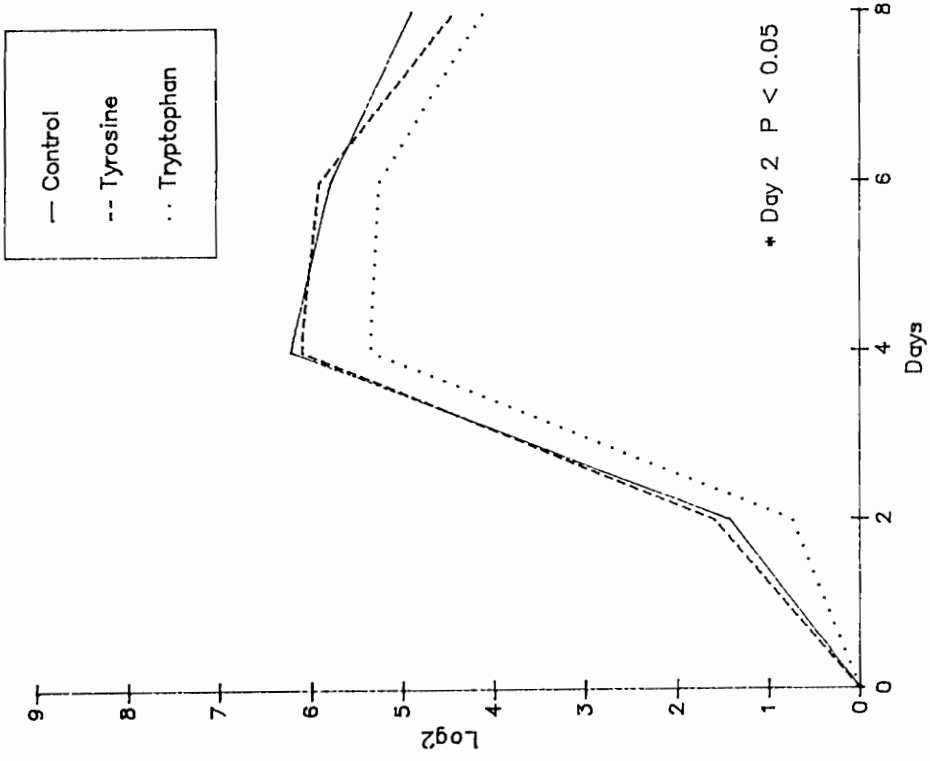


Figure 4. Secondary total antibody titers in Leghorns and broilers after pooling the two dietary addition levels of a given amino acid

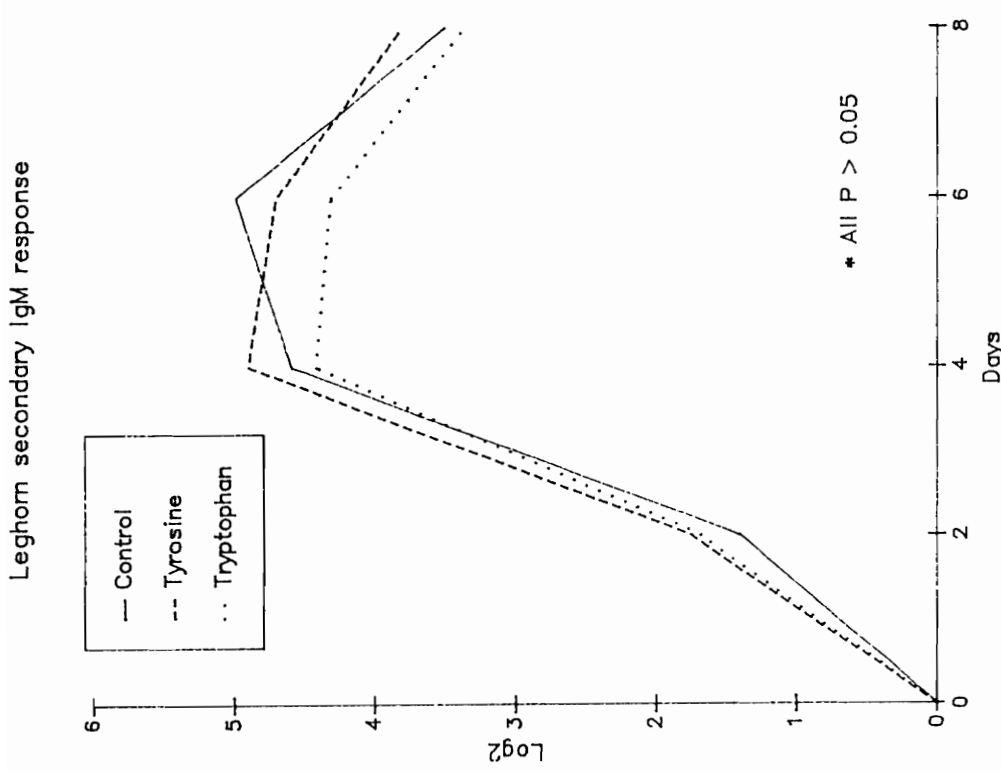
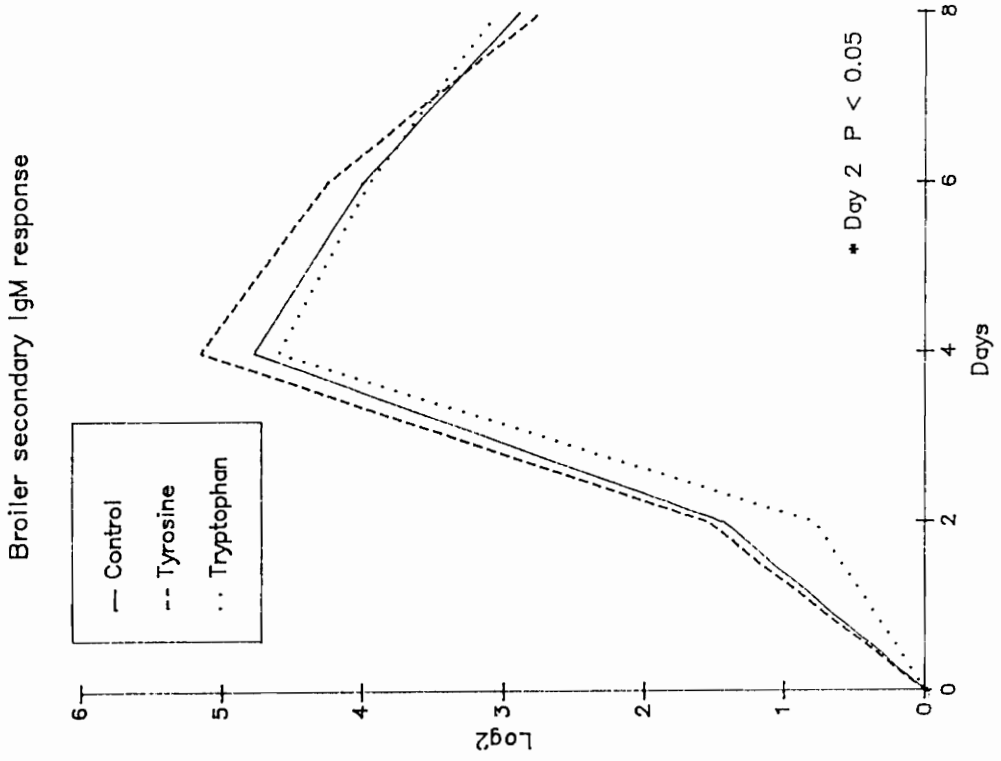


Figure 5. Secondary IgM titers in Leghorns and broilers after pooling the two dietary addition levels of a given amino acid

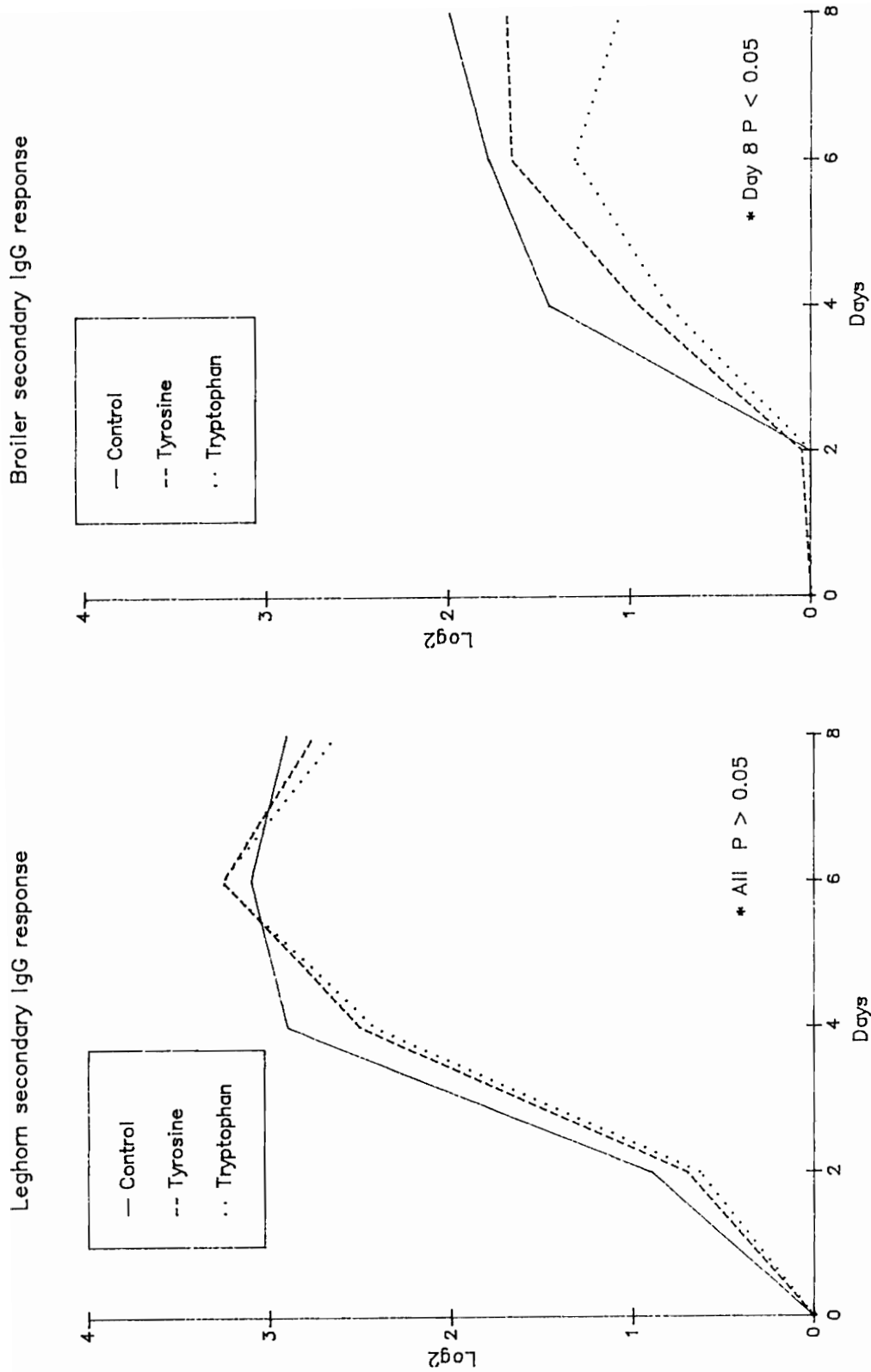
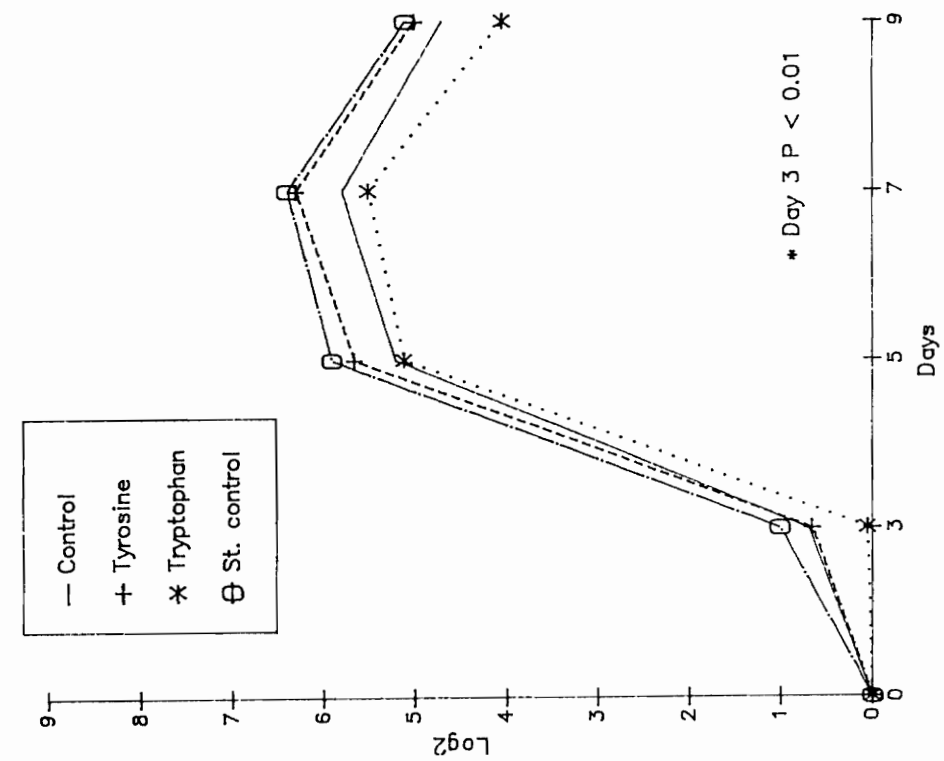


Figure 6. Secondary IgG titers in Leghorns and broilers after pooling the two dietary addition levels of a given amino acid

Stressed Broiler primary total antibody response



Stressed broiler total antibody response

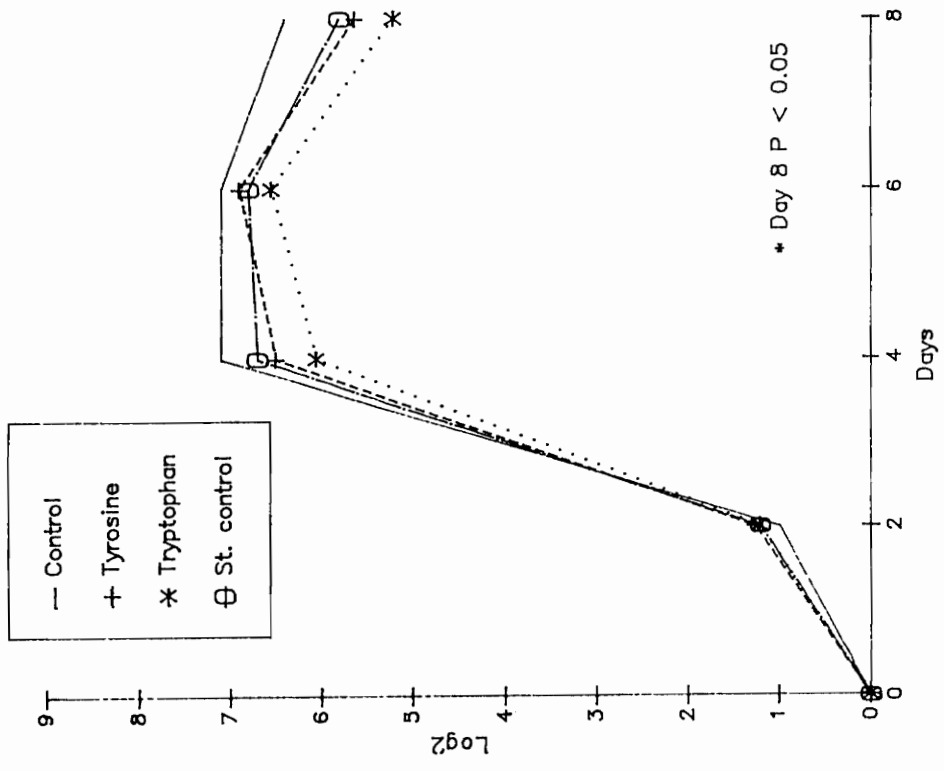


Figure 7. Primary and secondary total titers in Leghorns and broilers after pooling the two dietary addition levels of a given amino acid

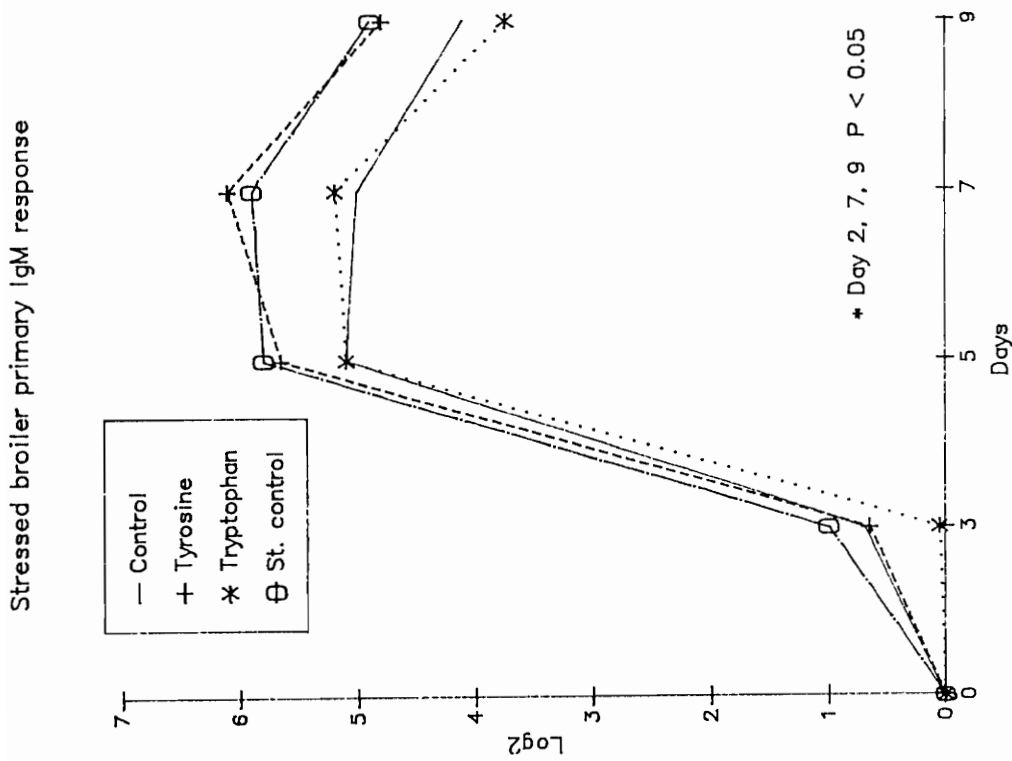
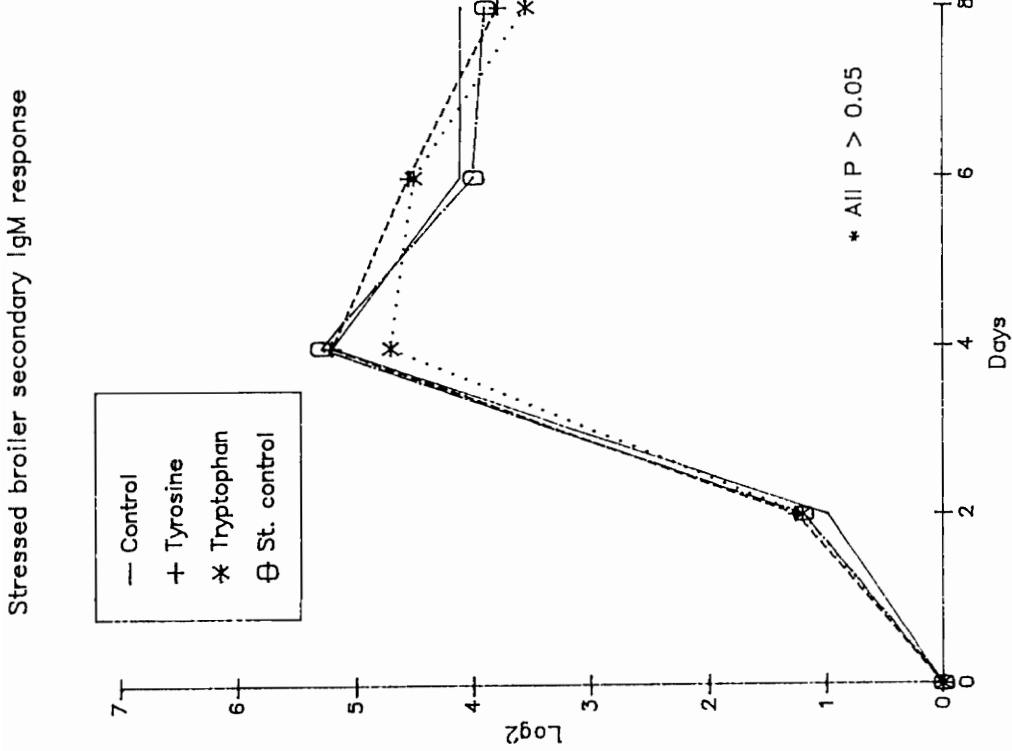
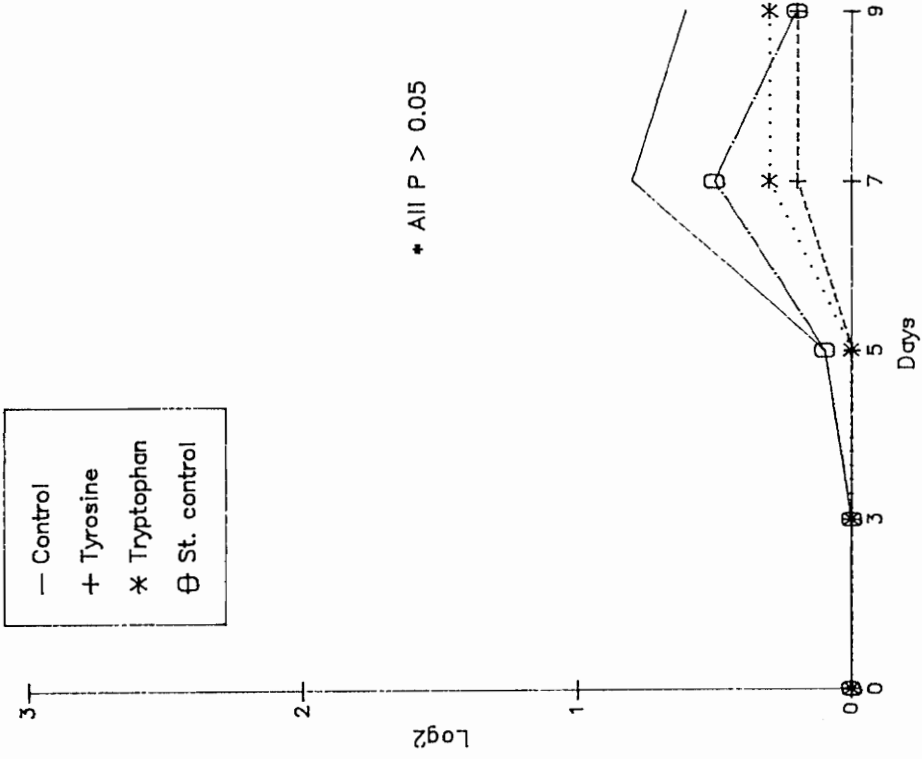


Figure 8. Primary and secondary IgM titers in stressed broilers after pooling the two dietary addition levels of a given amino acid

Stressed broiler primary IgG response



Stressed broiler primary IgG response

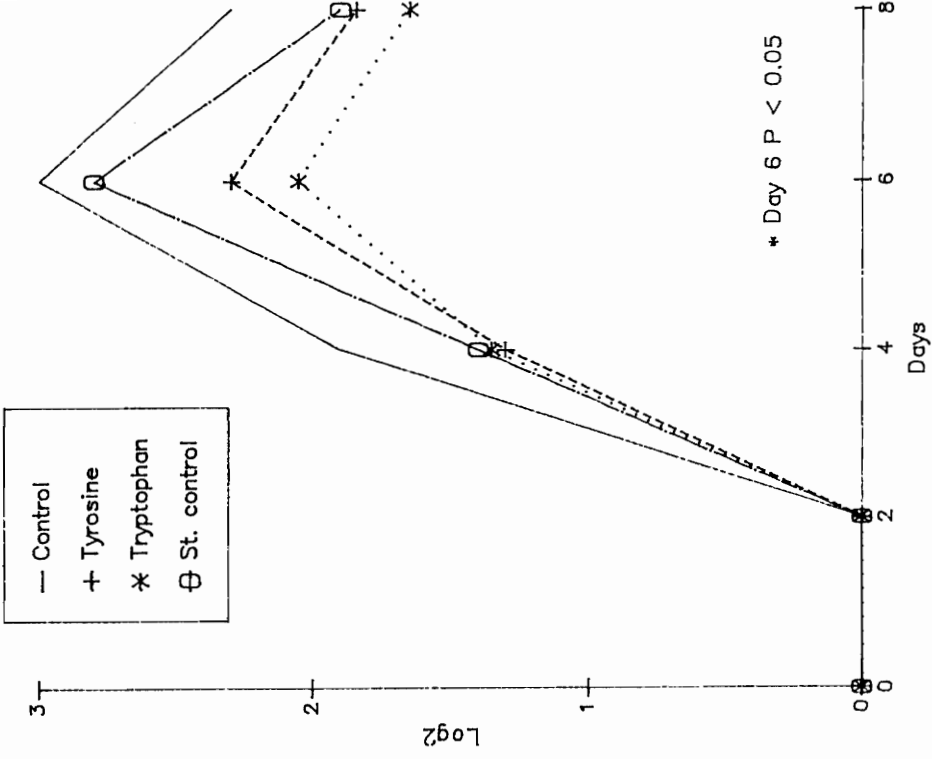


Figure 9. Primary and secondary IgG titers in stressed broilers after pooling the two dietary addition levels of a given amino acid

Appendix A. ASSAY PROCEDURES

Antibody Titers for Sheep Red Blood Cells

Materials:

Syringes (1cc, 60cc)
Needle (18 and 26 gauge)
96 well microtiter plates with "V" bottoms
25 μ l dispenser
25 μ l diluter
Incubator
Centrifuge

Chemicals:

Dextrose
Sodium citrate dihydrate
Citric acid monohydrate
Sodium chloride
2-Mercaptoethanol

Collecting SRBC

1. Using an 18 gauge needle and a syringe or vacutainer tube, collect blood from a sheep. Use an anticoagulant to prevent clotting. If blood is to be stored for a period of time, then everything must be sterilized.
2. Centrifuge the blood at 2000 g for 15 min and remove plasma. Wash the cells two times with saline and then a third time with Alsever's. Cells can be stored refrigerated for 2 weeks, 3-4 weeks if kept sterile.

* Preparation of Alsever's solution

Chemicals	g/l	g/100 ml	g/200 ml
dextrose	20.5	2.05	4.10
sodium citrate dihydrate	8.0	0.80	1.60
citric acid monohydrate	0.55	0.055	0.11
NaCl	4.2	0.42	0.84

pH = 6.1

Autoclave for 10 min.

3. Determine the hematocrit of the SRBC solution and use the following formula to dilute to the proper strength:

$$x = \frac{(Y \text{ ml of desired soln.}) \times (\% \text{ desired soln.})}{(\% \text{ stock soln.})}$$

X = ml of stock soln. needed to make Y ml of desired soln.

Y - X = ml of saline needed.

Microagglutination test for total antibody titers

1. Immunization and serum collection

- a. Inject birds with 1 ml of a 5% (or 0.1 ml of 0.5%) solution of SRBC's using a 26 gauge needle.
- b. At some period(s) of time after inoculation, usually 7 days, take 0.3-0.4 cc of blood per bird.
DO NOT USE AN ANTICOAGULANT.
- c. Let the blood stand, usually 1-2 hr, until a clot forms. Then centrifuge the samples and collect the serum. If necessary, blood can be kept in the refrigerator overnight and the serum collected the next day.
- d. If measuring both IgG and IgM titers, it is necessary to heat inactivate the serum at 56°C for 1 hr. If only total titers are to be measured, then it is not necessary to heat inactivate the serum.

* Preparation of 0.2M 2-mercaptoethanol (Used only for determining IgM)

- 1.4 ml 2-mercaptoethanol + 98.6 ml saline
- Make solution fresh daily.
- Store 2-ME refrigerated and in the dark.

2. Procedure for total antibody titers

- a. Add 25 μ l of saline to each well of microtiter plate. If also doing IgM titers, only add saline to the first well, do step 6, cover plates and incubate at 37 C for 30 min. Then add saline to the remaining wells.
- b. Add 25 μ l of antiserum to the first well of each row.
- c. Mix the first well and then, using a 25 μ l wand, transfer 25 μ l to the next well, mix and continue procedure to the final well.
- d. Add 25 μ l of antigen (2% SRBC) to each well, excluding the last well. Add saline to the last well to use as a control.
- e. Incubate at 37 C for 30 minutes.
- f. Read immediately. Values represent \log_2 .

NOTE: If measuring both IgM and IgG, need 30 ml of 2% SRBC per 30 blood samples.

3. Procedure for IgM titers

- a. Prepare a second set of microtiter plates by placing 25 μ l of 0.2 M 2-ME in the first well.
- b. Place 25 μ l of antiserum in the first well, cover with Saran wrap and incubate for 30 minutes at 37 C.
- c. Add 25 μ l of saline to remaining wells.

- d. Repeat steps c-f above.
- e. Titers are read and recorded as ME-resistant (ME-R) titers. The ME-sensitive (ME-S) titer is the difference between the total titers and the ME-R titers.
 - * The ME-S Ig is associated with levels of IgM and ME-R Ig is IgG levels.

References

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- Wegman, T. G., and O. Smithies. 1966. A simple hemagglutination system requiring small amounts of red cells and antibodies. *Transfusion* 6:67-73.
- Delhanty, J. J., and J. B. Solomon. 1966. The nature of the antibodies to goat erythrocytes in the developing chicken. *Immunol.* 11:103-113.

Brain Catecholamine and Indoleamine Assay for EC-HPLC

Materials:

Volumetric Flasks
1 1 Liter
1 250 ml
7 100 ml
1 25 ml
7 10 ml
Balances (0.00 mg and 0.000 g)
Millipore Filtering Apparatus
PII Meter
Stir Plate
Brinkman Polytron
Microfilters (Bioanalytical System)
Refrigerated Centrifuge
Ultralow Freezer(-70)

Chemicals:

70% Perchloric Acid
diNaEDTA
Sodium Hydroxide
Sodium Octyl Sulfate
Citric Acid
Acetonitrile
Triethanolamine
Norepinephrine Bitartrate (NE)
L-dopa (DOPA)
Dopamine (DA)
Serotonin (5HT)
5-Hydroxyindole Acetic Acid (5-HIAA)
HPLC Grade Water
Buffer Solution PH 4.0 and 7.0
Epinephrine (E)
3,4 Dihydroxybenzylamine (DHIBA)

Procedure:

Plan how many samples you can run that day. Each sample takes about 20 minutes to run. In addition to samples plan to run a standard every 4 samples. Keep brains and all solution on ice.

1. Weight brain sections in 10 ml tubes and record.
2. Add brain buffer:
 - a. 3ml for diencephalon, brain stem, and crebellum
 - b. 6 ml for telencephalon
3. Add 30 ul of 10 ug/ml solution of DHIBA in 3ml buffer and 60 ul in 6 ml buffer
4. Homogenize using Brinkman polytron.
5. Centrifuge (0°C) at 11,000 g for 15 minutes.
6. Pour off supernatant into BAS filters and centrifuge 5 minutes at 3000 g.
7. Freeze samples immediately and keep until ready to inject.
8. Inject 100 ul of filtered supernatant into HPLC.
9. Calibrate HPLC by injecting 100 ul of mixed standard

HPLC Mobile Phase for Brain Catecholamine and Indolamine Assay

1. 150 mg diNaOH
2. 350 mg NaOH
3. 19.2 g Citric acid
4. 225 mg Sodium octyl sulfate (variable)
5. 85 ml Acetonitrile
6. 1.8 ml Triethylamine
7. Bring up to 1 liter with HPLC grade water
8. Set PH to 3.1 using NaOH or citric acid
9. Filter and degas by passing through degaser before it enters HPLC

* Sodium octyl sulfate increases retention time; acetonitrile decreases retention time; triethylamine helps decrease "tail-in" of last peaks.

Brain Buffer (for 250 mls)

1. 37.5 mg diNa EDTA
2. 1.075 mls of 70% perchloric acid (0.5M)
3. Bring to 250 mls with HPLC grade water and refrigerate

Standards:

All standards are made with 0.1M HClO₄ (8.6 ml of 70% HClO₄/liter)

1. Preparation of Stock solutions (50 mg/100 mls)
 - a. NE - 94.36 mg per 100 mls of 0.1M HClO₄
 - b. E - 90.97 mg per 100 mls of 0.1M HClO₄
 - c. DOPA - 50.00 mg per 100 mls of 0.1M HClO₄
 - d. DA - 61.91 mg per 100 mls of 0.1M HClO₄
 - e. 5HT - 110.56 mg per 100 mls 0.1M HClO₄
 - f. 5HIAA - 50.0 mg per 100 mls 0.1M HClO₄
 - g. DHBA - 80 mg per 100 mls of 0.1M HClO₄
2. Preparation of working stock solution (10 µg/ml)
 - a. For all the above standards, add 200 µl of the stock solution to a 10 ml volumetric flask.
 - b. Bring to 10 ml using 0.1M HClO₄
3. Preparation of mixed standard for analysis
 - a. For all standard, except DHBA, add 125 µl of the working stock solution to a 25 ml volumetric flask.
 - b. Bring to 25 mls using the brain buffer. Make the mixed standard daily.
 - c. Refrigerate between uses.

HPLC Settings and Specifications:

Column specifications - Biophase ODS 5 μm ; 250 x 4.6 mm (Bioanalytical Systems - Part # MF6017)

Set pump at 1.5 ml/min

Set detector at 600 mV potential and 10 nA/V or 20 nA/V

Sample Calculation ($\mu\text{g/g}$)

$$\frac{\frac{\text{pk ht of X}}{\text{pk ht of DHBA unknown}} \times \text{std. conc.} \times \text{dil. factor}}{\frac{\text{pk ht of Y}}{\text{pk ht of DHBA known}}}$$

Brain Part Weight

pk ht = Peak height

X = catecholamine or indoleamine of interest from sample run

DHBA unknown = peak height of DHBA from the sample run

Y = catecholamine or indoleamine of interest from calibration run

DHBA known = peak height of DHBA from calibration run

std. conc. = concentration of the standard used for calibration

NOTE: The dilution factor takes into account the amount injected into the HPLC versus the amount of extract. For example if 100 μl are injected and one had 3 mls of extract, the dilution factor would be 3000 $\mu\text{l}/100 \mu\text{l}$ or 30.

The integrator and the computer can be programmed to calculate the numerator of the above equation.

Appendix B FEED FORMULATION OF STARTER, GROWER, AND SUPPLEMENTED DIET

Appendix B table. Feed formulation ingredient (%) for starter and grower diet

Ingredients	Starter	Grower
Corn	64.70	68.60
Soybean meal	24.00	20.00
Fish meal	1.00	-
Meat and bone meal	6.50	7.60
Limestone	0.55	0.55
Stabilized Fat	2.50	2.50
DL-methionine	0.20	0.20
Iodized salt	0.25	0.25
Vitamin premix ¹	0.25	0.25
Trace mineral premix ²	0.05	0.05

* Supplemented diet was formulated by adding 0.5% or 1% of L-tyrosine or L-tryptophan into grower diet.

¹ Vitamin Premix supplied (per kilogram feed): vitamin A, 2200 IU; vitamin D₃, 1100 IU; Vitamin E, 0.55 IU; choline chloride, 125 mg; menadione sodium bisulfite, 0.88 mg; calcium D-pantothenate, 1.65 mg; vitamin B₁₂, 2.75 µg; selenium, 0.1 mg; riboflavin, 1.65 mg; folic acid, 0.138 mg; niacin, 16.5 mg.

² Trace mineral premix supplied (per kilogram feed) manganese (min.) and zinc, 60 mg; iron, 20 mg; copper, 2.5 mg; iodine, 1 mg; cobalt, 0.225 mg and calcium (min.), 120 mg to (max.) 160 mg.

Appendix C

ANALYSIS OF VARIANCE TABLES

Appendix C Table 1. Analysis of variance for primary antibody titers in Leghorns fed 0.1% tyrosine or tryptophan supplemental diets at different days after SRBC injection

Parameter	Days	Source of variance	df	Sum of squares	F value	P
5% SRBC Injection						
Total	3	Trmt	2	1.267	0.98	0.3873
		Error	27	17.400		
		Total	29	18.667		
	5	Trmt	2	2.067	1.04	0.3682
		Error	27	26.900		
		Total	29	28.967		
	7	Trmt	2	3.267	0.89	0.4213
		Error	27	49.400		
		Total	29	52.667		
	9	Trmt	2	2.600	1.34	0.2788
		Error	27	26.200		
		Total	29	28.800		
IgG	3	Trmt	2	0	.	.
		Error	27	0		
		Total	29	0		
	5	Trmt	2	0.467	2.10	0.1420
		Error	27	3.000		
		Total	29	3.467		
	7	Trmt	2	2.600	1.24	0.3040
		Error	27	28.200		
		Total	29	30.800		
	9	Trmt	2	0.600	0.46	0.6376
		Error	27	17.700		
		Total	29	18.300		
IgM	3	Trmt	2	1.267	0.98	0.3873
		Error	27	17.400		
		Total	29	18.667		
	5	Trmt	2	0.600	0.30	0.7424
		Error	27	26.900		
		Total	29	27.500		
	7	Trmt	2	5.067	0.81	0.4552
		Error	27	84.400		
		Total	29	89.467		
	9	Trmt	2	0.800	0.35	0.7066
		Error	27	30.700		
		Total	29	31.500		

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Appendix C Table 1. continued

Parameter	Days	Source of variance	df	Sum of squares	F value	P
0.5% SRBC Injection						
Total	3	Trmt	2	4.267	2.29	0.1201
		Error	27	25.100		
		Total	29	29.367		
	5	Trmt	2	8.267	1.39	0.2668
		Error	27	80.400		
		Total	29	88.667		
	7	Trmt	2	1.400	0.38	0.6878
		Error	27	49.800		
		Total	29	51.200		
	9	Trmt	2	7.467	2.29	0.1205
		Error	27	44.000		
		Total	29	51.467		
IgG	3	Trmt	2	0	.	.
		Error	27	0		
		Total	29	0		
	5	Trmt	2	0.200	1.08	0.3538
		Error	27	2.500		
		Total	29	2.700		
	7	Trmt	2	1.400	0.92	0.4116
		Error	27	20.600		
		Total	29	22.000		
	9	Trmt	2	0.267	0.22	0.8075
		Error	27	16.700		
		Total	29	16.967		
IgM	3	Trmt	2	4.267	2.29	0.1201
		Error	27	25.100		
		Total	29	29.367		
	5	Trmt	2	10.466	1.59	0.2225
		Error	27	88.900		
		Total	29	99.367		
	7	Trmt	2	4.200	1.16	0.3295
		Error	27	49.000		
		Total	29	53.200		
	9	Trmt	2	8.267	3.29	0.0526
		Error	44	44.300		
		Total	48	48.000		

Appendix C Table 2. Analysis of variance for the secondary antibody titers in leghorns fed 0.1% tyrosine and tryptophan supplemental diets at different days after SRBC injection

Parameter	Days	Source of variance	df	Sum of squares	F value	P
5% SRBC Injection						
Total	3	Trmt	2	1.667	0.80	0.4606
		Error	27	28.200		
		Total	29	29.867		
	5	Trmt	2	4.666	0.17	0.8440
		Error	27	36.900		
		Total	29	37.367		
	7	Trmt	2	2.400	2.19	0.1315
		Error	27	14.800		
		Total	29	17.200		
	9	Trmt	2	3.800	2.95	0.0695
		Error	27	17.400		
		Total	29	21.200		
IgG	3	Trmt	2	0.600	0.48	0.6244
		Error	27	16.900		
		Total	29	17.500		
	5	Trmt	2	2.467	0.60	0.5571
		Error	27	55.700		
		Total	29	58.167		
	7	Trmt	2	0.800	0.30	0.7457
		Error	27	36.400		
		Total	29	37.200		
	9	Trmt	2	8.467	5.36	0.0111
		Error	27	21.400		
		Total	29	29.867		
IgM	3	Trmt	2	0.267	0.10	0.9029
		Error	27	35.100		
		Total	29	35.367		
	5	Trmt	2	0.800	0.22	0.8015
		Error	27	48.400		
		Total	29	49.200		
	7	Trmt	2	0.800	0.28	0.7613
		Error	27	39.200		
		Total	29	40.000		
	9	Trmt	2	14.066	6.21	0.0061
		Error	27	30.600		
		Total	29	44.667		

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Appendix C Table 2 continued

Parameter	Days	Source of variance	df	Sum of squares	F value	P
0.5% SRBC Injection						
Total	3	Trmt	2	0.467	0.16	0.8517
		Error	27	39.000		
		Total	29	39.467		
	5	Trmt	2	0.200	0.05	0.9485
		Error	27	51.000		
		Total	29	51.200		
	7	Trmt	2	0.267	0.12	0.8911
		Error	27	31.100		
		Total	29	31.367		
	9	Trmt	2	0.473	0.12	0.8834
		Error	26	49.389		
		Total	28	49.862		
IgG	3	Trmt	2	3.467	3.35	0.0429
		Error	27	13.200		
		Total	29	16.667		
	5	Trmt	2	3.467	1.38	0.2686
		Error	27	33.900		
		Total	29	37.367		
	7	Trmt	2	3.467	1.40	0.2646
		Error	27	33.500		
		Total	29	36.967		
	9	Trmt	2	8.267	3.58	0.0419
		Error	27	31.200		
		Total	29	39.467		
IgM	3	Trmt	2	4.067	1.21	0.3141
		Error	27	45.400		
		Total	29	49.467		
	5	Trmt	2	2.467	0.52	0.5988
		Error	27	63.700		
		Total	29	66.167		
	7	Trmt	2	5.600	1.27	0.2975
		Error	27	59.600		
		Total	29	65.200		
	9	Trmt	2	13.186	3.17	0.0586
		Error	26	54.056		
		Total	28	67.241		

Appendix C Table 3. Analysis of variance for primary antibody titers in broilers fed 0.1% tyrosine or tryptophan supplemental diets at days after SRBC injection

Parameter	Days	Source of variance	df	Sum of squares	F value	P
5% SRBC Injection						
Total	3	Trmt	2	4.267	1.22	0.3102
		Error	27	47.100		
		Total	29	51.367		
	5	Trmt	2	2.067	0.74	0.4874
		Error	27	37.800		
		Total	29	39.867		
	7	Trmt	2	1.267	0.71	0.5022
		Error	27	24.200		
		Total	29	25.467		
	9	Trmt	2	1.267	0.72	0.4951
		Error	27	23.700		
		Total	29	24.967		
	11	Trmt	2	0.800	0.51	0.6065
		Error	27	21.200		
		Total	29	22.000		
IgG	3	Trmt	2	0	.	.
		Error	27	0		
		Total	29	0		
	5	Trmt	2	0	.	.
		Error	27	0		
		Total	29	0		
	7	Trmt	2	1.400	0.52	0.5983
		Error	27	36.100		
		Total	29	37.500		
	9	Trmt	2	0.600	0.24	0.7907
		Error	27	34.200		
		Total	29	34.800		
	11	Trmt	2	0.867	0.81	0.4567
		Error	27	14.500		
		Total	29	15.367		
IgM	3	Trmt	2	4.267	1.22	0.3102
		Error	27	47.100		
		Total	29	51.367		
	5	Trmt	2	2.067	0.74	0.4874
		Error	27	37.800		
		Total	29	39.867		
	7	Trmt	2	2.467	1.17	0.3261
		Error	27	28.500		
		Total	29	30.967		
	9	Trmt	2	0.467	0.16	0.8513
		Error	27	38.900		
		Total	29	39.367		
	11	Trmt	2	3.267	1.57	0.2266
		Error	27	28.100		
		Total	29	31.367		

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Appendix C Table 3 continued

Parameter	Days	Source of variance	df	Sum of squares	F value	P
0.5% SRBC Injection						
Total	3	Trmt	2	4.467	1.70	0.2012
		Error	27	3.700		
		Total	29	4.167		
	5	Trmt	2	0.600	0.18	0.8349
		Error	27	44.600		
		Total	29	45.200		
	6	Trmt	2	2.726	0.76	0.4785
		Error	26	46.722		
		Total	28	49.448		
	7	Trmt	2	9.030	2.19	0.1318
		Error	26	53.522		
		Total	28	62.552		
	9	Trmt	2	5.130	1.35	0.2770
		Error	26	49.422		
		Total	28	54.552		
IgG	3	Trmt	2	0	0	1.0000
		Error	27	19.200		
		Total	29	19.200		
	5	Trmt	2	1.067	1.13	0.3394
		Error	27	12.800		
		Total	29	13.867		
	6	Trmt	2	0.341	0.18	0.8378
		Error	27	24.900		
		Total	29	25.241		
	7	Trmt	2	0.718	0.25	0.7814
		Error	26	37.489		
		Total	28	38.207		
	9	Trmt	2	0.293	0.53	0.5938
		Error	26	7.156		
		Total	28	7.448		
IgM	3	Trmt	2	0.467	0.33	0.7194
		Error	27	18.900		
		Total	29	19.367		
	5	Trmt	2	2.466	0.56	0.5753
		Error	27	59.000		
		Total	29	61.467		
	6	Trmt	2	1.139	0.31	0.7358
		Error	27	47.689		
		Total	29	48.828		
	7	Trmt	2	11.252	2.74	0.0829
		Error	26	53.300		
		Total	28	64.552		
	9	Trmt	2	7.203	2.25	0.1252
		Error	26	41.556		
		Total	28	48.758		

Appendix C Table 4. Analysis of variance for the secondary antibody titers in broilers fed 0.1% tyrosine and tryptophan supplemental diets at days after SRBC injection

Parameter	Days	Source of variance	df	Sum of squares	F value	P
5% SRBC Injection						
Total	3	Trmt	2	8.467	0.92	0.4125
		Error	27	124.900		
		Total	29	133.367		
	5	Trmt	2	1.400	0.19	0.8279
		Error	27	99.400		
		Total	29	100.800		
	7	Trmt	2	2.600	0.47	0.6282
		Error	27	74.200		
		Total	29	76.800		
	9	Trmt	2	6.467	0.82	0.4529
		Error	27	107.000		
		Total	29	113.467		
IgG	3	Trmt	2	0.200	0.05	0.9473
		Error	27	49.800		
		Total	29	50.000		
	5	Trmt	2	0.067	0.01	0.9867
		Error	27	67.400		
		Total	29	67.467		
	7	Trmt	2	6.179	2.00	0.1568
		Error	27	38.678		
		Total	29	44.857		
	9	Trmt	2	0.467	0.11	0.8978
		Error	27	58.200		
		Total	29	58.667		
IgM	3	Trmt	2	10.067	1.67	0.2068
		Error	27	81.300		
		Total	29	91.367		
	5	Trmt	2	6.067	4.85	0.0159
		Error	27	16.900		
		Total	29	22.967		
	7	Trmt	2	1.003	0.34	0.7139
		Error	27	36.711		
		Total	29	37.714		
	9	Trmt	2	10.067	5.35	0.0110
		Error	27	25.400		
		Total	29	35.467		

Appendix C Table 5. Analysis of variance for fold differences of wattle thickness in response to PIIA injection in Leghorns and broilers fed 0.1% tyrosine or tryptophan supplemental diets

Parameter	Source of variance	df	Sum of squares	F value	P
Leghorns					
Fold differences	Trmt	2	3.838	2.18	0.1223
	Error	27	50.163		
	Total	29	54.001		
Broilers					
	Trmt	2	0.141	0.21	0.8082
	Error	50	16.448		
	Total	52	16.588		

Appendix C Table 6. Analysis of variance for relative body weight changes in Leghorns fed 0.1% tyrosine and tryptophan supplemental diets at days after *E. coli* challenge

Parameter	Days	Source of variance	df	Sum of squares	F value	P
Weight change	1	Trmt	2	0.001	0.24	0.7874
		Error	33	0.074		
		Total	35	0.075		
	2	Trmt	2	0.002	0.90	0.4173
		Error	33	0.036		
		Total	35	0.038		
	3	Trmt	2	0.005	2.50	0.0977
		Error	33	0.032		
		Total	35	0.036		
	4	Trmt	2	0.002	0.81	0.4527
		Error	33	0.049		
		Total	35	0.051		

Appendix C Table 7. Analysis of variance for transformed scores of lesion of *E. coli* challenge in Leghorns fed 0.1% tyrosine or tryptophan supplemental diets

Parameter	Source of variance	df	Sum of squares	F value	P
Total	Trmt	2	0.097	0.11	0.8955
	Error	24	10.533		
	Total	26	10.630		

Appendix C Table 8. Analysis of variance for primary and secondary antibody titers in Leghorns fed 0.5% or 1% tyrosine and tryptophan supplemental diets at days after SRBC injection

Parameter	Days	Source of variance	df	Sum of squares	F value	P
Primary antibody response						
Total	3	Trmt	4	4.170	1.36	0.2641
		Error	44	33.789		
		Total	48	37.960		
	5	Trmt	4	3.285	0.94	0.4492
		Error	44	38.389		
		Total	48	41.673		
	7	Trmt	4	6.508	1.28	0.2921
		Error	44	55.900		
		Total	48	62.408		
	9	Trmt	4	3.875	0.95	0.4419
		Error	44	44.656		
		Total	48	48.531		
	11	Trmt	4	3.042	0.68	0.6122
		Error	44	49.489		
		Total	48	52.530		
IgG	3	Trmt	4	0	.	.
		Error	44	0		
		Total	48	0		
	5	Trmt	4	1.739	0.99	0.4230
		Error	44	19.322		
		Total	48	21.061		
	7	Trmt	4	2.076	1.33	0.2743
		Error	44	17.189		
		Total	48	19.265		
	9	Trmt	4	2.813	1.39	0.2536
		Error	44	22.289		
		Total	48	25.102		
	11	Trmt	4	5.276	2.28	0.0762
		Error	44	25.500		
		Total	48	30.776		
IgM	3	Trmt	4	4.170	1.36	0.2641
		Error	44	33.78		
		Total	48	37.959		
	5	Trmt	4	7.672	2.44	0.0634
		Error	44	35.022		
		Total	48	42.061		
	7	Trmt	4	12.244	1.62	0.1873
		Error	44	69.756		
		Total	48	80.000		

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Appendix C Table 8 continued

Parameter	Days	Source of variance	df	Sum of squares	F value	P
	9	Trmt	4	4.965	1.13	0.3544
		Error	44	48.300		
		Total	48	53.265		
	11	Trmt	4	8.800	2.91	0.0323
		Error	44	33.322		
		Total	48	42.122		
		Secondary antibody response				
Total	2	Trmt	4	0.432	0.21	0.9331
		Error	44	22.956		
		Total	48	23.388		
	4	Trmt	4	7.759	1.47	0.2286
		Error	44	58.200		
		Total	48	65.959		
	6	Trmt	6	4.794	1.20	0.3238
		Error	44	43.900		
		Total	48	48.694		
	8	Trmt	4	6.353	1.45	0.2324
		Error	44	48.056		
		Total	48	54.408		
IgG	2	Trmt	4	1.078	1.33	0.2743
		Error	44	8.922		
		Total	48	10.000		
	4	Trmt	4	2.467	1.54	0.2081
		Error	44	17.656		
		Total	48	20.122		
	6	Trmt	4	1.531	1.12	0.3582
		Error	44	15.000		
		Total	48	16.531		
	8	Trmt	4	1.662	1.32	0.2788
		Error	44	13.889		
		Total	48	15.551		
IgM	2	Trmt	4	1.046	0.41	0.8002
		Error	44	28.056		
		Total	48	29.102		
	4	Trmt	4	6.246	1.30	0.2848
		Error	44	52.856		
		Total	48	59.102		
	6	Trmt	4	3.533	0.88	0.4830
		Error	44	44.100		
		Total	48	47.633		
	8	Trmt	4	3.700	0.92	0.4616
		Error	44	44.300		
		Total	48	48.000		

Appendix C Table 9. Analysis of variance for primary and secondary antibody titers in broilers fed 0.5% or 1% tyrosine and tryptophan supplemental diets at days after SRBC injection

Parameter	Days	Source of variance	df	Sum of squares	F value	P
Primary antibody response						
Total	3	Trmt	4	1.000	0.59	0.6701
		Error	45	19.000		
		Total	49	20.000		
	5	Trmt	4	4.666	0.62	0.6502
		Error	45	83.400		
		Total	49	88.000		
	7	Trmt	4	11.320	1.73	0.1594
		Error	45	73.500		
		Total	49	84.820		
	9	Trmt	4	11.000	1.90	0.1263
		Error	45	65.000		
		Total	49	76.000		
IgG	3	Trmt	4	0.120	0.75	0.5632
		Error	45	1.800		
		Total	49	1.920		
	5	Trmt	4	0.080		0.4175
		Error	45	0.900		
		Total	49	0.980		
	7	Trmt	4	2.520	2.25	0.0785
		Error	45	12.600		
		Total	49	15.120		
	9	Trmt	4	1.520	1.41	0.2450
		Error	45	12.100		
		Total	49	13.620		
IgM	3	Trmt	4	0.720	0.55	0.7019
		Error	45	14.800		
		Total	49	15.520		
	5	Trmt	4	5.680	0.82	0.5178
		Error	45	77.700		
		Total	49	83.380		
	7	Trmt	4	11.480	1.80	0.1462
		Error	45	71.900		
		Total	49	83.380		
	9	Trmt	4	10.120	2.32	0.0715
		Error	45	49.100		
		Total	49	59.220		

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Appendix C Table 9 continued

Parameter	Days	Source of variance	df	Sum of squares	F value	P
Secondary antibody response						
Total	2	Trmt	4	7.971	4.26	0.0053
		Error	44	20.560		
		Total	48	28.531		
	4	Trmt	4	12.587	1.90	0.1269
		Error	44	72.760		
		Total	48	85.347		
	6	Trmt	6	11.055	3.52	0.0142
		Error	44	34.578		
		Total	48	45.633		
	8	Trmt	4	5.496	1.29	0.2884
		Error	43	40.909		
		Total	47	49.979		
IgG	2	Trmt	4	0.080	0.97	0.4321
		Error	44	0.900		
		Total	48	0.980		
	4	Trmt	4	3.560	1.08	0.3794
		Error	44	36.359		
		Total	48	39.918		
	6	Trmt	4	3.248	0.97	0.4354
		Error	44	36.957		
		Total	48	40.204		
	8	Trmt	4	9.070	2.38	0.0661
		Error	43	40.909		
		Total	47	49.979		
IgM	2	Trmt	4	7.100	3.14	0.0234
		Error	44	24.860		
		Total	48	31.959		
	4	Trmt	4	6.062	1.28	0.2909
		Error	44	51.938		
		Total	48	58.000		
	6	Trmt	4	4.628	1.64	0.1813
		Error	44	31.045		
		Total	48	35.673		
	8	Trmt	4	2.665	0.60	0.6652
		Error	43	47.814		
		Total	47	50.479		

Appendix C Table 10. Analysis of variance for primary and secondary antibody titers in broilers fed 0.5% or 1% tyrosine and tryptophan supplemental diets at days after SRBC injection

Parameter	Days	Source of variance	df	Sum of squares	F value	P
Primary antibody response						
Total	3	Trmt	5	9.483	18.62	0.0001
		Error	54	5.500		
		Total	59	14.983		
	5	Trmt	5	8.933	2.11	0.0786
		Error	54	45.800		
		Total	59	54.733		
	7	Trmt	5	12.333	1.63	0.1672
		Error	54	81.600		
		Total	59	93.933		
	9	Trmt	5	13.950	1.48	0.2112
		Error	54	101.700		
		Total	59	115.650		
IgG	3	Trmt	5	0	.	.
		Error	54	0		
		Total	59	0		
	5	Trmt	5	0.133	0.80	0.5546
		Error	54	1.800		
		Total	59	1.933		
	7	Trmt	5	3.083	1.58	0.1819
		Error	54	21.100		
		Total	59	24.183		
	9	Trmt	5	2.200	1.29	0.2813
		Error	54	18.400		
		Total	59	20.600		
IgM	3	Trmt	5	9.483	18.62	0.0001
		Error	54	5.500		
		Total	59	14.983		
	5	Trmt	5	8.600	2.12	0.0769
		Error	54	43.800		
		Total	59	52.400		
	7	Trmt	5	14.283	2.33	0.0551
		Error	54	66.300		
		Total	59	80.583		
	9	Trmt	5	18.150	2.29	0.0582
		Error	54	85.500		
		Total	59	103.650		

(continued on next page)

Appendix C Table 10

Parameter	Days	Source of variance	df	Sum of squares	F value	P
Secondary antibody response						
Total	2	Trmt	5	1.314	0.88	0.5038
		Error	52	15.600		
		Total	57	16.914		
	4	Trmt	5	10.505	1.24	0.3032
		Error	52	87.978		
		Total	57	98.483		
	6	Trmt	5	5.384	0.71	0.6202
		Error	50	76.044		
		Total	55	81.429		
	8	Trmt	5	9.885	1.96	0.1002
		Error	51	51.378		
		Total	56	61.263		
IgG	2	Trmt	5	0	.	.
		Error	52	0		
		Total	57	0		
	4	Trmt	5	2.468	0.64	0.6728
		Error	51	39.567		
		Total	56	42.035		
	6	Trmt	5	7.898	2.10	0.0812
		Error	50	37.656		
		Total	55	45.554		
	8	Trmt	5	3.033	0.62	0.6846
		Error	51	49.844		
		Total	56	52.877		
IgM	2	Trmt	5	1.314	0.88	0.5038
		Error	52	15.600		
		Total	57	16.914		
	4	Trmt	5	7.686	1.07	0.3870
		Error	51	73.156		
		Total	56	80.842		
	6	Trmt	5	3.536	0.52	0.7576
		Error	50	67.589		
		Total	55	71.125		
	8	Trmt	5	2.368	0.35	0.8820
		Error	51	69.667		
		Total	56	72.035		

Appendix C Table 11. Analysis of variance for 5-HIAA, 5-HT levels and their ratios in the brain stem of male Leghorns fed 0.5% or 1% supplemental tyrosine and tryptophan

Parameter	Days	Source of variance	df	Sum of squares	F value	P
5-HT	2	Trmt	4	890671.1	8.60	0.0003
		Error	20	517900.4		
		Total	24	1408571.6		
	4	Trmt	4	496964.6	1.26	0.3182
		Error	20	1971192		
		Total	24	2468156.8		
	6	Trmt	4	704718.1	4.42	0.0108
		Error	19	757583.1		
		Total	23	1462301.2		
	8	Trmt	4	185912.5	1.96	0.1397
		Error	20	474253.8		
		Total	24	660166.4		
	10	Trmt	4	151537.7	1.01	0.4258
		Error	19	711002.4		
		Total	23	862540.1		
5-HIAA	2	Trmt	4	244694.4	21.13	0.0001
		Error	20	57888.4		
		Total	24	302582.8		
	4	Trmt	4	282506.6	6.60	0.0015
		Error	20	204121.9		
		Total	24	496528.5		
	6	Trmt	4	138163.0	14.48	0.0001
		Error	19	45311.5		
		Total	23	183474.5		
	8	Trmt	4	13061.8	0.70	0.5980
		Error	20	92676.8		
		Total	24	105738.6		
	10	Trmt	4	11647.4	0.71	0.5966
		Error	19	78187.3		
		Total	23	89834.7		
5-HIAA/5-HT	2	Trmt	4	0.031	1.21	0.3381
		Error	20	0.128		
		Total	24	0.159		
	4	Trmt	4	0.074	7.87	0.0006
		Error	20	0.047		
		Total	24	0.121		
	6	Trmt	4	0.029	3.77	0.0200
		Error	19	0.036		
		Total	23	0.064		
	8	Trmt	4	0.064	1.12	0.3762
		Error	20	0.287		
		Total	24	0.351		
	10	Trmt	4	0.016	0.77	0.5602
		Error	19	0.010		
		Total	23	0.116		

Appendix C Table 11. Analysis of variance for 5-HIAA, 5-IIT levels and their ratios in the brain stem of male Leghorns between days within each dietary treatment

Parameter	Trmt	Source of variance	df	Sum of squares	F value	P
5-IIT	Control	Trmt	4	115398.1	0.93	0.4660
		Error	20	619851.1		
		Total	24	735249.2		
	0.5% Tyr	Trmt	4	587838.0	2.00	0.1361
		Error	19	1398984.6		
		Total	23	1986822.6		
	1% Tyr	Trmt	4	260198.6	1.85	0.1581
		Error	20	701685.1		
		Total	24	961883.6		
	0.5% Trp	Trmt	4	522112.3	2.86	0.0521
		Error	19	867870.4		
		Total	23	1389982.7		
	1% Trp	Trmt	4	631950.9	3.75	0.0197
		Error	20	843526.6		
		Total	24	1475477.5		
5-HIAA	Control	Trmt	4	14207.1	1.17	0.3549
		Error	20	60856.8		
		Total	24	75063.9		
	0.5% Tyr	Trmt	4	38217.5	1.95	0.1433
		Error	19	93070.1		
		Total	23	131287.6		
	1% Tyr	Trmt	4	41860.2	4.80	0.0071
		Error	20	43618.1		
		Total	24	85478.3		
	0.5% Trp	Trmt	4	90243.8	2.84	0.0532
		Error	19	151035.8		
		Total	23	241279.6		
	1% Trp	Trmt	4	289108.3	10.36	0.0001
		Error	20	139493.9		
		Total	24	428602.1		
5-HIAA/5-IIT	Control	Trmt	4	0.0294	1.71	0.1873
		Error	20	0.0861		
		Total	24	0.1155		
	0.5% Tyr	Trmt	4	0.0764	1.43	0.2612
		Error	19	0.2531		
		Total	23	0.3296		
	1% Tyr	Trmt	4	0.0441	4.01	0.0151
		Error	20	0.0550		
		Total	24	0.0991		
	0.5% Trp	Trmt	4	0.0318	0.98	0.4420
		Error	19	0.1543		
		Total	23	0.1861		
	1% Trp	Trmt	4	0.0545	5.56	0.0036
		Error	20	0.0491		
		Total	24	0.1036		

Appendix C Table 12. Analysis of variance for 5-HIAA, 5-IIT levels and their ratios in the diencephalon of male Leghorns fed 0.5% or 1% supplemental tyrosine and tryptophan at selected days after SRBC injection

Parameter	Days	Source of variance	df	Sum of squares	F value	P
5-IIT	2	Trmt	4	278537.1	0.83	0.5206
		Error	18	1501364.4		
		Total	22	1779899.5		
	4	Trmt	4	374988.0	4.08	0.0140
		Error	20	459117.2		
		Total	24	834105.2		
	6	Trmt	4	722594.6	8.32	0.0005
		Error	19	412364.2		
		Total	23	1134958.8		
	8	Trmt	4	44635.7	0.44	0.7798
		Error	20	509863.0		
		Total	24	554498.6		
10	Trmt	4	20567.5	0.20	0.9341	
	Error	19	4483388.6			
	Total	23	503956.0			
5-HIAA	2	Trmt	4	178497.3	3.82	0.0193
		Error	19	222240.5		
		Total	23	400737.8		
	4	Trmt	4	99403.8	4.54	0.0090
		Error	20	109359.5		
		Total	24	208763.3		
	6	Trmt	4	122830.8	2.97	0.0460
		Error	19	196243.0		
		Total	23	319073.8		
	8	Trmt	4	12103.0	1.42	0.2626
		Error	20	42513.1		
		Total	24	54616.2		
10	Trmt	4	11894.7	0.94	0.4619	
	Error	19	60067.8			
	Total	23	71962.1			
5-HIAAA/5-IIT	2	Trmt	4	0.0919	4.98	0.0070
		Error	18	0.0831		
		Total	22	0.1750		
	4	Trmt	4	0.0521	6.20	0.0021
		Error	20	0.0420		
		Total	24	0.0941.7		
	6	Trmt	4	0.0181	1.14	0.3695
		Error	19	0.0757		
		Total	23	0.0938.5		
	8	Trmt	4	0.0202	2.44	0.0803
		Error	20	0.04132		
		Total	24	0.0614		
10	Trmt	4	0.0128	0.62	0.6537	
	Error	19	0.0977			
	Total	23	0.1104			

Appendix C Table 12. Analysis of variance for 5-HIAA, 5-IIT levels and their ratios in the diencephalon of male Leghorns between days within each dietary treatment

Parameter	Trmt	Source of variance	df	Sum of squares	F value	P
5-HT	Control	Trmt	4	465229.2	2.61	0.0684
		Error	19	847564.2		
		Total	23	1312793.4		
	0.5% Tyr	Trmt	4	453089.2	3.08	0.0398
		Error	20	736320.9		
		Total	24	1189410.0		
	1% Tyr	Trmt	4	495136.5	3.82	0.0216
		Error	17	551332.1		
		Total	21	1046468.7		
	0.5% Trp	Trmt	4	1360496.1	8.27	0.0005
		Error	19	781061.4		
		Total	23	2141557.5		
	1% Trp	Trmt	4	1381726.3	15.52	0.0001
		Error	19	422894.0		
		Total	23	1804620.3		
5-HIAA	Control	Trmt	4	15507.4	0.67	0.6224
		Error	19	110355.5		
		Total	23	125862.9		
	0.5% Tyr	Trmt	4	28511.1	1.60	0.2136
		Error	20	89171.5		
		Total	24	117682.6		
	1% Tyr	Trmt	4	42063.5	2.82	0.0578
		Error	17	63291.4		
		Total	21	105354.9		
	0.5% Trp	Trmt	4	108653.9	3.89	0.0171
		Error	20	139735.9		
		Total	24	248389.8		
	1% Trp	Trmt	4	1381726.3	15.52	0.0001
		Error	19	422894.0		
		Total	23	1804620		
5-HIAAA/5-HT	Control	Trmt	4	0.0188	0.97	0.4459
		Error	19	0.0918		
		Total	23	0.1105		
	0.5% Tyr	Trmt	4	0.0863	6.63	0.0014
		Error	20	0.0651		
		Total	24	0.1515		
	1% Tyr	Trmt	4	0.0781	6.91	0.0017
		Error	17	0.0481		
		Total	21	0.1262.5		
	0.5% Trp	Trmt	4	0.0357	3.20	0.0363
		Error	19	0.0531		
		Total	23	0.0888		
	1% Trp	Trmt	4	0.0373	1.99	0.1363
		Error	19	0.0888		
		Total	23	0.1261		

Appendix C Table 13. Analysis of variance for 5-HIAA, 5-HT levels and their ratios in the telencephalon of male Leghorns fed with 0.5% or 1% supplemental tyrosine and tryptophan

Parameter	Days	Source of variance	df	Sum of squares	F value	P
5-HT	2	Trmt	4	236496.5	3.20	0.0347
		Error	20	369227.2		
		Total	24	605723.7		
	4	Trmt	4	456852.0	2.48	0.0768
		Error	20	920670.7		
		Total	24	1377522.7		
	6	Trmt	4	60021.1	0.91	0.4772
		Error	20	329863.4		
		Total	24	389884.6		
	8	Trmt	4	148952.5	0.61	0.6611
		Error	20	1223613.2		
		Total	24	1372565.7		
10	Trmt	4	70089.3	0.35	0.8381	
	Error	20	989646.5			
	Total	24	1059735.8			
5-HIAA	2	Trmt	4	27661.2	7.84	0.0006
		Error	20	17649.4		
		Total	24	45310.6		
	4	Trmt	4	15012.8	1.57	0.2202
		Error	20	47725.4		
		Total	24	62738.2		
	6	Trmt	4	7415.5	5.89	0.0027
		Error	20	6290.5		
		Total	24	13706.0		
	8	Trmt	4	800.9	0.82	0.5317
		Error	17	4165.2		
		Total	21	4966.0		
10	Trmt	4	78.8	0.11	0.9793	
	Error	20	3736.4			
	Total	24	3815.2			
5-HIAA/5-HT	2	Trmt	4	0.008	3.33	0.0304
		Error	20	0.012		
		Total	24	0.020		
	4	Trmt	4	0.013	0.93	0.4645
		Error	20	0.070		
		Total	24	0.083		
	6	Trmt	4	0.0062	4.43	0.0100
		Error	20	0.070		
		Total	24	0.0132		
	8	Trmt	4	0.0014	4.25	0.0145
		Error	17	0.014		
		Total	21	0.0029		
10	Trmt	4	0.0013	0.16	0.9579	
	Error	20	0.0408			
	Total	24	0.0420			

Appendix C Table 13. Analysis of variance for 5-HIAA, 5-HT levels and their ratios in the telencephalon of male Leghorns between days within each dietary treatment

Parameter	Trmt	Source of variance	df	Sum of squares	F value	P
5-HT	Control	Trmt	4	1208259.6	6.24	0.0018
		Error	21	1016370.4		
		Total	25	2224630.0		
	0.5% Tyr	Trmt	4	1214819.5	9.65	0.0002
		Error	20	629237.8		
		Total	24	1844057.3		
	1% Tyr	Trmt	4	733622.3	5.00	0.0059
		Error	20	733731.8		
		Total	24	1467354.1		
	0.5% Trp	Trmt	4	1533589.8	12.63	0.0001
		Error	20	606985.8		
		Total	24	2140575.6		
	1% Trp	Trmt	4	2244030.0	7.83	0.0006
		Error	20	1433273.6		
		Total	24	3677303.6		
5-HIAA	Control	Trmt	4	9056.8	2.25	0.1001
		Error	20	20146.8		
		Total	24	29203.7		
	0.5% Tyr	Trmt	4	2409.9	1.84	0.0533
		Error	19	4035.8		
		Total	23	6445.7		
	1% Tyr	Trmt	4	6712.5	1.13	0.3710
		Error	20	29723.4		
		Total	24	36436.0		
	0.5% Trp	Trmt	4	18055.4	5.63	0.0033
		Error	20	16042.3		
		Total	24	34097.7		
	1% Trp	Trmt	4	35936.6	10.26	0.0001
		Error	19	16643.0		
		Total	23	52579.6		
5-HIAA/5-HT	Control	Trmt	4	0.0025	1.07	0.3967
		Error	20	0.0118		
		Total	24	0.0143		
	0.5% Tyr	Trmt	4	0.0034	1.98	0.1381
		Error	19	0.0082		
		Total	23	0.0116		
	1% Tyr	Trmt	4	0.0010	0.86	0.5038
		Error	20	0.0580		
		Total	24	0.0680		
	0.5% Trp	Trmt	4	0.0177	2.42	0.0826
		Error	20	0.0367		
		Total	24	0.0545		
	1% Trp	Trmt	4	0.0164	4.39	0.0111
		Error	19	0.0177		
		Total	23	0.0341		

Appendix C Table 14. Analysis of variance for 5-HIAA, 5-HT levels, and 5-HIAA/5-HT ratios in the cerebellum of male Leghorns fed 0.5% or 1% supplemental tyrosine and tryptophan

Parameter	Days	Source of variance	df	Sum of squares	F value	P
5-HT	2	Trmt	4	25176.2	2.15	0.1162
		Error	18	52662.9		
		Total	22	77839.1		
	4	Trmt	4	90670.4	1.01	0.4318
		Error	17	383097.1		
		Total	21	473767.6		
	6	Trmt	4	126825.6	2.54	0.0737
		Error	19	237276.3		
		Total	23	364101.9		
	8	Trmt	4	158874.9	1.83	0.1645
		Error	19	412054.2		
		Total	23	570929.1		
	10	Trmt	4	41361.9	1.16	0.3577
		Error	19	168782.7		
		Total	23	210144.6		
5-HIAA	2	Trmt	4	5522.3	5.21	0.0114
		Error	12	3177.6		
		Total	16	8700.0		
	4	Trmt	4	1021.3	1.04	0.4236
		Error	13	3189.9		
		Total	17	4211.2		
	6	Trmt	4	1354.9	0.91	0.4765
		Error	20	7434.5		
		Total	24	8789.4		
	8	Trmt	4	5550.1	1.00	0.4350
		Error	18	25071.5		
		Total	22	30621.6		
	10	Trmt	4	961.1	0.80	0.5393
		Error	19	5697.6		
		Total	23	6658.7		
5-HIAAA/5-HT	2	Trmt	4	0.0477	4.73	0.0160
		Error	12	0.0302		
		Total	16	0.0779		
	4	Trmt	4	0.0100	0.87	0.5056
		Error	13	0.0371		
		Total	17	0.0470		
	6	Trmt	4	0.0280	3.44	0.0281
		Error	19	0.0386		
		Total	23	0.0665		
	8	Trmt	4	0.0191	0.12	0.9751
		Error	18	0.7403		
		Total	22	0.7594		
	10	Trmt	4	0.0086	0.92	0.4743
		Error	19	0.0446		
		Total	23	0.0533		

Appendix C Table 14. Analysis of variance for 5-HIAA, 5-HT levels, and 5-HIAA/5-HT ratios in the cerebellum of male Leghorns between days within each dietary treatment

Parameter	Trmt	Source of variance	df	Sum of squares	F value	P
5-HT	Control	Trmt	4	147561.8	3.56	0.0238
		Error	20	207094.9		
		Total	24	354656.7		
	0.5% Tyr	Trmt	4	82214.9	1.12	0.3762
		Error	19	348526.8		
		Total	23	430741.7		
	1% Tyr	Trmt	4	26452.5	1.11	0.3874
		Error	16	95613.7		
		Total	20	122066.2		
	0.5% Trp	Trmt	4	68253.5	2.12	0.1177
		Error	19	152627.2		
		Total	23	220979.7		
	1% Trp	Trmt	4	39798.6	0.14	0.8074
		Error	18	449911.8		
		Total	22	489710.4		
5-HIAA	Control	Trmt	4	4657.6	1.14	0.3737
		Error	15	15265.9		
		Total	19	19923.5		
	0.5% Tyr	Trmt	4	1066.0	1.59	0.2288
		Error	15	2517.8		
		Total	19	3583.8		
	1% Tyr	Trmt	4	4126.8	1.91	0.1586
		Error	16	8662.0		
		Total	20	12788.8		
	0.5% Trp	Trmt	4	11399.1	6.04	0.0026
		Error	19	8957.3		
		Total	23	20356.4		
	1% Trp	Trmt	4	8289.9	3.92	0.0197
		Error	17	8999.2		
		Total	21	17289.1		
5-HIAAA/5-HT	Control	Trmt	4	0.1777	1.14	0.3752
		Error	15	0.5844		
		Total	19	0.7621		
	0.5% Tyr	Trmt	4	0.0984	5.63	0.0057
		Error	15	0.0655		
		Total	19	0.1639		
	1% Tyr	Trmt	4	0.1163	6.97	0.0022
		Error	15	0.0626		
		Total	19	0.1789		
	0.5% Trp	Trmt	4	0.0831	3.60	0.0241
		Error	19	0.1098		
		Total	23	0.1929		
	1% Trp	Trmt	4	0.0737	4.57	0.0109
		Error	17	0.0686		
		Total	21	0.1423		

Appendix C Table 15. Analysis of variance for L-dopa and catecholamine levels in the brain stem of male Leghorns fed 0.5% or 1% supplemental tyrosine and tryptophan

Parameter	Days	Source of variance	df	Sum of squares	F value	P
L-Dopa	2	Trmt	4	4.170	1.36	0.2641
		Error	44	33.789		
		Total	48	37.960		
	4	Trmt	4	3.285	0.94	0.4492
		Error	44	38.389		
		Total	48	41.673		
	6	Trmt	4	3.285	0.94	0.4492
		Error	44	38.389		
		Total	48	41.673		
	8	Trmt	4	6.508	1.28	0.2921
		Error	44	55.900		
		Total	48	62.408		
10	Trmt	4	3.875	0.95	0.4419	
	Error	44	44.656			
	Total	48	48.531			
Dopamine	2	Trmt	4	0	.	.
		Error	44	0		
		Total	48	0		
	4	Trmt	4	1.739	0.99	0.4230
		Error	44	19.322		
		Total	48	21.061		
	6	Trmt	4	1.739	0.99	0.4230
		Error	44	19.322		
		Total	48	21.061		
	8	Trmt	4	2.076	1.33	0.2743
		Error	44	17.189		
		Total	48	19.265		
10	Trmt	4	2.813	1.39	0.2536	
	Error	44	22.289			
	Total	48	25.102			
Norepinephine	2	Trmt	4	4.170	1.36	0.2641
		Error	44	33.789		
		Total	48	37.959		
	4	Trmt	4	7.672	2.44	0.0634
		Error	44	35.022		
		Total	48	42.061		
	6	Trmt	4	12.244	1.62	0.1873
		Error	44	69.756		
		Total	48	80.000		
	8	Trmt	4	4.965	1.13	0.3544
		Error	44	48.300		
		Total	48	53.265		
10	Trmt	4	8.800	2.91	0.0323	
	Error	44	33.322			
	Total	48	42.122			

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Appendix C Table 15 continued

Parameter	Days	Source of variance	df	Sum of squares	F value	P
Epinephine	2	Trmt	4	3.285	0.94	0.4492
		Error	44	38.389		
		Total	48	41.673		
	4	Trmt	4	3.285	0.94	0.4492
		Error	44	38.389		
		Total	48	41.673		
	6	Trmt	4	6.508	1.28	0.2921
		Error	44	55.900		
		Total	48	62.408		
	8	Trmt	4	3.875	0.95	0.4419
		Error	44	44.656		
		Total	48	48.531		
	10	Trmt	4	3.042	0.68	0.6122
		Error	44	49.489		
		Total	48	52.530		

Appendix C Table 15. Analysis of variance for L-dopa and catecholamine levels in the brain stem of male Leghorns between days within each dietary treatment

Parameter	Trmt	Source of variance	df	Sum of squares	F value	P
L-Dopa	Control	Trmt	4	248295.3	4.11	0.0154
		Error	18	271910.5		
		Total	22	520205.9		
	0.5% Tyr	Trmt	4	300627.4	9.22	0.0003
		Error	19	154898.3		
		Total	23	455525.7		
	1% Tyr	Trmt	4	270769.4	12.41	0.0001
		Error	19	103631.8		
		Total	23	374401.2		
	0.5% Trp	Trmt	4	455097.5	14.57	0.0001
		Error	18	140576.0		
		Total	22	595673.5		
	1% Trp	Trmt	4	324535.2	7.30	0.0011
		Error	18	199987.8		
		Total	22	524522.9		
Dopamine	Control	Trmt	4	6744.6	3.13	0.0375
		Error	20	10771.1		
		Total	24	175157		
	0.5% Tyr	Trmt	4	34310.8	0.91	0.4788
		Error	19	179366.6		
		Total	23	213677.4		
	1% Tyr	Trmt	4	2049.0	1.32	0.2964
		Error	20	7756.1		
		Total	24	9805.0		
	0.5% Trp	Trmt	4	6094.9	2.84	0.0530
		Error	19	10193.1		
		Total	23	16288.0		
	1% Trp	Trmt	4	11688.8	4.34	0.0109
		Error	20	13473.7		
		Total	24	25162.5		
Norepinephrine	Control	Trmt	4	423802.3	3.17	0.0361
		Error	20	669351.3		
		Total	24	1093153.6		
	0.5% Tyr	Trmt	4	165912.5	3.34	0.0314
		Error	19	236113.8		
		Total	23	402026.3		
	1% Tyr	Trmt	4	294941.9	6.10	0.0022
		Error	20	241809.1		
		Total	24	536751.0		
	0.5% Trp	Trmt	4	808477.8	12.42	0.0001
		Error	19	309183.8		
		Total	23	1117661.6		
	1% Trp	Trmt	4	1413043.9	17.69	0.0001
		Error	19	379453.5		
		Total	23	1792497.4		

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Appendix C Table 15 continued

Parameter	Trmt	Source of variance	df	Sum of squares	F value	P
Epinephine	Control	Trmt	4	2563.1	1.30	0.3087
		Error	17	8365.7		
		Total	21	10928.8		
	0.5% Tyr	Trmt	4	13366.8	5.47	0.0051
		Error	17	10388.1		
		Total	21	23754.9		
	1% Tyr	Trmt	4	8534.3	1.03	0.4199
		Error	17	35190.1		
		Total	21	43724.3		
	0.5% Trp	Trmt	4	115639.7	3.47	0.0303
		Error	17	141773.4		
		Total	21	257413.1		
	1% Trp	Trmt	4	85691.2	1.12	0.3799
		Error	18	345659.6		
		Total	22	431350.7		

Appendix C Table 16. Analysis of variance for L-dopa and catecholamine levels in the diencephalon of male Leghorns fed 0.5% or 1% supplemental tyrosine and tryptophan at selected days after SRBC injection

Parameter	Days	Source of variance	df	Sum of squares	F value	P
L-Dopa	2	Trmt	4	177557.6	2.16	0.1176
		Error	17	349421.3		
		Total	21	526978.9		
	4	Trmt	4	38320.3	12.13	0.0001
		Error	19	15008.7		
		Total	23	53328.0		
	6	Trmt	4	30087.5	2.03	0.1310
		Error	19	70441.9		
		Total	23	100529.4		
	8	Trmt	4	68596.3	0.68	0.6168
		Error	16	404414.7		
		Total	20	473011.0		
10	Trmt	4	202195.5	2.10	0.1212	
	Error	19	458058.5			
	Total	23	660254.0			
Dopamine	2	Trmt	4	180607.2	1.98	0.1390
		Error	19	433910.1		
		Total	23	614517.3		
	4	Trmt	4	310982.1	15.98	0.0001
		Error	20	97305.9		
		Total	24	408288.0		
	6	Trmt	4	147896.3	9.70	0.0002
		Error	19	72429.2		
		Total	23	220325.5		
	8	Trmt	4	7098.3	1.02	0.4213
		Error	20	34824.6		
		Total	24	41922.9		
10	Trmt	4	5409.6	0.57	0.6855	
	Error	19	44846.8			
	Total	23	50256.3			
Norepinephine	2	Trmt	4	33958.2	0.55	0.7004
		Error	19	292598.5		
		Total	23	326556.8		
	4	Trmt	4	20104.2	0.24	0.9121
		Error	20	418173.4		
		Total	24	437277.7		
6	Trmt	4	37651.1	0.52	0.7188	
	Error	19	340775.4			
	Total	23	378426.5			

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Appendix C Table 16 continued

Parameter	Days	Source of variance	df	Sum of squares	F value	P
Epinephrine	8	Trmt	4	214490.5	0.67	0.6232
		Error	20	1610926.2		
		Total	24	1825416.6		
	10	Trmt	4	407561.6	1.60	0.2153
		Error	19	1210152.4		
		Total	23	1617714.0		
	2	Trmt	4	60597.2	4.16	0.0272
		Error	11	40077.9		
		Total	15	100675.1		
	4	Trmt	4	4502.7	0.77	0.5563
		Error	20	29170.9		
		Total	24	33673.7		
	6	Trmt	4	8432.2	1.65	0.2057
		Error	18	23018.1		
		Total	22	31451.3		
	8	Trmt	4	67496.2	0.54	0.7055
		Error	20	620622.7		
		Total	24	688119.0		
10	Trmt	4	177701.2	1.60	0.2210	
	Error	17	473326.5			
	Total	21	651027.7			

Appendix C Table 16. Analysis of variance for L-dopa and catecholamine levels in the diencephalon of male Leghorns between days within each dietary treatment

Parameter	Trmt	Source of variance	df	Sum of squares	F value	P
L-Dopa	Control	Trmt	4	1751536.8	14.96	0.0001
		Error	18	526934.7		
		Total	22	2278471.5		
	0.5% Tyr	Trmt	4	2686415.3	41.86	0.0001
		Error	19	304867.7		
		Total	23	2991283.0		
	1% Tyr	Trmt	4	1094714.7	25.40	0.0001
		Error	17	183195.5		
		Total	21	1277910.2		
	0.5% Trp	Trmt	4	1272061.5	25.66	0.0001
		Error	17	210702.1		
		Total	21	1482763.6		
	1% Trp	Trmt	4	1568619.0	95.01	0.0001
		Error	17	70165.9		
		Total	21	1638784.9		
Dopamine	Control	Trmt	4	66775.5	1.01	0.4286
		Error	19	315053.1		
		Total	23	381828.6		
	0.5% Tyr	Trmt	4	30903.1	1.93	0.1453
		Error	20	80198.7		
		Total	24	111101.8		
	1% Tyr	Trmt	4	26331.5	3.07	0.0449
		Error	17	36435.5		
		Total	21	62767.0		
	0.5% Trp	Trmt	4	129590.9	4.99	0.0059
		Error	20	129787.1		
		Total	24	259378.0		
	1% Trp	Trmt	4	380775.2	14.07	0.0001
		Error	19	128517.8		
		Total	23	509303.0		
Norepinephine	Control	Trmt	4	1473160.9	9.25	0.0003
		Error	19	756775.7		
		Total	23	2229936.6		
	0.5% Tyr	Trmt	4	2918723.6	12.05	0.0001
		Error	20	1210664.1		
		Total	24	4129387.7		
	1% Tyr	Trmt	4	1353047.3	5.40	0.0054
		Error	17	1063967.9		
		Total	21	2417015.3		

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Appendix C Table 16.

Parameter	Trmt	Source of variance	df	Sum of squares	F value	P
Epinephine	0.5% Trp	Trmt	4	696605.0	6.76	0.0013
		Error	20	515361.9		
		Total	24	1211966.8		
	1% Trp	Trmt	4	1624071.5	23.62	0.0001
		Error	19	326652.5		
		Total	23	1950724.2		
	Control	Trmt	4	106866.8	6.02	0.0033
		Error	17	75461.4		
		Total	21	182328.2		
	0.5% Tyr	Trmt	4	354651.8	5.01	0.0063
		Error	19	336439.4		
		Total	23	691091.2		
	1% Tyr	Trmt	4	224147.8	1.35	0.2937
		Error	17	708075.1		
		Total	21	932222.9		
	0.5% Trp	Trmt	4	30325.0	2.38	0.0920
		Error	17	54048.0		
		Total	21	84373.0		
1% Trp	Trmt	4	41625.7	7.87	0.0015	
	Error	14	18511.8			
	Total	18	60137.4			

Appendix C Table 17. Analysis of variance for L-dopa and catecholamine levels in the telencephalon of male Leghorns fed 0.5% or 1% supplemental tyrosine and tryptophan

Parameter	Days	Source of variance	df	Sum of squares	F value	P
Dopamine	2	Trmt	4	63615.1	1.54	0.2300
		Error	20	207116.8		
		Total	24	270731.9		
	4	Trmt	4	447633.9	4.70	0.0078
		Error	20	476622.7		
		Total	24	924256.0		
	6	Trmt	4	14258.0	0.13	0.9683
		Error	20	534979.9		
		Total	24	549238.0		
	8	Trmt	4	240086.0	2.59	0.0678
		Error	20	463332.1		
		Total	24	703418.1		
10	Trmt	4	40377.5	1.79	0.1696	
	Error	20	112523.2			
	Total	24	152900.6			
Norepinephine	2	Trmt	4	6197.7	1.33	0.3087
		Error	14	16366.7		
		Total	18	22564.4		
	4	Trmt	4	15163.1	0.52	0.7200
		Error	20	144967.2		
		Total	24	160130.3		
	6	Trmt	4	24214.3	2.36	0.0877
		Error	20	51218.0		
		Total	24	75432.3		
	8	Trmt	4	9641.4	0.60	0.6678
		Error	20	80512.7		
		Total	24	90154.1		
10	Trmt	4	618.0	0.09	0.9860	
	Error	20	36249.0			
	Total	24	36867.1			

Appendix C Table 17. Analysis of variance for L-Dopa and catecholamine levels in the telencephalon of male Leghorns between days within each dietary treatment

Parameter	Trmt	Source of variance	df	Sum of squares	F value	P
Dopamine	Control	Trmt	4	36533.3	0.66	0.6278
		Error	21	291412.1		
		Total	25	327945.5		
	0.5% Tyr	Trmt	4	99439.8	1.90	0.1493
		Error	20	261279.9		
		Total	24	360719.7		
	1% Tyr	Trmt	4	76525.2	1.22	0.3334
		Error	20	313451.1		
		Total	24	389976.4		
	0.5% Trp	Trmt	4	199994.8	2.29	0.0951
		Error	20	436059.2		
		Total	24	636053.9		
1% Trp	Trmt	4	535110.0	5.31	0.0044	
	Error	20	503528.6			
	Total	24	1038638.6			
Norepinephine	Control	Trmt	4	27596.6	3.69	0.0209
		Error	20	37426.9		
		Total	24	65023.5		
	0.5% Tyr	Trmt	4	99439.8	1.90	0.1493
		Error	20	261279.9		
		Total	24	360719.7		
	1% Tyr	Trmt	4	16040.5	0.77	0.5601
		Error	20	104769.3		
		Total	24	120809.8		
	0.5% Trp	Trmt	4	13317.5	0.92	0.4754
		Error	18	65344.0		
		Total	22	78661.5		
1% Trp	Trmt	4	9670.4	1.10	0.3905	
	Error	16	35184.3			
	Total	20	44854.6			

Appendix C Table 18. Analysis of variance for norepinephrine in the cerebellum of male Leghorns fed 0.5% or 1% supplemental tyrosine and tryptophan at selected days after SRBC injection

Parameter	Days	Source of variance	df	Sum of squares	F value	P
Norepinephrine	2	Trmt	4	85647.6	2.60	0.0712
		Error	18	148478.3		
		Total	22	234125.9		
	4	Trmt	4	52797.8	2.79	0.0576
		Error	18	85041.5		
		Total	22	137839.2		
	6	Trmt	4	56663.6	1.02	0.4209
		Error	20	277763.6		
		Total	24	334427.2		
	8	Trmt	4	23282.2	1.10	0.3833
		Error	19	100162.5		
		Total	23	123444.7		
10	Trmt	4	29623.1	0.60	0.6654	
	Error	19	233516.1			
	Total	23	263139.2			

Appendix C Table 18. Analysis of variance for norepinephrine in the cerebellum of male Leghorns between days within each dietary treatment

Parameter	Trmt	Source of variance	df	Sun of squares	F value	P
Norepinephrine	Control	Trmt	4	157720.2	5.50	0.0037
		Error	20	143425.2		
		Total	24	301145.4		
	0.5% Tyr	Trmt	4	4079.6	0.11	0.9792
		Error	19	183632.9		
		Total	23	187712.5		
	1% Tyr	Trmt	4	66803.6	2.06	0.1289
		Error	18	146014.3		
		Total	22	212817.9		
	0.5% Trp	Trmt	4	26585.1	0.71	0.5932
		Error	19	177099.2		
		Total	23	203684.3		
	1% Trp	Trmt	4	85345.5	1.97	0.1422
		Error	18	194790.4		
		Total	22	280135.9		

Appendix C Table 19. Analysis of variance for average daily feed intake and body weight gain of Leghorns and broilers during the dietary supplementation period

Parameter	Source of variance	df	Sum of squares	F value	P
Leghorns					
Feed intake	Trmt	4	9.83	0.34	0.8420
	Error	10	71.36		
	Total	14	81.19		
Body weight gain	Trmt	4	147.66	0.39	0.8099
	Error	10	941.77		
	Total	14	1089.43		
Broilers					
Feed intake	Trmt	4	79.34	0.99	0.4898
	Error	5	100.20		
	Total	9	179.54		
Body weight gain	Trmt	4	18.68	0.40	0.8002
	Error	5	57.90		
	Total	9	76.59		

Appendix C Table 20. Analysis of variance for average daily feed intake and body weight gain of stressed broilers during the dietary supplementation period

Parameter	Source of variance	df	Sum of squares	F value	P
Stressed broilers					
Feed intake	Trmt	5	173.79	0.77	0.6023
	Error	6	269.76		
	Total	9	430.55		
Body weight gain	Trmt	5	18.682	0.40	0.8002
	Error	6	57.904		
	Total	9	76.586		

Appendix B Table 21. Analysis of variance for the titers of Leghorn primary and secondary antibody titer following SRBC injection by pooling the two addition levels with the same amino acid supplementation

Parameter	Days	Source of variance	df	Sum of squares	F value	P
Primary antibody response						
Total	3	Trmt	2	4.083	2.77	0.0730
		Error	46	33.876		
		Total	48	37.959		
	5	Trmt	2	0.666	0.37	0.6905
		Error	46	41.008		
		Total	48	41.673		
	7	Trmt	2	2.0320	0.77	0.4671
		Error	46	60.376		
		Total	48	62.408		
	9	Trmt	2	3.796	1.95	0.1536
		Error	46	44.734		
		Total	48	48.531		
	11	Trmt	2	2.954	1.37	0.2641
		Error	46	49.576		
		Total	48	52.531		
IgG	3	Trmt	2	0	.	.
		Error	46	0		
		Total	48	0		
	5	Trmt	2	0.306	0.34	0.7142
		Error	46	20.755		
		Total	48	21.061		
	7	Trmt	2	0.615	0.76	0.4740
		Error	46	18.650		
		Total	48	19.265		
	9	Trmt	2	0.368	0.34	0.7121
		Error	46	24.734		
		Total	48	25.102		
	11	Trmt	2	2.954	2.44	0.0981
		Error	46	24.821		
		Total	48	30.776		
IgM	3	Trmt	2	4.083	2.77	0.0730
		Error	46	33.876		
		Total	48	37.959		
	5	Trmt	2	0.704	0.39	0.6821
		Error	46	41.989		
		Total	48	42.694		
	7	Trmt	2	2.874	0.86	0.4311
		Error	44	77.126		
		Total	48	80.000		
	9	Trmt	2	2.865	1.31	0.2803
		Error	46	50.400		
		Total	48	53.265		
	11	Trmt	2	5.515	3.46	0.0397
		Error	48	42.122		
		Total	48	42.122		

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Appendix C Table 21 continued

Parameter	Days	Source of variance	df	Sum of squares	F value	P
Secondary antibody response						
Total	2	Trmt	2	0.232	0.23	0.7947
		Error	46	23.155		
		Total	48	23.388		
	4	Trmt	2	4.133	1.54	0.2258
		Error	46	61.826		
		Total	48	65.959		
	6	Trmt	2	2.212	1.09	0.3432
		Error	46	46.482		
		Total	48	48.694		
	8	Trmt	2	3.058	1.37	0.2643
		Error	46	51.350		
		Total	48	54.408		
IgG	2	Trmt	2	0.479	1.16	0.3234
		Error	46	9.521		
		Total	48	10.000		
	4	Trmt	2	1.591	1.97	0.1504
		Error	46	28.122		
		Total	48	20.122		
	6	Trmt	2	0.196	0.28	0.7596
		Error	46	16.334		
		Total	48	16.531		
	8	Trmt	2	0.480	0.73	0.4862
		Error	46	15.071		
		Total	48	15.551		
IgM	2	Trmt	2	0.847	0.69	0.5070
		Error	46	28.255		
		Total	48	29.102		
	4	Trmt	2	2.270	0.92	0.4062
		Error	46	56.832		
		Total	48	59.102		
	6	Trmt	2	3.327	1.73	0.1891
		Error	46	44.305		
		Total	48	47.633		
	8	Trmt	2	1.879	0.94	0.3991
		Error	46	46.121		
		Total	48	48.000		

Appendix B Table 22. Analysis of variance for the titers of the broiler primary and secondary antibody titers following SRBC injection by pooling the two levels with the same amino acid supplementation

Parameter	Days	Source of variance	df	Sum of squares	F value	P
Primary antibody response						
Total	3	Trmt	2	0.500	0.60	0.5516
		Error	47	19.500		
		Total	49	20.000		
	5	Trmt	2	0.900	0.24	0.7854
		Error	47	87.100		
		Total	49	88.000		
	7	Trmt	2	11.270	3.60	0.0351
		Error	47	73.550		
		Total	49	84.820		
	9	Trmt	2	10.350	3.70	0.0321
		Error	47	65.650		
		Total	49	76.000		
IgG	3	Trmt	2	0.070	0.89	0.4178
		Error	47	1.850		
		Total	49	1.920		
	5	Trmt	2	0.030	0.74	0.4816
		Error	47	0.950		
		Total	49	0.980		
	7	Trmt	2	1.220	2.06	0.1385
		Error	47	13.900		
		Total	49	15.120		
	9	Trmt	2	1.470	2.84	0.0683
		Error	47	12.150		
		Total	49	13.620		
IgM	3	Trmt	2	0.270	0.42	0.6620
		Error	47	15.250		
		Total	49	15.520		
	5	Trmt	2	1.230	0.35	0.7053
		Error	47	82.150		
		Total	49	83.380		
	7	Trmt	2	9.630	3.07	0.0559
		Error	47	73.750		
		Total	49	83.380		
	9	Trmt	2	9.720	4.61	0.0148
		Error	47	49.500		
		Total	49	59.220		

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Appendix C Table 22 continued

Parameter	Days	Source of variance	df	Sum of squares	F value	P
Secondary antibody response						
Total	2	Trmt	2	7.758	8.59	0.0007
		Error	46	20.772		
		Total	48	28.531		
	4	Trmt	2	7.441	2.20	0.1227
		Error	46	77.906		
		Total	48	85.347		
	6	Trmt	2	4.527	2.53	0.0904
		Error	46	46.106		
		Total	48	45.633		
	8	Trmt	2	3.929	1.87	0.1661
		Error	45	47.320		
		Total	47	51.250		
IgG	2	Trmt	2	0.0296	0.72	0.4939
		Error	46	0.950		
		Total	48	0.980		
	4	Trmt	2	2.996	1.87	0.1662
		Error	46	36.922		
		Total	48	39.918		
	6	Trmt	2	1.899	1.14	0.3287
		Error	46	38.306		
		Total	48	40.204		
	8	Trmt	2	6.924	3.62	0.0349
		Error	45	43.055		
		Total	47	49.979		
IgM	2	Trmt	2	7.037	6.49	0.0033
		Error	46	24.922		
		Total	48	31.959		
	4	Trmt	2	3.094	1.30	0.2834
		Error	44	54.906		
		Total	48	58.000		
	6	Trmt	2	0.973	0.65	0.5292
		Error	46	34.700		
		Total	48	35.674		
	8	Trmt	2	0.956	0.43	0.6504
		Error	45	49.523		
		Total	47	50.479		

Appendix B Table 23. Analysis of variance for the titers of the stressed broiler primary and secondary antibody response following SRBC injection by pooling the two addition levels with the same amino acid supplementation

Parameter	Days	Source of variance	df	Sum of squares	F value	P
Primary antibody response						
Total	3	Trmt	3	6.983	16.29	0.0001
		Error	56	8.000		
		Total	59	14.983		
	5	Trmt	3	5.883	2.25	0.0927
		Error	56	48.850		
		Total	59	54.733		
	7	Trmt	3	8.733	1.91	0.1379
		Error	56	85.200		
		Total	59	93.933		
	9	Trmt	3	11.700	2.10	0.1104
		Error	56	103.950		
		Total	59	115.650		
IgG	3	Trmt	3	0		
		Error	56	0		
		Total	59	0		
	5	Trmt	3	0.133	1.38	0.2575
		Error	56	1.800		
		Total	59	1.933		
	7	Trmt	3	2.683	2.33	0.0842
		Error	56	21.500		
		Total	59	24.183		
	9	Trmt	3	1.200	1.15	0.3352
		Error	56	19.400		
		Total	59	20.600		
IgM	3	Trmt	3	6.983	16.29	0.0001
		Error	56	8.000		
		Total	59	14.983		
	5	Trmt	3	5.550	2.21	0.0968
		Error	56	46.850		
		Total	59	52.400		
	7	Trmt	3	12.683	3.49	0.0215
		Error	56	67.900		
		Total	59	80.583		
	9	Trmt	3	14.900	3.13	0.0326
		Error	56	88.750		
		Total	59	103.650		

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Appendix C Table 23 continued

Parameter	Days	Source of variance	df	Sum of squares	F value	P
Secondary antibody response						
Total	2	Trmt	3	0.430	0.47	0.7051
		Error	54	16.484		
		Total	57	16.914		
	4	Trmt	3	7.807	1.55	0.2122
		Error	54	90.676		
		Total	57	98.483		
	6	Trmt	3	2.256	0.49	0.6880
		Error	52	79.172		
		Total	55	81.429		
	8	Trmt	3	9.864	3.39	0.0245
		Error	53	51.399		
		Total	56	61.263		
IgG	2	Trmt	3	0	.	.
		Error	54	0		
		Total	57	0		
	4	Trmt	3	2.269	1.01	0.3966
		Error	53	39.766		
		Total	56	42.035		
	6	Trmt	3	7.048	3.17	0.0317
		Error	52	38.506		
		Total	55	45.554		
	8	Trmt	3	2.912	1.03	0.3870
		Error	53	49.965		
		Total	56	52.877		
IgM	2	Trmt	3	0.430	0.47	0.7051
		Error	54	16.484		
		Total	57	16.914		
	4	Trmt	3	3.929	0.90	0.4462
		Error	53	76.913		
		Total	56	80.842		
	6	Trmt	3	2.792	0.71	0.5515
		Error	52	68.333		
		Total	55	71.125		
	8	Trmt	3	2.149	0.54	0.6548
		Error	53	69.886		
		Total	56	72.035		

VITA

Jiangtao Zhu, son of Renshui Zhu and Xuchu Zhu, was born on July 5, 1962 in Fujian, P. R. of China. He attended elementary and high school at Reiken. Upon graduation in 1979, he enrolled at Fujian Agricultural College, Fuzhou, from which he received his Bachelor of Science in veterinary medicine in July, 1983. After graduation, he worked in the college for five years, then entered graduate school of Nanjing Agricultural University beginning a Master of Science in veterinary immunology in September, 1988. In August, 1989, he transferred his graduate studies to Virginia Polytechnic Institute and State University in Poultry Science.

Jiangtao Zhu