

Development of Cellulose-Titanium dioxide-Porphyrin Nanocomposite Films with High-barrier, UV-blocking, and Visible Light-Responsive Antimicrobial Features

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## ABSTRACT

The packaging does not serve as a mere containment but also can be designed to play a key role in preserving the product from quality-deteriorating factors, including oxygen, light irradiation, and foodborne pathogenic microorganisms (e.g., *Escherichia coli*). There has been a growing interest in employing ultra-porous metal-organic frameworks (MOF) with visible light-responsive antibacterial mechanisms to generate reactive oxygen species (ROS) that can eliminate bacteria via an oxidative burst. MOF is made of inorganic metal ions/nodes/clusters/secondary building units linked by organic bridge ligands, where titanium dioxide (TiO<sub>2</sub>) and tetrakis(4-carboxyphenyl)porphyrin (TCPP) were selected for these components, respectively. TiO<sub>2</sub> is an exceptional UV-A/B/C-blocker; meanwhile, TCPP dye performs a remarkable photocatalytic ability even under visible light, on top of its macro-heterocyclic structure that is ideal as a MOF linker. Both have good compatibility but suffer from the notorious tendency to self-quench/aggregate. The incorporation of MOF-based conjugates into a polymeric matrix, like cellulose, is among the proven-successful solutions. Cellulose is the Earth's most abundant and naturally biodegradable, and cellulose nanofibril (CNF) was particularly chosen for its high specific surface area and surface activity. However, a straightforward, cheap, and environmentally friendly approach of multicycle homogenization (0-25 passes) was conducted to solve neat cellulose's challenge of natural hydrophilicity, where low pressure (<10 MPa) was applied to prevent the common over-shearing effect. The antibacterial efficacy of CNF films functionalized with TiO<sub>2</sub>-TCPP conjugate on inhibiting *E. coli* growth was analyzed with and without light of different intensities (3000 and 6000 lux). The positive impacts of CNFs' promoted fibrillation and subsequent inter/intra-molecular hydrogen bonding post-homogenization were evidenced in an array of functional properties, i.e., crystallinity, TiO<sub>2</sub>-TCPP conjugate dispersion, surface smoothness, mechanical properties, thermal stability, hydrophobicity, oxygen barrier (comparable to ethylene-vinyl alcohol (EVOH), a commercial high-barrier polymer), and 100%-antibacterial rate (under 6000 lux after 72 hours). Varying optimum cycles of homogenization demonstrated the prospect of the proposed homogenization approach in preparing CNF with diverse processability and applicability. These findings also exhibited a promising potential for a myriad of high-barrier, UV-blocking, and/or visible light-responsive antibacterial film applications, including food packaging and biomedical.

# Development of Cellulose-Titanium dioxide-Porphyrin Nanocomposite Films with High-barrier, UV-blocking, and Visible Light-Responsive Antimicrobial Features

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## GENERAL AUDIENCE ABSTRACT

Packaging is useful not only as a container but can also be designed to help prevent products from being spoiled due to various reasons such as oxidation, light, and bacterial contamination. Researchers have discovered the promising antibacterial feature of the metal-organic framework (MOF). Packaging made with MOF technology can harness light and oxygen in the environment to produce a special form of oxygen called reactive oxygen species (ROS) that can kill unwanted bacteria. MOF is an extremely porous sponge-like material made of two ingredients: an inorganic metal cluster and an organic linker; in this study, titanium dioxide ( $\text{TiO}_2$ ) and a porphyrin called TCPP were selected, respectively.  $\text{TiO}_2$  is an excellent ultraviolet blocker, while TCPP has a unique, ring-like geometry that is ideal for use as a linker and an antimicrobial feature that works well under the visible light spectrum. The pair are compatible but still suffer from MOF's notorious challenge, where it tends to clump together because of its tiny size. To resolve this problem,  $\text{TiO}_2$ -TCPP MOF can be deposited evenly in a cast made of polymer. Cellulose has been proven to work effectively as a polymeric cast; moreover, it is natural, biodegradable, and in abundant supply. A type of nanosized cellulose—cellulose nanofibril (CNF)—was specifically chosen because its high surface area and activity are useful when blended with other materials. However, cellulose is naturally a poor water-repellant that is not ideal for packaging applications. As a solution, cellulose can be treated with a homogenization technique by passing the material through a very narrow hole under high pressure. Homogenization can be problematic as it possibly damages the cellulose's structure, and its high pressure can also be expensive and energy-consuming. Therefore, low pressure with multiple cycles was applied in this work. CNF- $\text{TiO}_2$ -TCPP films were tested for their ability to slow down *E. coli* bacteria growth with and without light of varying brightness to compare its light-sensitive antimicrobial feature. Homogenization was found helpful in producing higher-quality CNF, which improved several of the film's final characteristics, including an even material dispersion, structural order, smoothness, strength, heat resistance, and water repellency. Most importantly, it produced films with oxygen barrier ability comparable to commercial high-barrier plastics and completely eliminated bacteria after 72 hours. The optimum number of homogenization cycles was found to be dependent on the desired characteristics and application. Overall, these findings carry a promising potential for a variety of applications, including food packaging and the biomedical field.

## Dedication

To my (الله يرحمها) late, beloved Grandma (Uci): **Ranimah** (1930-2024).

The one who made me love schools. The one who believed in me the most.

You passed right the moment I passed my defense; I knew you were by my side regardless. As promised, I dedicate my last degree to you, Uci.

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## List of Abbreviations

CNF	cellulose nanofibril
TiO <sub>2</sub>	titanium dioxide
TCPP	tetrakis(4-carboxyphenyl)porphyrin
MOF	metal-organic framework
N-film	neat, homogenized-cellulose film
F-film	functionalized-cellulose film
HPLC	high-performance liquid chromatography
T	temperature
RH	relative humidity
FTIR	Fourier transform infrared
ATR	attenuated total reflectance
XRD	x-ray diffraction
CI	crystallinity index
SEM	scanning electron microscope
Sa	arithmetical mean height
UV-vis	ultraviolet-visible
OTR	oxygen transmission rate
OP	oxygen permeability
ASTM	American Society for Testing and Materials
UTS	ultimate tensile strength
TS	tensile strength
E	tensile or Young's modulus
EAB	elongation-at-break
DMA	dynamic mechanical analyzer
E'	storage or elastic modulus
E''	loss or imaginary modulus
Tan(δ)	loss or damping factor
TGA	thermogravimetric analyzer
T <sub>d,5%</sub>	temperature at 5%-weight loss
W	weight
CFU	colony-forming units

## CHAPTER 1

### General Introduction

Product quality/freshness can worsen after many factors, primarily oxygen, light irradiation, and contamination; hence, packaging advancement to tackle this constraint is of profound significance. Lately, photodynamic inactivation has been an emerging topic of an antimicrobial modality, one of which is through metal-organic frameworks (MOFs). MOF combines photosensitizer materials with light of a specific wavelength and molecular oxygen in the environment to generate reactive oxygen species (ROS) with a remarkable ability to kill microorganisms via an oxidative burst with no high heat nor harmful substances required and/or released. Made of metal ions/nodes (as inorganic building blocks) and ligands (as organic linkers), MOF is an ultra-porous sponge-like system with versatile functionalities. Porphyrin-based MOF (where the organic porphyrin acts as the bridging ligand) is among the extensively studied MOF types with feasible structural tunability, stable crystalline structure, open channels, and high ROS yield, mainly due to its heterocyclic structure. Among many popular porphyrin derivatives, tetrakis (4-carboxyphenyl) porphyrin (TCPP) has been gaining interest after its exceptional visible light excitation-induced photodynamic feature and efficient coordination with metal ions (Castro et al. 2019; M. Chen et al. 2020; Min et al. 2021; J. Park et al. 2016; X. Zhao et al. 2023; K. Lu, He, and Lin 2014; Zheng et al. 2018; Ahmadi et al. 2015; Q. Wang et al. 2018; Tian et al. 2020; Jian Zhao et al. 2007).

Titanium (Ti) precursors like titanium dioxide ( $\text{TiO}_2$ ) are among the most studied metal ions to be paired with porphyrin.  $\text{TiO}_2$  is cheap, ubiquitous, and FDA-approved, with proven antibacterial activity against an array of microorganisms under UV. By pairing it with TCPP, photocatalytic activity and ROS production under UV and visible light spectrums can be achieved. Despite their biocompatibility together, the notorious challenge of MOFs on self-aggregation/quenching tendency remains, and incorporating them into a polymeric carrier has been considered to be one of the viable options (El-Wakil et al. 2015; An et al. 2016; Castro et al. 2019; Gao et al. 2019; Roberta Grazia Toro et al. 2020; Wan et al. 2016; M. Chen et al. 2020; Tu et al. 2017; Cherian and Wamser 2000). Cellulose has been evidenced to be one of the ideal carriers for MOF conjugates due to its plentiful hydroxyl groups that have the potential to be employed as a stable and robust anchoring site or matrix through ex-situ loading. Cellulose-based composite also provides large surface areas, mechanical strength, and barrier properties (Adel et al. 2021; Roberta Grazia Toro et al. 2020; M. Chen et al. 2020; G.-Q. Liu et al. 2021; Min et al. 2021; J. Park et al. 2016; X. Zhao et al. 2023; An et al. 2016). Many have discovered the photosensitivity and antimicrobial potentials of one or more combinations of cellulose, TCPP, and  $\text{TiO}_2$  with other materials intended for varying applications from food/biomedical packaging, gas storage/separation, pollutant removal, to energy (G.-Q. Liu et al. 2021; Rathod et al. 2018; Roberta Grazia Toro et al. 2020; Gao et al. 2019; Min et al. 2021; J. Park et al. 2016; El-Wakil et al. 2015); yet, no report on the prospect of cellulose-based packaging incorporated with  $\text{TiO}_2$ -TCPP. This study's objective was to determine the optimum preparation of the cellulose-based matrix with desired functional properties before exploring the prospect of cellulose- $\text{TiO}_2$ -TCPP nanocomposite packaging, especially with its high barrier, UV-blocking, and visible-light responsive antimicrobial features.

## CHAPTER 2

### Literature Review

#### 2.1. Active packaging

The rapidly increasing world population in the past centuries (Ritchie et al. 2023) has led to hikes in global food supplies—an estimated 50-60% between 2010 and 2050 (Guillard et al. 2018; van Dijk et al. 2021). Likewise, the rising food packaging demands, followed by the constantly upgraded/updated and strict safety guidelines, became the pairing constraint to the food security aspect (Kusuma et al. 2023). The urbanization-derived global dietary change and rising reliance on processed foods (World Packaging Organisation 2008) also create urgency in the need for modernized packaging technology to be progressively applied in the industries. Though the vital functions of food packaging are to protect (contain and preserve), convey information, and transport (Ahmed et al. 2022), it plays a crucial role in the global burden of food contamination and foodborne pathogens that are recorded to be more than 600 million foodborne illnesses cases and 420,000 deaths could occur in a year (WHO 2022). Packaging plays an even greater role in keeping freshness/quality and preventing spoilage from microorganisms, water content, oxygen, humidity, temperature, and light (Magoulas 2016), thereby minimizing food waste (Almenar 2023; Vergheze et al. 2015). In the United States alone, 30-40% of the food is wasted, which costs the nation roughly 1.3% of GDP (EPA 2016a; FAO 2013a, 2021a). Together, food and its packaging contribute almost half of the solid waste in American landfills (EPA 2015a). Many waste management hierarchies (European Commission 2023; EPA 2016a; FAO 2022b) stated their highest preference for “prevention” over any other options; hence, the exceptionally growing field of food packaging advancing in food waste prevention through shelf life extension—one of which is through active packaging.

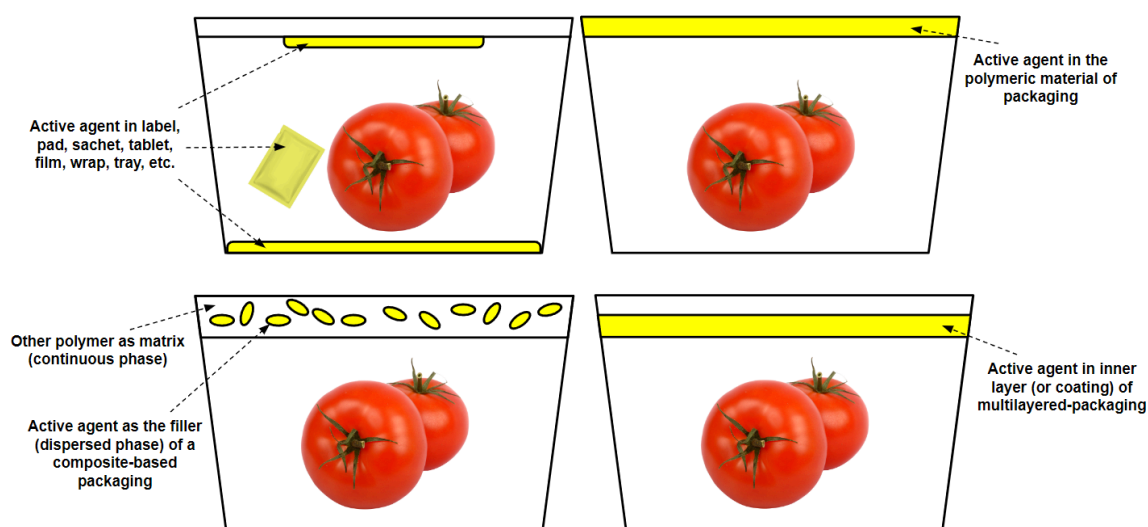
Active packaging is defined as a modified packaging system that incorporates subsidiary constituents (“active agent”) in or on either the packaging material or headspace that intentionally releases spoilage-preventive compounds and/or absorbs spoilage-causing compounds from the product or packaging headspace, aiming to preserve and/or improve shelf-life, quality/freshness, sensory, and food safety (Ahmed et al. 2022; Sand 2020; Janjarasskul and Suppakul 2018; Robertson 2012; Monção et al. 2022). Other possible properties of the added active agents include blocking and buffering (Kuswandi and Jumina 2020; Rehman et al. 2020). Having been developed as early as the 1980s, active packaging can be classified generally into two types based on how it functions in keeping the product quality or freshness (Contreras et al. 2017; Kusuma et al. 2023; Firouz, Mohi-Alden, and Omid 2021; Yildirim et al. 2018; Dainelli et al. 2008; Wyrwa and Barska 2017; Ahmed et al. 2022; F. Charles, Sanchez, and Gontard 2006; D. S. Lee 2016):

- Non-migratory (scavengers/absorbers): It absorbs or reacts with unwanted components (e.g., oxygen, carbon dioxide, ethylene, moisture, light, flavors, odor) that negatively impact the product and removes them from the packaging environment without intentional migration.

- Oxygen accelerates spoilage through fat/lipid and vitamin oxidation, which promotes molds, aerobic bacteria, and yeast growth and causes bad aroma, unpleasant taste/off-flavor, fats/oils rancidity, color changes, oxidative browning, and/or nutrient damage, especially in baked goods, fresh dark meats, and fruits/vegetables.
- Carbon dioxide is the main product of catabolic reactions in the biological systems of most non-pasteurized fermented or roasted foods, causing physiological injury (discoloration, off-flavor, and internal tissue breakdown). Two types of agents are used: physical (zeolite and active carbon powder) and chemical (calcium hydroxide, sodium carbonate, magnesium hydroxide).
- Moisture/humidity/water vapor affects texture, appearance, and microbial activity, especially in high-water-activity products like meat and produce.
- Light-absorbing agents (e.g., hydroxy benzophenone, benzophenone, benzotriazole, triazine, etc.) provide effective light-transmission reduction or stabilization, especially to light-sensitive foods that are particularly susceptible to oxidation and nutritional degradation, such as fruit juices, tea, sports drinks, and extra virgin oils.
- Migratory (emitters/releasers/diffusion): It allows controlled migration of desired substances (e.g., carbon dioxide, ethanol, sulfites, flavors, antioxidants, antimicrobials) that positively impact the product into the packaging environment.
  - Carbon dioxide benefits from its high solubility in food, ability to reduce physiological activities (respiration and ethylene production), and ability to balance pressure/volume between the packaging's inner headspace and external environment. It preserves freshness, reduces microorganisms' growth, and inhibits spoilage-associated odor formation, especially in fresh meat, fish, poultry, and non-climacteric fruits.
  - Ethanol vapor in the packaging headspace maintains the original textural quality and suppresses the growth of yeasts, anaerobic bacteria, and molds, especially in fruits and vegetables.
  - Sulfites (e.g., sulfur dioxide, sodium sulfite, sodium bisulfite, sodium metabisulfite, potassium metabisulfite, potassium sulfite), classified as major allergens, prevent (non)enzymatic browning or coloration during processing and storage by acting against oxidative decay, polyphenol oxidase, fungi and bacteria responsible for malo-lactic fermentation.
  - Flavor addition overcomes oxidation-derived flavor scalping.
  - Antioxidant agent limits oxidation of the fatty components.
  - Antimicrobials prevent microbe's growth by utilizing agents such as essential oils, metals (silver, silver zeolite, gold, copper, titanium dioxide, zinc oxide), triclosan, glucose oxidase, chlorine dioxide, natamycin, and allyl isothiocyanate.

Several mechanisms can be applied to incorporate the aforementioned active agents and obtain an active packaging system (Vidal et al. 2022; Patiño Vidal et al. 2022; Ahmed et al. 2022; Firouz, Mohi-Alden, and Omid 2021; Bahrami et al. 2020). Some of the examples are listed and illustrated below (**Fig. 2.1.**).

- Active agent-loaded external device (e.g., label, pad, sachet, tablet, film, wrap, tray) placed inside the packaging together with the product or attached/glued on the packaging's inner surface that contacts the headspace.
- The active agent is immobilized with a polymer through ion or covalent linkages by utilizing certain functional groups depending on the interactions/bondings.
- The active agent is added either in the form of filler (as the dispersed phase of a composite) or in the form of the matrix itself (as the continuous phase). In the case of multilayered films, the composite is positioned as the packaging's inner layer that contacts the headspace.
- The active agent is sprayed/coated in the packaging's inner surface that contacts the headspace, which would suffice for a compound that is non-compatible or immiscible with the desired polymer, either through immobilization or matrix-filler composite formation route.



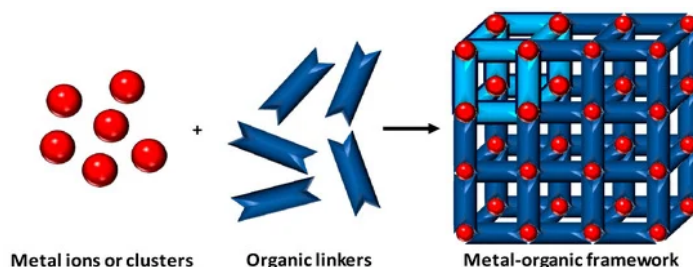
**Fig. 2.1.** Different mechanisms in the active packaging system

The growing interest in active food packaging research was evidenced by the fivefold increase in publications in the past decade, amounting to over ten thousand in 2021 (Ahmed et al. 2022). This prompts the regulatory agencies to set the legal basis for requirements and standards in active packaging production and use, especially in regard to the added substances and their potential impact on the packaged product. European Food Safety Authority acts as the European Union's regulator; meanwhile, the Food and Drug Administration (FDA) is responsible for American markets (Misko 2022).

## 2.2. Metal-organic framework (MOF)

Active agents contained in the highly stable metal-organic framework (MOF) have been one of the emerging topics among active packaging researchers. MOF is a synthetic polymeric hybrid material characterized by ultra-highly porous coordination, high surface area, crystalline structure, low density, tunable morphology, fully exposed active sites, abundant coordination-unsaturated metal sites, and substitutable components (Carrasco 2018; Furukawa et al. 2013; Omar M. Yaghi et al. 2003). The terminology defined by the International Union of Pure and Applied Chemistry (IUPAC) (Batten et al. 2013) is stated

as a further subset of coordination networks (a subset of coordination polymers) that contains potential voids. Similar terminologies exist besides MOF when describing hybrid inorganic and organic framework materials or the nonmolecular/extended solid-state structures containing metal ions and organic spacer ligands. “Coordination polymers” term was the popular term up until the year of 2000 when the trend of “MOF” term use took over ever since (Biradha, Ramanan, and Vittal 2009). As illustrated in **Fig. 2.2**, It is constructed by joining inorganic, metal ions/nodes/clusters (secondary building units (SBUs)) and organic linkers (bridge ligands) by strong bonds to create open crystalline frameworks with permanent porosity through what is termed reticular synthesis. Reticular synthesis is a process of assembling judiciously designed rigid molecular building blocks into predetermined ordered structures (networks), which are held together by strong bonding. Coordination polymerization and cross-linking occur between the inorganic and organic precursors (Furukawa et al. 2013; Omar M. Yaghi et al. 2003). Besides, MOF possesses high compatibility with both organic and aqueous solvents as the mixing media (Burtch, Jasuja, and Walton 2014). Aluminum, iron, zirconium, titanium, copper, and zinc are among the commonly used metal components in fabricating MOFs (Sultana, Kathuria, and Gaikwad 2022).

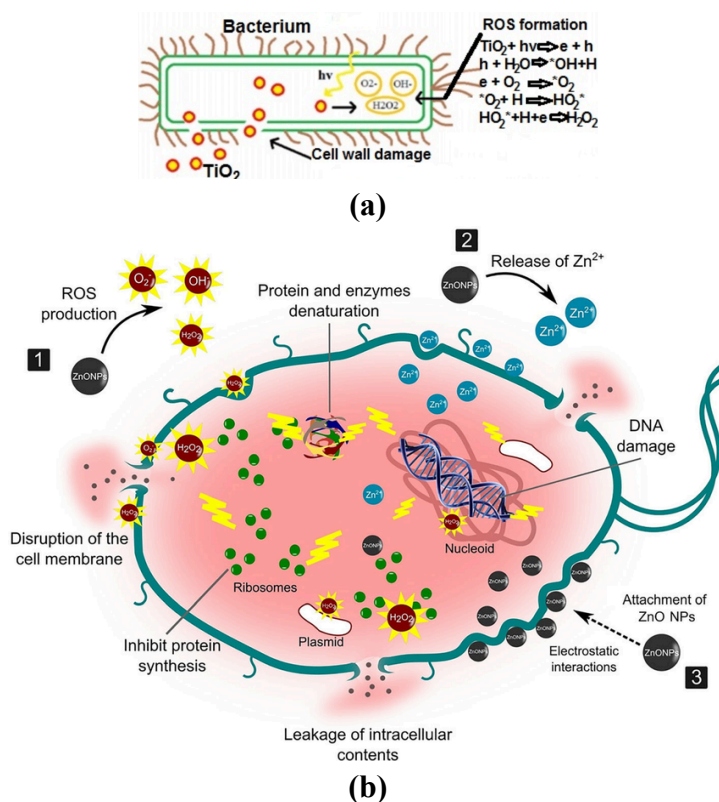


**Fig. 2.2.** The schematic metal-organic frameworks (MOFs) (Carrasco 2018)

There are numerous ways to categorize types of MOFs. For example, based on synthesis stages, first-generation MOFs have the basic structure of organic and inorganic moiety, while the second-generation undergoes surface modification. Lastly, the MOFs that include biomolecules in their structural design are called the third-generation. Based on stimuli, there are single-stimuli and multi-stimuli MOFs (Sultana, Kathuria, and Gaikwad 2022). Since both of the building components of MOF are easily tailorable, where specifically, the metal is easily doped, and the organic linker is easily replaced/modified, this increases the applicability. The abundant surface active groups also can facilitate modifications to enable synergistic or dual effects (Zacher et al. 2009). Since first introduced in 1995 (O. M. Yaghi, Li, and Li 1995), the number of MOF-themed publications has been facing popularity, reaching almost five thousand in 2019 (J. Meng et al. 2020). MOF is applicable in a broad range of fields depending on the properties, e.g., catalysis (Bavykina et al. 2020), gas storage-separation (Jia, Gu, and Li 2022), purification (Jun et al. 2020), drug delivery (He et al. 2021), sensors (Weiwei Cheng et al. 2021), energy (X. Li et al. 2020).

However, lately, MOF-based materials have also become a hotspot for antibacterial research that involves photocatalytic abilities, particularly by employing Reactive Oxygen Species (ROS) as the primary antibacterial mechanism. MOF can be designed to contain a photosensitive component that is able to combine visible light irradiation of a

specific wavelength and molecular/diatomic oxygen ( $O_2$ ) in the air to generate ROS (Castro et al. 2019; I. Kim et al. 2022). More specifically, there is a gap in molecular orbitals of MOF called the Highest Occupied Molecular Orbital—Least Unoccupied Molecular Orbital (HOMO-LUMO) gap where electron transition occurs (from the HOMO to LUMO, leaving holes ( $h^+$ ) in the HOMO) and become conducive and responsible for the formation of varied photogenerated ROS that targets multiple sites in bacteria (R. Li, Chen, and Pan 2021; Vaishampayan and Grohmann 2021). Though bacteria deploy defense mechanisms against oxidative stress caused by ROS, excess accumulation of ROS eventually leads to an oxidative burst that allows ROS first to penetrate through and disrupt the outer cell envelope (cell wall and plasma membrane) and destroy the cell integrity/viability, then damage DNA, lower the coenzyme A (an essential cofactor of cellular metabolism) level, increase plasma membrane permeability, oxidize membrane fatty acids/lipids and proteins, depolarize the membrane, carbonylate protein, and eventually leak the intracellular content—effectively killing the bacteria (Vaishampayan and Grohmann 2021; Bogdan, Zarzyńska, and Pławińska-Czarnak 2015; Sultana, Kathuria, and Gaikwad 2022; Y. Hong et al. 2019). Chelation, or coordination bonds between organic molecules and metals (Sears 2013), also plays a role in MOF-mediated microbial inhibition. It reduces the polarity of the metal component and enhances lipophilicity, which is beneficial in facilitating penetration to bacterial cell membranes (Tweedy 1964). MOF antibacterial function by the ROS formation and valorization using different metal components ( $TiO_2$  and  $ZnO$ ) are illustrated in **Fig. 2.3(a)** and **Fig. 2.3(b)**, respectively.



**Fig. 2.3.** The antibacterial mechanisms by reactive oxygen species (ROS) using different metal components: (a)  $TiO_2$  (Maryani et al. 2020) and (b)  $ZnO$ : (Gomaa 2022)

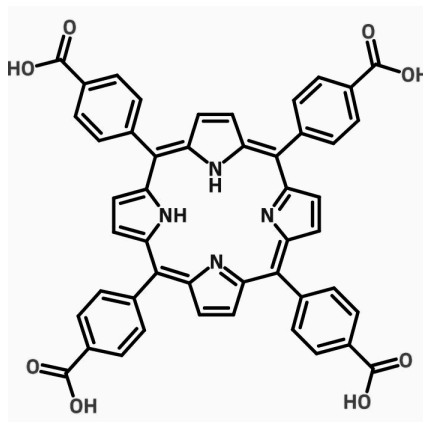
Bacteria physiology shall be taken into consideration when assessing the bactericidal efficiency of ROS since the bacterial structure (complexity and thickness) and chemical composition govern susceptibilities to oxidative damage. For example, the Gram-positive bacteria cell wall is made of a thick and porous peptidoglycan layer with attached teichoic acids, while the Gram-negative bacteria have a thin peptidoglycan layer with surrounding, asymmetric, highly-impermeable bilayer, as well as inner and outlet leaflets of phospholipids and strongly-negatively charged lipopolysaccharides, respectively. Hence, the permeation limitation of neutral/negatively charged photosensitizers (in MOF) against Gram-negative bacterial cells, in contrast to the positively charged ones, which are capable of advancing a strong electrostatic interaction and achieving a considerable bactericidal inhibition performance (Castro et al. 2019; Delcour 2009; Bogdan, Zarzyńska, and Pławińska-Czarnak 2015). In addition, peptidoglycan content in the targeted bacteria cell wall is another factor affecting bacteria resistance to oxidative damage and ROS activity. Though peptidoglycan degradation does not necessarily occur during bacteria inactivation, neither with Gram-negative nor with Gram-positive bacteria, peptidoglycan provides an efficient physical barrier from ROS gaining entry into the bacteria cell (Bogdan, Zarzyńska, and Pławińska-Czarnak 2015; Dalrymple et al. 2010). ROS formation is initiated by electron transfer reactions from the added photosensitizer's triplet state, which are followed by two (2) possible oxidative mechanisms or pathways: Type I, to surrounding substrates, generating radical ions or cytotoxic oxygen species (i.e., superoxide anion, hydroxyl, and lipid-derived radicals); and/or Type II, to ground state molecular oxygen ( $^3\text{O}_2$ ), generating excited-state singlet oxygen ( $^1\text{O}_2$ ) (Castro et al. 2019; Vaishampayan and Grohmann 2021)—to be employed in bacteria inactivation. Overall, the order of ROS ability in various infectious agent inactivation is reported as follows (from the most susceptible to ROS-derived oxidative damages to the least): viruses > prions > Gram-negative bacteria > Gram-positive bacteria > yeasts > molds (Bogdan, Zarzyńska, and Pławińska-Czarnak 2015).

### 2.3. Titanium dioxide ( $\text{TiO}_2$ ) in MOF

Titanium (Ti)-clusters have been among the most appealing choices as a multifunctional metal component of metal-organic frameworks (MOFs) due to their excellent optical response (photoactivity), (photo)chemical stability, varying structure, redox activity (transition between  $\text{Ti}^{3+}$  and  $\text{Ti}^{4+}$ ), and biocompatibility (D. Pinto et al. 2015; Schneider et al. 2014; J. Zhu et al. 2018). The first recorded success of exceptionally porous and crystalline Ti-based MOF fabrication was by a photochromic study in 2019 (Dan-Hardi et al. 2009). As the naturally occurring oxide of Ti that can be prepared in various degrees of purity,  $\text{TiO}_2$  continuously gains attention for its many advantageous properties, such as spectral, structural, mechanical, optical, dielectric/electronic, catalytic, and anti-corrosion. In addition, nano-engineering modification of the pairing component in MOFs—organic ligands—allows researchers to accomplish tunable optical band gap of  $\text{TiO}_2$  with added functional moieties (Fang et al. 2017; Fang, Zhang, and Zhang 2016; J.-X. Liu et al. 2016; Santiago Portillo et al. 2017; D. Pinto et al. 2015). Its photocatalytic applicabilities extend from being used in fields of UV protection (sunscreens, paints, ointments, toothpaste, pigments, plastics, photo-/electrochromics, thermal sprays), energy and electricity (dye-sensitized solar cells or photovoltaics, photo-conductor, lithium batteries, piezoelectric capacitors, fuel cell),

food/biomedical (antimicrobial, sensors, nanomembrane, gene therapy), and environmental (water splitting, pollutant degradation, gas storage/separation, including H<sub>2</sub> generation and CO<sub>2</sub> reduction) (D. Pinto et al. 2015; Assi et al. 2017; D. Sun and Li 2017; Schneider et al. 2014; J. Zhu et al. 2018). Specifically in food packaging applications, TiO<sub>2</sub> is proven to possess exceptional antimicrobial and UV rays-blocking performances (Varghese et al. 2023) due to its promising light absorbance (at wavelengths of 275-405 nm) and high refractive index; moreover, for nanosized TiO<sub>2</sub> (Ghamarpour, Fallah, and Jamshidi 2023; Khorasaninejad et al. 2017). When selecting the right metal component, it is no less crucial to correlate between investigations of their bacteria photocatalytic inactivation with the reactive oxygen species (ROS) generation under both light and dark conditions. TiO<sub>2</sub> was found to perform better than its biggest metal ion contender—zinc oxide (ZnO)—in regards to ROS production and loss of cell viability assessed on representative bacteria to both Gram-positive (*S. aureus* and *B. subtilis*) and Gram-negative (*E. coli*). The same study also evidenced the more prevalent self-agglomeration issue in ZnO specimens (Barnes et al. 2013). An earlier, similar study confirmed the notion as well (Qamar and Muneer 2009). Despite their nearly identical band gap energy, ZnO suffers from worsened photosensitivity in aqueous media (e.g., water) due to photo-corrosion tendency under UV illumination (Hariharan 2006; van Dijken et al. 1998; Neppolian et al. 1999).

#### 2.4. Tetrakis(4-carboxyphenyl)porphyrin (TCPP) in MOF

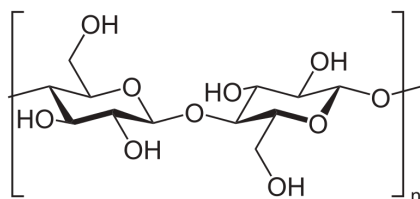


**Fig. 2.4.** Chemical structure of tetrakis(4-carboxyphenyl)porphyrin (TCPP)

Porphyrins are among the classes of organic chromophores and N-heterocycles, consisting of macromolecular compounds of porphin (C<sub>20</sub>H<sub>14</sub>N<sub>4</sub>), whose meso/ $\beta$ -position can be substituted by varieties of functional groups (Gottfried 2015). In fact, porphyrin naturally exists in many vital functions of living organisms, such as hemoglobin (blood), chlorophylls, catalase, heme, cytochromes, and vitamin B12 (J. Chen, Zhu, and Kaskel 2021; H. Huang et al. 2015; Jasat and Dolphin 1997). The free-base porphyrin (center or periphery) can be bound with metal ions to produce metalloporphyrins (metal complexes) with various tunable substitution patterns, molecular cages, and framework solids due to its intricate binding modes. Other reasons behind the metalized porphyrin derivatives versatility are their large  $\pi$ -aromatic system, chemical-thermal stability, energy transfer, fluorescence characteristic, chemical catalytic activity, photophysical, and

electrochemical properties (Adler et al. 1970; Longevial et al. 2018; Durot, Taesch, and Heitz 2014). Porphyrins (derivatives) famously possess electromagnetic spectrum absorption in the visible light section, which is the strongest around 400–450 nm (Soret band) with gradually reduced intensity at 500-700 nm (Q-bands) (Gottfried 2015). Their applications include light harvesting, solar cells, molecular electronics, biomedical (photodynamic therapy, tumor treatment), and (photo-/electro-/or biomimetic) catalysts (Steer 2018; Rezaeifard and Jafarpour 2014). However, the drawbacks limit their further development, one of which is the self-quenching/aggregation tendencies. The growing solution nowadays is through proposing multifunctional carriers (including incorporation into metal-organic frameworks or MOFs) to encapsulate, physically adsorb, or covalently bind porphyrins (derivatives), such as micelles, liposomes, carbon nanotubes, inorganic nanoparticles, and polymer nanoparticles (J. Chen, Zhu, and Kaskel 2021). This advanced concept also leads to improved physicochemical behaviors, maximum highly active and functional building units, and minimum inactive constituents (Freund et al. 2018). Particularly in the past two decades (A. Li et al. 2020), porphyrin and derivatives as neutral organic linkers in MOFs have garnered popularity among researchers. Owing to their unique macro-heterocyclic geometry (**Fig. 2.4**), the advantages include exposed metal sites, long-lived excited states, and high triplet quantum yield, which subsequently leads to their pronounced entire-spectrum visible-light responsive photocatalytic and redox-active photosensitizer abilities (C. Liu et al. 2020; J. Liu et al. 2018; X. Zhang et al. 2021; C. Y. Lee et al. 2011). Another promising finding was also obtained on photo-oxidation involving ROS (singlet oxygen) formation by porphyrin-based MOFs (Xie et al. 2011; J. A. Johnson et al. 2014; Z. W. Jiang et al. 2020), which is described more in detail in **Fig. 2.3**. in regards to ROS generation and antibacterial mechanism in MOFs. The carboxyl group-substituted 5,10,15,20-tetrakis (4-carboxyphenyl) porphyrin (TCPP), or  $C_{48}H_{30}N_4O_8$ , is a derivative and a member of the porphyrin family with remarkable prospects, especially after their visible light excitation-induced photosensitization (Y. Wang et al. 2018; Ahmadi et al. 2015). TCPP is especially compatible with  $TiO_2$  as it adsorbs strongly onto  $TiO_2$  nanoparticles as photosensitizer (Cherian and Wamser 2000; Kalyanasundaram et al. 1987; Dabestani et al. 1988; Vlachopoulos et al. 1987). Other advantageous characteristics of TCPP over other porphyrin-based photosensitizers are its superior simplicity, efficient coordination with the metal ions, reduced self-quenching behavior, and preserved photosensitivity even under a high dose (Tian et al. 2020; Jian Zhao et al. 2007).

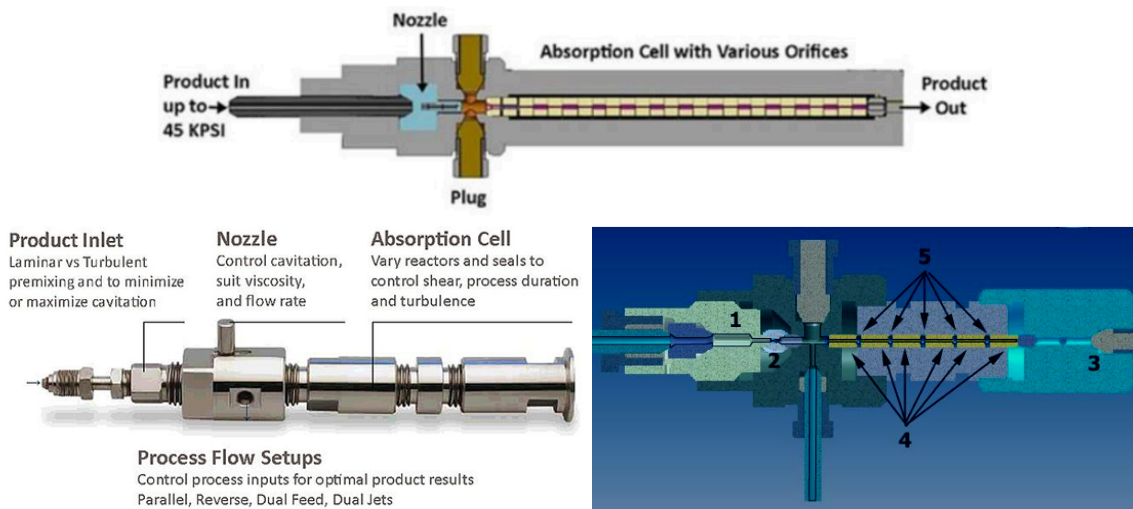
## 2.5. Cellulose-MOF composite



**Fig. 2.5.** Chemical structure of cellulose

Cellulose is named the Earth's most abundant and naturally biodegradable polymer (Klemm et al. 1998) and can be extracted from a myriad of sources (e.g., wood, plant,

algae, tunicate, bacteria) naturally or through controlled synthesis. Cellulose is made of D-glucose monomers linked via  $\beta(1\rightarrow4)$  glycosidic bonds (**Fig. 2.5**), which creates the chemical formula of  $(C_6H_{10}O_5)_n$  (where  $n$  is the degree of polymerization ranging from 10,000-15,000 depending on the source). Cellulose is synthesized biologically in nature by the joining of glucose molecules with the loss of 1  $H_2O$  molecule for each bond formed (Werpy and Petersen 2004; Manahan 2011). Its multifaceted applicability is mainly due to its ease of modification and high biocompatibility (Abdelhamid and Mathew 2022); moreover, it is able to be incorporated with organic and/or inorganic conjugates (Erlantz Lizundia et al. 2020). Nanocellulose is a ‘top-down’ nanoscaled product of cellulose that is commonly preferred for the further valorization of cellulose materials due to its increased surface area and extensive network structure (H. Lee, Sundaram, and Mani 2017). There are two types: 1) the rigid and rod-like cellulose nanocrystals or CNCs, which are prepared chemically, e.g., acid hydrolysis, oxidation, and cationization, by removing amorphous region and leaving purely-crystallines; and 2) the flexible cellulose nanofibrils or CNFs, which are prepared mechanically, e.g., micro-fluidization, homogenization, grinding process, sonication, by preserving both amorphous and crystalline structure (Dias et al. 2020; Ibrahim et al. 2019; Sofla et al. 2016). However, despite its nano-size, neat cellulose still suffers from various issues such as natural insolubility, non-dispersibility, moisture absorption, and hydrophilic/hygroscopicity, which all lead to weakened interfibrillar hydrogen bonding and many other functional properties (Nechyporchuk, Belgacem, and Bras 2016; Xia et al. 2018; B. Yang et al. 2019); therefore, modification is often necessary. Mechanical modification—especially through homogenization employing pressure, velocity, cavitation, shear, and impact forces for solution passed through a very small vessel/orifice—is among the favored solutions. It is owing to its simplicity and environmental-friendliness for having no requirement of high temperatures and mostly requiring sole water as mixing media (L. Wang et al. 2021; Deepa et al. 2019; W. Huang 2018). Homogenization for years has proven its success in promoting fibrillation, surface area, hydrogen bonding, 3D network strength, and structural stability (Nagarajan et al. 2021; H. Lee, Sundaram, and Mani 2017; Chun et al. 2011; Wenjian Cheng et al. 2010; Flores et al. 2021; Usenko et al. 2013). However, the recurring issues with the traditional homogenizer approach are the use of high conditioning pressure, meaning high energy consumption, cost, and risk of over-shearing/over-heating/over-pressure (which leads to material destruction and/or agglomeration), on top of clogging (H. Lee, Sundaram, and Mani 2017; Nagarajan et al. 2021; Boulemkahel et al. 2021). The use of lower pressure and/or multiple passes/cycles of homogenization has been explored as an effort to resolve these limitations (Boulemkahel et al. 2021; Fu et al. 2011; Yu et al. 2021). Moreover, several reports highlighted the potential of a homogenizer equipped with novel “Emulsifying Cell” technology (BEE International 2015) at multiple passes in preparation of a variety of materials, including cellulose. As illustrated in **Fig. 2.6**, such cell design allows the multiple passes effect, thus optimizing the number of cycles and operating time the solution has to pass through the cell. Varieties of nozzle diameter are available as well to customize the overall impacts.



**Fig. 2.6.** Schematic general principle of homogenizer equipped with emulsifying cell technology (BEE International 2015)

Cellulose, just like many other polymers, has been proven to be a noteworthy pairing material (either carrier, support, modulator, binder, or filler) of metal-organic frameworks (MOF) to solve its notorious constraint of low stability (X.-F. Zhang et al. 2021; Xiongli Liu et al. 2021; X.-M. Liu, Xie, and Wu 2020; S. Wang et al. 2020). Cellulose contains advantageous functional (carboxyl/hydroxyl) groups that can interact with the positive charge of MOF's metal nodes. It is also reported that the fruitful use of photosensitizer dye in maximizing the final product's adsorption capacity (C. Zhu, Monti, and Mathew 2020). Non-cytotoxicity yet possessing excellent antibacterial efficacy (with or without stimuli such as light) is another appealing factor of cellulose being commonly used in antibacterial product developments. Many developed cellulose-MOFs offered remarkable porosity (pore volume), mechanical properties (e.g., compression strength, flexibility, shapeability), thermal stability, and low density (Y. Wu et al. 2020; L. Zhu et al. 2018; Lei et al. 2019). Moreover, it is also possible to combine characteristics of both cellulose and MOFs in order to acquire the final product with multifaceted functions, e.g., UV-blocking, hydrophobicity, photoluminescence, and biodegradability, including lowered cytotoxicity derived from the neat-MOF (L. Lu et al. 2018; Ozer and Hinestroza 2015; D. Huang et al. 2020). The advantageous functional groups of cellulose also play a crucial role in preventing the leaching of metal nodes of the MOF conjugate out of the system. In addition, MOF is beneficial as well in maintaining nano-cellulosic structure and preventing aggregation by inhibiting the interaction between cellulose surface and the abundant hydroxyl groups, especially at high-content cellulose (D. Huang et al. 2020). There are two (2) distinct mechanisms to prepare cellulose-MOF:

- **In-situ synthesis:** MOF precursors (either inorganic metals and/or organic linkers) are added to the cellulose to allow MOF to 'in-situ' grow into the cellulose fibers. Fiber modification (e.g., esterification (Abdelhameed, Rehan, and Emam 2018)) is often necessary to ensure high metal-fiber interaction. The conformational structure of cellulose is possibly altered post-MOF incorporation (Valencia et al. 2020).

- Ex-situ synthesis: Multi-step procedures of loading/doping the separately and initially synthesized MOFs into cellulose (which plays a role as carrier/support). It allows the more precise loading control of MOF conjugate. The most notorious challenge is MOFs' severe aggregation and uneven dispersion within the cellulose network (Tan et al. 2020).

**Table 2** below lists the examples of developed cellulose-MOF in the past decade specifically intended for high-performance antibacterial application use against one of the main pathogens *Escherichia coli* or *E. coli* (da Silva Pinto, Sierra-Avila, and Hinestroza 2012; Abdelhamid and Mathew 2022). Detailed description on antimicrobial test and *E. coli* are at the next sections.

**Table 2.** Application of cellulose-MOF as antibacterial films against *E. coli*

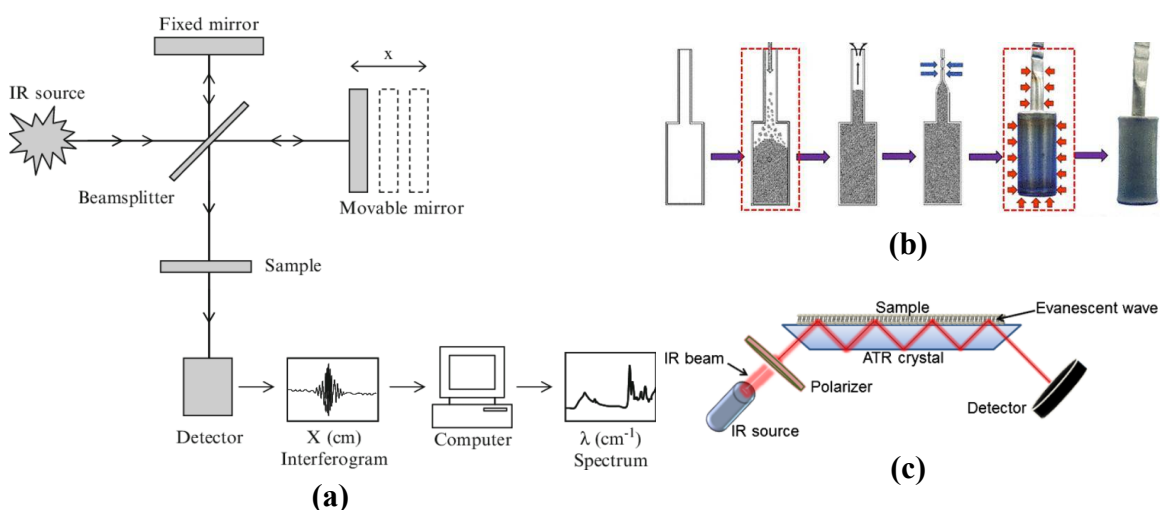
Reference	Building components	Efficiency	Contact duration
(Y. Yang et al. 2020)	cellulosic cotton fabric; zeolitic imidazolate framework (ZIF-8) as MOF	100%	18 h
(Qian et al. 2018)	cellulose paper; cellulose nanofiber; zeolitic imidazolate framework-67 (ZIF-67) as MOF	100%	24 h
(Rodríguez et al. 2014)	cellulosic woven cotton fabrics; Cu-BTC or copper benzene-1,3,5-tricarboxylate (MOF-199 or HKUST-1)	100%	24 h
(Rubin, Neufeld, and Reynolds 2018)	carboxymethylated cellulosic cotton fiber; $\text{Cu}_3(\text{NH}_2\text{BTC})_2$ as MOF	99%	24 h
(Nie et al. 2020)	cellulosic woven cotton fabrics; porphyrinic MOF (PCN-224)	99%	24 h
(L. Lu et al. 2018)	cellulosic cotton fabric; ZnBDC as MOF (zinc(II)acetate+1,4-benzenedicarboxylic acid)	99%	24 h
(Duan et al. 2019)	oxidized cellulose-rich corn cob; copper/zinc-based MOF	90%	24 h
(Noorian, Hemmatinejad, and Navarro 2019)	cellulosic cotton fabric; zinc glutamate BioMOF; nitric oxide; 5-fluorouracil (5FU)	90%	24 h

## 2.6. Characterization

### 2.6.1. Fourier transform infrared (FTIR) analysis

Fourier transform infrared (FTIR) spectroscopy, as illustrated in **Fig. 2.7(a)**, is a form of vibrational spectroscopy that employs infrared radiation and is often used as a physicochemical quantitative analytical method to 'fingerprint' substances and determine their identity, quality, impurities, and changes based on the molecular composition and bonding mechanisms. A specific wavelength of infrared is produced by the instrument to be absorbed/transmitted through the sample to the detector that scans the 'fingerprints.' The functional groups existing in the specimen's composition are identified by detecting their vibrations (either through stretching or bending in various ways) and their intensity (either in %transmission or absorbance unit), which are plotted against the light

frequency ( $\text{cm}^{-1}$ ) to produce an FTIR spectrum. Molecular vibration types can be related directly to molecules' symmetry, which is helpful in precisely identifying how a molecule is bonding on surfaces or as a component in a solid phase from its infrared spectrum. The bands' position and intensity are related to changes in surface groups (Hamad et al., 2018; Wade, 2003). The resulting spectra are characteristic of the bonds in molecular species (or extended bonded structures such as in solid lattices) and are further identified by referring to the spectral library. The advantages include the wide spectral and wavelength ranges ( $400\text{--}4000\text{ cm}^{-1}$ ), no required derivatization, and the fact that it is non-destructive yet still considerably a rapid and low-cost technique. FTIR is among the vital characterizations when analyzing materials as it addresses one's chemical composition (Peak 2005; Daéid 2005; Tranter 2017; Bhunia 2008; Ercolini 2014; Dwivedi et al. 2017; Pednekar et al. 2017).

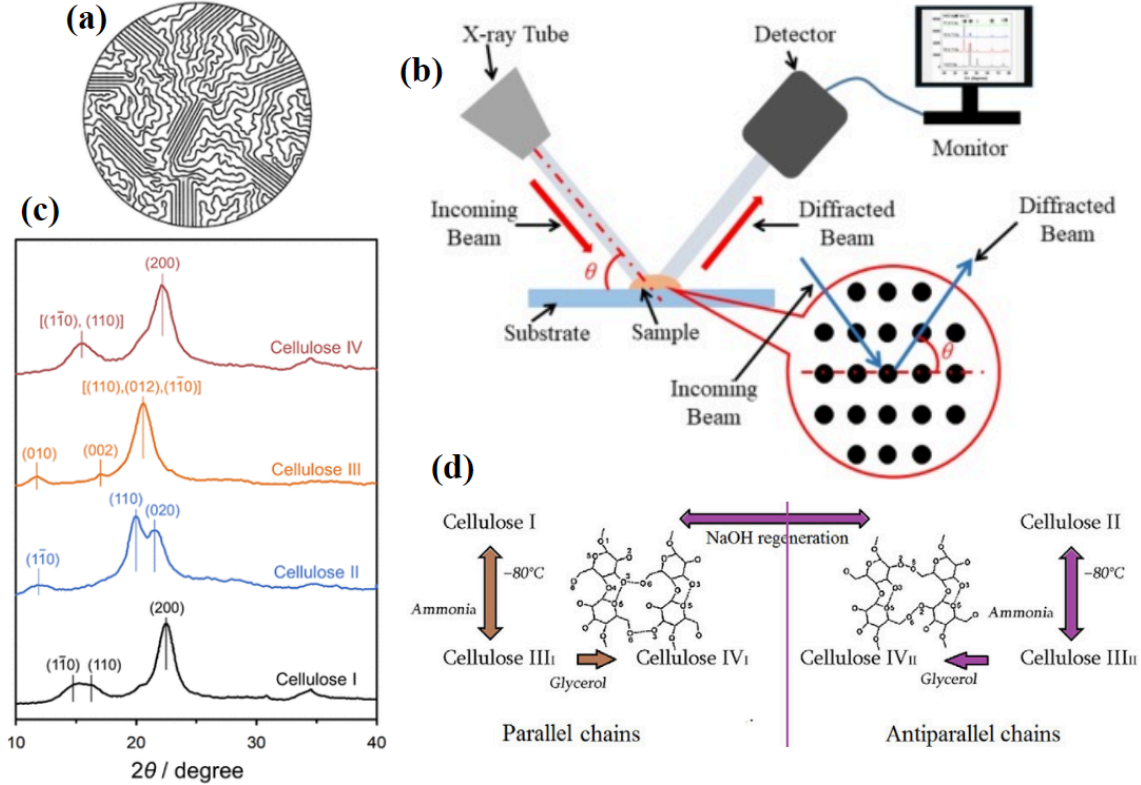


**Fig. 2.7.** Schematic general principles: (a) basic FTIR (Ojeda and Dittrich 2012); (b) sample pellet preparation (Kintek 2024); (c) ATR-FTIR (Ausili, Sánchez, and Gómez-Fernández 2015)

The two most common FTIR approaches are 1) the pellet (**Fig. 2.7(b)**), which is placed in the path of the infrared beam, and 2) Attenuated Total Reflectance (ATR) (**Fig. 2.7(c)**). Potassium bromide (KBr) is among the most universal salts/crystals to be used as specimen carriers when preparing FTIR pellets due to being simple and optically transparent or having a 100%-transmittance or no absorption bands in the range of FTIR tested wavenumber ( $4000\text{--}400\text{ cm}^{-1}$ ) as well as good reproducibility (Goriletsky et al. 2001; Gendreau and Burton 1979). On the other hand, in the ATR approach, the sample is placed on top of the ATR crystal lens (fully covered), where the infrared beam passes through, and the resulting evanescent wave penetrates and gets absorbed into the specimen. The polarizer generates the parallel-perpendicular polarized incident beams (Ausili, Sánchez, and Gómez-Fernández 2015). Optical contact between the specimen and the ATR crystal lens shall be maintained using stable and constant pressure in order to generate data with decent reproducibility. The ATR-FTIR approach is sometimes preferable as it does not require the traditional KBr pellet preparation, enhancing its simplicity (especially beneficial for sample types unable to be incorporated evenly in

pellets) and the ability for real-time decision-making (Keshavarzi et al. 2020; Sauer et al. 2019; Tiernan, Byrne, and Kazarian 2020).

### 2.6.2. Crystallinity



**Fig. 2.8.** Schematic: (a) illustration of polymers' crystalline-amorphous regions (Uzun 2023); (b) general principle of XRD (C. Zhao et al. 2023); (c) and (d) cellulose's crystalline allomorphs (Y. Liu et al. 2022; Nasir et al. 2017)

Polymers structurally comprise crystalline (ordered/aligned) and amorphous (less ordered/aligned) regions (Nisizawa 1973), as illustrated in **Fig. 2.8(a)**. Crystallinity represents the mass fraction of the crystalline regions and has an important effect on many physical and chemical properties of polymers, especially in governing one's hardness and density, by defining the material's crystal structure and microstructure. The degree of crystallinity, often called the Crystallinity Index (CI), can be determined by X-ray diffraction (XRD) (**Fig. 2.8(b)**). X-rays are high-energy electromagnetic waves at a wavelength ranging from  $10^{-3}$ - $10^1$  nm (Spiess et al. 2009). The crystalline region in the tested specimen scatters the incoming X-ray beam and produces sharp reflections, leading to beam diffraction at an angle ( $2\theta$ ) that obeys Bragg's Law (Bragg 1913). Bragg's Law accounts for the relationship between the electromagnetic radiation wavelength and the specimen-diffraction angle's lattice spacing. On the other hand, the amorphous region generates a diffuse background in the resulting XRD pattern. XRD spectra plot the diffraction signal intensity against either the produced diffraction angle ( $2\theta$ ) or spacing between the atomic planes or  $d$  (nm). The most common wavelength used

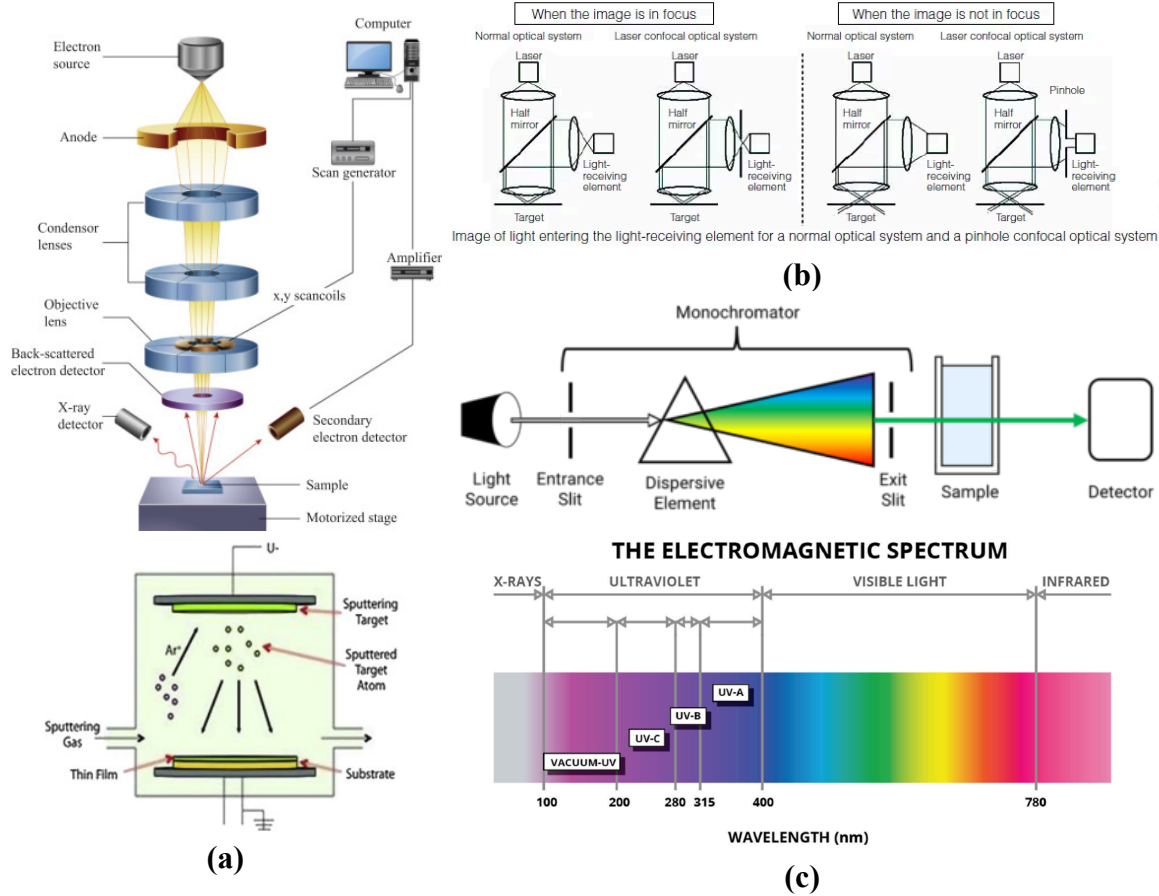
is 1.54 Å (Cu K $\alpha$ ) (Uzun 2023; C. Zhao et al. 2023; Sharma et al. 2012; H. Khan et al. 2020; Evans and Radosavljevic Evans 2004; Epp 2016). XRD is often considered among the most relevant characterizations when analyzing materials as the determined crystallinity implies mass fraction or microstructure of one's crystalline regions, which governs the ability to form a uniform network and subsequent physical-mechanical properties (Siti Syazwani et al. 2022; I. S. Goldstein 2004). Crystallinity signifies an active and stabilized surface charge of a suspension and a more homogenized mixture, which correlates with the ability to form a uniform network (Hafid et al. 2023). It is also an inherent property that governs the mechanical property, affinity for water, and accessibility to chemical reagents. The amorphous region, with its higher porosity than crystallines, allows water to penetrate through and increases reactivity to either chemical (acids and alkalines) or enzymatic hydrolysis (Nunes 2017; Pérez and Samain 2010).

Several methods, instruments, and equations can be employed in characterizing crystallinity; however, the Segal et al. method (Segal et al. 1959) is named the most popular due to its superior ease of use and time efficiency (S. Park et al. 2010). Also known as the "XRD peak height" method, it utilizes the produced XRD patterns to calculate CI using Segal's empirical equation, or the intensity ratio between the specimen's crystalline and amorphous peaks. The crystalline peak (major peak, or  $I_{total}$ ) is located around  $2\theta = 23^\circ$ , which measures the maximum intensity of the crystalline peak. Meanwhile, the amorphous peak is also known as the minima between the major (indices (200) at  $2\theta = 22.5^\circ$ ) and secondary peaks (indices (10 $\bar{1}$ ) at  $2\theta = 14.7^\circ$ – $16.1^\circ$ ), or  $I_{am}$ , located around  $2\theta = 18^\circ$ , which is the minimum intensity of the amorphous peak or the maximum point where the entirely amorphous cellulose would be (Segal et al. 1959; Nurhadi et al. 2022). In these equations,  $n$  is an integer (1, 2, 3, 4, ...);  $d_{hkl}$  is interplanar spacing generating diffraction between atomic planes;  $\theta$  is X-ray incident angle; CI is crystallinity index;  $I_{total}$  is the crystalline (major) peak, at  $2\theta = 23^\circ$ ; and  $I_{am}$  is the amorphous (minor) peak, at  $2\theta = 18^\circ$ .

$$n \lambda = 2 d \sin\theta \quad ; \quad CI = \left(1 - \frac{I_{am}}{I_{total}}\right) \times 100\%$$

Furthermore, another crystallinity profile of cellulosic specimens identifiable through XRD spectra is crystalline allomorphs. Among the four (4) crystalline allomorphs introduced, Cellulose I is the naturally obtained or parent/native form, thus being the most abundant. Cellulose I is cellulose chains assembly with interchain hydrogen-bonding networks to form van der Waals-bonded, densely-stack sheets. Cellulose I can be converted to Cellulose II through either mercerization (alkali treatment) or regeneration (solubilization-recrystallization). By treating Cellulose I and II with liquid ammonia, Celluloses III (either III<sub>I</sub> or III<sub>II</sub>, depending on the cellulose source) and IV can be further obtained, respectively. Cellulose III can also be transformed to cellulose IV (either IV<sub>I</sub> or IV<sub>II</sub>) using thermal treatment (Hayashi et al. 1975; Nishiyama et al. 2003; Nishiyama, Langan, and Chanzy 2002; Isogai et al. 1989; Z. Wu et al. 2018; Kulshreshtha 1979). **Fig. 2.8(c)** and **Fig. 2.8(d)** demonstrated the comparison between four different crystalline allomorphs of cellulose as well as how to identify each type by analyzing their characteristic peaks and patterns in the resulting XRD spectra.

### 2.6.3. Morphology, surface roughness, and transparency



**Fig. 2.9.** Schematic general principles: (a) SEM and ion sputter coater (Inkson 2016; Kafle 2020); (b) roughness (Keyence 2021), and (c) UV-vis spectroscopy and ranges of different electromagnetic spectrums (Houck and Siegel 2015; Forte 2014)

Morphology, topography (surface roughness), and optical behavior (including optical transparency) are among the crucial characterizations whose impacts often correlate when analyzing a material's functional properties and applicability. Morphology describes the structural conformation and molecular arrangement, extending from the nanoscale (nanoparticles and crystalline lamella) to the microscale (homogeneity and domains) to the macroscale (surface and mechanical properties). The defining factors include molecular weight, crystallinity, branching, and crosslinking. Morphological findings are of great importance to the product's final properties, such as mechanical, chemical, electrical, thermal, adhesiveness, and wettability (Khalifeh 2020; Popelka, Zavahir, and Habib 2020; Araki, Shibayama, and Tran-Cong 1998; Gnanasekaran et al. 2016) Morphological analysis can be conducted either directly or indirectly. The material's surface and internal structures can be directly observed by employing a variety of microscopic techniques depending on measurable sizes, attainable resolutions, and magnifications, which can be combined when necessary (Maxfield 1994). Electron microscopes are among the most popular advanced imaging tools, even compared to the most common choice—light microscopes. Despite its low cost and non-destructiveness,

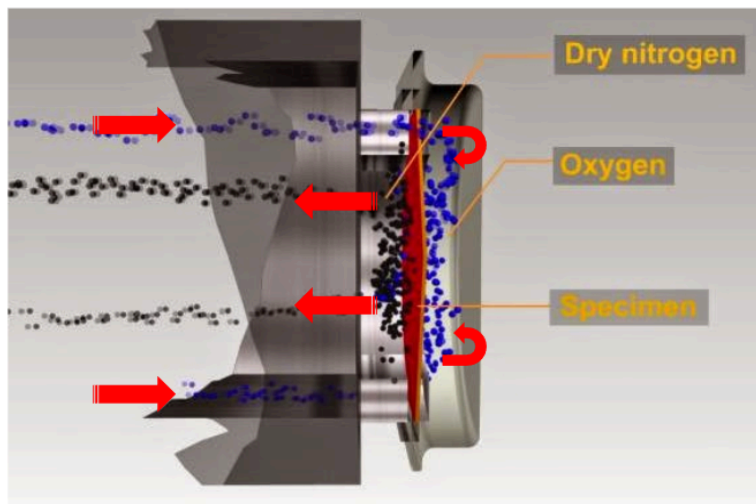
the light microscope has limited imaging resolution at visible light radiation wavelength (400–700 nm), where radiation with shorter wavelengths would be more susceptible to interact with nanomaterials, producing higher resolution (J. C. H. Spence 2013). Factors that can affect wavelengths include voltage and electrons' momentum and acceleration (Vinet and Zhedanov 2011; L. Reimer 2000). The use of electrons is also beneficial after their negative charge, hence the extremely strong interaction with atoms and subsequently produced emission signals (even for specimens sized under 15 nm). Another advantage offered by electron microscopy is the ultra-high magnification ranges. Scanning Electron Microscopy (SEM) is one of two generic types of electron microscopes, which is primarily designed for material surface testing, unlike Transmission Electron Microscopy (TEM), which is preferable when observing the internal structure (Inkson 2016; Popelka, Zavahir, and Habib 2020). As demonstrated in **Fig. 2.9(a)**, SEM scans a few micrometers of the surface where the produced images are based on the reflected electron beams that are ejected out of a probe. Since the electron beam charging leads to high-voltage burning, specimens must be non-conductive; therefore, low accelerating voltage (1–4 kV) is required at times (Egerton 2005; J. I. Goldstein et al. 2003). In addition, during specimen preparation, ion sputter coating with an electrically conductive material (e.g., gold, platinum, carbon, palladium, or combination) is also among the common solutions, usually by utilizing argon as the sputtering gas of choice due to its high atomic weight (meaning higher kinetic energy), inert chemistry, and non-destructiveness, unlike oxygen/air that possess risks of surface oxidation (Kleiner, Matthes, and Rößler 2021; Hatano et al. 2022; West and Kelly 2011). The specimen is kept in a vacuum chamber, with the probe on the top that emits electrons (accelerated based on voltage difference ranging from 0.1–50 keV) focused on a small spot to be scanned sequentially across the surface. Operating vacuum pressure may range from  $0.1\text{--}10^{-4}$  Pa, to be maintained low to eliminate electrons-air interaction (J. I. Goldstein et al. 2017; B. Reimer 2006; Popelka, Zavahir, and Habib 2020). Imaging can be conducted on either the surface or (fractured or microtomed) cross-section of the sample (Mohan et al. 2016).

Sometimes referred to as surface morphology in 3D (topography), surface texture and roughness quantify surface irregularities (Mirabal et al. 2023). Roughness property is of a great importance as it affects structural integrity & interaction with environment e.g., wettability, wear behavior, stresses (G. Song, Wang, and Tan 2022; Antonov, Islamova, and Strizhak 2023). Surface roughness can be analyzed by observing 3D microscopic and Z-stack or focus-stack images. The latter is produced by combining multiple images taken at different focal distances to generate a composite image with a greater field depth (e.g., the plane of focus' thickness) than the individual source images (Ray 2002; D. Johnson 2008). In-focus images are accomplished through filtering by referring the reference plane to the films' real surface to reduce height variability. This step would solve the notorious issue of lowered depth-of-field for numerical and high-magnifying apertures (Larson and Banks 2022). As illustrated in **Fig. 2.9(b)**, digital microscopes can be employed to observe topography/roughness, which is sometimes classified as surface morphology in 3D. The instrument is equipped with a camera and wide-range zoom lens connected to the free-angle observation stand where the Z-axis lens can be moved around (followed by the changing focus level), and then the surface irregularities height can be measured. Topographically, the highest points of an object come into focus first, while

the lowest points come into focus last. The height positions of the individual focal planes are simultaneously recorded to generate the final 3D images that can be used to analyze the specimen's surface roughness/waviness. This surface micro-geometry and irregularity characteristic is expressed by the arithmetic mean deviation of the height of a line, also expressed as Ra (Woch et al. 2022). Furthermore, the extension of Ra to a surface, Sa, is the difference in height compared to the surface's arithmetical mean. Simply put, Ra is the average of the surface heights along a measurement trace, while the Sa is the average roughness over a measurement area (Adamčík et al. 2023; Keyence 2021; Mike 2022).

Lastly, transparency or optical polarization functionality is among the optical properties heavily associated with morphology findings when analyzing a material. A matter's color is governed by its light-emitting ability (Owen 1996). A spectrophotometer (**Fig. 2.9(c)**) can be used for this testing by employing an artificial UV light source to determine the percentage transmission/absorbance of UV regions/spectrums (Gadgil 1981), whose wavelength ranges from 100-400 nm and is divided into three (3) bands: UV-A (320–400 nm), UV-B (280–320 nm), and UV-C (200–280 nm) (Parit et al. 2018; WHO 2016; Forte 2014). Furthermore, the instrument can also measure the transmission/absorbance of visible light whose wavelength goes up to 800 nm. Light at a certain wavelength passes through the specimen, then reflects off both opaque and transparent surfaces and is refracted by crystals (Rocha et al. 2018). The instrument is equipped with a mirror as a light source selector and the desired light wavelength to analyze is set by a monochromator through a diffraction grating. Two coupled detectors are used to subtract the resulting light absorption from the tested specimen (Houck and Siegel 2015; Reusch 1999). Speaking of the significant impacts by UV, its radiation exposure under sunlight brings a myriad of risks, such as biological damage, organic compound degradation, discoloration, weathering, and mechanical loss (cracking); thus, UV-protection stability influences material's shelf and service life (Sadeghifar et al. 2017; Parit et al. 2018), solidifying the significance of UV-vis spectrophotometry characterization on many relevant applications, other than solely determining transparency or opacity.

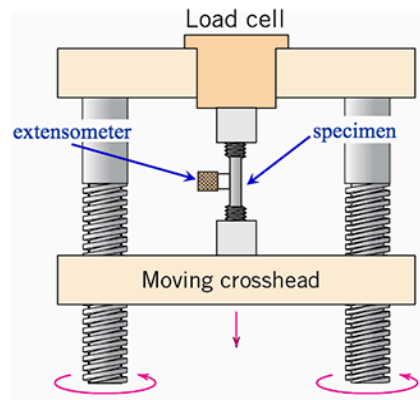
#### 2.6.4. Barrier property



**Fig. 2.10.** Schematic general principle of oxygen barrier testing (Labthink 2018)

Barrier properties are vital when fabricating materials with sensitiveness of water vapor or humidity, gases (e.g., oxygen), and/or temperature so that degradation can be delayed and shelf life extended. In particular, oxygen permeability dictates how easily oxygen passes through (Ehrmann 2024) and is among the most studied main permeants, one of which is due to its ability to move from/to the internal/external environment (Siracusa 2012; Germain 1997). The molecular diameter of oxygen is  $2.98 \times 10^{-8}$  cm; thus, specimens with pore size larger than that would possess higher permeation. Oxygen has a major role in a myriad of degradation processes, such as lipid oxidation, microorganism growth, enzymatic browning, ethylene production acceleration, and losses in organoleptic qualities such as flavor, color, and nutrients (Brown 1992; S.-I. Hong and Krochta 2006; Ayranci and Tunc 2003; Oms-Oliu, Soliva-Fortuny, and Martín-Belloso 2008; Rojas-Graü et al. 2007; Russo, Simon, and Incarnato 2006). Testing relative humidity and temperature can be set as desired when measuring oxygen permeability. Other influencing factors include cohesive energy density, polarity, crystallinity, and chemical structure, which dictate the material's free volume (Abdul Khalil et al. 2019; Miller and Krochta 1997; Koontz 2016). **Fig. 2.10.** demonstrated the schematic principle of one of the oxygen permeability testing instruments (Labthink 2018). The machine employs a dual-chamber system where one flows oxygen or air (as desired) while purged nitrogen is in the other chamber, which creates a concentration difference that allows the gases to flow between chambers. In it, a pre-conditioned sample film is mounted at ambient atmospheric pressure. A coulometric sensor scans the generated electrical signals after oxygen permeates through the sample film before flowing into the inner chamber full of dry nitrogen, which plays the role of the gas carrier for the permeated oxygen.

### 2.6.5. Mechanical properties



**Fig. 2.11.** Schematic general principle of stress-strain test (Callister and Rethwisch 2019)

Mechanical properties depict how a material behaves with the principal phenomena under the applied force of a load, making them of great significance. Deformation, disintegration, fractures, shape/dimension change, or any other subsequent macroscopic responses are heavily influenced by the material's structure. The resistance depends on both the material's interatomic forces and the subjected external forces. The most important terms in regard to material characterizations are "stress" (the acting force per unit area) and "strain" (the displacement per unit distance) (Pelleg 2012). The

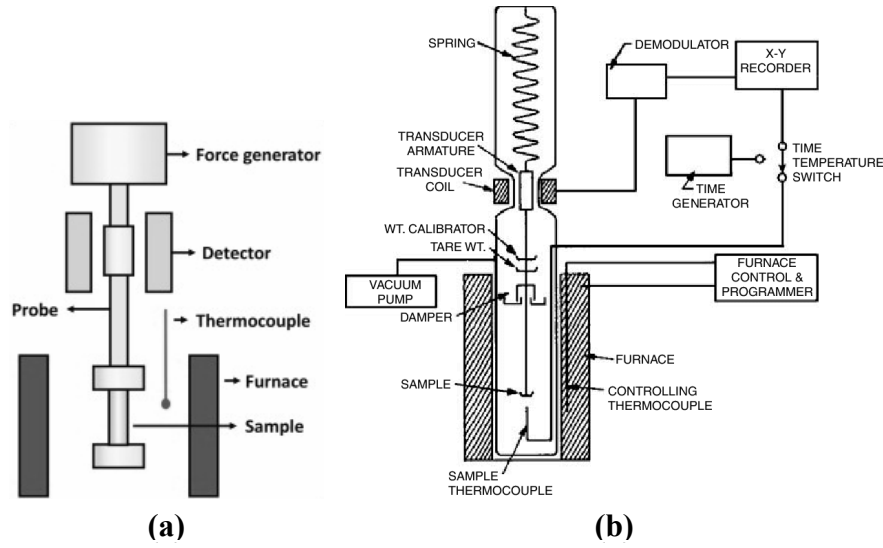
stress-strain relationship is determined by applying tension or compression in the form of a steadily increasing axial force to the sample. As illustrated in **Fig. 2.11**, as the load is increased, the deflection occurs, which is dependent on the material's elastic modulus and geometry (area and length), where, at last, the load-deflection curve is plotted. The effect of geometry can be removed by converting the values, i.e., from load to stress and/or from deflection to strain (Budynas and Nisbett 2006; Dowling 2007; Gere 2003; Hibbeler 2016; Lindeburg 2013).

Many principal mechanical behaviors can be explained by stress-strain findings such as tensile strength (or ultimate strength), elastic modulus (or modulus of elasticity; or Young's modulus; or tensile modulus), and elongation-at-break (or fracture strain or ductility). The tensile test is standardized based on the American Society for Testing and Materials (ASTM), specifically ASTM D638 (ASTM 2022), which corresponds to the material's strength and stiffness. Sample films' prior conditioning at a controlled temperature and humidity for 40 hours aims to decrease inconsistency. A tensile load is applied to the sample mounted between two crossheads and stopped once it breaks (Jain, Pandey, and Rao 2009; Koo et al. 2017). From the generated stress-strain curve, the tensile strength value can be obtained, which points to the maximum stress value. Post-ultimate stress, 'necking'—significant decrement of the cross-sectional area in a localized region—occurs in the case of ductile/stretchy samples. Tensile strength is important for its ability to determine the maximum stresses one can withstand before falling. Furthermore, the elastic modulus is the next behavior that can be determined from the stress-strain graph. Below the curve's proportionality limit portion, there is a linear stress-strain relationship whose slope locates the elastic modulus value (hence, the "constant of proportionality" term), which is overall expressed by Hooke's law. Elastic modulus depicts the material's relative stiffness or rigidity, whose formula is the ratio of stress to strain. The stiffer one material, the higher its elastic modulus, and vice versa for flexible materials (Vaidya and Pathak 2019; V. C. Pinto et al. 2015). Elastic modulus' importance is after its definition of how much material would deflect, hence displacement of structure, under an applied load (Pavlik, Pavlíková, and Záleská 2019; Jurowski and Grzeszczyk 2015). In the case of composite-based specimens, composition, matrix-filler interface, and characteristics of the incorporated filler (e.g., shape, concentration, diameter, content, dispersion) play a vital role (Wazzan, Al-Turaif, and Abdelkader 2006). The other point of interest is elongation at break, which is calculated by the ratio between length difference (post-breakage) and original length. This percentage is an important characteristic as it expresses the material's resistance to shape changes without crack formation (Djafari Petroudy 2017). A material can be classified as ductile when it possesses the capability to withstand large strains even after it has begun to yield (Budynas and Nisbett 2006; Dowling 2007; Gere 2003; Hibbeler 2016; Lindeburg 2013). The relationships between these mechanical behaviors are described through these equations below:

$$\sigma = \frac{P}{A_0} \quad ; \quad \epsilon = \frac{L-L_0}{L_0} = \frac{\delta}{L_0} \quad ; \quad \sigma = E \epsilon$$

where  $\sigma$  is stress;  $\epsilon$  is strain;  $P$  is load;  $A_0$  is the sample's initial cross-sectional area;  $L$  is the sample's current length;  $L_0$  is the sample's initial length;  $E$  is elastic modulus.

### 2.6.6. Thermal properties



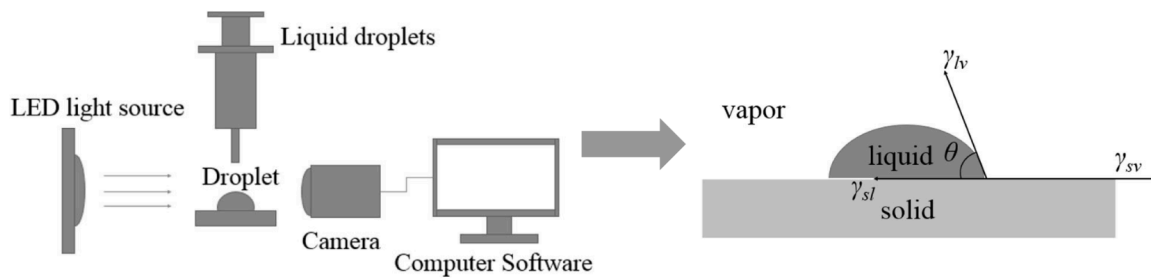
**Fig. 2.12.** Schematic general principles: **(a)** DMA (Tripathy, Padhiari, and Hota 2023) and **(b)** TGA (Rowell and LeVan-Green 2005)

Thermal analysis investigates a material's physical property as a function of an externally applied and programmed temperature at a predetermined rate (Ewing 1997). Phenomena that can be observed include endothermic (i.e., melting, boiling, sublimation, vaporization, desolvation, solid-solid phase transitions, and chemical degradation) and exothermic processes (i.e., crystallization and oxidative decomposition) (Brittain 2011). Mechanical behavioral responses (stress/strain) of a material over changing temperature can be determined by dynamic mechanical analysis (DMA). As seen in its schematic (**Fig. 2.12(a)**), the instrumental probe subjects a sinusoidal force to the sample clamped in the measuring head so the force-deformation relationship can be studied afterward. Viscoelastic variables (e.g., storage modulus or elastic modulus; loss modulus or imaginary modulus; and loss tangent or loss factor or damping factor) can be obtained (Roodbar Shojaei et al. 2019; Tripathy, Padhiari, and Hota 2023). In analyzing these properties, two common reference temperatures to highlight the transition of interest are: 1) room temperature ( $\sim 25^{\circ}\text{C}$ ) to  $32^{\circ}\text{C}$ ; and 2) boiling points at  $\geq 100^{\circ}\text{C}$  (Kwan 1998). Associated with strength, stiffness, and rigidity, the storage/elastic modulus typically decreases as temperature increases, back rises at some point before reaching a maximum value due to the altered network structure post-curing, and decreases again post-glass transition temperature, indicating thermal degradation and devitrification (Candan, Gardner, and Shaler 2016; Sanyang et al. 2016). Meanwhile, the loss/imaginary modulus property represents the material's viscous response and is usually related to internal friction. The value is obtained as the energy loss under stress/deformation (Jawaid et al. 2015; Saba et al. 2017). Lastly, the loss tangent or loss/damping factor, being the  $E''$  to  $E'$  ratio, depicts molecular movement. When exposed to varying temperatures and the sample distortion occurs, the measured recoverable strain energy indicates internal energy dissipation. The higher the temperature, the higher the damping; it is maximized in the transition region and decreases in the rubbery region. The higher the peak intensity, the higher the molecular

mobility degree and the faster the energy dissipation process, suggesting better dispersion and the eventual polymer movement/rotation inhibition. In contrast, aggregation/agglomeration allows more vacant space for the polymer chain movement/rotation, hence the visco-elastic damping factor increment (W. Chen et al. 2018; Saba et al. 2017; Sanyang et al. 2016).

Another simple and popular thermoanalytical technique is thermogravimetric analysis (TGA), as seen in **Fig. 2.12(b)**. A small amount of sample on a metal pan is positioned inside a furnace which is heated at a desired rate, with nitrogen and oxygen as the employed gases. Afterward, the thermally induced weight loss as a function of temperature is plotted (Rowell and LeVan-Green 2005; Duval 1963; Keatch and Dollimore 1975). TGA is useful for studying the material's total volatile content, desolvation, decomposition (or other composition change), and, ultimately, thermal stability—crucial as kinetic parameters when designing the product's processability and applicability (Brittain 2011; Tripathy, Padhiari, and Hota 2023). It is likewise important when analyzing degradability (Brunšek et al. 2023).

### 2.6.7. Contact angle or wettability



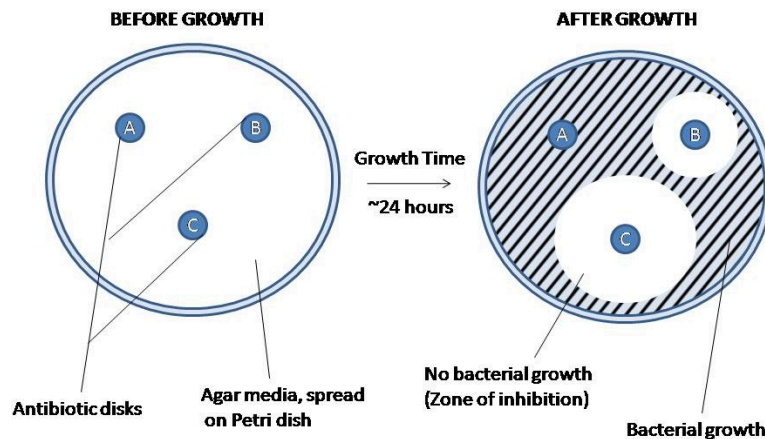
**Fig. 2.13.** Schematic general principle of optical tensiometer (J. Zhang et al. 2023)

Surface wettability/hydrophilicity (or hydrophobicity), also known as contact angle property, corresponds to a material solid surface behavior and intermolecular interactions, heavily linked to surface/adhesion energy, wetting/non-wetting, and absorption abilities, making it an essential criterion. Wetting is simply described as the possibility of a liquid to flow or, in contrast, form droplets when subjected to a solid's surface (L. Feng et al. 2014; Good 1992; Deville and Cojocaru 2006). As illustrated in **Fig. 2.13**, an optical topography instrument in the form of a drop-shape analyzer, also called a tensiometer, is employed. A dropper containing the testing liquid suspension is set; meanwhile, a camera and LED light source are adjusted to illuminate and record the droplet's contact with the solid surface of the tested specimen. At a point on the solid-liquid-vapor three-phase contact line and phase interface, a tangent ( $\gamma$ ) vapor-liquid contact line is created whose the formed angles of  $\gamma_{lv}$ ,  $\gamma_{sl}$ , and  $\gamma_{sv}$  represent the surface tension/energy between liquid-vapor, solid-liquid and solid-vapor, respectively. Interfacial tensions between these phases govern the aforesaid three (3) angles and can be calculated by the Young–Laplace equation (de Laplace 1805; Young 1805) to determine the sample's contact angle. Hydrophilic material is categorized for contact angle lower than  $90^\circ$ ; and hydrophobic for one higher than  $90^\circ$  (M. Q. Khan et al. 2019; Tripathy, Padhiari, and Hota 2023; J. Zhang et al. 2023; Río and Neumann 1997). This technique is named among the easiest and cheapest for regular evaluation on material surfaces (Selva, Selva, and Prata 2023; Cobb,

Ayres, and Macpherson 2018; Macpherson 2015; B. Yang et al. 2019; Deville and Cojocar 2006).

### 2.6.8. Antibacterial activity

Food safety becomes an issue once pathogenic microorganisms contaminate. Named among the main foodborne pathogens that cause recurrent annual outbreaks (Koutsoumanis et al. 2020; FDA 2022, 2019a), *Escherichia coli* (*E. coli*) is a common aerobic Gram-negative bacilli in a normal intestinal tract and regarded as a fecal pollution indicator. Despite being an important member of mammals' normal intestinal microflora, the species contains pathotypes/toxins that may cause an array of health issues, both intestinal and extraintestinal, e.g., diarrhea/dysentery, vomiting, (intra)abdominal infections, septicemia, hemolytic uremic syndrome, urinary tract infections, meningitis, necrosis, end-stage renal disease, hemorrhagic colitis, atherosclerosis, and immunological diseases (reactive and rheumatoid arthritis). This may also include one of the most notorious of all, Shiga intoxication symptoms, which can range from mere discomfort to death. There are a variety of strains depending on virulence factors, from commensal to lethal (Majowicz et al. 2014; S.-C. Yang et al. 2017; Obrig 2010; J. B. Kaper 2005; Munekata et al. 2020; Murray, Rosenthal, and Pfaller 2009; Yen et al. 2011; Olsvik et al. 1991; James B. Kaper, Nataro, and Mobley 2004).



**Fig. 2.14.** Schematic antimicrobial susceptibility testing: disc diffusion (EUCAST 2021)

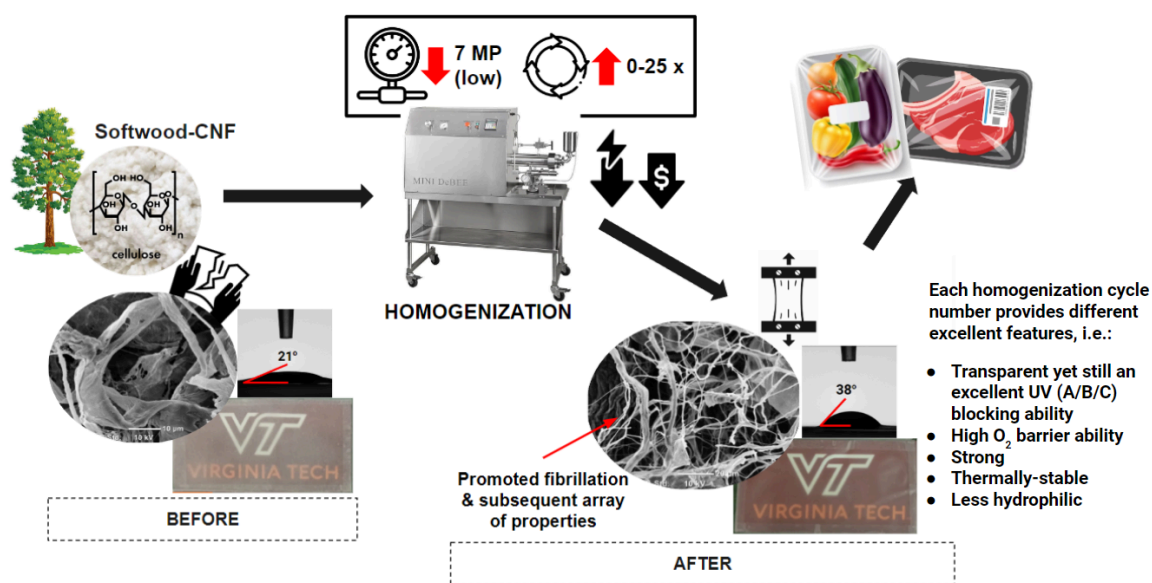
Among the several antimicrobial susceptibility tests in line with the Clinical and Laboratory Standards Institute guidelines (CLSI 2024), the agar disc-diffusion approach is very popular. Developed in the 1940s (Heatley 1944), it permits the testing of a myriad of critical bacterial pathogens. It is excellent because of its straightforwardness and cost-effectiveness (Fourati-Ben Fguira et al. 2005; Das, Tiwari, and Shrivastava 2010; Konaté et al. 2012). As seen in **Fig. 2.14**, standardized inoculum of the microorganism of choice is prepared to inoculate on an agar plate before placing on it a disc at a certain diameter made of the antimicrobial agent-incorporated specimen. The contained petri dishes are then incubated under the proper conditions to allow the said microorganism to grow. The growth inhibition zone surrounding the disc can be observed afterward, which indicates the inhibition efficacy of the antimicrobial agent (Hossain et al. 2022).

## CHAPTER 3

### Impacts of cycles of a novel homogenization process on nanocellulose and its film properties

#### ABSTRACT

The abundant and naturally biodegradable cellulose nanofibrils (CNF) have been among the most researched materials for their myriad advantages, yet are still facing challenges toward advanced developments due to their natural hydrophilicity affecting a broad range of properties. A simple, cheap, and plasticizer-free novel homogenization approach ('emulsifying cells'—equipped, low pressure at <math><10\text{ MPa}</math>, for 0-25 cycles) was proposed, and its effects on the Northern bleached softwood based-CNF films' properties were investigated. XRD results showed a maintained high crystallinity (CI=64%). Promoted fibrillation and hydrogen bonding were also observed morphologically through surface SEM images, aligned with the 44% smoother surface of homogenized CNF films ( $S_a=2.637\mu\text{m}$ ) compared to the untreated CNF films. The resulting 48% decrease in oxygen permeability ( $0.250\text{ cc}\cdot\mu\text{m}^2\cdot\text{day}\cdot\text{kPa}$  at 50%RH) is comparable to ethylene-vinyl alcohol (EVOH) and other oxygen barrier plastics. Significant improvements in mechanical (tensile strength, tensile/Young's modulus, and elongation-at-break) and thermal (storage/elastic modulus, loss/imaginary modulus, loss/damping factor, and degradation temperature) properties were confirmed. Hydrophobicity improvement (0-60 seconds) was also obtained. With varying optimum cycles of homogenization, this work demonstrates the prospect of a straightforward, cheap, and environmentally friendly homogenization approach in preparing CNF with enhanced processability and applicability.



Graphical abstract

### 3.1. Introduction

Americans waste 40% of their food, costing 1.3% of GDP (FAO 2021b, 2013b; EPA 2016b). Food spoilage is mainly triggered by microorganisms, water content, oxygen, humidity, temperature, and light (Magoulas 2016), creating waste. Together, food and its packaging comprise roughly 45% of the solid wastes landfilled in the USA (EPA 2015b). To produce such food waste, one-third of the global agricultural land is used annually, releasing large amounts of greenhouse gas. Around 98% of single-use plastic products (including food packaging) are made of non-sustainable and/or non-biodegradable fossil fuel or "virgin" feedstock (D. Charles and Kimman 2023); thus, 80% have entered landfills and nature (Geyer, Jambeck, and Law 2017). Composting merely covers 6% due to limitations in infrastructure, upfront/expensive labor, material supply assurance, and contamination (ReFED 2018; NRDC 2020). Only 12% and 9% are incinerated and recycled, respectively, as they suffer from losses, degradation, reduced mechanical performance, high cost, complexity, released toxic chemicals, micro-/nano-plastics, volatile organic compounds, and carbon oxides. The remaining in landfills and oceans results in uncontrolled degradation and methane contaminating soil/water, especially when left indefinitely under anaerobic conditions (UNEP 2022; Geyer, Jambeck, and Law 2017; Birkett et al. 2017; Byun et al. 2015). Due to the urbanization-derived global dietary change and rising reliance on processed foods (World Packaging Organisation 2008), food packaging remains in high demand.

In response to these lingering constraints of food and food packaging waste, "prevention" (e.g., spoilage-prevention packaging) is largely encouraged over "recovery" and "recycling" actions (FAO 2022a; EPA 2016b; European Commission 2023); thus, studies of food primary packaging are continually advancing in shelf life extension through moisture and oxygen-barrier properties improvement, aiming for lower food spoilage rates (Verghese et al. 2015; Almenar 2023). Likewise, research on packaging materials with biodegradation characteristics in various environmental conditions has also been prioritized to help tackle the lingering issue of accumulating packaging waste. The Earth's most abundant naturally biodegradable polymer is cellulose (Klemm et al. 1998). It comprises several linear  $\beta$ -(1,4)-d-glucose chains, forming (micro)fibril structures whose individual chains are joined by a network of inter-/intramolecular hydrogen bonding and van der Waals forces (Xiaowen Liu et al. 2023). Cellulose can be converted into two (2) 'top-down' nanosized products that excel in ability to form more extensive network structures due to their nanoscale dimensions (H. Lee, Sundaram, and Mani 2017): the rigid rod-like cellulose nanocrystals/CNC (chemically, e.g., acid hydrolysis, oxidation, and cationization; removing amorphous region, leaving purely-crystallines) and the flexible cellulose nanofibrils/CNF (mechanically, e.g., micro-fluidization, homogenization, grinding process, sonication; preserving both amorphous and crystalline structure) (Dias et al. 2020; Ibrahim et al. 2019; Sofla et al. 2016). Sometimes referred to as cellulose nanofibers or nanofibrillated cellulose, CNF is among the most researched materials due to its high specific surface area, crystallinity, surface activity, oxygen barrier ability, and an even better aspect ratio than CNC, while having low porosity and density (B. Yang et al. 2019). In the past decades, many have researched the growing potential of CNF in improving barrier properties in packaging applications (Roilo et al. 2018; Fukuzumi et al. 2013, 2009; Aulin, Gällstedt, and

Lindström 2010) as well as its biodegradability in diverse environments (Arun et al. 2022; Niu et al. 2021; Jinchuan Xu et al. 2021; Babae et al. 2015). However, neat CNF film still possesses some challenges that hinder its further processability and applicability, including in the packaging field, especially if compared to conventional polymers. CNF is insoluble and non-redispersible in water and most organic solvents due to strong hydrogen bonding between the nanofibrils (Nechyporchuk, Belgacem, and Bras 2016). CNF is also hydrophilic/hygroscopic by nature, and moisture absorption weakens its interfibrillar hydrogen bonding and causes swelling, resulting in loss of strength, durability, oxygen barrier ability, and thermal stability (Xia et al. 2018; B. Yang et al. 2019).

Homogenization is among the most popular treatments for cellulose-based materials as it does not require high temperatures and organic solvents or chemicals other than simply water, highlighting its simplicity and environmental friendliness (L. Wang et al. 2021). Cellulose-based aqueous suspension is pumped through a spring-loaded valve assembly that opens and closes repetitively, allowing the suspension to be subjected to certain pressure drops, inducing high shear, cavitation, and impact forces in the narrow slit of the valve. It promotes fibrillation that eventually improves hydrogen bonding and 3-dimension network strength (Nagarajan et al. 2021; H. Lee, Sundaram, and Mani 2017; Chun et al. 2011). The structural stability would be improved since the optimized homogenization affects rheology, particle size, surface area contact, and distribution/dispersity, both before and after the film drying stage, especially for polysaccharides like CNF (Flores et al. 2021; Usenko et al. 2013; Wenjian Cheng et al. 2010). Homogenization is categorized based on the conditioning pressure range, i.e., ultra-high (>250 MPa), high (50-250 MPa), and low (<50 MPa) (Aguayo et al. 2017). However, the most notorious drawback of conventional homogenization is the high pressure that leads to high energy consumption and cost (Nagarajan et al. 2021; H. Lee, Sundaram, and Mani 2017). High-pressure homogenization also has a negative linear effect on CNF crystallinity due to surface crystallites peeling off under high shear and impact forces (Davoudpour et al. 2015). Therefore, homogenization under lower pressures (<50 MPa) is prospective for a significantly less costly homogenization (Boulemkahel et al. 2021), including for CNF processing, which has yet to be studied to date. In the case of a solely homogenization strategy in cellulose-based material treatment, multiple passes or cycles are usually required to obtain a well-controlled manner (Nakagaito and Yano 2004). The number of cycles determines the final homogenates' degree of fibrillation and subsequent properties (Davoudpour et al. 2015; Kalia et al. 2014). Some studies even emphasized the importance of the number of passes over other homogenization operating conditions, like pressure (Fu et al. 2011; Yu et al. 2021). Another common challenge in the homogenization technique is the clogging issue (Nagarajan et al. 2021; H. Lee, Sundaram, and Mani 2017) therefore, this work also explored the utilization of a homogenizer equipped with the novel, patented modular 'emulsifying cells' that allows easier control of pressure, impact, shear, cavitation force, flow mechanisms, and process duration to prevent clogging, agglomeration/aggregation, and destruction in the resultant CNF films due to overheating, over-shearing, and/or over-pressure (BEE International 2015). Such design has been evidenced to be fruitful in an array of materials, including CNF-based products where the effect of varying cycles/passes and/or pressures were investigated, e.g., TEMPO-based CNF/8 passes/151

MPa (V. C. F. Li et al. 2018), CNF aggregates/5 passes/69-207 MPa (Q. Cheng and Via 2017), CNF-polyamide/10-30 passes/70-310 MPa (J.-C. Lee et al. 2018), TEMPO-based CNF/3 passes/160 MPa (Du et al., 2016), and CNF-PLA/4-20 passes/110-290 MPa (T. Wang and Drzal 2012). However, to date, the utilization of the aforesaid ‘emulsifying cell’ based homogenization system at low pressure on neat-CNF preparation has yet to be found.

Softwood-based CNF, specifically one derived from northern bleached softwood kraft pulp, was selected for this study. Besides being the first raw material used in CNF manufacturing (Turbak, Snyder, and Sandberg 1983; Herrick et al. 1983) and today’s most popular (H. Lee, Sundaram, and Mani 2017), softwood-based CNF was more flexible and responsive to be fibrillated than hardwood, whose spirally-layered outer secondary wall reduces flexibility and accessibility to the inner secondary wall (Stelte and Sanadi 2009). Softwood is also easier to modify chemically due to glucomannan being the dominant component of its hemicellulose, which contains C6 primary hydroxyls easily convertible into other functional groups. In contrast, the hemicellulose of hardwood is mainly xylan, which does not have C6 primary hydroxyl groups (Fukuzumi et al. 2009; Sjostrom 2013). In addition, films produced using softwood-based CNF also possessed a higher light transmittance (Fukuzumi et al. 2009).

Accordingly, plasticizer addition to a film-forming solution for biopolymers is usually essential to increase film flexibility, although it may also lead to poor mechanical properties and low thermal stability. Plasticizer was not used in this study because of numerous concerns, such as aging, leaching, migration, volatility, extraction, exudation, evaporation, segregation phenomenon (phase separation), and anti-plasticization effects (Eslami et al. 2023; Paixão et al. 2019; Sothornvit and Krochta 2005). Plasticizer addition may also negatively affect the film’s permeability. Hydrophilic plasticizers affect water vapor permeability; hydrophobic plasticizers affect oxygen, aroma, and oil permeability (Daniels 1989). Even without any plasticizer, drying the colloidal aqueous CNF dispersions produces the typical CNF non-porous film, with random-in-plane orientation and slightly swirled conformation of nanofibrils (Henriksson et al. 2008). In this instance, capillary forces arise from the water, seeking to minimize the liquid/air interface. This force is the main adhesion mechanism through liquid bridges (Z. Zhou, Li, and Zhao 2006). Since the crystalline region of the cellulose surface is strongly hydrophilic, this capillary force can be strong enough to plastically deform the nanofibrils (Ogawa, Nishiyama, and Mazeau 2020). In addition, the plastic yielding (yield strength) in CNF films is related to interfibril shearing in the interphase region (C. Zhang et al. 2021; Sinko and Keten 2014), showing the significance of abundant H-bonding sites on the nanofibril surfaces. As the water between the fibers evaporates, the capillary pressure increases, pulling the fibers together, whereas when still wet, the delignified fiber wall is highly hydrated, and the water acts as a strong plasticizer (Wohlert et al. 2022).

Ultimately, the development of plasticizer-free softwood-based CNF film treated with a novel, milder-conditioned homogenization in varying cycles was conducted. This study aimed to determine the influence of homogenization in the aforesaid operating conditions on the functional properties of the resulting CNF. The effect of different numbers of homogenization passes was hypothesized to properly promote the fibrillation of CNF that governs the final product’s properties, including, but not limited to, physical, mechanical, morphological, and thermal stability.

## 3.2. Materials and Method

### 3.2.1. Materials

CNF slurry with 3% solids derived from northern bleached softwood kraft pulp was obtained from the Process Development Center of the University of Maine (Orono, ME, USA). HPLC-grade water was purchased from EMD Millipore (Burlington, MA, USA).

### 3.2.2. Homogenization process

CNF slurry (3% solids) was diluted with water (1:3) by mixing under constant stirring (30 mins) to produce neat-CNF solution. The solution was further treated with a Mini DeBEE 30 homogenizer (BEE International, South Easton, MA, USA) with a 0.20 mm (0.008 inch) nozzle at 7 MPa, repeated for a different number of cycles (0, 5, 10, 15, 20, 25; labeled as N-0, N-5, N-10, N-15, N-20, N-25, respectively). Homogenization cycles were maintained under 30 passes according to the preliminary studies, as well as referring to optimized results of previous studies on homogenized cellulose prepared at a medium/low conditioning pressure range (Chun et al. 2011; V. C. F. Li et al. 2018; Q. Cheng and Via 2017; Du et al. 2016; J.-C. Lee et al. 2018; T. Wang and Drzal 2012). On top of the internal cooling system, the homogenizer was equipped with an Isotemp Refrigerated Circulating Bath 910 chiller (Fisher Scientific, Waltham, MA, USA).

### 3.2.3. Film production

The homogenized CNF solution (30 mL) was poured into a 9 cm x 1.5 cm petri dish using a solvent casting technique and left to dry in the Forma Environmental Chamber (Thermo Scientific, Waltham, MA, USA) (50 %RH, 23 °C, 48 hours). The same preconditioning parameters were completed prior to each test of the films. The schematic is presented in **Fig. S.1**.

### 3.2.4. Characterizations

#### 3.2.4.1. Fourier transform infrared (FTIR) analysis

FTIR spectra indicating functional groups were determined by the the Nicolet 8700 spectroscopy equipped with Attenuated Total Reflectance (ATR) (Thermo Scientific, Waltham, MA, USA) at a wave number range of 400–4000  $\text{cm}^{-1}$  and a total of 32 scans. The results were reported as an average of three (3) replicates.

#### 3.2.4.2. Crystallinity

X-ray diffraction (XRD) pattern and relative crystallinity were investigated by the D8 (Bruker Corporation, Billerica, MA, USA) equipped with twin/twin optics and a cooler (15-20 °C). Using the diffrac.commander software, the “Motorized Slit: Slitwidth” setting was applied for both primary (0.6 mm) and secondary (5 mm) twins. The sample was irradiated in the range of  $2\theta = 5\text{--}35^\circ$  with total scans of 1181 steps and  $0.025^\circ$  increment for 0.5 s. The generator was set with 40 kV energy, 40 mA current, and Cu  $K\alpha$  irradiation ( $\lambda = 1.54056 \text{ \AA} = 0.154 \text{ nm}$ ). The resulting data in *.brml* format were processed using PowDLL (Kourkoumelis 2013) and Profex (Doebelin and Kleeberg 2015) software. The crystallinity index (CI) was evaluated using the “height XRD” method or Segal’s empirical equation (Segal et al. 1959), which is deemed the easiest and most

time-efficient, thus being the most popular (S. Park et al. 2010). It measures the ratio of the intensity between the crystalline and amorphous peaks, which are located around  $2\theta = 23^\circ$  and  $2\theta = 18^\circ$ , respectively. The crystalline peak can also be called the major peak or  $I_{total}$ . Meanwhile, the amorphous peak is also known as the minima between the major (indices (200) at  $2\theta = 22.5^\circ$ ) and secondary peaks (indices (10 $\bar{1}$ ) at  $2\theta = 14.7^\circ$ – $16.1^\circ$ ), or  $I_{am}$ . Ultimately,  $I_{total}$  measures the maximum intensity of the crystalline peak, while  $I_{am}$  is the minimum intensity of the amorphous peak or the maximum point where the entirely amorphous cellulose would be (Segal et al. 1959; Nurhadi et al. 2022). The results were reported as an average of three (3) replicates.

$$CI = \left(1 - \frac{I_{am}}{I_{total}}\right) \times 100\%$$

#### 3.2.4.3. Morphology, surface roughness, and transparency

Surface morphology was characterized by the NeoScope JCM 5000 Scanning Electron Microscope (SEM) (JEOL Ltd., Tokyo, Japan). For surface SEM observation, the sample (25 mL) was initially prepared by lyophilization or freeze-drying (Millrock Technology, Kingston, NY, USA) after being kept in a  $-80^\circ\text{C}$  Forma 88000 ultra-low freezer (Thermo Scientific, Waltham, MA, USA). It was then mounted onto a pin stub mount using copper tape before being coated with roughly 100 Å of gold in an ion sputter coater (Denton Vacuum, Moorestown, NJ USA) using argon gas (10-15 psi) at pressure and current of 50 mTorr and 45 milliamps, respectively, for 45 seconds. Specimens were subsequently imaged at different magnifications under a vacuum at an operating voltage of 10 kV. Furthermore, the surface roughness of films was investigated using the Keyence VK-X3000 3D surface profilometer (Keyence Corporation, Osaka, Japan) referring to ISO 25178. Three-dimensional images of the film surfaces were analyzed with the Keyence VK-X3000 MultiFileAnalyzer (Keyence Corporation, Osaka, Japan) software to perform filtering according to the reference plane to the films' real surface to smooth out height variability before the roughness parameters measurement. Afterward, arithmetical mean height ( $S_a$ ) or the average absolute height value at each point in the definition area was reported as an average of three (3) replicates. Lastly, to determine transparency, light transmission spectra were analyzed using a Genesys 10S UV-vis spectrometer (Thermo Scientific, Waltham, MA, USA) in the wavelength range of 190–900 nm with 0.5 nm interval, in addition to the visual observation

#### 3.2.4.4. Barrier property

According to ASTM D 3985-95, Oxygen Permeability (OP) was characterized using the C230 Oxygen Transmission Rate Testing System (Labthink International Inc., Medford, MA, USA). The 700-118-20 Quick Mini digital thickness gauge (Mitutoyo, Kawasaki, Japan) was used to measure ten random readings to the nearest 0.001 mm, which were then averaged. The plate was then inserted into the cell chamber. The “Film-ASTM” test type without compensation was selected. To maintain purity, cycle process mode with rezero purging for  $\text{N}_2$  gas in the cells was performed for 30 seconds with one cycle interval. The system was preheated for 2 hours. The flow rate, test humidity, and test temperature were adjusted to 100 mL/min, 50%RH, and  $23^\circ\text{C}$  receptively. Six (6) cycles under standard test mode were completed at 30-minute intervals in each cell. The results were reported as an average of six (6) replicates.

#### 3.2.4.5. Mechanical properties

Mechanical properties were measured with Universal Testing Machines (UTM) Material Test Systems 10-GL 27-00112 (MTS Systems Corporation, Eden Prairie, MN, USA). The films' tensile strength (TS), tensile/Young's modulus (E), and elongation-at-break percentage (%EAB) were measured with 0.1 N preload at a test speed of 0.40 mm/s until the films (2 × 8 cm) were ruptured. TS was calculated from the ratio of maximum load to the cross-sectional area of the film. Meanwhile, %EAB was the ratio between the increased length pre-rupture and the film's initial length. Lastly, E was the ratio between stress and strain from the linear portion of the stress-strain curve. The results were reported as an average of twenty (20) replicates.

#### 3.2.4.6. Thermal properties

Thermomechanical properties were measured using the Discovery Q850 Dynamic Mechanical Analyzer (DMA) (TA Instruments, New Castle, DE, USA). The film strip (13 × 7 mm) was loaded using the dual screw film clamp type. The multi-frequency strain was employed at a % oscillatory strain of 0.05%, and a static force of 0.01 N was applied in tension. Using temperature ramp mode (-60–150 °C), conditions were maintained at 5 °C/min. Force track and soak times were adjusted to 125 % and 5 minutes, respectively. The data was then recorded at 1 Hz. E' (elastic/storage modulus), E'' (loss/imaginary modulus), and Tan( $\delta$ ) (loss/damping factor;  $\text{Tan}\delta = E''/E'$ ) plots were observed. Furthermore, thermal degradation analysis was performed using a Thermogravimetric Analyzer (TGA) 550 (TA Instruments, New Castle, DE, USA). Pan loading-taring was conducted using the auto sampling technique using a platinum (100  $\mu\text{L}$ ) pan. An amount of freeze-dried CNF (5-15 mg) was spread in the pan to maximize surface area. It was equilibrated at 50 °C, then ramped from 10 °C/minute to 800 °C. Temperature was recorded when the sample's weight loss reached 5% ( $T_{d,5\%}$ ). Data analysis using the TRIOS or TA Universal Analysis (TA Instruments, New Castle, DE, USA) software was then performed. The results were reported as an average of six (6) replicates.

#### 3.2.4.7. Contact angle or wettability

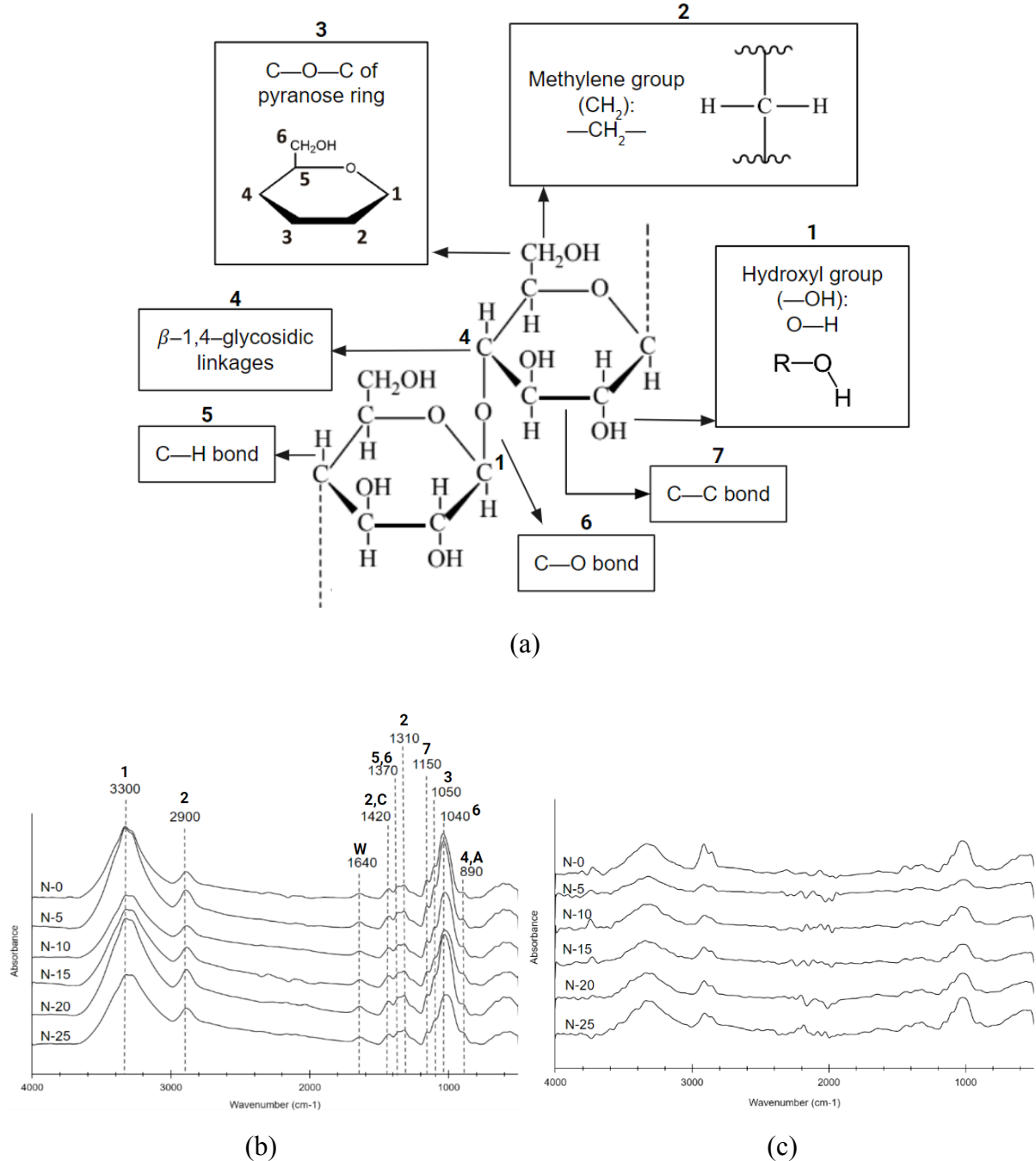
The water contact angle or wettability was tested by the OneAttension® Theta Flow optical tensiometer (Biolin Scientific, Stockholm, Sweden). Using the “sessile drop” experiment type equipped with a 200  $\mu\text{L}$  pipette tip at 25  $\mu\text{L}/\text{min}$  dispenser speed, a drop (5  $\mu\text{L}$ ) of HPLC-grade water was deposited onto the film surface to measure the contact angle in varying regions. “Light” and “heavy” phases were set as air and water (20 °C), approximately, which are selected to determine the density difference value for the surface tension measurement. The drop images and subsequent contact angles were obtained with a total duration of 60 seconds at ten-second intervals and reported as an average of three (3) replicates.

#### 3.2.5. Statistical analysis

Data were analyzed by a one-way variance analysis (ANOVA) followed by Tukey's test and Student's t-test, using the Microsoft Excel 2019 (Microsoft Corporation, Redmond, WA, USA) and/or JMP 16 Pro (SAS Institute, Cary, NC, USA) software to establish significant differences ( $p\text{-value} \leq 0.05$ ). The use of different letters on graphs data denotes statistically significant differences.

### 3.3. Results and Discussion

#### 3.3.1. Fourier transform infrared (FTIR) analysis



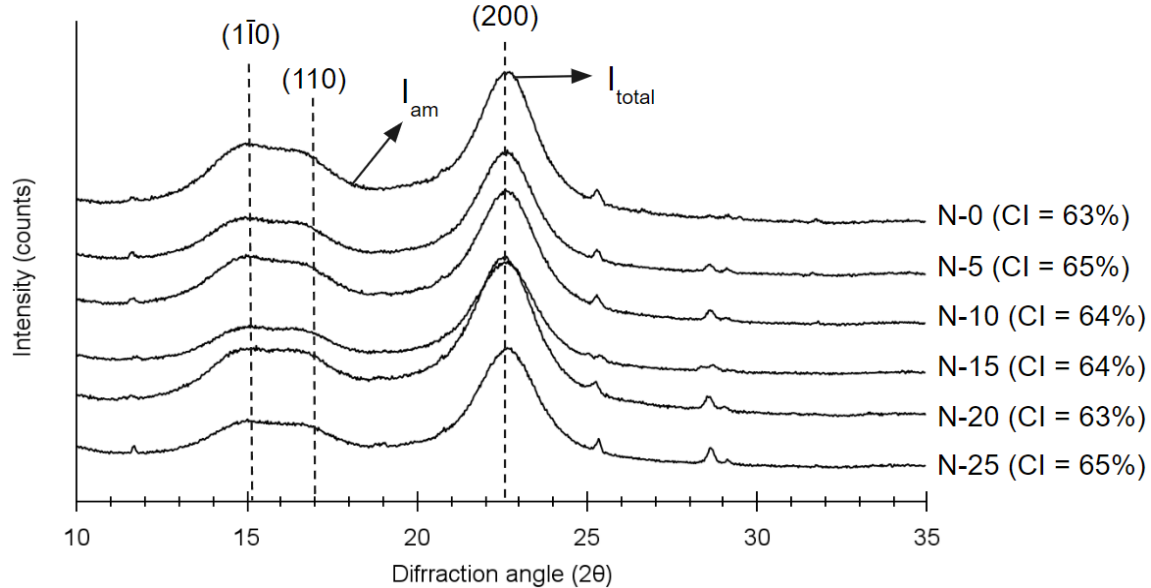
**Fig. 3.1.** (a) Chemical structure of cellulose and its major functional groups; (b) FTIR spectra of freeze-dried CNFs and (c) CNF films

**Table 3.1.** FTIR peaks assignment (functional group and molecular vibration type) of freeze-dried CNFs

Approximate wavenumber (cm <sup>-1</sup> )	Functional group and molecular vibration type
3300–3340	stretching of O—H hydroxyl groups
2900	(anti)symmetric stretching of C—H (of CH <sub>2</sub> or methylene group)
1640	bending of O—H and H—O—H (absorbed of H <sub>2</sub> O or water)
1420	scissoring of C—H (of CH <sub>2</sub> or methylene group); cellulose's crystalline region
1370	bending of C—H and C—O bond
1310	wagging of C—H (of CH <sub>2</sub> or methylene group)
1150	stretching of C—C bond
1050	antisymmetric stretching of C—O—C (of pyranose ring)
1040	stretching of C—O bond
890	cellulosic β-1,4-glycosidic linkages; cellulose's amorphous region

**Fig. 3.1(a)** illustrates the chemical structure and major functional groups of cellulose, whose presence was confirmed and discussed in this section through FTIR analysis. **Fig. 3.1(b)** showcases the resultant FTIR spectra of the freeze-dried CNFs, and **Table 3.1** lists the approximate wavenumber (cm<sup>-1</sup>) of dominant peaks or spectral bands observed in each spectrum and their assigned functional group (Soni, Hassan, and Mahmoud 2015; Morán et al. 2008). Firstly, the intramolecular and intermolecular hydrogen (—OH) bonds are characterized by the observed O—H stretching vibrations that belong to hydroxyl groups. These bonds were observed in all CNFs, likewise for the methylene (CH<sub>2</sub>) groups, C—H, C—C, C—O, and C—O—C bonds. These bonds occurred not only as glycosidic linkages but also within each of the pyranose rings of glucose units. The spectral bands indicating the water (“W”) O—H and H—O—H bending were present in all CNFs spectra. This finding indicates the presence of remaining moisture even after the drying process due to the cellulose-water interaction (Morán et al. 2008; Korhonen et al. 2011). FTIR analysis in this study used freeze-dried CNFs (**Fig. 3.1(b)**) instead of CNF films (**Fig. 3.1(c)**). The eliminated water content produced FTIR spectra with more obvious peaks, making it easier to observe the dominant spectral bands. Besides water, other volatile degradation products emitted during the freeze-drying could be hydrocarbons (3100–2600 cm<sup>-1</sup>), carbon dioxide (2420–2240 cm<sup>-1</sup>), carbon monoxide (2240–2040 cm<sup>-1</sup>), carboxyl and carbonyl functional groups: C=O (1800–1600 cm<sup>-1</sup>) and C—O functional groups (1130–1050 cm<sup>-1</sup>) (Quiévy et al. 2010)—which are all indeed present in CNF films spectra, but unapparent in freeze-dried CNF's. All spectra discussed were similarly observed in several other studies on homogenization impact on cellulose-based materials (Quiévy et al. 2010; Rahbar Shamskar, Heidari, and Rashidi 2019; D. Huang et al. 2020). Conclusively, no new bonds were formed after homogenization, indicating no chemical composition change, as seen in similar studies (D. Huang et al. 2020; Ilyas et al. 2019). In addition, the resulting FTIR spectra also confirmed that, despite the mechanical treatments, the desired high crystallinity of CNF was successfully maintained, whose more in-depth analysis is discussed in the next section.

### 3.3.2. Crystallinity



**Fig. 3.2.** XRD diffraction spectra and crystallinity index (CI) of CNF films

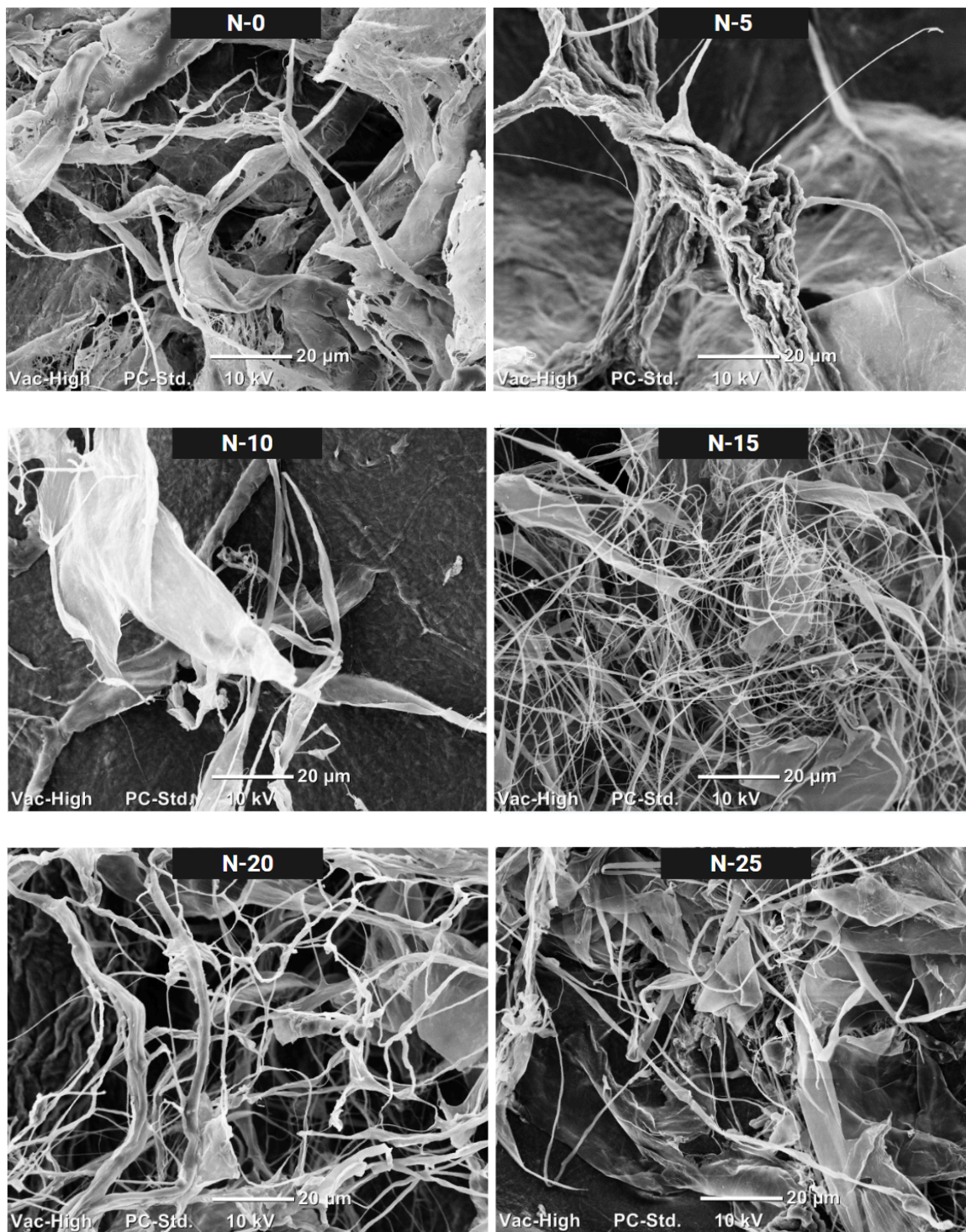
**Fig. 3.2** presents diffraction peaks of each CNF films, which is described further by referring to several other reports (Manimaran et al. 2019; Siti Syazwani et al. 2022; SriBala et al. 2016; Hafez, Amini, and Tajvidi 2020; G. Zhao et al. 2019). Overall, referring to the characteristic peaks of each cellulose type (Y. Liu et al. 2022), all of the resulting diffraction patterns evidenced the Cellulose I type. The peaks typically found in Cellulose I are the Miller indices of  $(1\bar{1}0)$  at  $2\theta=15^\circ$ ,  $(110)$  at  $2\theta=17^\circ$ , and  $(200)$  at  $2\theta=22.5^\circ$ , and  $(004)$  at  $2\theta=35^\circ$ —which are similarly displayed for each CNFs. It indicates that, despite the mechanical treatment, CNFs retained the crystallinity profile and structure of the native cellulose contained in the rawest form of softwood, as found in nature. Lastly, minuscule peaks arising at  $2\theta = 25^\circ$ ,  $2\theta = 28.5^\circ$  and  $2\theta = 32^\circ$  are likely attributed to the crystalline phase of impurities/trace elements or residual salts.

**Fig. 3.2** also displays the crystallinity index (CI) that defines the mass ratio of the crystalline substance in the total dry sample based on the crystallographic two-phase model (Daicho et al. 2018). CI is measured by determining two peaks: the sharp, high, and obvious major/crystalline/monoclinic peak ( $I_{total}$ , around  $2\theta = 20-25^\circ$ ); and the broad secondary/amorphous peak ( $I_{am}$ , around  $2\theta = 12-20^\circ$ ). As seen in the XRD spectra comparison, the results showed no significant difference in CI values, averaging 64%. It signifies that the applied milder-conditioned homogenization (7 MPa for 0-25 cycles) did not significantly alter the molecular structure and crystalline rearrangement/reorganization of the homogenized CNFs. Similar findings of low-pressured homogenizations causing only slight crystalline structure loss were reported in previous studies, e.g., 30-50 MPa for 0-3 cycles (Boulemkahel et al. 2021), 30-60 MPa for one cycle (Kasemwong et al. 2011), and 60 MPa for one cycle (B. Wang et al. 2008). In comparison, some researchers investigated the considerable effects of homogenization on CNF's crystallinity profiles under higher pressure and/or cycles, e.g., 20-50 MPa for 20-40 cycles (Davoudpour et al. 2015), 40 MPa for four cycles (Xiaowen

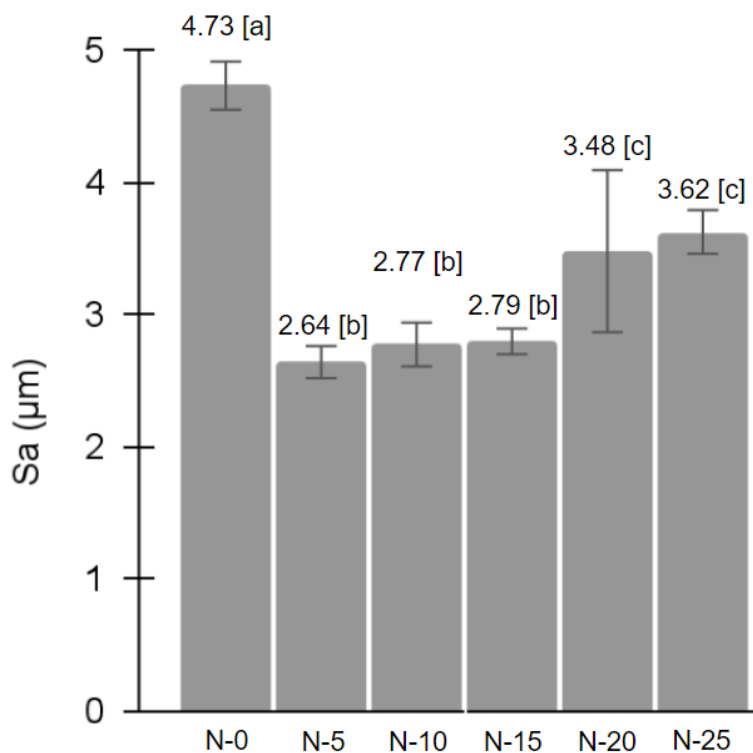
Liu et al. 2023), 140 MPa for 4-20 cycles (Chun et al. 2011), and 150 MPa for 0-20 cycles (Ueda et al. 2022). Though mechanical treatment promotes water binding capacity and gel-like structure formation (Ulbrich and Flöter 2014), harsh conditions (e.g., excessive pressure and number of passes) during homogenization may possibly lead to the destruction of (sub)crystal zones and covalent bonds within and between molecules (Hoover and Ratnayake 2002). Particles passing through such a "high energy zone" results in aggravated particle structure and compact layer structure damage, decreased amorphous region stability, and even branched molecules in the crystal region to be cut into short straight-chain molecules (Guo et al. 2019; S. Meng et al. 2014). This highlighted the influence of the applied lower pressured homogenization, despite multiple passes, in maintaining the crystallinity profile. This technique would suit certain CNF processing where the preserved long cellulose chain is essential to the final product, e.g., reinforcement agent (Yano et al. 2018) and thermal-conductive filler (Lou et al. 2021).

### *3.3.3. Morphology, surface roughness, and transparency*

The surface SEM images of the freeze-dried CNFs post-homogenization are presented in **Fig. 3.3**. Prior homogenization (N-0) images showed a compact but rough and uneven appearance of CNF, consisting of strongly entangled fibrils forming thick bundles with an average width of 8-10  $\mu\text{m}$ . The degree of fibrillation was improved after homogenization due to the provided shear impact as the suspensions flew through extremely small gaps under intensive shear (L. Wang et al. 2021), even after only five (5) cycles (N-5). As shown in N-5 images, fibrils began to loosen and detach from each other, significantly reducing the bundles' dimensions and releasing separated, individual fibrils with nanosized diameter. The increasing specific surface areas eventually promoted hydrogen bonding with water (Kose et al. 2011). Images also showed less roughness due to the reduced number of big fibril bundles. Though much more undetached, some thick fibril bundles could still be seen in N-5 surface micrograph images. Furthermore, it was investigated that the more homogenization passes, the higher the degree of fibrillations, the more hydrogen bonds with water, and ultimately, a web-like network of thin and long fibrils was formed. Furthermore, more obvious and drastic fibrillation was observed after 20 homogenization cycles (N-20). A complex web was formed, made of numerous fine fibrils connected with each other, indicating the successfully promoted hydrogen bonds. However, the over-shearing occurrence was discovered in the higher number of homogenization cycles, indicated by the individual fibrils that began to agglomerate at N-15, N-20, and even worse at N-25. The apparent aggregation caused by such excessive mechanical treatment was observed in other reports (Malucelli et al., 2018; Pelissari et al., 2017). Overall, identical behavior was observed in previous studies that prepared nanocellulose using other homogenization approach (multiple passes with high-pressure/shear-speed) coupled with other physical/chemical treatments (Kose et al. 2011; Dilamian and Noroozi 2019), showing the effectiveness of the conducted method in this study despite the much milder conditions applied (ultra-low pressure).



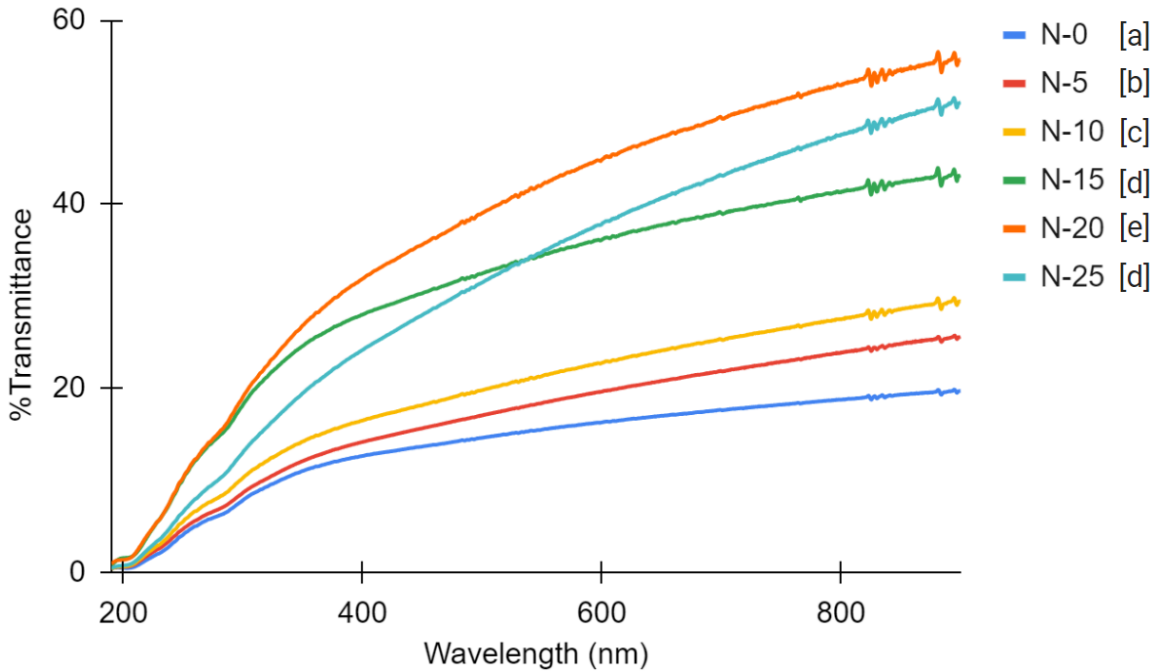
**Fig. 3.3.** Surface SEM images of freeze-dried CNFs at 1000x magnification



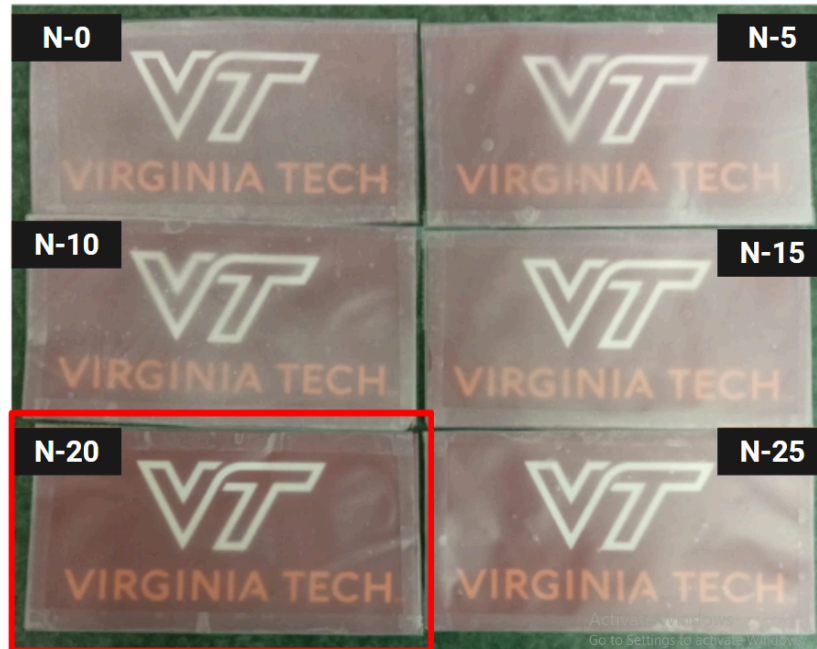
**Fig. 3.4.** Surface roughness of CNF films: arithmetical mean height or  $S_a$

The roughness characteristics of the CNF films' top surface were presented in both microscopic images (**Fig. S1**) and  $S_a$  values (**Fig. 3.4**), which can be defined as the arithmetic mean deviation of the height of a line, where this basic parameter depicts the surface micro-geometry of the specimen (Woch et al. 2022). The obtained range of values was comparable to that of other CNF reports (Miettinen, Chinga-Carrasco, and Kataja 2014). It can be seen that solely five (5) cycles of homogenization managed to successfully decrease the roughness of the CNF control film, making it approximately 44% smoother, from 4.73  $\mu\text{m}$  (N-0) to 2.64  $\mu\text{m}$  (N-5). However, the effect of over-shearing was evidenced after 20 passes (N-20), where the surface became rougher. Although, N-20 is still significantly smoother, specifically 27% smoother, than the surface of the unhomogenized CNF film (N-0). A similar finding of the recovered roughness was investigated in other studies confirming protuberance prominences on the material's surface, which might occur whether aggregation ensued (Xue et al. 2021) or not (Żołek-Tryznowska et al. 2023). Evidently, the effect of such protuberance on the film's roughness was more obtrusive despite the promoted fibrillation and no aggregation (as observed in morphological results in **Fig. 3.3**). Additionally, other roughness profiles investigated were z-stack and three-dimensional microscopic images (**Fig. S1**). Z-stacking or focus-stacking combines multiple images taken at different focal distances to provide a composite image with a greater depth of field, including the thickness of the plane of focus, than any of the individual source images (Ray 2002; D. Johnson 2008). During analysis, filtering referred the reference plane to the CNF films' real surface to reduce height variability. Especially when conducting characterization with such high magnification and numerical apertures that suffer from decreasing depth-of-field, this

image processing can capture in-focus images (Larson and Banks 2022). The obtained images confirmed the findings on roughness value analysis, where the surface of N-5 showed a significantly smoother appearance compared to N-0 and was even better at N-20. These roughness results were supported by the morphological findings (**Fig. 3.3**) of surface examination, as the surface roughness is highly influenced by residual fibers and the morphology of fibrils (Chinga-Carrasco et al. 2014). Micro-pores likely existed in locations along the thicker parts of the CNF films, which are usually attributed to containing more residual fibers (Miettinen, Chinga-Carrasco, and Kataja 2014). Residual fibers are the non-fibrillated microscopic fibers and fiber fragments whose amount dictates the morphology of a CNF (Nechyporchuk, Belgacem, and Bras 2016; Chinga-Carrasco et al. 2014). CNF films with smooth features are commonly used as gloss finishes; meanwhile, the roughly-surfaced CNF films possess noteworthy implications on electrical behavior when used in organic field-effect transistors (Lehmann and Zahn 2009).



(a)



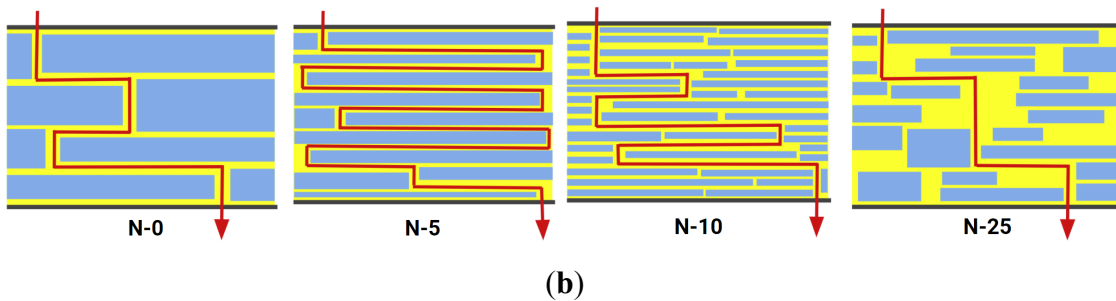
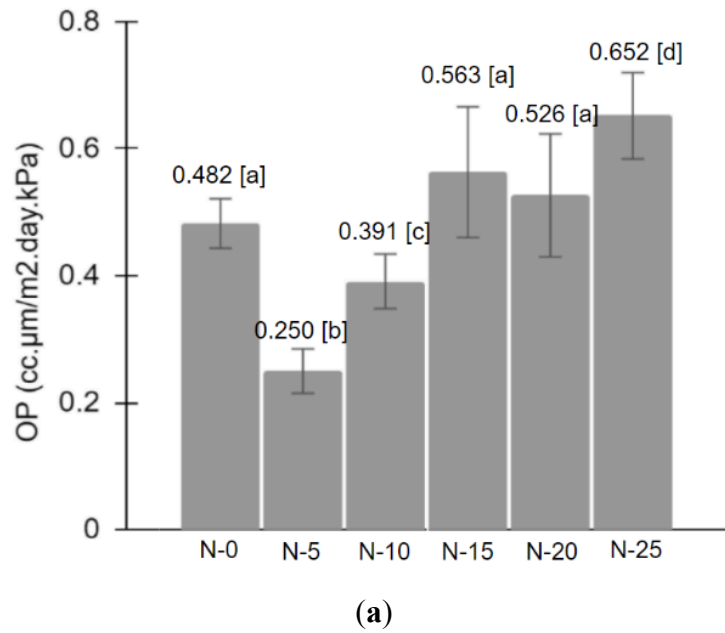
(b)

**Fig. 3.5.** Transparency of CNF films: (a) UV-vis spectra and (b) visual observation

The homogenization effect on the transparency of the CNF films was also analyzed, supporting the morphological and surface roughness results. Optical properties help determine major appearance quality that governs suitability for use in myriad applications. Transparency is directly related to the degree of homogeneity, internal microstructure, porosity, surface roughness, materials size, and distribution/dispersity (S. Cheng et al. 2015; Leppänen et al. 2022; Toivonen et al. 2018; H. Zhu et al. 2013). As seen in **Fig. 3.5**, either through UV-vis spectra (**Fig. 3.5(a)**) or simple visual observation (**Fig. 3.5(b)**), it is evident the homogenized films showcased better transparency due to the enhanced fibrillation (Dinesh and Kandasubramanian 2022). N-20 displayed the best transparency among all films, whose transmittance reached 56% at 900 nm. It is nearly thrice as transparent as the control film (N-0), where merely 20% of the transmittance of light is managed to pass through. This finding is superior to other nanocellulose reports (X. Sun et al. 2018; M. Herrera 2015; M. A. Herrera, Mathew, and Oksman 2012). Decreasing transparency was observed after 25 passes of homogenizations (N-25) due to the over-shearing effect. The fibrils agglomerated to the point where the bundles' diameter or width was significantly increased. It led to more scattered light and eventually decreased optical transmittance (E. Lizundia et al. 2016). Such a negative effect of agglomeration on nanocellulose films' transparency was also discussed in other reports (S. Cheng et al. 2015). The produced films exhibited high transmittance not only in the visible spectrum but also high absorption in the UV spectrum. UV-blocking ability is important because excessive exposure to UV sunlight radiation increases biological damage and degradation of organic compounds. This property is responsible for the discoloration, weathering, and loss of mechanical properties (cracking) (Sadeghifar et al. 2017). The UV region covers the wavelength range 100-400 nm and is divided into three (3) bands: UV-A (320–400 nm), UV-B (280–320 nm), and UV-C (200–280 nm) (Parit et

al. 2018; WHO 2016). The most transparent film (N-20) partially blocked UV-A, B., and C, where the resulting transmittances at the aforementioned ranges were 9-15%, 15-23%, and 23-32%, respectively. On the other hand, the resulting transmittances of the least transparent film (N-0) in the three (3) UV spectrums were 9-13%, 6-9%, and 0.5-6% for UV-A, B, and C, respectively. These findings are superior to previous cellulose studies (Sadeghifar et al. 2017; Guzman-Puyol et al. 2022; Bian et al. 2022), showing the excellence of homogenized films in UV-blocking features despite the accomplished transparency. Both transparent and opaque CNFs have potential in a variety of applications, e.g., transparent wound dressing (Baş et al. 2023) and opaque thermal insulation film (Grabowska and Kasperski 2018; Pan et al. 2023) or opaque mulching film (N. Yang et al. 2015).

### 3.3.4. Barrier property



**Fig. 3.6.** (a) Oxygen permeability (OP) of CNF films at 50%RH; and (b) illustration of oxygen diffusion paths comparison (where blue blocks represents cellulose fibrils and red arrow represents oxygen flow)

**Fig. 3.6(a)** displays the oxygen permeability (OP) of the homogenized CNF films at 50%RH. The varying homogenization cycles and their influences on the oxygen diffusion path are illustrated in **Fig. 3.6(b)**. OP significantly decreased by 48% after five (5) cycles of homogenization, from 0.482 (N-0) to 0.250 (N-5) cc.μm/m<sup>2</sup>.day.kPa at 50%RH. This showed the impact of promoted fibrillation post-homogenization that provides a more hydrogen-bonding network between fibrils, creating a more densely packed structure (Peresin et al. 2017), as evidenced in morphological findings. The obtained OP values were compared with ethylene vinyl alcohol (EVOH), a commercial polymer with exceptional oxygen permeability. EVOH is usually applied in multilayer structures with other polymers through blow molding, thermoforming, and orientation techniques. Its composition dictates its performance, and 25–45 mol % ethylene content is required for high barrier resin standards (Artzi, Narkis, and Siegmann 2005). At 50%RH, it was revealed that the best oxygen barrier–CNF (N-5) possessed a noteworthy barrier performance compared to EVOH, falling between EVOH38 and EVOH44 (EVOH with 38 and 44 mol% ethylene, respectively), in particular H and E type of the EVAL™ brand (approximately 0.099–0.138 and 0.197–0.375 cc.μm/m<sup>2</sup>.day.kPa), respectively (Kuraray, 1972). OP of N-5 is also notably superior against several common synthetic oxygen barriers–plastics, i.e., PVA, MXD6, PET, and PVC at 0.197–9.869, 0.493, 10–50, and 20–79 cc.μm/m<sup>2</sup>.day.kPa, respectively, even some PVDC (0.099–2.961 cc.μm/m<sup>2</sup>.day.kPa), at 50%RH (Michiels, Van Puyvelde, and Sels 2017). This result is comparable to the findings of the latest CNF report (Z. Cheng et al. 2024). Nonetheless, the OP increased when more cycles of homogenization were applied, which is presumably after fibril length reduction as the aftermath of “over-shearing.” Similar phenomena were reported where the initially observed long fibrils at small numbers disappeared with continued mechanical treatment, alongside the appearance of newly-produced fibrils (Redlinger-Pohn, Petkovšek, et al. 2022; Y. Zhou et al. 2018). Instead of the long fibrils with a high aspect ratio turning and entangling into a web structure, the hydrogen-bonded shorter fibrils and subsequent more void volumes made a significantly less tortuous diffusion path for oxygen (L. Wang et al. 2020). Therefore, despite the promoted fibrillation, N-10 and beyond suffered from lower oxygen barrier ability.

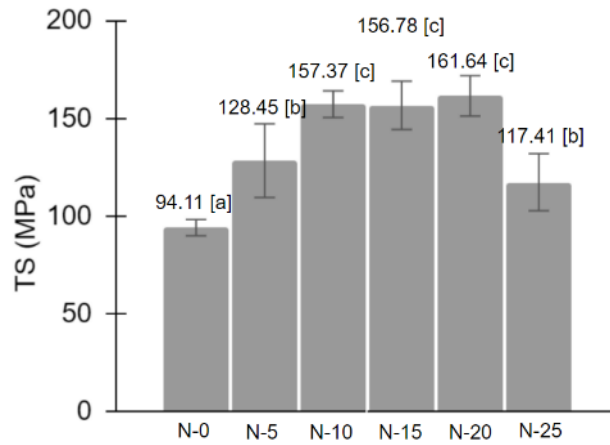
The high oxygen barrier property is among cellulose’s diverse advantages that have gained attention for centuries. Oxygen permeability (OP) is one of the most important functionalities in food/pharmaceutical packaging as it dictates the material’s ability to reduce/prevent oxygen permeation, resulting in responsible for many degradation processes (e.g., lipid oxidation, microorganism growth, enzymatic browning, and losses in flavor, color, and nutrients)—it is even identified as the most common cause of packaged food’s quality loss (Brown 1992; S.-I. Hong and Krochta 2006; Ayranci and Tunc 2003). Despite the necessary presence of oxygen for any living tissue’s respiration, oxygen also accelerates ethylene production, leading to senescence (Oms-Oliu, Soliva-Fortuny, and Martín-Belloso 2008; Rojas-Graü et al. 2007). The oxygen barrier characteristics provide the degree of protection necessary to achieve the desired shelf life of a product whose oxygen sensitivity may vary (Koontz 2016). Moreover, under high humidity, the hydrophilic nature of cellulose leads to swelling and excessive moisture-absorbing tendencies of hydroxyl groups, allowing the water to replace hydrogen bonds between cellulose. This phenomenon eventually loosens the film’s

structure and allows gas, such as oxygen, to penetrate and diffuse. Several techniques have been explored aiming to solve this issue, but each is faced with different limitations. Combinations with other polymers through the coating (Bayer et al. 2011; Stanssens et al. 2011) or composite formation (Benhamou et al. 2015; D. Liu et al. 2014) may reduce biodegradability (Xia et al. 2018) or matrix-filler compatibility issues that might require surface modification (Aziz et al. 2023). Whereas the introduction of a hydrophobic group chemically, especially those reacting with hydroxyl groups (Jonoobi, Harun, Mathew, Hussein, et al. 2010; Missoum et al. 2013), is potentially retarding the existing hydrogen bonding in cellulose, subsequently affecting an array of major properties (Xia et al. 2018). Such an approach involving solvent exchange can also be tedious, reagent-intensive, and time-consuming (Peresin et al. 2017). Thermal modification was also reported (Xia et al. 2018), though the required high temperatures are likely destructive to some extent (L. Wang et al. 2021; Neto et al. 2021). Therefore, the effectiveness of a milder-conditioned homogenization approach in retaining the low oxygen transmission rate of cellulose, especially under humid conditions, was explored. OP measurement involves two (2) different gas streams, oxygen, and nitrogen, each at one side of the film. Nitrogen acts as the carrier gas of the oxygen, and a dedicated analyzer monitors the transmitted oxygen. OP is highly dependent on relative humidity (%RH), which dictates the amount of water molecules interacting with the tested sample, and temperature, which affects gas transference. To favor mobility and extensive mass transfer across the sample, controlled %RH and temperature shall be conducted (Bonilla et al. 2012; S.-I. Hong and Krochta 2006). Temperature at 23°C and 50%RH were chosen, which considerably deviated from the realistic chilled storage conditions (Jakobsen et al. 2005).

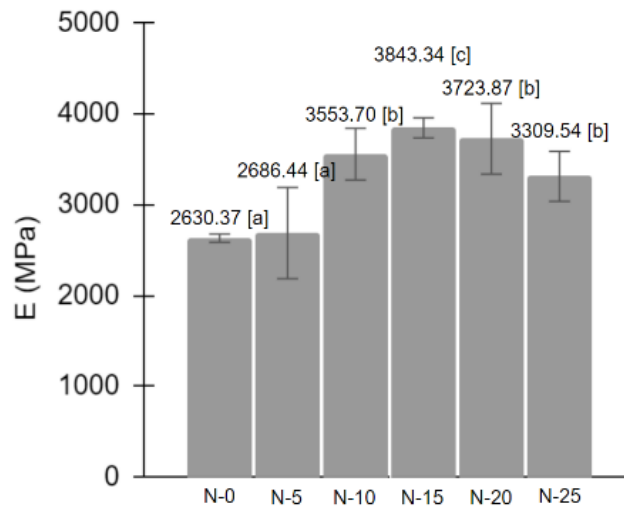
### 3.3.5. Mechanical properties

The mechanical properties of CNF are directly dependent on individual fibrils' strength. Fibril-fibril hydrogen bonding interaction is also important since it facilitates stress transfer and is influenced by dimensions, orientation, surface functionalities, counterions, and porosity. Strong fibril-fibril interfacial adhesion eases the stress distribution during deformation, improving the final film's strength due to the repeated breakage/reformation of hydrogen bonds between neighboring fibrils and the energy dissipation under stress (Mokhena et al. 2021). Other factors include preparation method, source, degree of polymerization, porosity, moisture content, fibril alignment, surface functionality, and counterions presence, which are often interdependent, thus making it difficult to separate their individual effects (Fukuzumi et al. 2013). **Fig. 3.7** displays the mechanical properties of CNF films: tensile strength or TS (**Fig. 3.7(a)**); tensile/Young's modulus or E (**Fig. 3.7(b)**); and elongation-at-break or EAB (**Fig. 3.7(c)**). The highest TS and E were achieved by N-10 (157.4 MPa) and N-15 (3843.3 MPa), respectively, which produced a significant 67% and 46% increase from the performances of untreated CNF (N-0). These optimum results proved the enhanced nanofibrils association within the polymer network and the increased resistance from rupturing. Furthermore, EAB was found the lowest at N-25 (3.90%) or a 31% decrease from N-0 (5.69%). EAB is highly influenced by aspect ratio; thus, CNFs with a lower aspect ratio post-homogenization are rigid and easy to pull out from the aggregated fibers (Pelissari et al. 2017; Saïd Azizi Samir et al. 2004). These mechanical analysis results are comparable with values

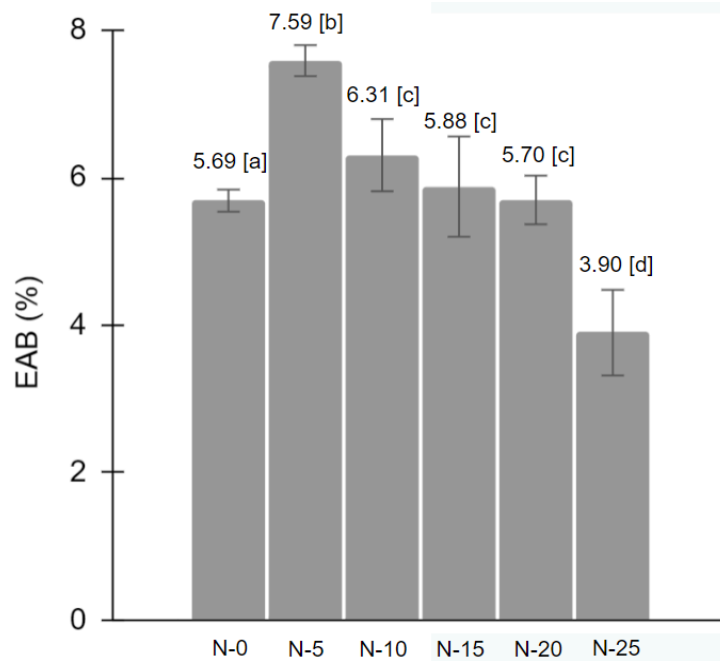
obtained in other reports: softwood CNF (TS = 98 MPa, E = ~3000 MPa) (Fein, Bousfield, and Gramlich 2021), kenaf CNF (TS = 75 MPa and E = ~1190 MPa) (Atiqah et al. 2019), eucalyptus CNF after 25 grinding passages (E = ~3600 MPa and tensile strain = 6.3%) (Malucelli et al. 2018), and banana starch composite reinforced with banana peel-CNF (TS = 11.1 MPa, E = 1047.7 MPa) (Pelissari et al. 2017). The observed ‘plateauing’ of mechanical properties after ten passes of homogenization (N-10) was similar to the phenomenon found in the previous homogenized cellulose reports (Naderi, Lindström, and Sundström 2015; Josset et al. 2014; K. L. Spence et al. 2010; Iwamoto, Nakagaito, and Yano 2007). Overall, N-25 showcased the effect of ‘over-shearing’ whose effects were highlighted as well in other properties such as morphological, surface roughness, transparency, and thermal stability (Fig. 3.3, Fig. 3.4, Fig. 3.5, Fig. 3.8(a-b), respectively) and discussed in each section. The apparent agglomeration after an excessive mechanical treatment impaired contact among the fibrils, weakening mechanical properties and making it more prone to rupture. Similar behavior was evident in other studies investigating the extent or severity of mechanical treatment to the CNFs' properties (Malucelli et al. 2018; Pelissari et al. 2017).



(a)



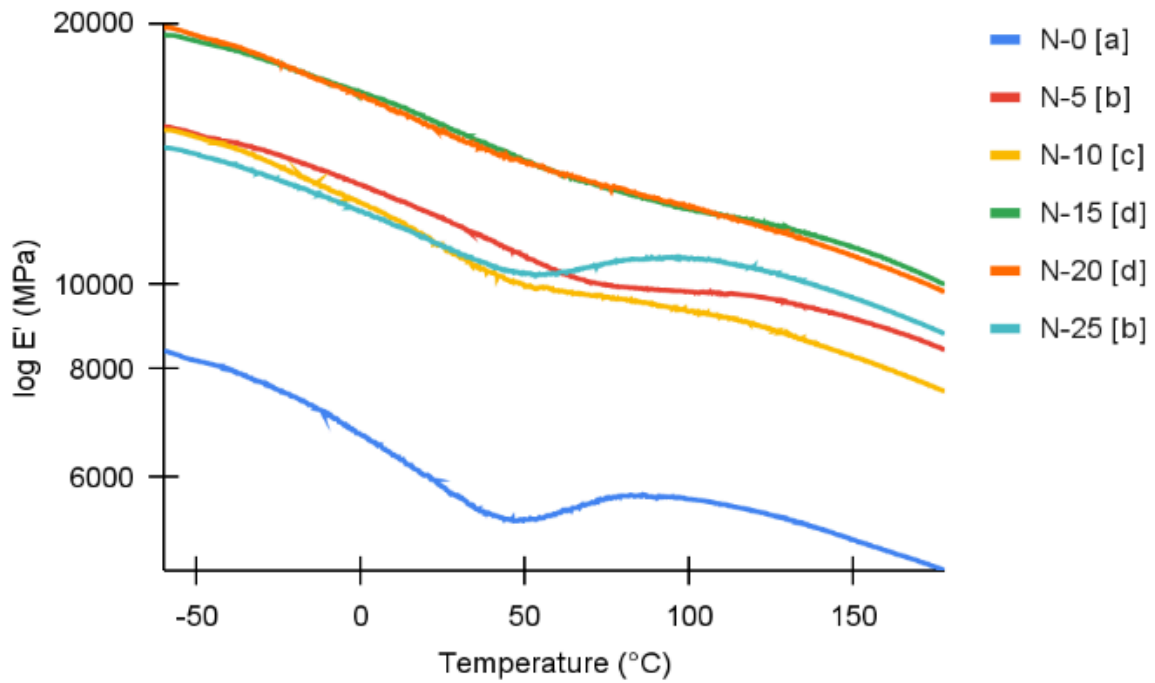
(b)



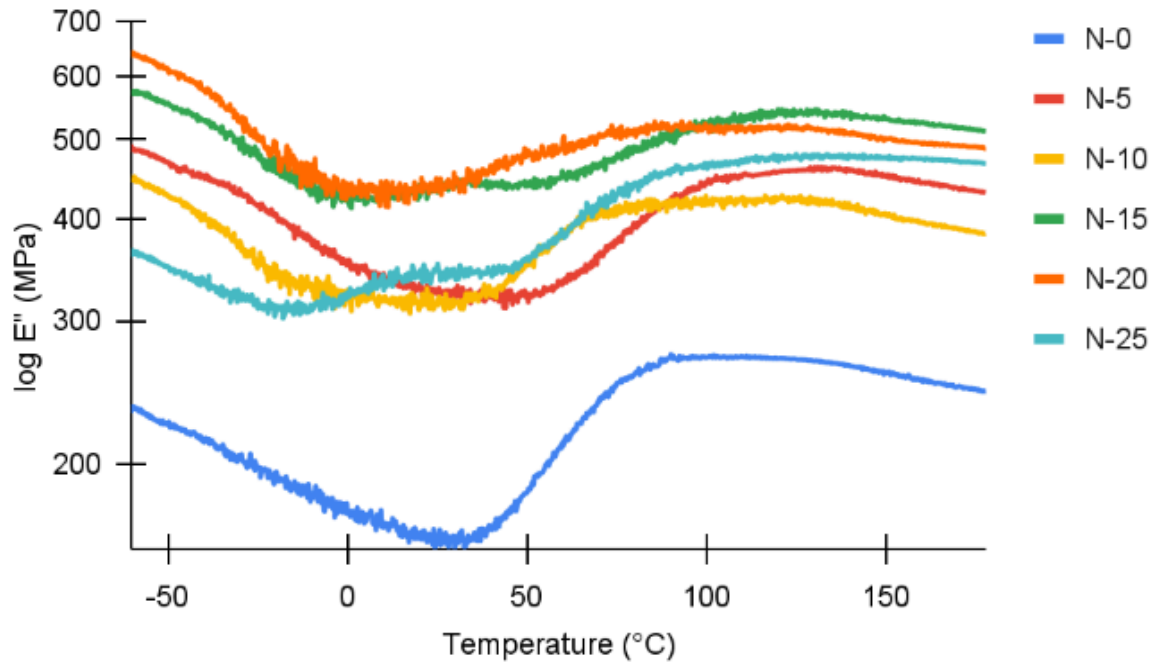
(c)

**Fig. 3.7.** Mechanical properties of CNF films: (a) tensile strength or TS, (b) tensile/Young's modulus or E, (c) elongation-at-break or EAB

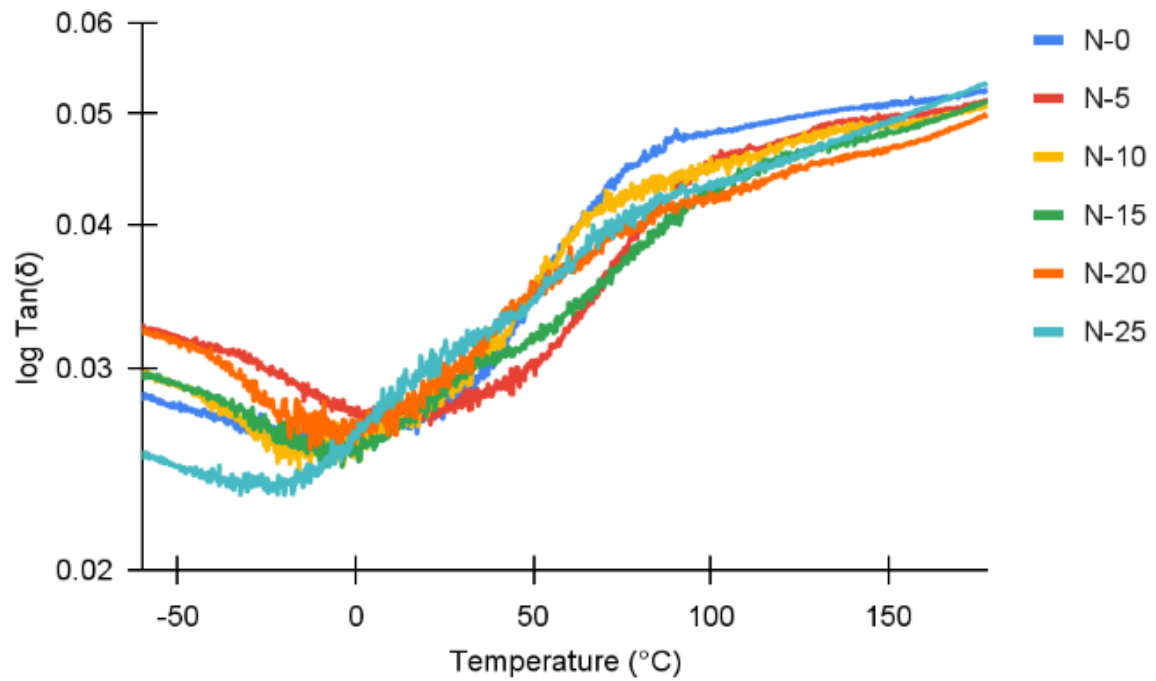
### 3.3.6. Thermal properties



(a)



(b)



(c)

**Fig. 3.8.** Thermomechanical properties of CNF films: (a) storage/elastic modulus [ $E'$ ], (b) loss/imaginary modulus [ $E''$ ], and (c) loss/damping factor [ $\text{Tan}(\delta) = E''/E'$ ]

**Table 3.2.** Thermomechanical properties of CNF films at 25 and 100 °C

CNF	T (°C)	E' (MPa)	E'' (MPa)	Tan( $\delta$ )
N-0	25	5855	160	0.028
	100	5668 [a]	270 [a]	0.048 [a]
N-5	25	11,916	324	0.028
	100	9811 [b]	444 [b]	0.046 [b]
N-10	25	11,092	314	0.028
	100	9315 [c]	427 [c]	0.046 [b]
N-15	25	15,265	434	0.029
	100	12,199 [d]	520 [d]	0.043 [c]
N-20	25	14,953	449	0.030
	100	12,268 [d]	520 [e]	0.043 [c]
N-25	25	11,073	347	0.031
	100	10,722 [e]	468 [f]	0.044 [c]

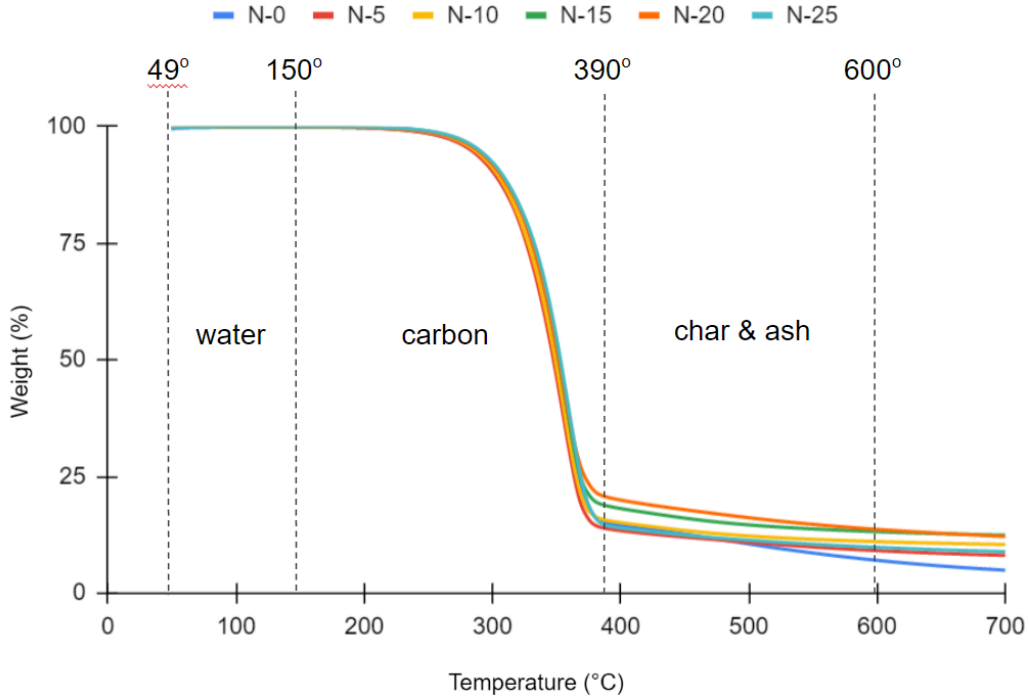
Thermal stability, be it thermomechanical properties (**Fig. 3.8**) or thermogravimetric weight loss (**Fig. 3.9**), makes a considerable difference to not only the material's processability and applicability but also degradability. Dynamic mechanical analysis (DMA) plots the storage/elastic modulus or  $E'$  (**Fig. 3.8(a)**), loss/imaginary modulus or  $E''$  (**Fig. 3.8(b)**), and loss/damping factor or  $\text{Tan}(\delta)$  (**Fig. 3.8(c)**), where the values obtained specifically at 25 and 100 °C were highlighted in **Table 3.2**.  $E'$  defines the material's strength, stiffness, and rigidity. It typically decreases as temperature increases, and at some point increases again, reaching a maximum value because of a change in network structure, before decreasing back after reaching the final cure due to thermal degradation and devitrification after glass transition temperature (Sanyang et al. 2016; Candan, Gardner, and Shaler 2016). An in-depth thermal characterization study on CNF also reported that, at low temperatures,  $E'$  remains roughly constant ( $\sim 2\text{--}3$  GPa) with temperature before a sharp modulus drop corresponding to the glass-rubber transition at  $-3\text{--}27$  °C. This drop corresponds to an energy dissipation phenomenon during the concomitant relaxation (Saïd Azizi Samir et al. 2004). Such a pattern is often also found in the  $E''$  plots (Ilyas et al. 2018), to be discussed in the next paragraph. At the N-0  $E'$  plot (**Fig. 3.8(a)**), a large improvement after the relaxation was observed, with a formed 'plateau' indicating entangled nanofibers (Jonoobi, Harun, Mathew, and Oksman 2010; Saïd Azizi Samir et al. 2004), as confirmed by morphological images (**Fig. 3.3(a)**). As the cycle of homogenization increased, the plateau began to fade due to the promoted fibrillation, hence the less and less tangling effect. However, at N-25, 'over-shearing' occurred, resulting in significantly shorter fibrils, in addition to some new fibrils broken apart from the initial structure. At this state, CNF became highly prone to be back entangled—hence, the reappearance of the 'plateau.' A similar phenomenon on barrier ability (**Fig. 3.6**) was discussed. Regardless, N-25 still produced a much higher thermal resistance than N-0, showing much higher  $E'$  values than N-0 at the same temperatures due to the enhanced hydrogen bonding. Additionally, it can be noted that all  $E'$  plots are significantly different, except the similarly-valued N-5–N-25 and N-15–N-20. After only five (5) cycles of

homogenization (N-5),  $E'$  was significantly increased by 104% (11,916 MPa) and 73% (9811 MPa) at 25 and 100 °C, respectively, compared to the untreated CNF (N-0), valued at 5855 MPa and 5668 MPa at 25 and 100 °C, respectively. Even better at N-15, the  $E'$  increase was by 161% (15,265 MPa) and 115% (12,199 MPa) at 25 and 100 °C, respectively. These evidenced the enhanced heat stability and incorporated rigidity due to stronger interfacial interactions post-homogenization. These values are on par with previously-researched materials, e.g., PLA-CNF (4400 MPa and 2500 MPa at 25 and 70 °C, respectively) (Jonoobi, Harun, Mathew, and Oksman 2010), PVAc-CNF (3485 MPa and 32 MPa at 30 and 70 °C, respectively) (Gong et al. 2011), CNF-epoxy (~2000 MPa at 25 °C) (Saba et al. 2017), and sugar palm-based CNC-starch composite (~1600 MPa and ~800 MPa at 25 and 100 °C, respectively) (Ilyas et al. 2018).

The value of  $E''$  (loss/imaginary modulus) represents the material's viscous response by measuring energy loss under stress or deformation as heat/cycle, hence its association with 'internal friction' (Saba et al. 2017; Jawaid et al. 2015). As mentioned, an identical trend between  $E'$  and  $E''$  plots was apparent. **Fig. 3.8(b)** shows  $E''$  plots that are all significantly different. After only five (5) cycles of homogenization (N-5),  $E''$  rose by 102% (324 MPa) and 64% (444 MPa) at 25 and 100 °C, respectively, compared to N-0, valued at 160 MPa and 270 MPa at 25 and 100 °C, respectively. The highest improvement was achieved after 20 cycles (N-20), with a 180% (449 MPa) and 93% (520 MPa) increase at 25 and 100 °C, respectively. These values are superior to the optimum results of other similarly-designed studies, such as sugar palm-based CNC-starch composite (~150 MPa and ~100 MPa at 25 and 100 °C, respectively) (Ilyas et al. 2018) and CNF-epoxy (~170 MPa at 25 °C) (Saba et al. 2017). Higher  $E''$  values emphasize a more homogenous dispersion/distribution and less visible agglomeration (Saba et al. 2017; S. Xu et al. 2013).

The loss/damping factor or  $\text{Tan}(\delta)$  (**Fig. 3.8(c)**) measures the ratio of  $E''$  to  $E'$ . Regarding the material's molecular movement, it characterizes the recoverable strain energy in the distorted specimen, providing valuable information about internal energy dissipation when exposed to varying temperatures (Sanyang et al. 2016). As the temperature increases, damping increases to its maximum in the transition region and decreases in the rubbery region (Saba et al. 2017). The higher the peak intensity, the higher the degree of molecular mobility; the lower the peak intensity, the slower the energy dissipation process (G. Chen et al. 2008), which suggests better dispersion that effectively inhibits polymer movement. Agglomeration increases the vacant space for the polymer chain to move/rotate, thereby increasing the visco-elastic damping behavior (Saba et al. 2017). The increase in  $E'$ , together with a positive shift in the  $\text{Tan}(\delta)$  peak, can be attributed to the promoted interaction that restricts the segmental mobility of the polymer chains. Viscoelastic behavior agrees with the mechanical properties (**Fig. 3.7**), where the reinforcing effect restricts molecular chain mobility (Bondeson, Syre, and Niska 2007; Jonoobi, Harun, Mathew, and Oksman 2010). These theories supported the findings, including N-0 showing the highest  $\text{Tan}(\delta)$  at high temperatures (~70-100 °C), indicating the most molecular mobility in the chains of the untreated CNFs that consisted of entangled fibrils. As the fibrillation was enhanced after more homogenization cycles, there became less moving space for the polymer chains, as confirmed by morphological images (**Fig. 3.3(a)**). Compared to the N-0  $\text{Tan}(\delta)$  peak at 100 °C (0.048), a significant decrease of 5% and 10% were observed at N-5 (0.046) and N-25 (0.044), respectively.

These results are comparable to other reports, e.g., PLA-CNF (~0.4 at 90 °C) (Jonoobi, Harun, Mathew, and Oksman 2010), PVAc-CNF (~0.4 at 90 °C) (Gong et al. 2011), CNF-epoxy (~0.5 at 100 °C) (Saba et al. 2017), and sugar palm-based CNC-starch composite (~0.2 at 100 °C) (Ilyas et al. 2018).



**Fig. 3.9.** Thermogravimetric weight loss profile of freeze-dried CNFs

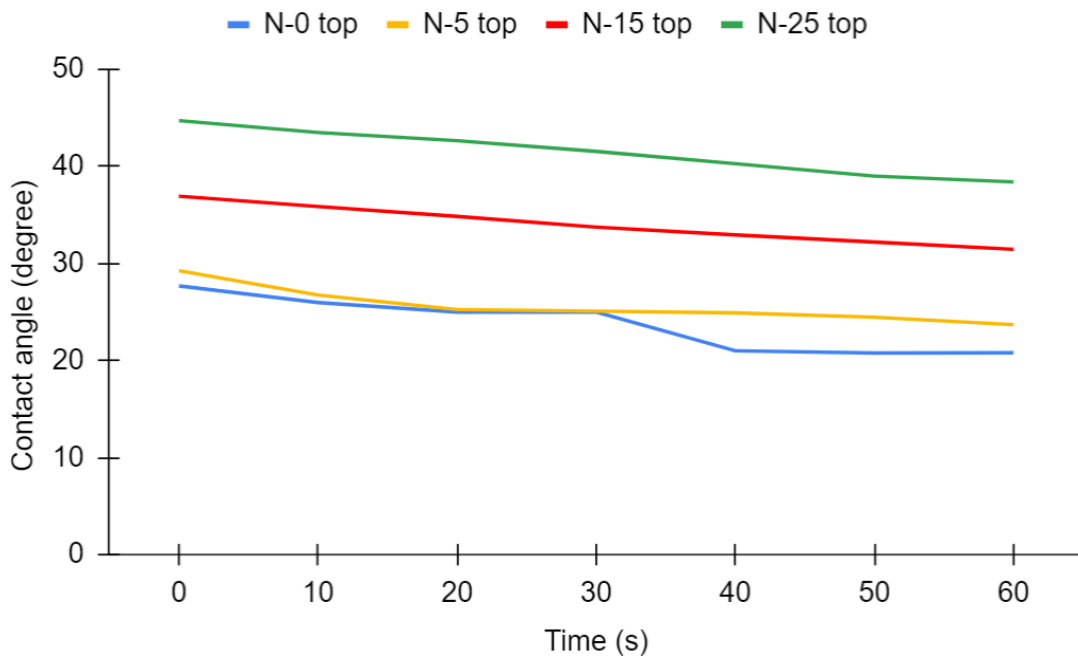
**Table 3.3.** Degradation temperature and weight loss profile of freeze-dried CNFs

CNF	$T_{d5\%}$ (°C)	%W at 700°C
N-0	281	5
N-5	281	8
N-10	283	11
N-15	287	12
N-20	286	12
N-25	287	9

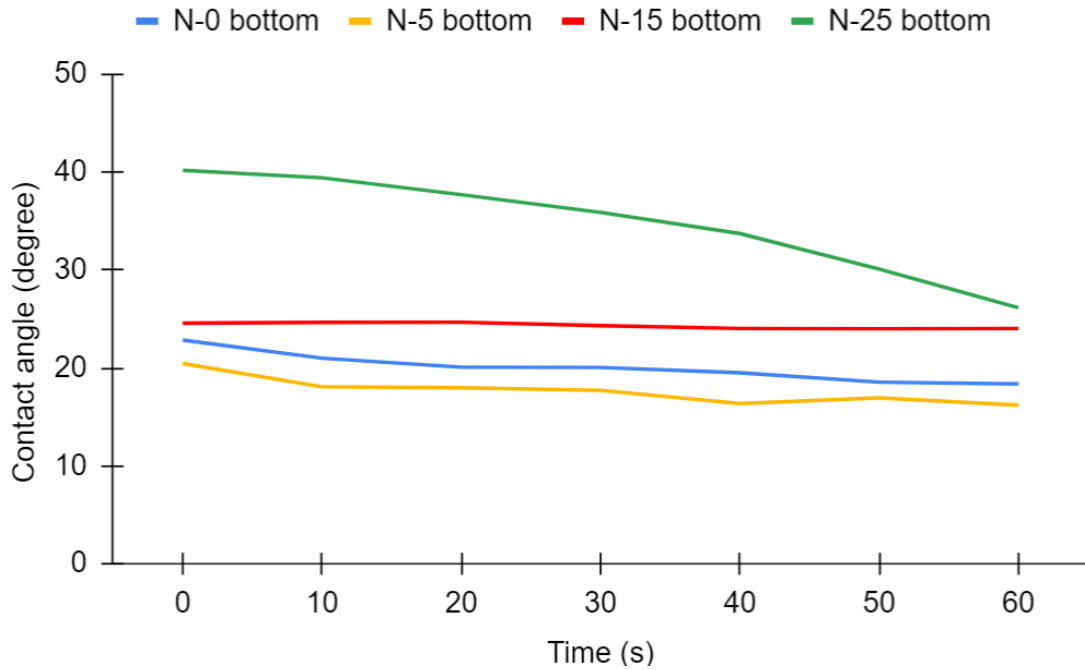
Thermogravimetric analysis (TGA) provides insights into thermal stability and decomposition behavior by measuring mass loss as a function of temperature or time. Thus, it can be crucial when determining a material's (bio)degradability (Brunšek et al. 2023). Regarding the TGA results (**Fig. 3.9** and **Table 3.3**), it is worth noting that the first decomposition of CNFs generally takes place at temperatures as low as 40-47 °C due to the absorbed moisture, typically through the evaporation or dehydration of loosely bound water contained in the CNF films. Since freeze-dried CNF was used for the measurement, the first decomposition was not obviously initiated until around the water boiling point at 100 °C (B. Yang et al. 2019; Ilyas et al. 2018). However, N-0 still displayed the earliest decomposition, implying the water's highest evaporation effect due to its still-natural high hydrophilicity (Abral et al. 2020), confirmed in the hydrophobicity discussion in the next

section. The second significant weight loss was next investigated, which is typically caused by cellulose decomposition at 260-360 °C, marking the start of pyrolysis (Abral et al. 2019; Araújo et al. 2018), followed by the third after 360 °C that can be attributed to the final, thermal-oxidative decomposition of char/ash (Asrofi et al. 2018; Jiangqi Zhao et al. 2013). An improvement in thermal resistance was evident in the TGA results, where post-homogenization, CNFs did not fully degrade even after reaching 800 °C and maintained 8-12% of their initial weight. It is a substantial resistance compared to the untreated CNF that reached the same weight loss at a far lower temperature and only maintained a little over 5% of its initial weight at 800 °C. This is owing to the enhanced amount of interlinking hydrogen bonding between the polymer chains post-homogenization, forcing a higher energy requirement to be decomposed (A. Islam, Yasin, and Rehman 2014; U.-J. Kim, Eom, and Wada 2010). Temperatures, when the samples endured the 5%-weight loss (Td,5%), were enhanced by over 10% post-homogenization (281-287 °C). Such finding was a significant improvement from other similar studies, including other softwood-based CNF (227.8 °C) (B. Yang et al. 2019), softwood-based raw cellulose (275 °C) (Fukuzumi et al. 2010), softwood-based TEMPO-oxidized cellulose (222 and 211 °C) (Fukuzumi et al. 2010), and oil palm empty-fruit-bunch based CNF (201 °C) (Supian et al. 2020). For heat-resistant applications, it is known that the recommended ambient temperature should not exceed 250 °C to ensure fit-for-use strength retention. Therefore, the obtained CNFs would suffice for the aforementioned applications (B. Yang et al. 2019). In addition, such thermostable films exhibit high potential as conducting films in touch panels, displays, lighting devices, flexible printed circuit boards, solar cells, and medical devices (L. Sun et al. 2023).

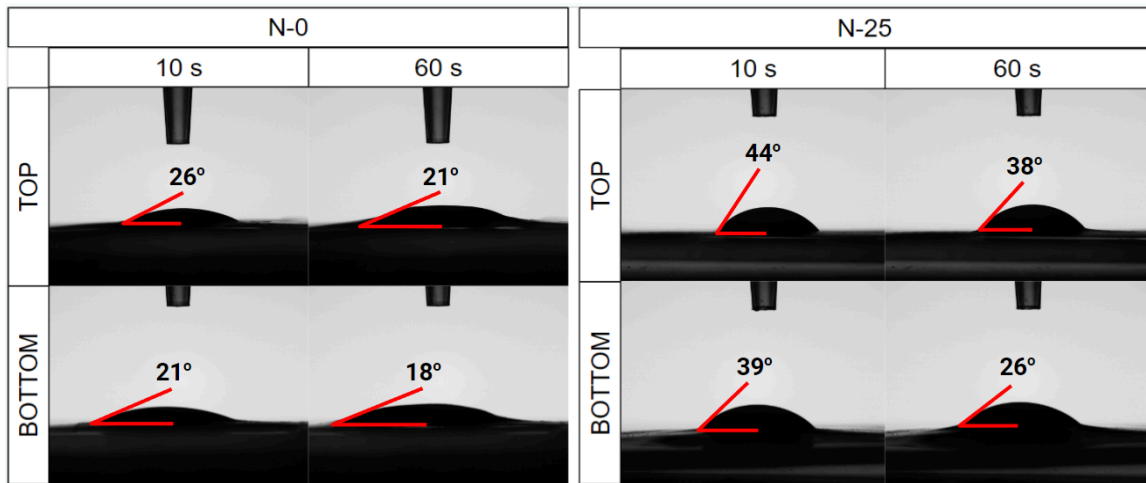
### 3.3.7. Contact angle or wettability



(a)



(b)



(c)

**Fig. 3.10.** Contact angle of the CNF film's surface : (a) top, (b) bottom, and (c) image comparison

Contact angle is the angle between the tangent to the droplet at the point of contact with the surface under test and that surface, as a measure of the hydrophobicity or wettability. The standard measuring liquid should be of low volatility so that its eventual evaporation does not affect the testing, not chemically reactive because that can cause the surface layer of the tested specimen to swell, and capable of producing droplets with sufficiently high wetting angle, identity, and repeatability, for the sake of testing ease and precision. The droplet volume should be small enough to eliminate the impact of

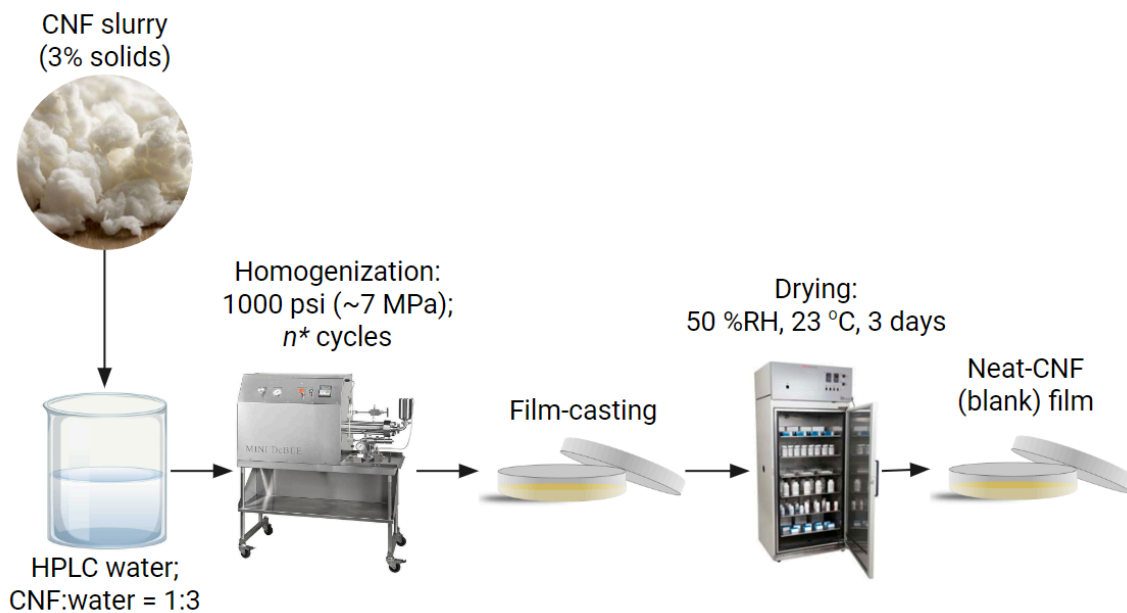
gravitational forces yet large enough to prevent the surface micro-roughness from obscuring the measurement. The sitting droplet method was chosen in this study due to its simplicity and low cost. Other influencing factors include physical/chemical homogeneity, roughness, contamination, droplet size, liquid dispenser speed, humidity, and ambient temperature (Jagodzińska et al. 2021; Wawro and Kazimierczak 2008). Studies have also discovered a variance in microstructure, porosity, and pore size between the top and bottom surface of the cellulose-based film due to the casting method; therefore, the top surface tends to be significantly more hydrophobic (Jagodzińska et al. 2021; McGaughey et al. 2020). **Fig. 3.10** showcased the effect of homogenization cycles on the contact angle of CNFs, where it indeed found that the top surface (**Fig. 3.10(a)**) was more hydrophobic than the bottom (**Fig. 3.10(b)**) regardless of the cycle of homogenization. It was observed that the more passes of homogenization, the higher the wettability. The highest hydrophobicity was performed by the top surface of CNF after 25 cycles of homogenization (N-25), with contact angles of 45° (after 10 s) and 38° (after 60 s), which is 62% (after 10 s) and 85% (after 60 s) increase from hydrophobicity of N-0 top surface. This wettability is comparable to that of other CNFs, for example, softwood (44.2°) and kenaf (33° after 10 s, 0° after 60 s) (Jonoobi, Harun, Mathew, Hussein, et al. 2010). Though surface roughness plays a crucial role in creating better hydrophobicity (Jonoobi, Harun, Mathew, Hussein, et al. 2010), **the high hydrophilicity nature of cellulose, owing to the plentiful polar groups, e.g., -OH and -COOH (Q. Wang et al. 2018), is also taken into account.** Studies have proven a negative relationship between the number of polar functional groups and the contact angle values (G. Zhou et al. 2015; X. Sun et al. 2021). Hence, despite possessing the roughest surface (as confirmed in **Fig. 3.4**), N-0 did not display the highest contact angle. Furthermore, N-5, being the least hydrophobic, is also aligned with its surface roughness findings that indicate its utmost smoothness. Hydrophobic CNFs are applicable for coating (S. Huang et al. 2022), while hydrophilic ones would be suitable for hydrogel/aerogel as a drug-delivery system (De France, Hoare, and Cranston 2017).

### 3.4. Conclusions

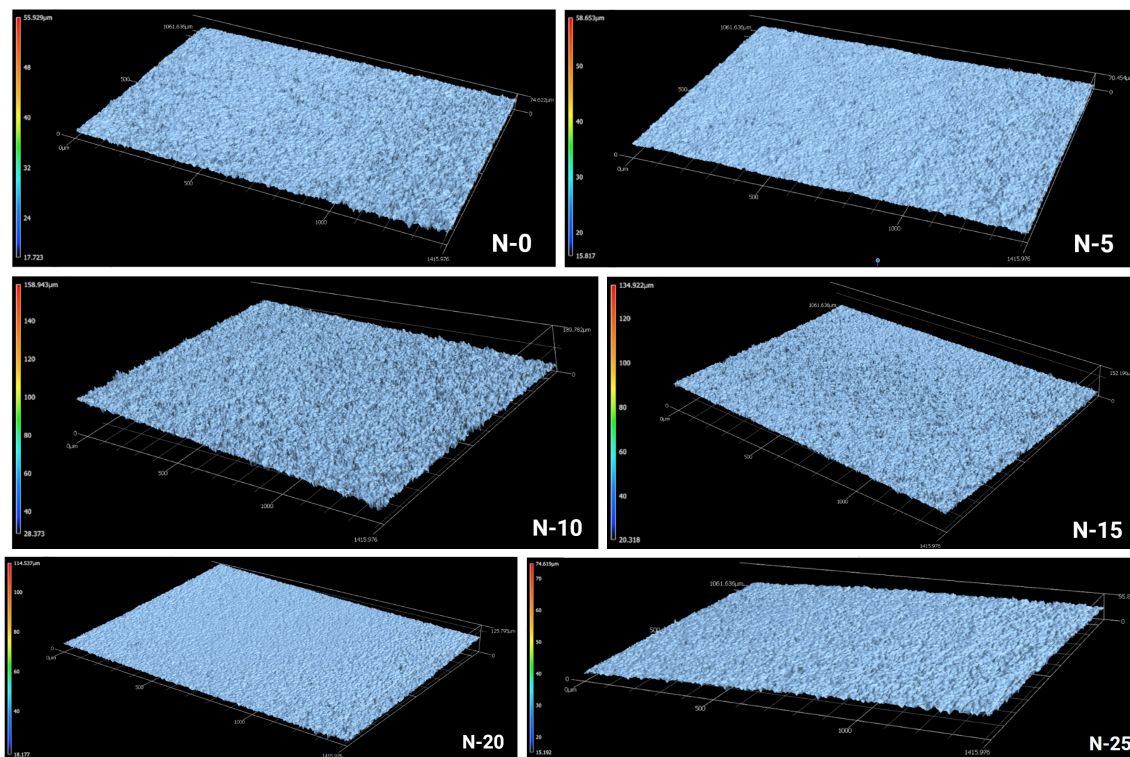
In summary, this work comprehensively investigated the effect of varying cycles of mild-conditioned homogenization on the properties of Northern bleached softwood-based CNF. This technique shows significant advantages over CNF in fibrillation and the subsequent hydrogen bonding, promoting several functional properties, such as surface smoothness (44% increase by N-5), OP (48% decrease by N-5), mechanical properties (TS increase of 67% by N-10; E increase of 46% by N-15; EAB decrease of 31% by N-25), thermal properties (E' increase of 161% at 25 °C and 115% at 100 °C by N-15; E'' increase 180% at 25 °C and 93% at 100 °C by N-20; Tan-δ increase of 10% at 100 °C by N-25; and T<sub>d,5%</sub> increase over 10% post-homogenization), hydrophobicity (62% after 10 s and 85% after 60 s by N-25), on top of the successfully-maintained high crystallinity (averaged CI = 64%). Those impressive improvements confirmed the prospect of the developed novel approach in preparing CNF, which was homogenization equipped with an 'emulsifying cell,' low pressure, and no plasticizer. These operating conditions would overcome the recurring complications in myriad other techniques in preparing CNF, e.g.,

costly and clogging issues in conventional high-pressure homogenizers that also destructs crystallinity; hydrophilicity nature of CNF that suffers from declining oxygen barrier ability over time under humidity; polymer blending through coating or composites formation that may retard biodegradability while still having to address the matrix-filler compatibility; chemical modifications that may disrupt the existing hydrogen bonding; solvent methods that can be tedious, reagent-intensive, and time-consuming; and a thermal process that can cause partial degradation. With the offered characteristics, the resultant CNFs prepared using a simple, fast, and cheap approach would be comparable to conventional oxygen barrier polymers like EVOH. As the optimum homogenization cycle differs per property, further development can be designed according to the intended applications, e.g., high oxygen-barrier and UV-blocking food/pharmaceutical packaging, smooth/glossy finishes, rough-surfaced semiconductor, transparent wound dressing, opaque thermal insulation or mulching film, strong/elastic/stretchy film, thermostable display/electronic conducting film, hydrophobic coating, and hydrophilic drug-delivery hydrogel/aerogel.

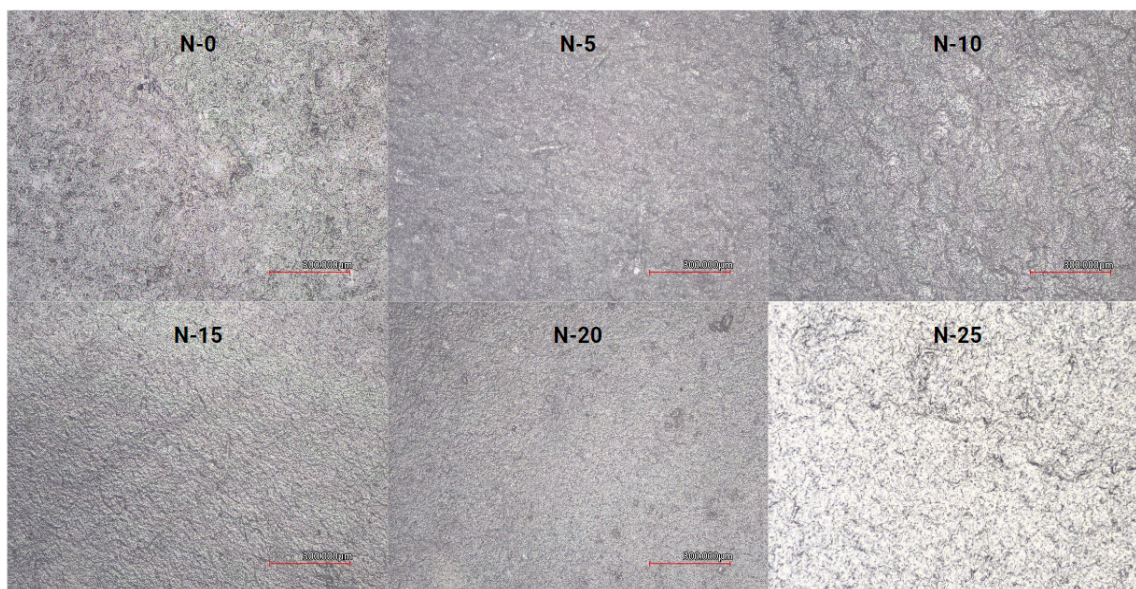
### 3.5. Supplemental Data



**Fig. S.1.** Schematic method of homogenized CNF films preparation



(a)



(b)

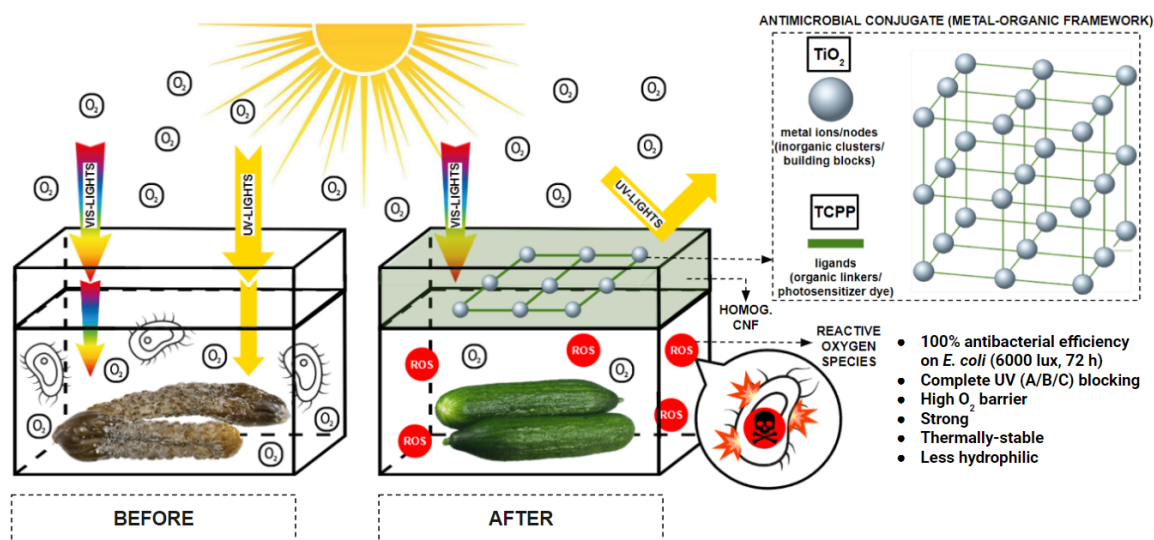
**Fig. S.2.** Surface roughness of CNF films: (a) 3D and (b) z-stack images

## CHAPTER 4

### Functionalization of nanocellulose film using $\text{TiO}_2$ -TCPP as photosensitive antibacterial agents

#### ABSTRACT

Freshness and quality of food may deteriorate due to infectious contamination of bacteria, including *Escherichia coli*, a common foodborne pathogen. Metal-organic frameworks (MOF) have drawn growing interest through their ability to combine light, oxygen, and photosensitizer to generate reactive oxygen species that can kill microorganisms via an oxidative burst. Titanium dioxide ( $\text{TiO}_2$ ) and tetrakis(4-carboxyphenyl)porphyrin (TCPP) were selected as the developed MOF's inorganic secondary building blocks and organic ligands/linkers, respectively, which were incorporated into a homogenized (0-25 cycles) cellulose nanofibril (CNF) matrix. TCPP's performance as a photosensitizer dye was proven by the resulting complete inhibition of *E. coli* growth (100% antibacterial rate) under 6000 lux-light irradiation after 72 hours by functionalized films prepared using homogenized CNF. Likewise,  $\text{TiO}_2$ 's UV-blocking ability was also evidenced by the complete UV-A/B/C absorption. In addition, surface smoothness, mechanical properties, thermal stability, and hydrophobicity of the functionalized films using homogenized CNF were improved, indicating the positive effect of enhanced fibrillation and the subsequent inter/intra-molecular hydrogen bonding post-homogenization. This work also certified cellulose's capability as a MOF carrier that solved its notoriously challenging self-aggregation/quenching tendency, as shown morphologically which confirmed the even dispersion of the  $\text{TiO}_2$ -TCPP conjugate. These findings overall exhibited a promising potential for a myriad of antibacterial film applications, including food packaging and biomedical.



Graphical abstract

## 4.1. Introduction

The freshness and quality of food can deteriorate due to infectious bacterial contamination. As it has been on the rise in recent years, advancement in the packaging field to tackle such concerns is of profound significance. Especially as using chemical disinfectants is not ecologically convenient after the occupational and operational risks that come with it. Traditionally, packaging is mainly made of synthetic materials derived from fossil fuel, meaning they are non-renewable and non-biodegradable. In this context, it has been increasingly encouraged to employ natural alternatives that are available in abundance, one of which is cellulose. It is named among the most abundant biopolymers in nature, with remarkable biocompatibility, low cost, and nontoxicity. In spite of the significant strengthening of the mechanical and barrier properties of nanocellulose-based packaging materials, the threat from pathogens may still remain. Nowadays, photosensitization or photodynamic inactivation has been proposed as an alternative antimicrobial modality. One of the emerging schemes is through metal-organic frameworks (MOFs); it combines light of a specific wavelength, molecular oxygen, and photosensitizer to generate reactive oxygen species (ROS), especially singlet oxygen ( $^1\text{O}_2$ ), that can kill microorganisms via an oxidative burst. The process involves electron-transfer reactions from the photosensitizer's triplet state to surrounding substrates, thereby facilitating cytotoxic species, such as superoxide anion, hydroxyl, and lipid-derived radicals (mechanism Type I) and/or energy transfer from the triplet state to ground state molecular oxygen ( $^3\text{O}_2$ ) to produce excited-state singlet oxygen ( $^1\text{O}_2$ ) (Type II) (Castro et al. 2019). No high heat nor harmful substances are required and/or released. MOF is a crystalline porous system consisting of ligands (as organic linkers) and metal ions/nodes (as inorganic clusters or building blocks) whose composition, structure, and porosity can be modulated to the desired functionality. It is also appealing for the ease of synthesis and surface modification, as well as the sustained releasing capacity. Porphyrin-based MOF is among the many photosensitive porous coordination networks with feasible structural tunability, stable crystalline structure, open channels, and biocompatibility. The highly-conjugated macromolecular and heterocyclic-structured porphyrin and its derivatives act as a photosensitizer to generate relatively high-yielding  $^1\text{O}_2$  for antibacterial packaging applications under simply visible light irradiation, as UV light may harm normal cells and tissues (Min et al. 2021; M. Chen et al. 2020; J. Park et al. 2016; X. Zhao et al. 2023). It can also help to avoid photosensitizer's quenching under the excited states or energy, as well as to enable facile accessibility of ROS diffusion over the rigid and porous network (K. Lu, He, and Lin 2014; Zheng et al. 2018).

Titanium (Ti) is one of the most studied examples of the inorganic metal ions in the case of porphyrin-based MOFs, where its high oxidative state leads to a preference to coordinate with the carboxyl functional group rather than the porphyrinic core. Titanium dioxide ( $\text{TiO}_2$ ) is among the common Ti precursors. As a photocatalyst, it has been utilized for its antimicrobial feature, including in food packaging, as it holds FDA approval as a "food contact substance." Its antibacterial activity is proven against a broad spectrum of microorganisms due to the high redox potential of the surface species formed by photoexcitation (under UV illumination), affording non-selective oxidative attack of microorganisms (El-Wakil et al. 2015).  $\text{TiO}_2$  is also attractive for its high availability, low price, safety (non-toxicity), non-corrosiveness, ease of production, excellent optical

properties, high surface areas, and stability in physical, chemical, and photoactivity aspects (Gao et al. 2019; Castro et al. 2019; Roberta Grazia Toro et al. 2020; An et al. 2016). Three (3) crystal structures of TiO<sub>2</sub> comprise rutile, brookite, and anatase; the latter possesses better photocatalytic activity and is most widely used in practical applications due to its wider band gap. However, the unsatisfactory photocatalytic activity of TiO<sub>2</sub> under visible light limits its applicability. Dye sensitization using porphyrin-based MOF as the organic bridging ligand is among the solutions, one of which is tetrakis(4-carboxyphenyl)porphyrin (TCPP). Some semiconductor studies highlighted the effect of incorporating such photosensitizer that enlarged the wavelength response range, enhanced the interfacial charge migration and separation, as well as inhibited the recombination of photo-generated electron-hole pairs in the photoreduction system—essentially improving the photocatalytic activity of TiO<sub>2</sub> under visible light region (Wan et al. 2016; Gao et al. 2019). Ti also boosts ROS generation, especially superoxide anion radicals, due to the improved photo-electron transfer from TCPP. The effectiveness was proven on Gram-positive/negative pathogens, besides the biocompatibility, minimal cytotoxicity, and minimal inflammatory effect (M. Chen et al. 2020; Tu et al. 2017). A study has determined the potential of TCPP dye-sensitized TiO<sub>2</sub> for solar-energy-converting electrodes. The strong adsorption of TCPP onto nano-TiO<sub>2</sub> was observed, as well as its efficiency as a photosensitizer. Effective coupling between the orbitals of the two materials was accomplished, allowing efficient electron transfer from the excited TCPP to the TiO<sub>2</sub> conduction band. Another advantage of porphyrins as one of the common photosensitizers is their strong absorption in both Soret (400–450 nm) and Q-bands (500–700 nm) (Cherian and Wamser 2000).

Cellulose has been explored as an ideal carrier for MOFs. Its repeat unit comprises a cyclic backbone of sugar rings with three hydroxyls on each ring alongside ether groups, where these high-density oxygenated functional groups provide the chemical functionalities of cellulose (Adel et al. 2021; Adel, El-Shafei, et al. 2019; Adel, Ibrahim, et al. 2019). The aforementioned hydroxyl groups present on the nanocellulose fibers' surface act as stable and robust anchoring sites as matrices for ex-situ loading and stabilization of pre-formed TiO<sub>2</sub> nanoparticles. Improved mechanical and barrier properties were also evidenced (Roberta Grazia Toro et al. 2020). In regards to MOFs (including TCPP) and TiO<sub>2</sub>, the problem lies in self-aggregation and self-quenching tendency, besides the remaining metal toxicity and complexity during recycling/reusing and separation from the reaction medium due to its nanoscale size (An et al. 2016). In this regard, incorporating them into polymeric nanofibers/nanotubes (as carriers) has attracted considerable attention in improving their processability and biosafety (Min et al. 2021; M. Chen et al. 2020; J. Park et al. 2016; X. Zhao et al. 2023; G.-Q. Liu et al. 2021). It can also help to avoid photosensitizer's quenching under the excited states or energy, as well as to enable facile accessibility of ROS diffusion over the rigid and porous network (K. Lu, He, and Lin 2014; Zheng et al. 2018). Embedding/immobilizing TiO<sub>2</sub> onto dispersive supporting materials with large surface areas (e.g., cellulose (An et al. 2016), graphene (Bhirud et al. 2015), and carbon nanofibers (M. Kim et al. 2014)), has been proven successful by previous studies in addressing TiO<sub>2</sub> agglomeration issue and subsequently enhancing the photocatalytic activity. The development of composite between nanocellulose (as matrix) and TiO<sub>2</sub>-TCPP (as photosensitive MOF) is expected

to promote a more efficacious dispersion of catalyst nanoparticles and enhance the functional properties of the as-synthesized composite (Roberta G. Toro et al. 2022).

Several reports found that Gram-positive and Gram-negative bacterial strains are susceptible to ROS produced by porphyrins; however, their efficacy highly depends on the used porphyrin structure and its charge. In particular, neutral, cationic, or anionic porphyrin derivatives can inactivate Gram-positive bacteria (Gomes et al. 2013), but an effective photo-inactivation of Gram-negative bacteria (without membrane disrupting agents) requires the presence of positive charges in the porphyrin structure. The relationship between the porphyrin surrounding charges and the capability to inactivate Gram-negative bacteria can be explained by considering the bacteria's physiology. While the Gram-positive bacteria cell wall is composed of a thick porous layer of peptidoglycan with attached teichoic acids, the Gram-negative bacteria cell wall includes a relatively thin layer of peptidoglycan surrounded by an asymmetric highly impermeable bilayer with an inner leaflet of phospholipids and outer leaflets of lipopolysaccharide. Each lipopolysaccharide imparts a strongly negative charge on the surface of Gram-negative bacterial cells, limiting the permeation of neutral and negatively charged photosensitizers (Delcour 2009). However, photosensitizers containing positive charges are able to promote a strong electrostatic interaction with the negatively charged sites at the outer surface of the bacterial cells, thus facilitating their accumulation in key subcellular compartments (Castro et al. 2019).

Many studies have explored the photosensitivity and antimicrobial potentials of CNF-TiO<sub>2</sub> (G.-Q. Liu et al. 2021; An et al. 2016), CNC-TiO<sub>2</sub> (Roberta G. Toro et al. 2022; Rathod et al. 2018), cellulose paper-TiO<sub>2</sub> (Roberta Grazia Toro et al. 2020), TiO<sub>2</sub>-TCPP (Cherian and Wamser 2000; Gao et al. 2019), pullulan-PVA-TCPP-thymol (Min et al. 2021), PP-CNC-TiO<sub>2</sub> (Zielińska et al. 2022), gluten-CNC-TiO<sub>2</sub> (El-Wakil et al. 2015), polycaprolactone-TCPP (X. Zhao et al. 2023), CNF-porphyrin (Fularz, Almohammed, and Rice 2021), CNC-porphyrin (Carpenter et al. 2012; Fularz, Rice, and Ballone 2021) and Zr-TCPP (J. Park et al. 2016) intended for varying applications from food packaging (El-Wakil et al. 2015; X. Zhao et al. 2023; Larrea et al. 2024; Min et al. 2021), solar-energy converter (Cherian and Wamser 2000), CO<sub>2</sub> photoreductor/converter (Gao et al. 2019), photodynamic therapy (J. Park et al. 2016; Carpenter et al. 2012), spectroscopy (Fularz, Almohammed, and Rice 2021), hydrogen generator (An et al. 2016), nanodevices (Fularz, Rice, and Ballone 2021), until micropollutants removal or dye wastewater treatment through adsorption/photodegradation (Rathod et al. 2018; Roberta Grazia Toro et al. 2020; G.-Q. Liu et al. 2021). Yet, no report on the photocatalytic disinfection properties of CNF films functionalized by TiO<sub>2</sub>-TCPP has been conducted. The objective of the present study is to be a step toward using active biobased packaging in food applications instead of petrochemical-based plastics by improving the functional properties, processability, and applicability of nanocellulose-based films. In this regard, the films' oxygen permeability, mechanical properties, thermal stability, morphology, roughness, opacity, wettability/contact angle measurements, and bactericidal inhibition rate against the pervasive pathogen *Escherichia coli* were investigated.

## 4.2. Materials and method

### 4.2.1. Materials

Cellulose nanofibrils (CNF) slurry with 3% solids derived from northern bleached softwood kraft pulp was obtained from the Process Development Center of the University of Maine (Orono, ME, USA). TCPP, labeled as meso-Tetra(4-carboxyphenyl)porphine, was purchased from Frontiers Scientific (Newark, DE, USA), while nano-titanium dioxide (TiO<sub>2</sub>, AEROXIDE® P25) was acquired from Acros Organics (Geel, Antwerp, Belgium). HPLC-grade water and 96%-ethanol (190 proof) were from EMD Millipore (Burlington, MA, USA) and Decon Laboratory (King of Prussia, PA, USA), respectively. Tryptic soy agar and broth (TSA, TSB) were purchased from Becton Dickinson (Franklin Lakes, NJ, USA). Phosphate Buffered Saline (PBS) was from VWR International (Radnor, PA, USA). All chemicals were analytical grade.

### 4.2.2. Homogenization process

The CNF slurry (3% solids) was diluted with water (1:3) by mixing under constant stirring (30 mins) to produce neat-CNF solution. The solution was further treated with a Mini DeBEE 30 homogenizer (BEE International, South Easton, MA, USA) with a 0.20 mm (0.008 inch) nozzle at 7 MPa, repeated for a different number of cycles (0, 5, 15, 25; or labeled as N-0, N-5, N-15, N-25, respectively). Homogenization cycles were maintained to be under 30 passes according to the preliminary studies, as well as referring to optimized results of previous studies on homogenized cellulose prepared at a medium/low conditioning pressure range (Chun et al. 2011; V. C. F. Li et al. 2018; Q. Cheng and Via 2017; Du et al. 2016; J.-C. Lee et al. 2018; T. Wang and Drzal 2012). On top of the internal cooling system, the homogenizer was equipped with an Isotemp Refrigerated Circulating Bath 910 chiller (Fisher Scientific, Waltham, MA, USA).

### 4.2.3. Antibacterial TiO<sub>2</sub>-TCPP conjugates preparation

TCPP (1 mmol) was mixed into 96%-ethanol before adding nano-TiO<sub>2</sub> (1% w/v of TCPP solution). The mixing was conducted while the solution was kept and covered in the dark overnight (75 °C). The antibacterial conjugate solution was prepared by mixing water and the TiO<sub>2</sub>-TCPP conjugate powder (0.2% w/v of water).

### 4.2.4. Film production

The homogenized neat-CNF (30 mL) and antibacterial conjugate solution (14 mL) were mixed using Mini Vortexer (VWR International, Radnor, PA, USA) on high speed for a few minutes to produce 2.2 mg/cm<sup>2</sup> functionalized-CNF film solution or labeled as F-0, F-5, F-15, and F-25. The mixture was then poured into a 9 cm x 1.5 cm petri dish using solvent casting technique and left to dry in the Forma Environmental Chamber (Thermo Scientific, Waltham, MA, USA) (50 %RH, 23 °C, 48 hours). The same preconditioning parameters were set before each test of the films.

#### 4.2.5. Characterizations

##### 4.2.5.1. Fourier transform infrared (FTIR) analysis

FTIR spectra indicating functional groups were determined by the Nicolet 8700 spectroscopy equipped with Attenuated Total Reflectance (ATR) (Thermo Scientific, Waltham, MA, USA) at a wave-number range of 400–4000  $\text{cm}^{-1}$  and a total of 32 scans. The results were reported as an average of three (3) replicates.

##### 4.2.5.2. Morphology, surface roughness, and opacity

Surface and cross-sectional morphology were characterized by the NeoScope JCM 5000 Scanning Electron Microscope (SEM) (JEOL Ltd., Tokyo, Japan). For surface SEM observation, the sample (25 mL) was initially prepared by lyophilization or freeze-drying after being kept in a -80 °C ultra-low freezer (Millrock Technology, Kingston, NY, USA). It was then mounted onto a pin stub mount using copper tape before being coated with roughly 100 Å of gold in an ion sputter coater (Denton Vacuum, Moorestown, NJ USA) using argon gas (10-15 psi) at pressure and current of 50 mTorr and 45 milliamps, respectively, for 45 seconds. Specimens were subsequently imaged at different magnifications under a vacuum at an operating voltage of 10 kV. Furthermore, the surface roughness of films was investigated using the Keyence VK-X3000 3D surface profilometer (Keyence Corporation, Osaka, Japan) referring to ISO 25178. Three-dimensional images of the film surfaces were analyzed with the Keyence VK-X3000 MultiFileAnalyzer (Keyence Corporation, Osaka, Japan) software to perform filtering according to the reference plane to the films' real surface to smooth out height variability before the roughness parameters measurement. Afterward, arithmetical mean height ( $S_a$ ) or the average absolute height value at each point in the definition area was reported as an average of three (3) replicates. Lastly, to determine opacity, transmission spectra of the films were carried out using a Genesys 10S UV-vis spectrometer (Thermo Scientific, Waltham, MA, USA) in the wavelength range of 190–900 nm with 0.5 nm interval, in addition to the visual observation

##### 4.2.5.3. Barrier property

Oxygen Permeability (OP) was characterized using the C230 Oxygen Transmission Rate Testing System (Labthink International Inc., Medford, MA, USA). The 700-118-20 Quick Mini digital thickness gauge (Mitutoyo, Kawasaki, Japan) to the nearest 0.001 mm was used to measure ten (10) random readings, which were then averaged. The plate was then inserted into the cell chamber and closed tight. The “Film-ASTM” test type without compensation was selected. To maintain purity, cycle process mode with rezero purging for  $\text{N}_2$  gas in the cells was performed for 30 seconds with one cycle interval. The system was preheated for 2 hours. The flow rate, test humidity, and test temperature were adjusted to 100 mL/min, 50%RH, and 23 °C respectively. Six (6) cycles under standard test mode were completed at 30-minute intervals in each cell. The results were reported as an average of six (6) replicates.

##### 4.2.5.4. Mechanical properties

Mechanical properties were measured with Material Test Systems 27-00112 (MTS Systems Corporation, Eden Prairie, MN, USA). The films' tensile strength (TS), tensile/Young's modulus (E), and elongation-at-break percentage (%EAB) were

measured with 0.1 N preload at a test speed of 0.40 mm/s until the films ( $2 \times 8$  cm) were ruptured. TS was calculated from the ratio of maximum load to the cross-sectional area of the film. Meanwhile, %EAB was the ratio between the increased length pre-rupture and the film's initial length. Lastly, E was the ratio between stress and strain from the linear portion of the stress-strain curve. The results were reported as an average of ten (10) replicates.

#### 4.2.5.5. *Thermal properties*

Thermal analysis was performed using a Thermogravimetric Analyzer (TGA) 550 (TA Instruments, New Castle, DE, USA). Pan loading-taring was conducted using the auto sampling technique using a platinum (100  $\mu$ L) pan. A sample (5-15 mg) was spread in the pan to maximize surface area. It was equilibrated at 50  $^{\circ}$ C, then ramped from 20  $^{\circ}$ C/minute to 700  $^{\circ}$ C. Temperature when the sample's weight loss reached 5% ( $T_{d,5\%}$ ) was recorded. Data analysis using the TRIOS or TA Universal Analysis (TA Instruments, New Castle, DE, USA) software was then performed. The results were reported as an average of five (5) replicates.

#### 4.2.5.6. *Contact angle or wettability*

The water contact angle or wettability was tested by the OneAttension<sup>®</sup> Theta Flow optical tensiometer (Biolin Scientific, Stockholm, Sweden). Using the "sessile drop" experiment type equipped with a 200  $\mu$ L pipette tip at 25  $\mu$ L/min dispenser speed, a drop (5  $\mu$ L) of HPLC-grade water was deposited onto the film surface to measure the contact angle in varying regions. "Light" and "heavy" phases were set as air and water (20  $^{\circ}$ C), approximately, which are selected to determine the density difference value for the surface tension measurement. The drop images and subsequent contact angles were obtained with a total duration of 60 seconds at ten-second intervals and reported as an average of three (3) replicates.

#### 4.2.5.7. *Antibacterial activity*

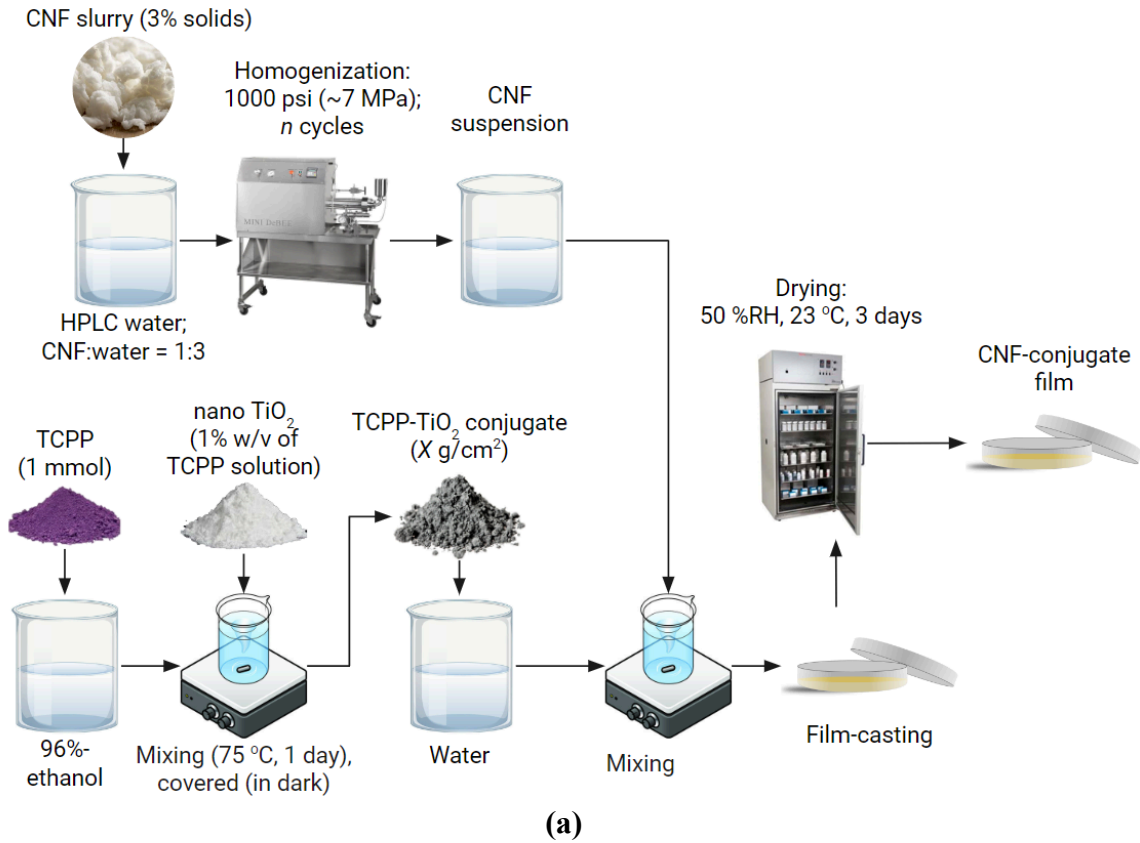
*Escherichia coli* (ATCC 25922) was selected as the representative bacteria to test the bactericidal efficacy of TiO<sub>2</sub>-TCPP conjugate-incorporated CNF films (F-films). The stock culture was inoculated in 10 mL of TSB in 15-mL centrifuge tubes and incubated at 37  $^{\circ}$ C for 16 hours until the Optical Density (OD) reached  $\sim 1.0$ . Bacteria were harvested by centrifugation at 5,524g for 5 minutes and re-suspended into 10 mL PBS solution (pH 7). After resuspension, 10-fold serial dilutions were performed to yield an *E. coli* concentration of  $\sim 10^5$  CFUs (colony-forming units)/mL. To start the antibacterial efficacy assessment, the initially prepared F-films were cut into 5/8-inch circular discs. The films were then placed into a sterile 24-well plate (VWR, Allentown, PA, USA), with a volume of 3.7 mL (diameter: 1.66 cm; height: 1.72 cm), completely submerged in 900  $\mu$ L of the prepared bacterial suspension on an orbital shaker (4  $^{\circ}$ C, 200 rpm) with an 800D LED 40W panel (GVM, Philadelphia, PA, USA) as the light source under different intensities (3000 and 6000 lux) and color temperature adjusted to 4100K, as confirmed using an SDL 4000 light meter (Extech Instruments, Nashua, NH, USA). Sampling was performed at a 24-hour interval for 72 to 96 hours. One duplicate dark control was also prepared by wrapping the 24-well plate with aluminum foil. After incubation, 100 mL of bacterial suspension was withdrawn from the well and plated onto TSA plates using an Autoplate 4000 spiral plater (Spiral Biotech Inc., Norwood, MA, USA). The TSA plates were

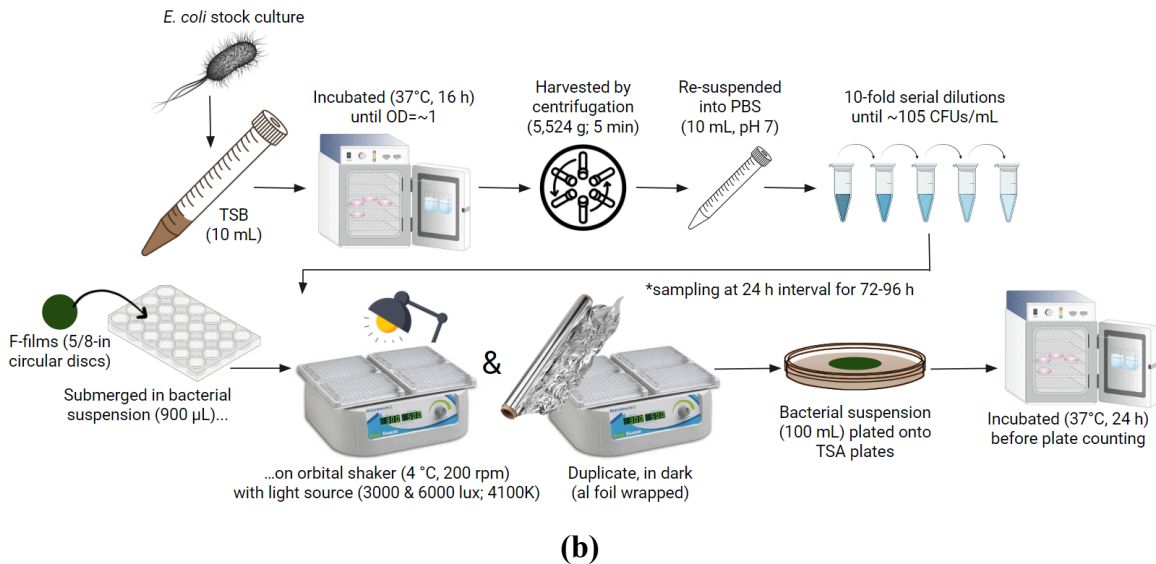
incubated at 37 °C for 24 hours, and plate counting was conducted to determine bacterial viability. The antibacterial rate was evaluated using the equation below (Mao et al. 2019; Tao et al. 2020; Min et al. 2021), where N is the average number of *E. coli* colonies calculated from control groups and experimental groups. The results were reported as an average of duplicates.

$$\text{Antibacterial rate (\%)} = \frac{N_{\text{control}} - N_{\text{sample}}}{N_{\text{sample}}} \times 100\%$$

#### 4.2.6. Statistical analysis

Data were analyzed by a one-way variance analysis (ANOVA) followed by Tukey's test and Student's t-test, using the Microsoft Excel 2019 (Microsoft Corporation, Redmond, WA, USA) and/or JMP 16 Pro (SAS Institute, Cary, NC, USA) software to establish significant differences (p-value < 0.05). The use of different letters on graphs data denotes statistically significant differences.

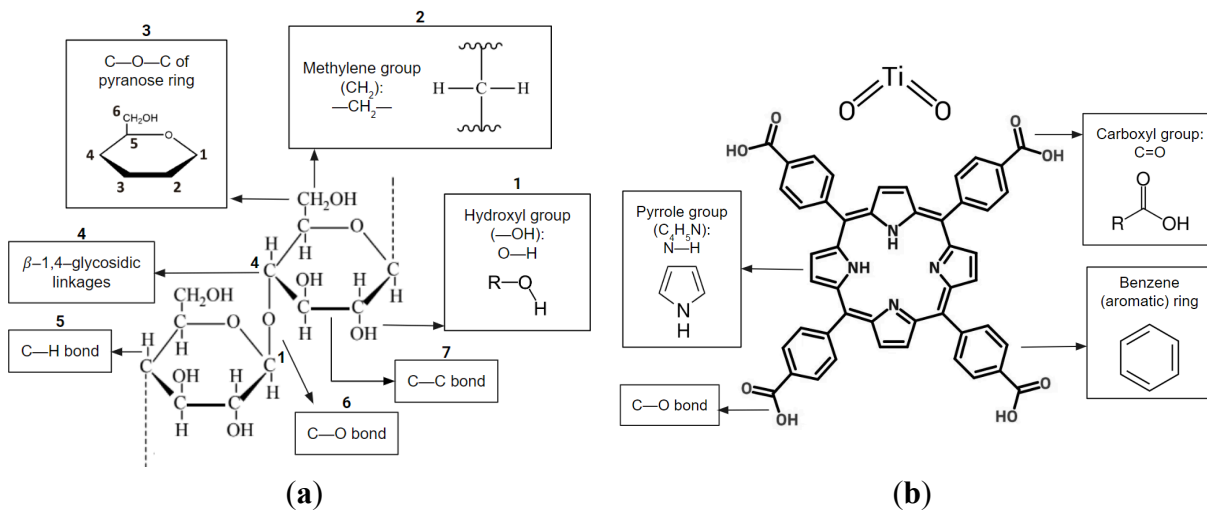




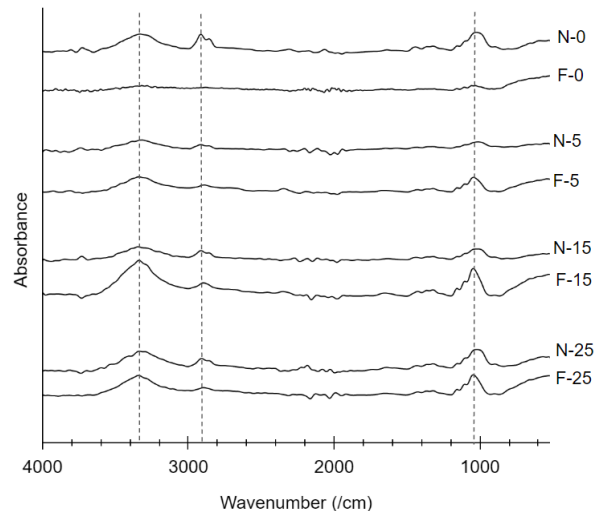
**Fig. 4.** Schematic methods: (a) CNF-conjugate preparation and (b) antibacterial activity

### 4.3. Results and discussion

#### 4.3.1. Fourier transform infrared (FTIR) analysis



**Fig. 4.1.** Chemical structure and major functional groups of (a) cellulose; (b) TiO<sub>2</sub> and TCPP



**Fig. 4.2.** FTIR spectra comparison between N-films and F-films

**Table 4.1.** FTIR peaks assignment (functional group and molecular vibration type)

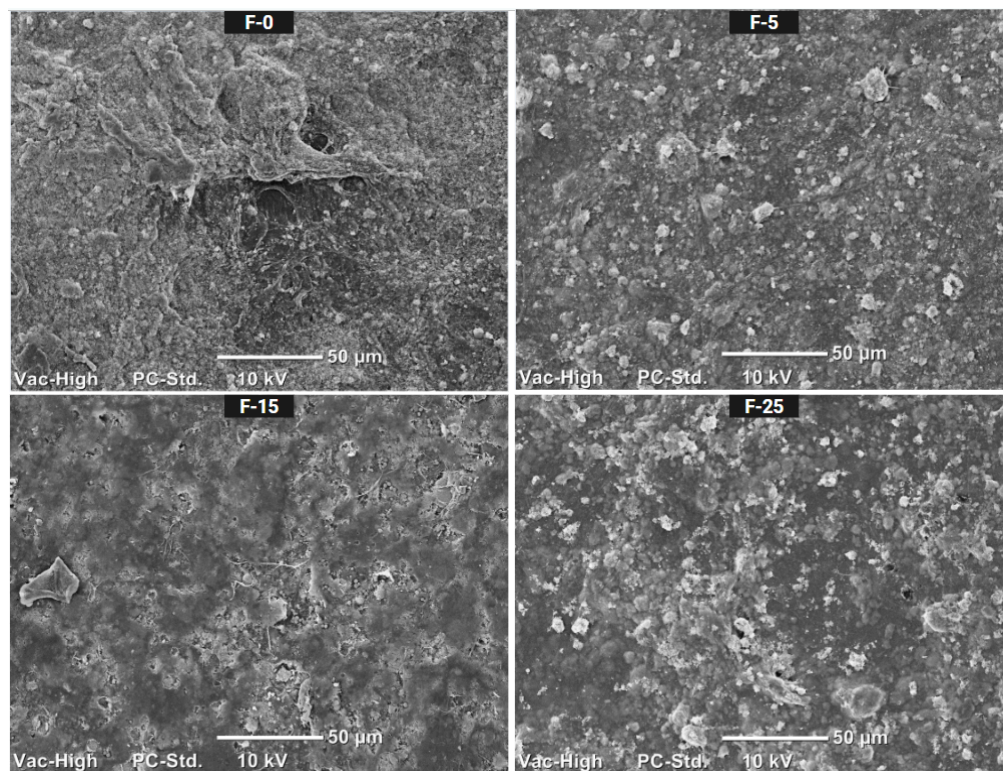
Approximate wavenumber (cm <sup>-1</sup> )	Functional group and molecular vibration type
3350	stretching of O—H (of —OH or hydroxyl groups)
2910	stretching/bending of N—H (of C <sub>4</sub> H <sub>5</sub> N or pyrrole group)
2900	(anti)symmetric stretching of C—H (of CH <sub>3</sub> /methyl or CH <sub>2</sub> /methylene group)
1650	bending of O—H and H—O—H (of H <sub>2</sub> O or absorbed water)
1600	benzene (aromatic) ring skeleton
1530	stretching of C=O or carboxyl group
1510	benzene (aromatic) ring skeleton
1450	benzene (aromatic) ring skeleton
1420	scissoring of C—H (of CH <sub>2</sub> or methylene group); cellulose's crystalline region
1395	benzene (aromatic) ring skeleton
1390	TiO <sub>2</sub> lattice
1370	bending of C—H and C—O
1310	wagging of C—H (of CH <sub>2</sub> or methylene group)
1160	stretching of C—O
1150	stretching of C—C
1130	Ti—OH
1050	antisymmetric stretching of C—O—C (of pyranose ring)
1045	Ti—O—C
1040	stretching of C—O
990	bending of C=C
970	in-plane bending of N—H (of C <sub>4</sub> H <sub>5</sub> N or pyrrole group)
890	cellulosic β-1,4-glycosidic linkages; cellulose's amorphous region
800	Ti—O—Ti
670	out-of-plane bending of C—OH
590	Ti—O—O

The chemical nature of surface functional groups in the functionalized film was investigated by FTIR spectroscopy. **Fig. 4.1** provides the chemical structure of the three components of the developed antimicrobial composite: cellulose nanofibrils (CNF) (**Fig. 4.1(a)**) and the conjugates (**Fig. 4.1(b)**), i.e., titanium dioxide ( $\text{TiO}_2$ ) and meso-Tetra(4-carboxyphenyl)porphine (TCPP). The presence of each of the highlighted functional groups was confirmed in the resulting FTIR spectra in **Fig. 4.2**; where the functional group and molecular vibration type assigned to each wavenumber are listed in **Table 4.1**. The identified stretching vibrations of (free) O—H or hydrogen bonds were present in phenolic and aliphatic structures (Md Salim, Asik, and Sarjadi 2021). These broad bands correlated to scission by the equatorially positioned hydroxyl groups of cellulose that form inter/intra-molecular hydrogen bonds between its sequential glucose units and with water, either as the solvent or existing moisture in the air (Wohlert et al. 2022; Tang et al. 2018). The same peak could also be ascribed to the O—H stretching vibrations, confirming the presence of hydroxyl groups on the  $\text{TiO}_2$  surface (Erdem et al. 2001). The (anti)symmetric stretching, scissoring, and wagging of C—H of methylene groups ( $\text{CH}_2$ ), as well as the C—H stretching of methyl groups ( $\text{CH}_3$ ) and C—O—C antisymmetric stretching, belonged to the pyranose ring of the cellulose's glucose units (Roberta Grazia Toro et al. 2020). Another band indicative of interstitial or adsorbed water was the bending of O—H and H—O—H bonds (Roberta Grazia Toro et al. 2020; An et al. 2016; Jiaoxing Xu et al. 2008). More typical absorption bands of cellulose were observed, contributing to  $\beta$ -1,4-glycosidic linkages and amorphous/crystalline regions (Maślana et al. 2021). These aforementioned bonds were present in all N-film and F-film samples, thus shown in their spectra. Furthermore, speaking of the conjugate incorporated into CNF,  $\text{TiO}_2$  characteristic wide bands are located below  $800\text{ cm}^{-1}$  wavelength and identified in the spectra, which attributed to the following bonds: Ti—O—Ti (Sui, Rizkalla, and Charpentier 2004; Choudhury and Choudhury 2012), Ti—O—O (Rajakumar et al. 2012), Ti—OH and Ti—O—C bonds (Kalaiarasi and Jose 2017; Maślana et al. 2021; Vasconcelos et al. 2011), and  $\text{TiO}_2$  lattice vibration (Nyquist and Kagel 2012). This finding verified the successful loading of  $\text{TiO}_2$  without destruction of chemical reactions between cellulose and  $\text{TiO}_2$ , as the building block of the developed metal-organic framework (MOF), onto the CNF substrate (An et al. 2016). Lastly, TCPP, which acts as the conjugate's bridging ligand, is a porphyrin type with four carboxyl groups on its exterior benzene rings, and the presence of such an aromatic ring skeleton (Reagen et al. 2022; L.-J. Feng et al. 2017; J. Wu, Wang, and Wang 2020) and carboxyl groups (Roberta Grazia Toro et al. 2020; J. Wu, Wang, and Wang 2020) were evidenced in the resulting spectra. Some other characteristic peaks of TCPP were too located in all the F-films spectra, which are assigned to the stretching and (in-plane) bending of the N—H bonds that belong to the pyrrole groups ( $\text{C}_4\text{H}_5\text{N}$ ) of TCPP (L.-J. Feng et al. 2017; J. Wu, Wang, and Wang 2020).

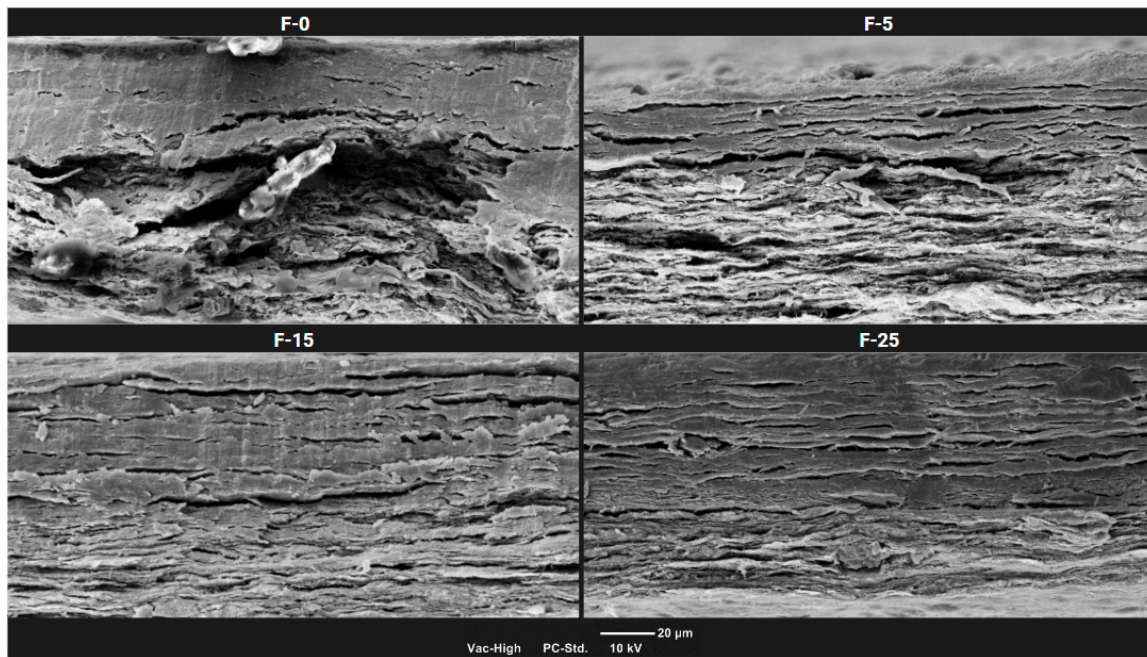
The position and intensity of FTIR bands are related to changes in the chemical surface groups (Hamad et al. 2018; Wade 2003). As seen in similar reports (D. Huang et al. 2020; Ilyas et al. 2019), the resulting spectra of N-films are identical, which indicates no chemical composition change occurred, meaning no new bonds formed post-homogenization. Meanwhile, for F-films, though mostly identical, different intensities were observed for the peaks around the  $3350\text{ cm}^{-1}$  region, which correspond to stretching vibrations of hydroxyl groups. Using unhomogenized CNF that contained

less-fibrillated cellulose fibrils, the TiO<sub>2</sub>-TCPP conjugates were not dispersed evenly. On top of the conjugate's self-aggregation and self-quenching tendency (An et al. 2016), this phenomenon led to much fewer hydrogen bonds being formed, either between CNF fibrils or between CNF and conjugates, as evidenced in the F-0 spectra. In contrast, F-5, F-15, and F-25 films produced stronger peaks in the said region due to the promoted hydrogen bonding from using the homogenized CNFs with enhanced fibrillation as the matrix; more specifically, strong interaction between —OH groups of CNF & Ti—O bonds in TiO<sub>2</sub> (Shu and Li 2012). Similar findings of strong hydrogen bond interaction between CNF and the loaded conjugates were reported (An et al. 2016; Zeng et al. 2010; Roberta Grazia Toro et al. 2020), which confirmed the fruitful immobilization in the matrix and low detachment tendency of the conjugate nanoparticles (Mohamed et al. 2015). In terms of the developed films' application as a visible light-responsive antimicrobial packaging, such an increase of —OH surface groups promoted hydroxyl radicals (•OH) formation during the photocatalytic process (de Almeida et al. 2020; Roberta G. Toro et al. 2022). As already known, hydroxyl radical is among the prominent reactive oxygen species (ROS) that exert microbicidal/bactericidal activity against a broad range of pathogens (Vatansever et al. 2013; Kohanski et al. 2007). In addition, the film's functionalization could also be proved by observing the C—H stretching vibration (2900 cm<sup>-1</sup>). Upon the incorporation of conjugates into CNF, the peak intensity decreased as a result of steric hindrance and blocking of C—H in-plane stretching (Roberta Grazia Toro et al. 2020). More in-depth analysis of these phenomena morphologically through the lens of microscopic images is discussed in the next section.

#### 4.3.2. Morphology, surface roughness, and transparency



(a)

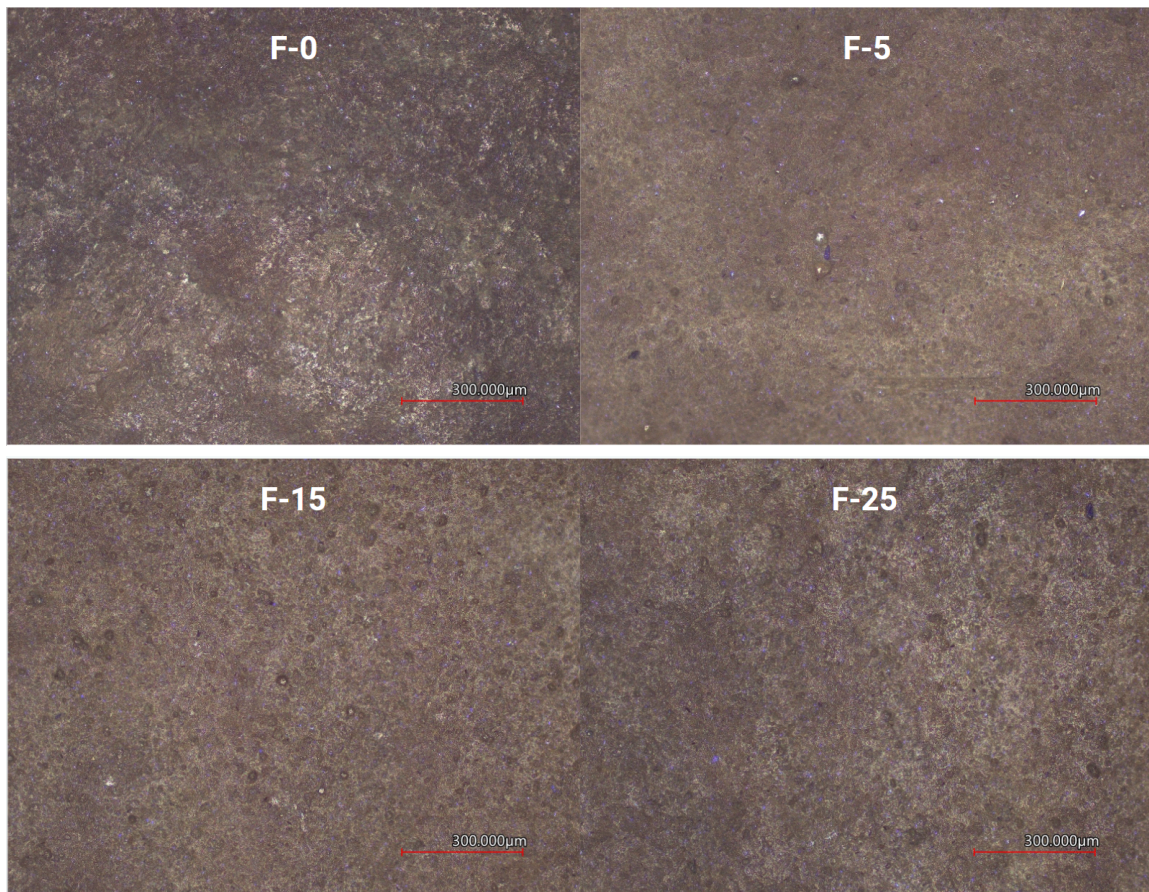


(b)

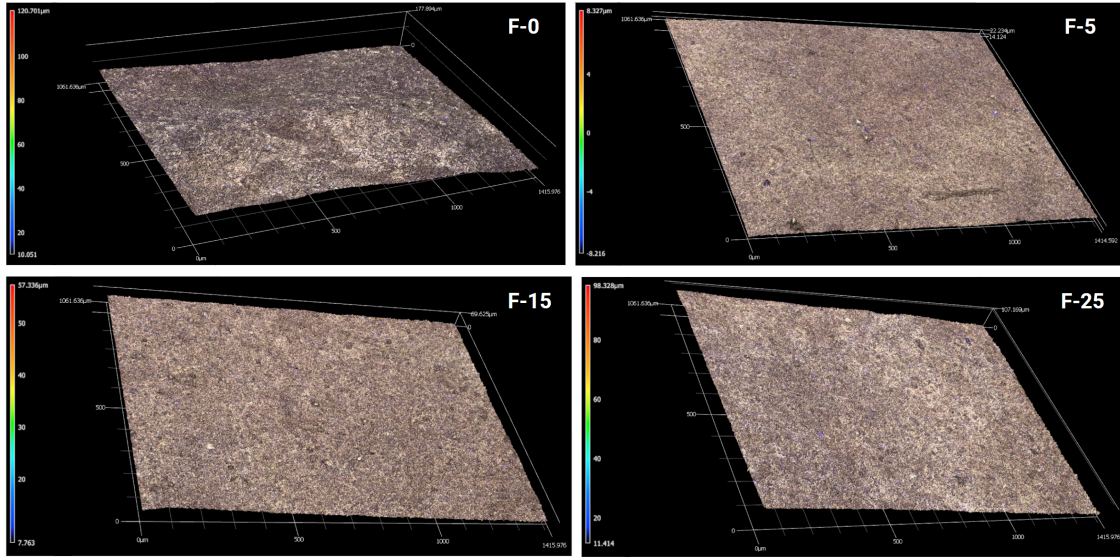
**Fig. 4.3.** SEM images of functionalized CNF films: (a) surface and (b) cross-section, at 500x and 1000x magnifications, respectively

The surface and cross-sectional SEM images of the functionalized CNF films post-homogenization are presented in **Fig. 4.3(a)** and **Fig. 4.3(b)**, respectively. Prior homogenization (F-0), surface observation (**Fig. 4.3(a)**) showed massive agglomerates of cellulose fibers and  $\text{TiO}_2$ -TCPP conjugates, creating a rough and coarse appearance of fragments (repeating units of cellulose) (Gautam et al. 2010; Redlinger-Pohn, Petkovšek, et al. 2022). This is led by the untreated cellulose fibrils pre-fibrillation that were still so strongly entangled, forming thick irregularly-shaped bundles that prevent an even dispersion of the incorporated conjugate granules throughout the matrix system, and many eventually aggregated since both  $\text{TiO}_2$  and TCPP, like the majority of MOFs, suffer from self-aggregation/quenching tendency issues, especially when nanosized (An et al. 2016). The naturally high surface energy of  $\text{TiO}_2$  also caused conjugates to agglomerate into big clusters (G.-Q. Liu et al. 2021). The effect of the hindered conjugate distribution could be explained better by the cross-section image (**Fig. 4.3(b)**). Conjugates could not maneuver through the not-fibrillated and compacted thick mass of cellulose fibrils and sank to the bottom of the petri dish when the film was being dried, creating two separate 'layers' where the bottom one was significantly rougher due to the accumulated conjugate aggregates. The conducted multi-cycled homogenization provides pressure drop, high shear, and impact forces that trigger mechanical release or fibrillation of the aforementioned cellulose bundles. The produced fine fibrils possess high surface areas, which means more inter-/intramolecular hydrogen bonding (Nagarajan et al. 2021; H. Lee, Sundaram, and Mani 2017; Chun et al. 2011). This allowed the formation of pores/cavities for inorganic nanoparticles like the conjugates to be dispersed more evenly and homogeneously (Zeng et al. 2010), as evidenced in the post-homogenization surface images (**Fig. 4.3(a)**), even after only five passes (F-5). Aggregates still remained,

although significantly less in number and size. At F-15, as the degree of fibrillation increased, so did the intermolecular hydrogen bonding and the high amount of conjugates dispersed within the numerous cavities that were responsible for the sheet/layer-like nature (Gautam et al. 2010; Zeng et al. 2010), as seen in F-15's surface image (Fig. 4.3(a)). Individual ultra-long-and-fine cellulose fibrils and evenly dispersed conjugate granules could be seen peeking out underneath. Cross-sectionally (Fig. 4.3(b)), 'layers' existing in F-0 became significantly less apparent the more homogenized the CNF was, indicating the more uniform scatter of conjugates throughout the matrix. The less aggregated conjugates also meant a smoother cross-sectional appearance. However, the 'over-shearing' effect was evident after 25 passes of homogenization (F-25). As also investigated in similar reports (L. Wang et al. 2020; Redlinger-Pohn, Brouzet, et al. 2022; Y. Zhou et al. 2018; Dilamian and Noroozi 2019; Kose et al. 2011), excessive mechanical treatment ended up dismantling the ultra-long-and-fine cellulose fibrils into shorter, smaller fibrils. The reduced aspect ratio, on top of the self-aggregation/quenching tendency, led to the return of agglomeration of cellulose fibrils and conjugates despite the promoted fibrillation, as seen in both surface and cross-sectional images of F-25 (Fig. 4.3(a) and (b)).



(a)

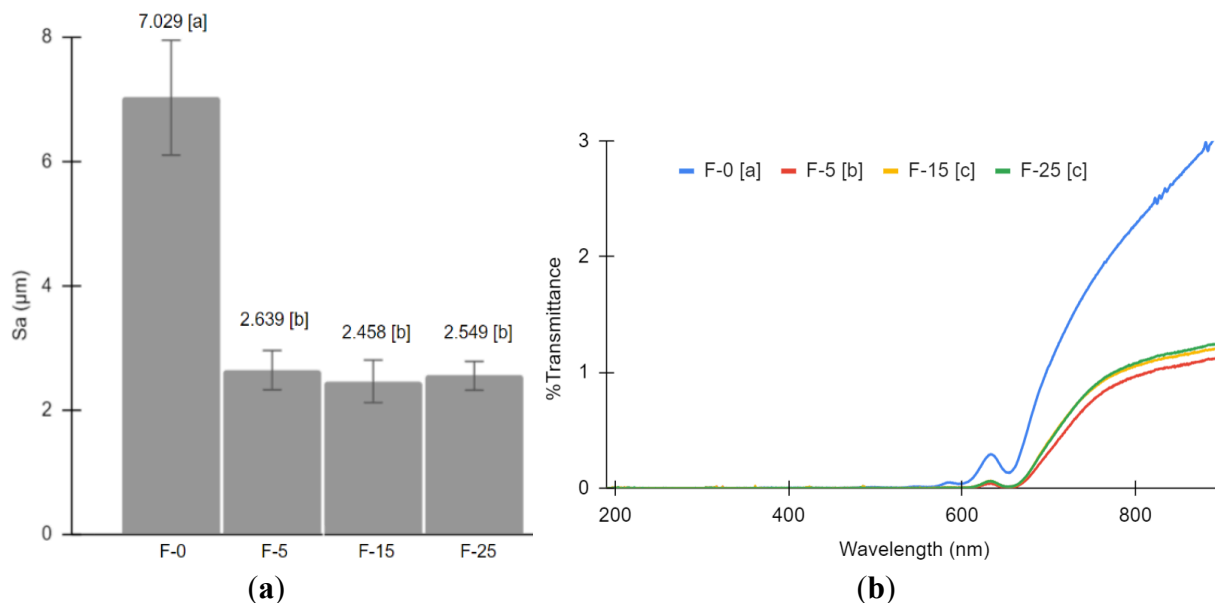


(b)

**Fig. 4.4.** Surface roughness of functionalized-CNF films: (a) z-stack and (b) 3D images

The images depicting the surface roughness property of the functionalized CNF films are presented in **Fig. 4.4**. On top of 3D microscopic images (**Fig. 4.4(a)**), Z/focus-stack images (**Fig. 4.4(b)**) were also taken to illustrate the roughness comparison through topography. It is by combining multiple images taken at different focal distances, producing a composite image with a greater depth of field (e.g., the plane of focus' thickness) than the individual source images (Ray 2002; D. Johnson 2008). Being a numerical and high-magnifying aperture whose major constraint is the decreasing depth-of-field, image processing through filtering (by referring the reference plane to the films' real surface to reduce height variability) could capture in-focus images (Larson and Banks 2022). The roughness values of the functionalized CNF films' top surface were expressed in Sa (**Fig. 4.5a**). This surface micro-geometry and irregularity characteristic is calculated by the arithmetic mean deviation ( $\mu\text{m}$ ) of the height of a line (Woch et al. 2022). Post-homogenization, the resulting range of optimum values (2.458-2.639  $\mu\text{m}$ ) was 65% smoother than the control film (F-0) (7.029  $\mu\text{m}$ ). This finding is comparable to similar studies (Miettinen, Chinga-Carrasco, and Kataja 2014; Xue et al. 2021; Żołek-Tryznowska et al. 2023). This phenomenon, similar to morphology (**Fig. 4.3**), indicates the relationship between the two characteristics—morphology and roughness (Chinga-Carrasco et al., 2014), and can be explained as the impact of the promoted fibrillation. There is no significant difference between the roughness of F-5, F-15, and F-25. Despite the starting phenomenon of oversharing at F-25 that produced shorter/smaller fibrils, the smoothness remained due to the inevitable effect of homogenization-induced fibrillation that still produced sufficient porosity and an even enough dispersion of the conjugates. It is important to note that the reason only the films' top surfaces were analyzed for roughness properties was due to the fact that the bottom surface conforms to the petri dishes' surface; hence, it would not suffice as well as the top surface as the roughness indicator (Miettinen, Chinga-Carrasco, and Kataja 2014). Regarding applicability, functionalized CNFs with desirable smoothness have the potential to be used as a glossy finish for packaging and laminating purposes. Meanwhile,

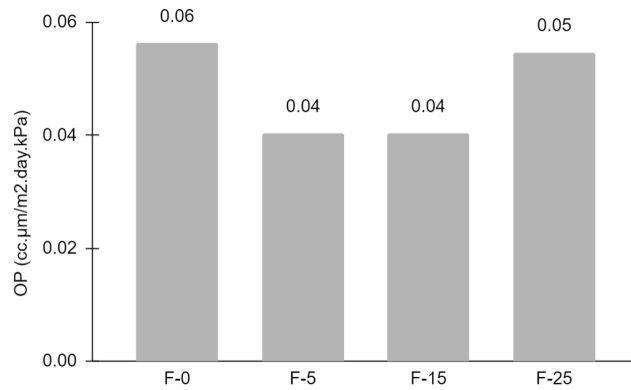
ones with rougher surfaces can be advantageous to be used in antimicrobial and self-cleaning smart textile surface coating with dye removal and water-oil separation purposes (Görgülüer, Çakıroğlu, and Özacar 2021).



**Fig. 4.5.** (a) Surface roughness (arithmetical mean height (Sa)); and (b) UV-vis spectra or opacity of functionalized CNF films

Lastly, following the morphological and surface roughness characteristics, the opacity of the developed functionalized CNF films was analyzed. Especially for films comprised of a matrix with incorporated conjugates, this property is often affiliated with the degree of distribution/dispersity and homogeneity (Alves et al. 2023; Dinesh and Kandasubramanian 2022). It is apparent from both UV-vis spectra results (**Fig. 4.5(b)**) and visual observations (**Fig. S.3**) that the functionalized films made with homogenized CNFs were far more opaque. This is after the effect of promoted fibrillation and inter-/intra-molecular hydrogen bonding post-homogenization—as proven in morphological results above (**Fig. 4.3**)—that allowed a better dispersion of the incorporated conjugates throughout the CNF matrix. The developed MOF consisted of  $\text{TiO}_2$  as the secondary building blocks linked and bound using dark-colored dye (TCPP); hence, the opaque appearance of the produced films. Generally, the functionalized film using F-5 was slightly more opaque than F-15 and F-20, which shows the effect of the number of homogenization cycles. The resulting spectra also suggested the complete UV-blocking-ability for all UV-A (320–400 nm), UV-B (280–320 nm), and UV-C (200–280 nm) (Parit et al. 2018; WHO 2016) by all functionalized films. Exposure to UV sunlight radiation increases the risk of biological damage, organic compound degradation, discoloration, weathering, and loss of mechanical properties (cracking) (Sadeghifar et al. 2017). The more transparent functionalized films would be suitable for packaging (X. Zhao et al. 2023), while other fields of applications, e.g., wastewater decontamination/disinfection (Maldonado-Carmona et al. 2020), would not be opposed to the opaque appearance.

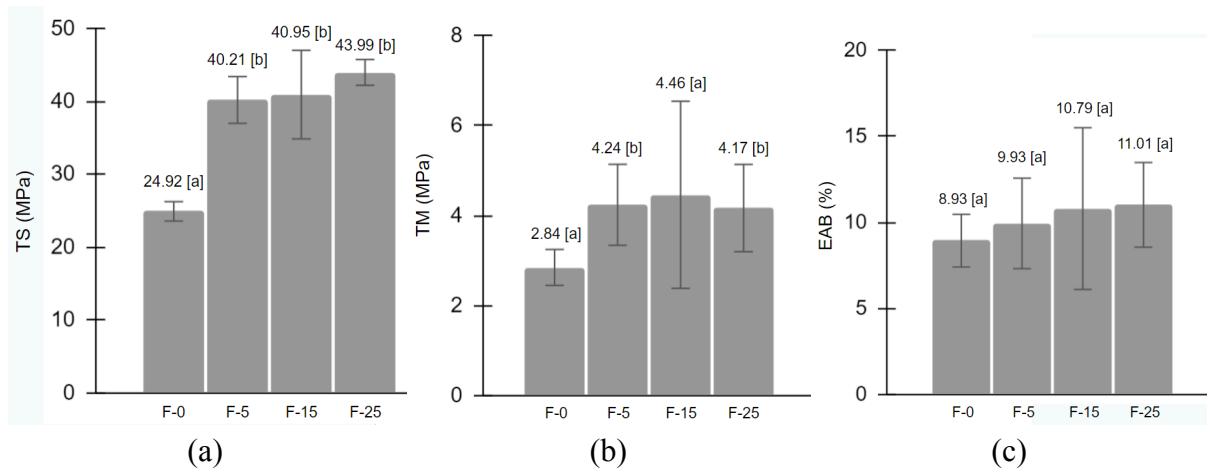
### 4.3.3. Barrier property



**Fig. 4.6.** Oxygen permeability (OP) of functionalized CNF films at 50%RH

**Fig. 4.6.** presents the oxygen permeability (OP) of the functionalized CNF films at 50%RH. There is no significant difference between films prepared at different numbers of homogenization cycles. However, the obtained values are comparable to numerous commercial high-barrier polymers often used in the packaging field, such as ethylene vinyl alcohol (EVOH), PVDC, PVA, MXD6, and PET (Michiels, Van Puyvelde, and Sels 2017; Artzi, Narkis, and Siegmann 2005; Kuraray 1972). These findings are also far superior to N-films (**Fig. 3.6**) owing to F-films' extremely dense composite structure after TiO<sub>2</sub>-TCPP conjugate incorporation, which could be compared morphologically with that of neat-CNF (**Fig. 4.3** and **Fig. 3.3**).

### 4.3.4. Mechanical properties

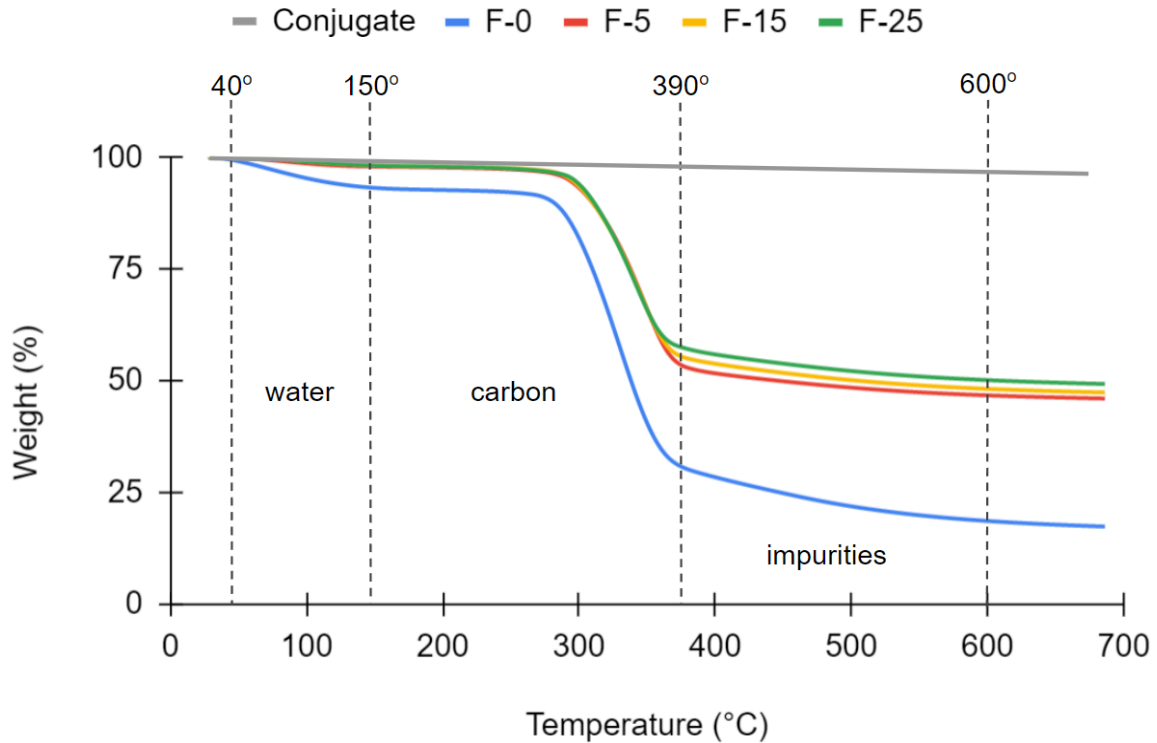


**Fig. 4.7.** Mechanical properties of functionalized CNF films: (a) tensile strength or TS, (b) tensile/Young's modulus or E, (c) elongation-at-break or EAB

**Fig. 4.7** displays three different mechanical properties of functionalized CNF films. There are likewise not many significant differences between films prepared at different numbers of homogenization cycles, except in regards to tensile strength and tensile modulus, where the only noticeable improvement was between the use of CNF before

and after homogenization. The enhanced fibrillation and hydrogen bonding post-homogenization (as seen morphologically in **Fig. 4.3**) (Mokhena et al. 2021; Peresin et al. 2017) played a key role in the even dispersion and less tendency of self-quenching/aggregation of conjugates, which are among the most detrimental challenges of MOF-based materials to not only their antibacterial activity but also functional properties like mechanical strength.

#### 4.3.5. Thermal properties



**Fig. 4.8.** Thermogravimetric spectra of conjugate and functionalized CNF films

**Table 4.2.** Degradation temperature and weight loss profile of conjugate and functionalized CNF films

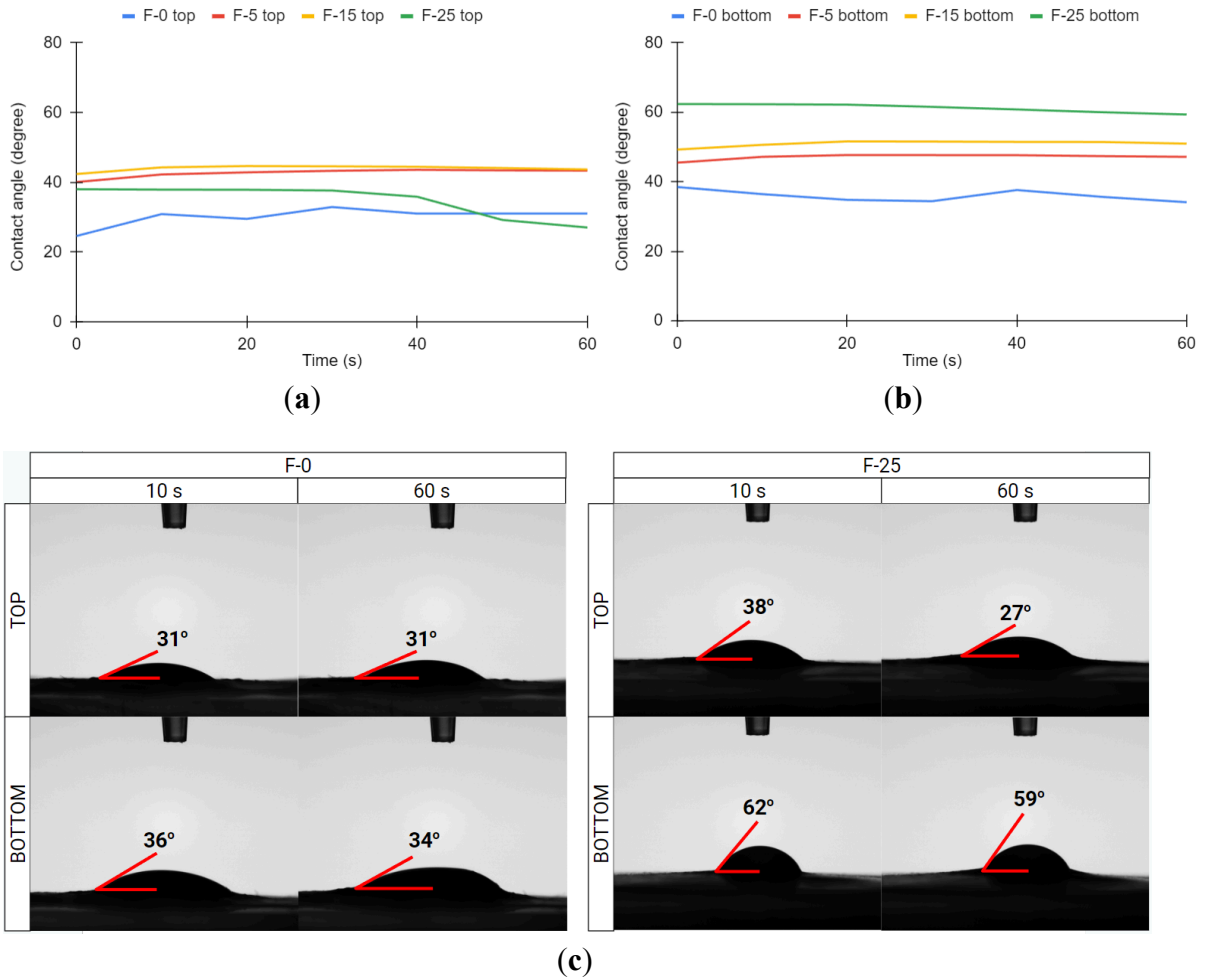
Sample	$T_{d5\%}$ (°C)	%W at 700°C
Conjugate	-	96
F-0	100	17
F-5	291	46
F-15	293	48
F-25	295	49

Thermogravimetric analysis (TGA) provides insights into thermal stability by measuring weight/mass (loss or gain) as a function of temperature or time. The decomposition behavior output may be crucial when determining one's (bio)degradability (Brunšek et al. 2023). **Fig. 4.8** presents TGA spectra of conjugate powder and all F-films, and **Table 4.2** lists each specimen's  $T_{d5\%}$  (the temperature at 5%-weight loss), and %weight at 700 °C, which is in the pyrolysis temperature range where usually material's

stagnant weight change is achieved (D. D. Jiang et al. 1999; M. A. Islam et al. 2016). As seen in the produced spectra, conjugate, being a metal-organic framework (MOF) that contains metal ions/nodes in the form of  $\text{TiO}_2$  that act as the inorganic cluster/building blocks, displaying a distinctly different thermal behavior compared to any of the organic, cellulose-based F-films. Due to the lack of carbon/hydrogen atoms and strong covalent bonds (Malakhov and Samsonov 1966; Johns, McElhill, and Smith 1962), conjugate retains the residual masses of 96.23% at 700 °C, a slight increment from that of neat- $\text{TiO}_2$  (X. W. Li et al. 2013) due to the linkage with TCPP, which as a monomer would lose roughly half of its mass at 700 °C (L.-J. Feng et al. 2017; Ji et al. 2024). In terms of F-films spectra, temperatures at 40-47 °C and 100 °C marked the first and least significant decompositions assigned to the absorbed moisture or loosely-bound water contained in the films, typically through evaporation or dehydration (B. Yang et al. 2019; Ilyas et al. 2018). F-0, due to the still-naturally high hydrophilicity (Abral et al. 2020), displayed the water's highest evaporation effect, meaning the earliest decomposition. Overall, cellulose only started to decompose extensively through pyrolysis at 260-360 °C (Abral et al. 2019; Araújo et al. 2018), where this time was shown by the identical spectrum of all three homogenized films (F-5, F-15, and F-25). Again, contrary to the homogenized films, F-0 displayed weight loss much earlier at around 200°, which was attributed to CNF's dehydration and decarboxylation that produced combustion gases (e.g., levoglucosan) (Cao et al. 2019). Finally, the most rapid thermal-oxidative decomposition of char/ash around 300-360 °C was the last decomposition stage (Asrofi et al. 2018; Jiangqi Zhao et al. 2013), where a slight improvement in thermal resistance was discovered at the increasing passes of homogenization. After 400 °C, the thermal degradation rates decreased, marking the start of a slow thermal decomposition (G.-Q. Liu et al. 2021). F-25 suffered a 5%-weight loss at the highest temperature (295 °C) and maintained the most weight (49%), followed by F-15 (293 °C and 48%), and lastly, F-5 (291 °C and 46%). Homogenized films meant fibrillated structure, higher surface area, and enhanced interlinking hydrogen bonding between the polymer chains that required higher decomposition energy (A. Islam, Yasin, and Rehman 2014; U.-J. Kim, Eom, and Wada 2010). These findings are comparable to those of other reports (Min et al. 2021). Post-homogenized, these resulting thermal stability indicators were far superior to the untreated film (F-0) (100 °C and 17%); where due to the lacking amount of hydrogen bonding present, when the temperature reached around 500 °C, CNF itself decomposed nearly completely, while the conjugate aggregates remained. The incorporated inorganic nanoparticles— $\text{TiO}_2$  as clusters or building blocks in the MOFs-based conjugates—into CNF possess catalytic and conductivity properties that negatively affect decomposition temperature (Zeng et al. 2010; Ma, Li, and Bai 2006), which can be proven by the decreasing  $T_{d,5\%}$  of F-0 (100 °C) compared to N-0 (257 °C) The same phenomenon occurred in other reports on cellulose incorporated with ZnO (S.-W. Zhao et al. 2017; Onyszko et al. 2022), a similar metal ion/node commonly used in MOFs as UV-blocker and antimicrobial, where its lower thermal decomposition temperature was caused by the direct interaction between cellulose and ZnO particles that allowed an easier heat flow within the composite. The resulting thermostable films possess a high potential to be used as medical devices (L. Sun et al. 2023) and personal thermal management (Gu et al. 2020). Despite heightened concerns on  $\text{TiO}_2$  use as food additives, which still deals with difficulties and uncertainties to date to reach a proper and retrospective safety assessment

(Winkler et al. 2018), there has yet any regulation on its use as packaging, especially when incorporated in composite structure like the developed CNF-MOF with negligible migration (W. Zhang and Rhim 2022). Moreover, when speaking of degradability, TiO<sub>2</sub> addition in different packaging systems were also proven advantageous in promoting photo/bio-degradation rate (Tu-morn et al. 2019; Bang, Bang, and Lee 2007; Iacovone et al. 2023).

#### 4.3.6. Contact angle or wettability



**Fig. 4.9.** Contact angle of functionalized CNF films: (a) top surface, (b) bottom surface, and (c) image comparison

Contact angle or wettability dictates if the material is either hydrophobic or hydrophilic, measured by the angle produced between the tangent to the droplet of liquid of choice, at the point of contact with the material's surface. The standard measuring liquid ought to be of low volatility to avoid evaporation as well as non-reactive to the tested surface. Some parameters can be customized, such as droplet size and liquid dispenser speed; meanwhile, many other factors may pertain to the resulting contact angle, including humidity, temperature, roughness (topography), morphology, and physical/chemical homogeneity (Jagodzińska et al. 2021; Wawro and Kazimierczak

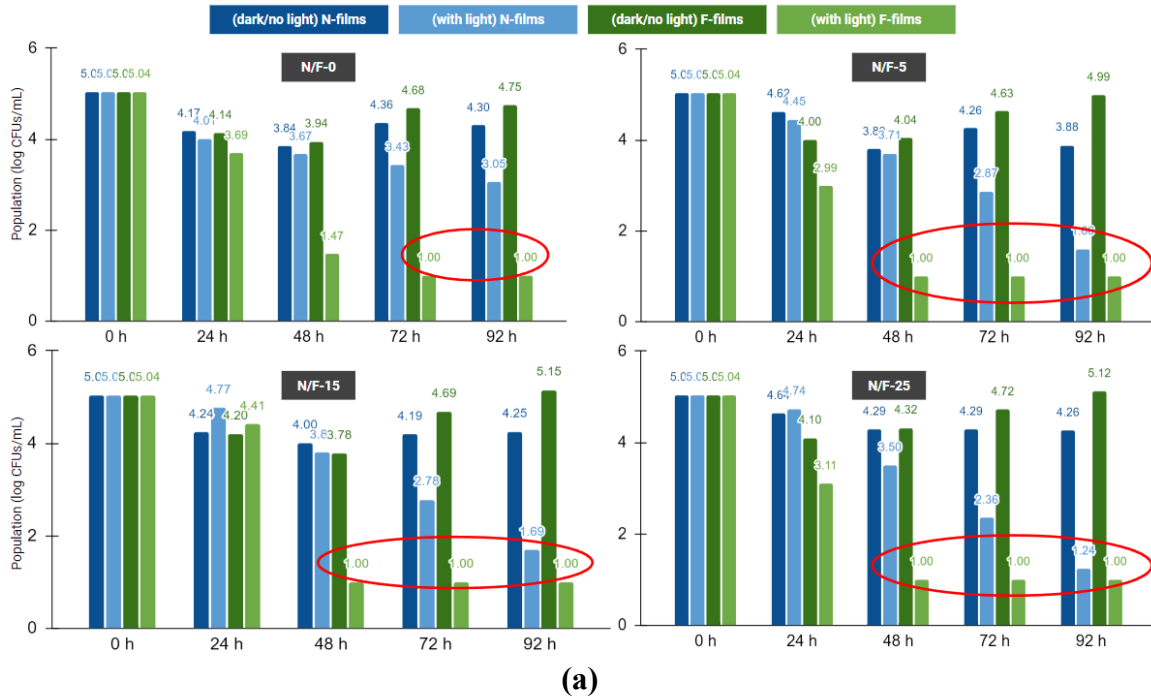
2008; Jonoobi, Harun, Mathew, Hussein, et al. 2010). Wenzel and Cassie–Baxter equations are commonly used to explain hydrophobicity, where the liquid cannot penetrate the grooves on a rough surface, leaving air gaps (J. Song and Rojas 2013; Alimohammadzadeh, Sanhueza, and Córdova 2023). It is important to note the significant difference between wettability tested on top and bottom surfaces when analyzing films prepared using casting techniques, which vary in microstructure, porosity, and pore size. In the case of a single-layered, non-composite film that comprises one singular polymeric ingredient, the top surface tends to possess a higher contact angle value (Jagodzińska et al. 2021; McGaughey et al. 2020). However, the developed films consisted of continuous and dispersed phases (CNF and conjugates, respectively), where the composite's degree of distribution and homogeneity (as seen in morphological, cross-sectional images in **Fig. 4.3(b)**) influence the surface roughness (**Fig. 4.4** and **Fig. 4.5(a)**) and subsequent wettability profile. **Fig. 4.9** displayed the contact angle of the functionalized CNF films plotted over time, which shows the top side of the film's surface (**Fig. 4.9(a)**) was more hydrophobic compared to the bottom (**Fig. 4.9(b)**), regardless of the cycle of homogenization. As described in the surface roughness analysis, F-0 was discovered to have the roughest surface; despite that, it appeared to be the most hydrophilic regardless of the surface sides. This phenomenon was caused by the unhomogenized CNF as the matrix of F-0 still suffers from the intrinsic, naturally high hydrophilicity of cellulose, thanks to their plentiful polar groups (e.g., hydroxyl and carboxyl groups) that negatively affects hydrophobicity (Q. Wang et al. 2018; X. Sun et al. 2021; G. Zhou et al. 2015; Oh and Park 2022). Furthermore, specifically on the bottom surface's contact angle results, it was found that films prepared using CNF with more passes of homogenization were more hydrophobic. The even dispersion of the conjugates within the fibrillated CNF matrix produced films with better hydrophobicity. The most hydrophobic result was achieved by F-25's bottom surface, which produced 62° after 10 seconds—a comparable finding to other reports on conjugates-incorporated composite (X. Zhao et al. 2023; El-Wakil et al. 2015). F-25's top surface is an exception due to the effect of over-shearing, where the lower aspect ratio on top of the conjugates' self-aggregation/quenching tendency (An et al. 2016) led to agglomeration and sinking of the produced agglomerates to the bottom of the film, resulting in a smoother top surface. The hydrophilicity of F-25 even surpassed that of F-0 after 50 seconds. Functionalized films with a hydrophilic profile would suffice as hydrogel/aerogel in drug delivery and tissue engineering applications (De France, Hoare, and Cranston 2017; Y. Sun et al. 2024); meanwhile, the acquired hydrophobicity would be beneficial for the development of active and intelligent/smart food packaging (X. Zhao et al. 2023; El-Wakil et al. 2015).

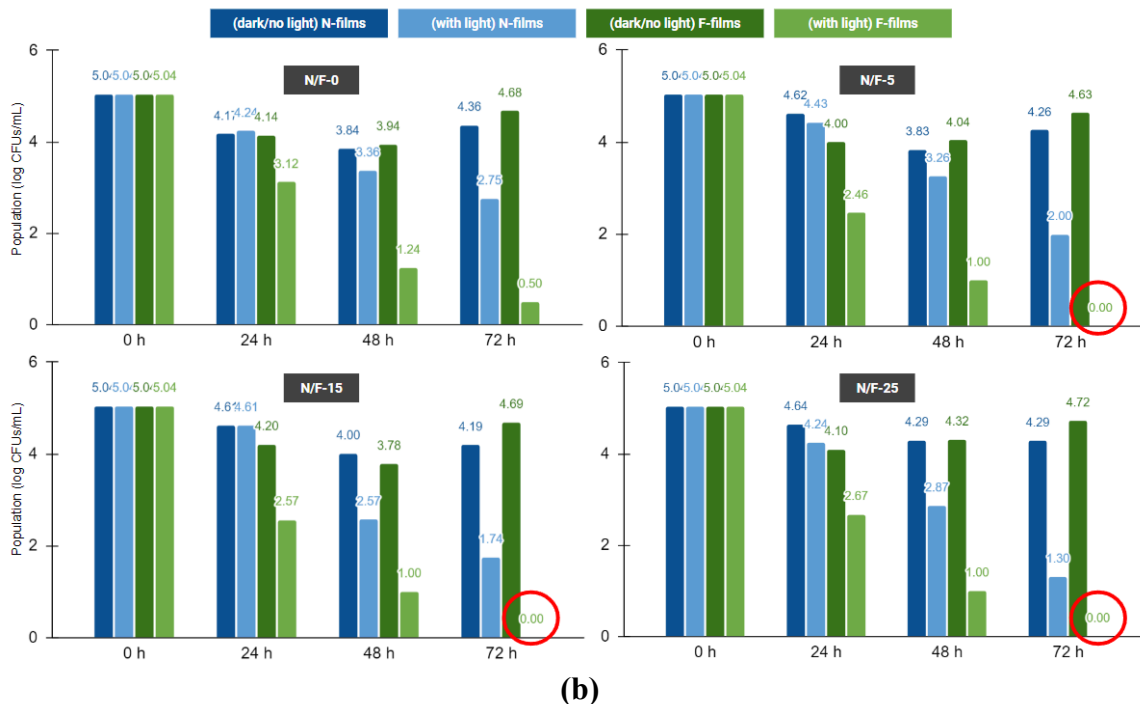
#### 4.3.7. Antibacterial activity

The antibacterial efficacy of functionalized CNF films was evaluated against *Escherichia coli* (*E. coli*)—a common foodborne pathogen found not only in polluted water and soil but also in wildlife, livestock, and humans as their carriers (FDA 2019b). *E. coli* was selected because cellulose and TiO<sub>2</sub> performed better antibacterial efficacy against Gram-negative bacteria, e.g., *E. coli*, than against Gram-positive bacteria (Maślana et al. 2021). The difference between Gram-negative/-positive bacteria is in their cell wall structure difference; the former contains a thin peptidoglycan layer in the cell wall, whereas the latter is much thicker, which provides cell resistance against Ti ions. Gram-negative bacteria's structure with lipopolysaccharide in their outer membrane favors an affinity in the adherence of the Ti ion to the cell membrane (Arularasu, Harb, and Sundaram 2020; Slavin et al. 2017). To evaluate such bactericidal activity between CNF films before and after functionalization (N-films and F-films, respectively), the viable bacterial counts recovered under different light intensities and contact durations are shown in **Fig. 4.10**. Time-kill test/curve is a standardized method for determining bactericidal activity of antimicrobial agents (Clinical and Laboratory Standards Institute 1999) and 0-96 hours are the usual contact duration applied for *E. coli* studies (Elliker and Frazier 1938). Speaking of the light intensity factor, 1000-10,000 lux is the range of illuminance used to activate photoactive antimicrobial surfaces (K. Wu et al., 2020). Two light intensities (3000 and 6000 lux) were selected after the optimum conditions obtained by several previous studies on light-activated antimicrobial surfaces for *E. coli* bactericidal inhibition (Noimark, Allan, and Parkin 2014; Noimark et al. 2013; Sehmi et al. 2016).

Antibacterial efficacy under 6000 lux light was reported in **Fig. 4.10(a)**. Under 24 hours, the functionalized film using five-passes homogenized-CNF (F-5) under light was the best-performing film with a 41% antibacterial rate. The next day, F-5 and F-15, under light, were the joint films with the highest antibacterial rate at 80%. After 72 hours, all homogenized CNF films functionalized with conjugates (F-5, F-15, F-25) under light succeeded in completely inhibiting *E.coli* growth (100% antibacterial rate). These findings were superior compared to similar studies (Min et al. 2021; X. Zhao et al. 2023; El-Wakil et al. 2015). A former report on porphyrin-based MOF (Min et al. 2021) further proved this exceptional antibacterial efficacy through SEM analysis of *E.coli* showing wrinkled/distorted morphology after light irradiation that led to bacteria death. This result certified the impact of TCPP as a photosensitizer for the high-efficiency generation of reactive oxygen species (ROS), especially singlet oxygen (<sup>1</sup>O<sub>2</sub>). By utilizing oxygen and light, the photosensitizer absorbs light energy to form an excited singlet state, then an excited triplet state, before the final product of ROS in the form of <sup>1</sup>O<sub>2</sub> (DeRosa and Crutchley 2002). Speaking of the antibacterial activity of neat-CNF films prior to conjugates incorporation (N-films) in the dark, five-passes homogenized-CNF film (N-5) performed the highest antibacterial rate at 24%, which was obtained after 48-hour-long storage. This finding is aligned with the result about N-5 possessing the lowest oxygen permeability (**Fig. 4.6**) among all N-films that helped to reduce the growth rate of food spoilage bacteria, including facultative anaerobes like *E. coli* (Couvert et al. 2023). Interestingly, after 72 hours under light irradiation, N-25 showed an exceptional antibacterial rate at 74%, exceeding other N-films stored for the same duration.

**Fig. 4.10(b)** and **Table 4.3/****Table 4.4** presents the films' antibacterial activity under 3000 lux light. After being stored for 24 hours under light, F-5 was once more the one exhibiting the highest antibacterial rate among all films, at 41%—identical to its performance under 6000 lux light. The next day, all homogenized CNF films functionalized with conjugates (F-5, F-15, F-25) under light became the joint top-performing films with an 80% antibacterial rate. After 72 and 92 hours, all F-films, including one prepared using unhomogenized CNF (F-0) under light, achieved the same antibacterial rate (80%). This best outcome under the 3000 lux light scenario was lower than that treated under 6000 lux, which performed a complete inhibition growth (100% antibacterial rate). This finding is in agreement with former reports (K. Wu et al. 2020; Macdonald et al. 2016) that determined 6000 lux as the minimum light intensity in order to reach the optimum *E. coli* growth inhibition rate. Regarding N-films, the best-performing specimen in the dark was N-5, with a 25% antibacterial rate after 48 hours. Meanwhile, under light, N-25 performed exceptionally well after 92 hours, exhibiting a 75% antibacterial rate. Both of these N-film results were identical to the ones under 6000 lux light, certifying the insignificant effect of higher light intensity on N-films' bactericidal efficacy, as similarly found in some photodynamic antimicrobial studies on *E. coli* (Ullah et al. 2018; Sehmi et al. 2016). The promising potential of the developed antimicrobial films ranges from food packaging (El-Wakil et al. 2015; Larrea et al. 2024; Min et al. 2021; X. Zhao et al. 2023), converter (Cherian and Wamser 2000; Gao et al. 2019), biomedical (Carpenter et al. 2012; J. Park et al. 2016), until micropollutants removal (G.-Q. Liu et al. 2021; Rathod et al. 2018; Roberta Grazia Toro et al. 2020).





**Fig. 4.10.** Antibacterial activity of neat CNF and functionalized CNF films on *E. coli* under varying light intensities: (a) 3000 and (b) 6000 lux

**Table 4.3.** The best performing films under 3000 lux-light

Contact period (h)	The best-performing film and its antibacterial rate (%) or <i>E. coli</i> population (log CFUs/mL)
24	F-5, under light = <b>41%</b> (2.99 log CFUs/mL)
48	F-5/15/25, under light = <b>80%</b> (1.00 log CFUs/mL)
72	F-0/5/15/25, under light = <b>80%</b> (1.00 log CFUs/mL)

**Table 4.4.** The best performing films under 6000 lux-light

Contact period (h)	The best-performing film and its antibacterial rate (%) or <i>E. coli</i> population (log CFUs/mL)
24	F-5, under light = <b>51%</b> (2.46 log CFUs/mL)
48	F-5/15/25, under light = <b>80%</b> (1.00 log CFUs/mL)
72	F-0/5/15/25, under light = <b>100%</b> (0.00 log CFUs/mL)

#### 4.4. Conclusions

Cellulose nanofibril (CNF) film functionalized with a metal-organic framework (MOF)-based photosensitive antibacterial agent involving reactive oxygen species (ROS) generation was successfully fabricated. CNF was initially prepared by different cycles of homogenization before being impregnated with TiO<sub>2</sub>-TCPP conjugates. Chosen for its excellent UV-blocking ability, TiO<sub>2</sub> potential was proven by its complete absorption of all UV spectrums (A, B, and C). Meanwhile, TCPP was selected as a photosensitizer for its photodynamic activity even under the visible light spectrum and verified likewise by the observed complete inhibition of *Escherichia coli* growth (100% antibacterial rate) under 6000 lux-light irradiation after 72 hours. Improvement in conjugate's even dispersion, surface smoothness, mechanical properties, thermal stability, and hydrophobicity on functionalized films was also obtained, indicating the positive effect of enhanced fibrillation and the subsequent inter/intra-molecular hydrogen bonding post-homogenization. This work overall certified TiO<sub>2</sub>-TCPP biocompatibility and collective prospect as a remarkable photosensitive antimicrobial agent, as well as the cellulose's fruitful performance taking the role of MOF conjugate's ex-situ carrier that solved the notoriously challenging self-aggregation and self-quenching tendency.

## CHAPTER 5

### Conclusions

To help the globally recurring food spoilage and food waste issue, nanocomposite films made of cellulose, titanium dioxide ( $\text{TiO}_2$ ), and porphyrin with high-barrier, UV-blocking, and visible light-responsive antimicrobial features were developed. Metal-organic framework (MOF) consisting of  $\text{TiO}_2$  and TCPP (a biocompatible porphyrin derivative) was chosen as the antibacterial modality due to its excellent photodynamic activity and generation of reactive oxygen species (ROS) that can kill bacteria under not only UV but also visible light spectrum. Other than being the Earth's most abundant and naturally biodegradable, cellulose was employed as an ex-situ carrier of the MOF conjugate to solve the common constraint of MOF-based conjugates on their self-quenching/aggregation tendency, and cellulose nanofibril (CNF)—a type of nanosized cellulose—was specially chosen for its high specific surface area and surface activity. To solve neat cellulose's challenge of natural hydrophilicity that limits its processability, a straightforward, cheap, and environmentally friendly approach of multicycle homogenization was conducted, where low pressure was applied to prevent the common over-shearing effect. Other than the functional properties, the antimicrobial efficiency of the functionalized films against *Escherichia coli* was analyzed with and without light of different intensities to confirm the photosensitivity effect. Complete UV-A/B/C blocking was observed, and so was the complete growth inhibition (100% antibacterial rate) under 6000 lux-light irradiation after 72 hours. Collectively, improvement in crystallinity, conjugate's even dispersion, surface smoothness, mechanical properties, thermal stability, hydrophobicity, and oxygen barrier (comparable to ethylene-vinyl alcohol (EVOH), a commercial high-barrier polymer) on the functionalized films was also obtained. These findings depicted the positive effect of enhanced fibrillation and the subsequent inter/intra-molecular hydrogen bonding post-homogenization, as well as the promising photosensitive antibacterial performance of  $\text{TiO}_2$ -TCPP. The applicability prospect of the developed films with notable features of high-barrier, UV-blocking, and/or visible light-responsive antibacterial ranges from food/drug packaging, biomedical (including photodynamic therapy), nanodevices, sensors, gas storage/separation, pollutant removal, and energy conversion.

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