



Complete Genome Sequence of *Providencia stuartii* CMC-4104, Isolated from a Human Splenic Abscess, Containing Multiple Copies of NDM-1 and PER-1 Carbapenem Resistance Genes

Jayasimha Rao,^{a,b,c} Nicholas K. Stornelli,^{b,d} Nathan A. Everson,^{b,d} Lauren F. McDaniel,^{b,d} Mariana Gomez De La Espriella,^{a,b} Jason R. Faulhaber,^{a,b,c} S. Michelle Todd,^e Kevin K. Lahmers,^e Roderick V. Jensen^f

^aInternal Medicine, Division of Infectious Disease, Carilion Medical Center, Roanoke, Virginia, USA

^bDivision of Infectious Disease, Virginia Tech Carilion School of Medicine, Roanoke, Virginia, USA

^cCenter for Emerging, Zoonotic, and Arthropod-Borne Pathogens, Virginia Tech, Blacksburg, Virginia, USA

^dDepartment of Pharmacy, Carilion Roanoke Memorial Hospital, Roanoke, Virginia, USA

^eDepartment of Biomedical Sciences and Pathobiology, VA-MD College of Veterinary Medicine, Virginia Tech, Blacksburg, Virginia, USA

^fDepartment of Biological Sciences, Virginia Tech, Blacksburg, Virginia, USA

Jayasimha Rao and Nicholas K. Stornelli contributed equally to this article. Jayasimha Rao contributed the majority of the work, including writing the manuscript.

ABSTRACT We report the complete genome sequence of a clinical isolate of *Providencia stuartii* strain CMC-4104, isolated from a splenic abscess. Oxford Nanopore Technologies (ONT) and Illumina sequencing reads were assembled using Geneious to generate a 4,504,925-bp circular chromosome containing multiple copies of the NDM-1 and PER-1 genes in a genomic resistance island.

The spread of carbapenemase-producing *Enterobacteriales* is increasing in health care facilities (1). We describe a multidrug-resistant (MDR) clinical isolate of *Providencia stuartii* recovered from an infected necrotizing pancreatitis patient, belonging to the class of difficult-to-treat resistance (2, 3) strains, panresistant to first-line antimicrobials, including newer β -lactam/ β -lactamase inhibitor combinations (4–6).

P. stuartii CMC-4104, isolated from a patient with splenic abscess aspirate, was inoculated onto sheep blood agar and MacConkey agar plates and incubated at 35°C aerobically for 24 to 72 h. Antibiotic susceptibility testing for 17 antimicrobial agents based on the CLSI M100 standards (7) was carried out by Quest Diagnostics at the Carilion Clinic (Table 1).

A single colony of *P. stuartii* CMC-4104 was grown in 25 mL lysogeny broth at 37°C and 200 rpm for 18 h. The cell pellet was used for genomic DNA isolation with a Genomic-tip 20/G kit (8, 9). The unsharded DNA was sequenced without size selection using the ONT 9.4.1 MinION flow cell with the SQK-LSK109 ligation sequencing kit. MiSeq Illumina paired-end sequencing (350-bp insert size) was performed using the TruSeq DNA PCR-free library prep kit.

The ONT sequencing generated 19,109 reads with a maximum length of 170,418 bp and an average length of 15,700 bp for a total of 306 Mbp. Base calling was performed using Guppy 4.4.1, and Porechop 0.2.4 (10) was used for adaptor trimming. The resulting fastq files were initially assembled using Flye 2.8 (11) in Geneious Prime 2022.0.2 to produce a large, closed 4.59-Mbp draft genome with $\sim 65\times$ coverage. The Illumina sequencing generated 2,290,766 paired-end reads 150 bp long for a total of 344 Mbp. The “Map to Reference” tool in Geneious Prime was used (with low sensitivity settings) to align all of the Illumina reads to the draft genome sequence and correct ONT sequencing errors. Variations in the Illumina read coverage were used to identify three circular plasmid sequences and a highly repeated set of MDR gene cassettes in the

Editor Julie C. Dunning Hotopp, University of Maryland School of Medicine

Copyright © 2022 Rao et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Jayasimha Rao, jr Rao@carilionclinic.org, or Roderick V. Jensen, rvjensen@vt.edu.

The authors declare no conflict of interest.

Received 25 May 2022

Accepted 18 July 2022

TABLE 1 Antimicrobial susceptibility for *P. stuartii* strain CMC-4104

| Antimicrobial | MIC ^a | Result |
|--------------------------------------|------------------|--------|
| Trimethoprim sulfamethoxazole | ≥ 320 | R |
| Ampicillin + sulbactam | ≥ 32 | R |
| Ertapenem | ≥ 8 | R |
| Imipenem | ≥ 16 | R |
| Piperacillin + tazobactam | ≥ 128 | R |
| Cefazolin | ≥ 64 | R |
| Cefepime | ≥ 32 | R |
| Ceftazidime | ≥ 64 | R |
| Ceftriaxone | ≥ 64 | R |
| Ceftazidime + avibactam ^b | ≥ 250 | R |
| Imipenem + relebactam ^b | ≥ 32 | R |
| Meropenem + vaborbactam ^b | ≥ 250 | R |
| Tobramycin/gentamicin | 4 | R |
| Levofloxacin | ≥ 16 | R |
| Cefiderocol ^c | 2 | S |

^a MICs were determined (7) using the Vitek and Verigene testing systems at the Quest Diagnostics microbiology laboratory at Carilion. The carbapenemase-producing strain was confirmed by the Commonwealth of Virginia Consolidated Laboratory Services, VDH.

^b Susceptibility testing was performed using the E-test for ceftazidime + avibactam, imipenem + relebactam, and meropenem + vaborbactam.

^c Cefiderocol susceptibility testing was completed by Associated Regional and University Pathologists Laboratories in Salt Lake City, UT.

main chromosome. Finally, Geneious Prime was used to align the ONT reads to the plasmid and genome sequences (with medium sensitivity settings) to confirm closure of the circular plasmid and main chromosome sequences, as well as the repetition of the MDR gene cassettes. Default parameters were used for all software unless otherwise specified.

Final assembly of the *P. stuartii* CMC-4104 genome resulted in a main chromosome (GenBank accession number [CP095443](#)) of 4,504,925 bp with 41.4% GC content and an average Illumina coverage (AIC) of 65×; a large, low-copy, circular plasmid ([CP095444](#)) of 278,489 bp with 47.3% GC content and an AIC of 85×; a small, high-copy, circular plasmid ([CP095445](#)) of 2,683 bp with 41.8% GC content and an AIC of 4,616×; and a phage-like circular sequence ([CP095442](#)) of 51,458 bp with 41.7% GC content and an AIC of 105×. In the main chromosome ([CP095443](#)), the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) 6.1 (12) was used to identify 4,066 protein coding genes, 79 tRNAs, 22 rRNAs, and 4 CRISPR arrays. Of particular note was a large 215-kbp genomic resistance island (*PsGRI*) integrated with a site-specific integrase into the 3' end of a tRNA modification GTPase gene (*mmE*), like the *Salmonella* genomic island (13, 14). Remarkably, the *PsGRI* contained 15 copies of the MDR gene cassettes containing *bla*_{NDM-1} and 2 copies of *bla*_{PER-1} shown in Fig. 1. The large plasmid ([CP095444](#)) also contained one complete copy each of the *bla*_{NDM-1} and *bla*_{PER-1} MDR cassettes and an additional *bla*_{OXA-10} MDR cassette (14–18). The small, high-copy plasmid ([CP095445](#)) also contained a fluoroquinolone resistance gene, *qnrD* (19). The assembly revealed a circular phage-like sequence ([CP095442](#)) that is identical to a prophage in the main chromosome and is very similar to a second prophage integrated into a CRISPR locus (20).

Data availability. The annotated complete genome assembly of strain *Providencia stuartii* CMC-4104 is available at GenBank under accession numbers [CP095442.1](#), [CP095443.1](#), [CP095444.1](#), and [CP095445.1](#), SRA accession numbers [SRR18691816](#) and [SRR18691817](#), BioProject accession number [PRJNA824933](#), and BioSample accession number [SAMN27484493](#).

ACKNOWLEDGMENTS

We acknowledge the faculty members of the Division of Infectious Disease, Carilion Clinic, and the Carilion clinical microbiology laboratory for their assistance in collecting patient isolates. Experiments were carried out at the Carilion Basic Science Research

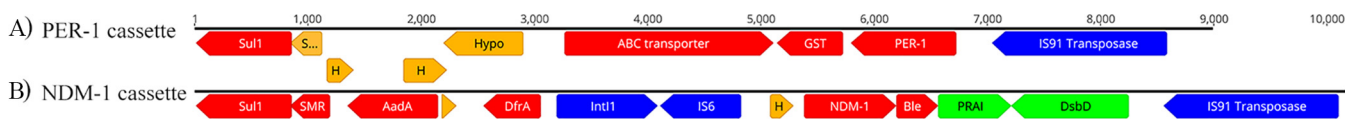


FIG 1 Gene organization in the repeated multidrug-resistant (MDR) cassettes. (A) The PER-1 cassette (8,986 bp) included genes for sulfonamide-resistant dihydropteroate synthase (*sulI*), small multidrug resistance (SMR) (*qacEΔ1*) (truncated), hypothetical proteins, ATP-binding protein/permease (ABC transporter), glutathione S-transferase (GST) family protein, class A extended-spectrum β -lactamase (*bla*_{PER-1}), and ISCR1 family transposase (IS91). (B) The New Delhi metallo β -lactamase (*bla*_{NDM-1}) cassette (10,494 bp) included genes for sulfonamide-resistant dihydropteroate synthase (*sulI*), small multidrug resistance (SMR) *qacEΔ1*, aminoglycoside 3'-O-nucleotidyltransferase (*aadA*), DUF1010 domain-containing hypothetical protein, trimethoprim-resistant dihydrofolate reductase (*dfrA12*), class 1 integron integrase (*int1*), IS26 family transposase (IS6), IS30 family transposase (truncated), subclass B1 β -lactamase (*bla*_{NDM-1}), bleomycin resistance protein (Ble), phosphoribosylanthranilate isomerase (PRAI), cytochrome c-type biogenesis protein/protein-disulfide reductase (*dsbD*), and ISCR1 family transposase (IS91). Red represents drug resistance genes; blue, transposases; green, metabolic genes; and yellow, hypothetical or pseudogenes (truncated or overlapped). Annotated genes were used from NCBI GenBank, and the scale indicates the number of base pair residues.

Laboratory, Carilion Roanoke Community Hospital. We thank the Carilion Clinic Healthcare Analytics Research Team for data storage and maintenance. We thank Sarah Cox, Radford University Carilion, for her valuable support in manuscript reading and suggestions.

This study was supported by the Division of Infectious Diseases, Carilion Clinic (J.R.). The Oxford Nanopore sequencing was completed with the support of an FDA CVM Vet-LIRN Veterinary Diagnostic Laboratory Program Infrastructure Grant (K.K.L.), and the Illumina sequencing was performed at the Virginia Tech Genomics Resource Center with the support of the Fralin Life Science Institute (R.V.J.).

REFERENCES

- Murray CJ, Ikuta KS, Sharara F, Swetschinski L, Robles Aguilar G, Gray A, Han C, Bisignano C, Rao P, Wool E, Johnson SC, Browne AJ, Chipeta MG, Fell F, Hackett S, Haines-Woodhouse G, Kashef Hamadani BH, Kumaran EAP, McManigal B, Agarwal R, Akech S, Albertson S, Amuasi J, Andrews J, Aravkin A, Ashley E, Bailey F, Baker S, Basnyat B, Bekker A, Bender R, Bethou A, Bielicki J, Boonkasidecha S, Bukosia J, Carvalho C, Castañeda-Orjuela C, Chansamouth V, Chaurasia S, Chiurchiù S, Chowdhury F, Cook AJ, Cooper B, Cressey TR, Criollo-Mora E, Cunningham M, Darboe S, Day NPJ, De Luca M, Dokova K, et al. 2022. Global burden of bacterial antimicrobial resistance in 2019: a systematic analysis. *Lancet* 399:629–655. [https://doi.org/10.1016/S0140-6736\(21\)02724-0](https://doi.org/10.1016/S0140-6736(21)02724-0).
- Abdallah M, Balshi A. 2018. First literature review of carbapenem-resistant *Providencia*. *New Microbes New Infect* 25:16–23. <https://doi.org/10.1016/j.nmni.2018.05.009>.
- Kadri SS, Adjemian J, Lai YL, Spaulding AB, Ricotta E, Prevots DR, Palmore TN, Rhee C, Klompas M, Dekker JP, Powers JH, Suffredini AF, Hooper DC, Fridkin S, Danner RL, National Institutes of Health Antimicrobial Resistance Outcomes Research Initiative (NIH-ARORI). 2018. Difficult-to-treat resistance in Gram-negative bacteremia at 173 US hospitals: retrospective cohort analysis of prevalence, predictors, and outcome of resistance to all first-line agents. *Clin Infect Dis* 67:1803–1814. <https://doi.org/10.1093/cid/ciy378>.
- Coppi M, Di Pilato V, Monaco F, Giani T, Conaldi PG, Rossolini GM. 2020. Cef-tazidime-avibactam resistance associated with increased bla KPC-3 gene copy number mediated by pKpQL plasmid derivatives in sequence type 258 *Klebsiella pneumoniae*. *Antimicrob Agents Chemother* 64:e01816-19. <https://doi.org/10.1128/AAC.01816-19>.
- Wang Y, Wang J, Wang R, Cai Y. 2020. Resistance to ceftazidime-avibactam and underlying mechanisms. *J Glob Antimicrob Resist* 22:18–27. <https://doi.org/10.1016/j.jgar.2019.12.009>.
- Lebreton F, Corey BW, McElheny CL, Iovleva A, Preston L, Margulieux KR, Cybulski RJ, Mc Gann P, Doi Y, Bennett JW. 2021. Characterization of KPC-82, a KPC-2 variant conferring resistance to ceftazidime-avibactam in a carbapenem-nonsusceptible clinical isolate of *Citrobacter koseri*. *Antimicrob Agents Chemother* 65:e0015021. <https://doi.org/10.1128/AAC.00150-21>.
- Clinical and Laboratory Standards Institute. 2020. Performance standards for antimicrobial susceptibility testing, 30th ed. Document M100. CLSI, Wayne, PA.
- Adenikinju A, Jensen RV, Kerkering TM, Garner DC, Rao J. 2020. Complete genome sequence of *Pseudomonas aeruginosa* CMC-115, a clinical strain from an acute ventilator-associated pneumonia patient. *Microbiol Resour Announc* 9:e00595-20. <https://doi.org/10.1128/MRA.00595-20>.
- Rao J, Susanti D, Childress JC, Mitkos MC, Brima JK, Baffoe-Bonnie AW, Pearce SN, Grgurich D, Fernandez-Cotarello MJ, Kerkering TM, Mukhopadhyay B. 2018. Tn2008-driven carbapenem resistance in *Acinetobacter baumannii* isolates from a period of increased incidence of infections in a southwest Virginia hospital (USA). *J Glob Antimicrob Resist* 12:79–87. <https://doi.org/10.1016/j.jgar.2017.08.017>.
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Completing bacterial genome assemblies with multiplex MinION sequencing. *Microb Genom* 3:e000132. <https://doi.org/10.1099/mgen.0.000132>.
- Kolmogorov M, Yuan J, Lin Y, Pevzner PA. 2019. Assembly of long, error-prone reads using repeat graphs. *Nat Biotechnol* 37:540–546. <https://doi.org/10.1038/s41587-019-0072-8>.
- Li W, O'Neill KR, Haft DH, DiCuccio M, Chetverin V, Badretin A, Coulouris G, Chitsaz F, Derbyshire MK, Durkin AS, Gonzales NR, Gwadz M, Lanczycki CJ, Song JS, Thanki N, Wang J, Yamashita RA, Yang M, Zheng C, Marchler-Bauer A, Thibaud-Nissen F. 2021. RefSeq: expanding the Prokaryotic Genome Annotation Pipeline reach with protein family model curation. *Nucleic Acids Res* 49:D1020–D1028. <https://doi.org/10.1093/nar/gkaa1105>.
- de Curraize C, Siebor E, Neuwirth C. 2021. Genomic islands related to *Salmonella* genomic island 1; integrative mobilisable elements in trmE mobilised in trans by A/C plasmids. *Plasmid* 114:102565. <https://doi.org/10.1016/j.plasmid.2021.102565>.
- Siebor E, de Curraize C, Neuwirth C. 2019. Identification of AGI1-A, a variant of *Acinetobacter* genomic island 1 (AGI1), in a French clinical isolate belonging to the *Enterobacter cloacae* complex. *J Antimicrob Chemother* 74:311–314. <https://doi.org/10.1093/jac/dky442>.
- Carraro N, Rivard N, Burrus V, Ceccarelli D. 2017. Mobilizable genomic islands, different strategies for the dissemination of multidrug resistance and other adaptive traits. *Mob Genet Elements* 7:1–6. <https://doi.org/10.1080/2159256X.2017.1304193>.
- McGann P, Hang J, Clifford RJ, Yang Y, Kwak YI, Kuschner RA, Lesho EP, Waterman PE. 2012. Complete sequence of a novel 178-kilobase plasmid carrying bla(NDM-1) in a *Providencia stuartii* strain isolated in Afghanistan. *Antimicrob Agents Chemother* 56:1673–1679. <https://doi.org/10.1128/AAC.05604-11>.
- Shin S, Jeong SH, Lee H, Hong JS, Park MJ, Song W. 2018. Emergence of multidrug-resistant *Providencia rettgeri* isolates co-producing NDM-1 carbapenemase and PER-1 extended-spectrum β -lactamase causing a first

- outbreak in Korea. *Ann Clin Microbiol Antimicrob* 17:20. <https://doi.org/10.1186/s12941-018-0272-y>.
18. Shen S, Huang X, Shi Q, Guo Y, Yang Y, Yin D, Zhou X, Ding L, Han R, Yu H, Hu F. 2021. Occurrence of NDM-1, VIM-1, and OXA-10 co-producing *Providencia rettgeri* clinical isolate in China. *Front Cell Infect Microbiol* 11: 789646. <https://doi.org/10.3389/fcimb.2021.789646>.
 19. Guillard T, Grillon A, de Champs C, Cartier C, Madoux J, Berçot B, Lebreil A-L, Lozniewski A, Riahi J, Vernet-Garnier V, Cambau E. 2014. Mobile insertion cassette elements found in small non-transmissible plasmids in Proteaceae may explain qnrD mobilization. *PLoS One* 9:e87801. <https://doi.org/10.1371/journal.pone.0087801>.
 20. Varble A, Campisi E, Euler CW, Maguin P, Kozlova A, Fyodorova J, Rostøl JT, Fischetti VA, Marraffini LA. 2021. Prophage integration into CRISPR loci enables evasion of antiviral immunity in *Streptococcus pyogenes*. *Nat Microbiol* 6:1516–1525. <https://doi.org/10.1038/s41564-021-00996-8>.