

THE COLLECTORLESS FLOTATION OF SPHALERITE

by

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(ABSTRACT)

The flotation of sphalerite has been demonstrated without the use of collectors. The effect of redox potential, pH, and copper-activation have been investigated in tests using samples of pure mineral. It has been found that in general, collectorless flotation of sphalerite can be accomplished at potentials greater than -200 mV, SHE, and is more readily carried out in acidic solutions. It has also been shown that although copper-activation was necessary to achieve flotation recoveries above 35%, an excessive addition of cupric ions may result in a decrease in floatability.

Batch flotation experiments conducted using Elmwood Mine sphalerite ore have shown that in addition to copper-activation, the addition of sodium sulfide was required to obtain high grades and recoveries. If the ratio of the addition of these reagents is maintained such that the atomic ratio of cupric ions to sulfide ions is 0.31, good flotation is observed over a range of reagent dosages.

X-ray photoelectron spectroscopy (XPS) was conducted on pure mineral samples after microflotation testing. Based on the sulfur species identified on highly floatable samples, possible mechanisms for collectorless flotation of sphalerite have been suggested. These include: i) elemental sulfur formed under oxidizing conditions is responsible for collectorless flotation; ii) polysulfides or

metal-deficient sulfides formed as a result of mineral oxidation are responsible for collectorless flotation; and iii) removal of  $\text{HS}^-$  ions, which may render the surface hydrophilic, under oxidizing conditions. The third mechanism is based on the assumption that clean, unoxidized sphalerite surfaces are naturally hydrophobic. Evidence has been presented to suggest that the first mechanism may be responsible for collectorless flotation in acidic solutions, while the second mechanism may be of greater importance in nearly neutral or basic solutions where elemental sulfur is thermodynamically less stable.

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## INTRODUCTION

### GENERAL

The process of froth flotation is of extreme importance to the mineral processing industry. Over  $2 \times 10^9$  tons of ore are processed using this technique annually (Leja, 1982).

This process relies on interfacial phenomena. The chemical environment is therefore extremely important in determining the interaction between the solid, liquid, and gas phases involved. The common method calls for particular surfactants, called collectors, to be adsorbed onto the solid surface, rendering it sufficiently hydrophobic to adhere to a rising stream of air bubbles.

As most minerals are naturally hydrophilic, the use of these collectors, as well as other reagents to promote collector adsorption is required. However, some minerals, such as graphite, sulfur and talc, do not require collector for their flotation (Gaudin, Miaw and Spedden, 1957). Most common sulfide minerals have long been thought to be hydrophilic and, have been treated as such in flotation practice.

However, some sulfides have been shown to float well without the addition of collector, suggesting that perhaps they are naturally hydrophobic. Another possible explanation for the observed collectorless flotation is that the chemical environment favors the formation of hydrophobic surface species which lead to good flotability.

This study attempts to examine the effect of pulp chemistry on the collectorless flotation of sphalerite, a zinc sulfide. The possible

flotation mechanisms for collectorless flotation have been examined with a focus on identifying the hydrophobic species formed during flotation. The collectorless flotation process studied in the present work might lead to a savings in reagent costs and to increased efficiency.

## LITERATURE REVIEW

There have long been questions about the specific character of sulfide minerals. Very early in the development of the science of mineral processing, it was noted that sulfide minerals had less affinity for water than the associated gangue minerals. The bulk oil flotation technique exploited this difference by inducing the sulfide minerals into an oil phase in the presence of water (Haynes, 1860 and Everson, 1886). This technique was soon abandoned due to the high consumption of oil.

Another early process that made use of the wettability characteristics of sulfides was known as the skin flotation technique (Bradford, 1885). In this method of separation fine ore powder was slowly introduced onto the surface of a quiescent water bath. The sulfide minerals floated on the water and the gangue, which was easily wetted, sank. These early processes became obsolete when the use of a rising stream of air bubbles to buoy oil-coated sulfide minerals to the pulp surface was developed (Ballot, Sulman, and Picard, 1905).

Strong evidence has been presented both for and against the natural flotability of sulfide minerals. Sulman (1930) reported that galena and chalcopyrite were floatable without collectors. Other investigators, such as Ravitz and Porter (1933), Ravitz (1940), and Herd and Ure (1941) specifically suggested that galena was naturally floatable, if cleaned of all oxidation products. Ravitz and Porter (1933) further suggested that the role of xanthate, the most common sulfide mineral collector, was to chemically clean the mineral surface, thus revealing its inherent

flotability.

Contradictory evidence was presented, however, by Hagihara (1952) who used the electron diffraction technique to show that xanthate did indeed adsorb on galena surfaces. Sutherland and Wark (1955) demonstrated that different xanthates produced different contact angles on the same sulfide mineral surfaces. These facts seem to overwhelmingly refute Ravitz and Porter's conjecture.

The natural flotability of sulfides was challenged in work done by Knoll and Baker (1941). They showed that clean galena did not float without collector, nor did it adsorb xanthate. Another study where sodium sulfide ( $\text{Na}_2\text{S}$ ) was used in an attempt to produce a clean galena surface indicated a need for collector (Wark, 1938). In addition the depressant effect of  $\text{Na}_2\text{S}$  was noted. Gaudin (1932) discovered that this effect was true for both the reducing agent,  $\text{Na}_2\text{S}$ , and the oxidizing agent, potassium permanganate ( $\text{KMnO}_4$ ) in a pure chalcopyrite system. This controversy was temporarily laid to rest by Taggart, del Giudice, and Ziehl (1934) who speculated that the samples had been contaminated by oily substances or that the frothers had collecting properties themselves. Others (Plaksin, 1959; Glembotskii, Klassen, and Plaksin, 1963) have suggested that natural hydrophobicity is a result of the adsorption of molecular oxygen. The speculation was that this adsorption led to dehydration of the mineral surface allowing air to displace water more readily at the surface.

Collectorless flotation has gained renewed interest in recent years. Boyce, Venter, and Adam (1970) reported that it was being

applied for galena and sphalerite recovery at the Tsumeb concentrator in South Africa. Lepetic (1974) showed that chalcopyrite was flotable using only a frother after dry, autogenous grinding. Pyrrhotite has also been reported to be naturally flotable (Hodgson and Agar, 1984). In addition, Rey and Formanek (1960), Mino (1957), and Plaksin, Khazinskaya, and Tyurnikova (1955) have observed flotability of sphalerite without collector.

Heyes and Trahar (1977) discovered that the flotability of chalcopyrite depended on the potential of the pulp. They also took precautions to eliminate possible contamination by organics. They showed, by using alkaline salts to promote frothing, that organic frothers were not responsible for the phenomenon of collectorless flotation. Gardner and Woods (1979) confirmed Heyes and Trahar's results using electrochemical techniques. Using potential sweep voltammetry, they identified the presence of elemental sulfur as one of the oxidation products in alkaline pH. Since the mineral floated only under oxidizing conditions, where it can be oxidized to form elemental sulfur on its surface, these investigators considered the elemental sulfur to be the hydrophobic entity responsible for the collectorless flotation. One thing to note however, is that in addition to the elemental sulfur, they also identified the presence of iron hydroxide which is hydrophilic.

Finklestein, Allison, Lovell and Stewart (1975) differed with this view, finding no correlation between elemental sulfur on the mineral surface and natural flotability. Other work in this area has been done

by Clifford, Purdy, and Miller (1974), Trahar (1983), and others, who found evidence demonstrating a correlation between elemental sulfur and flotation. Heyes and Trahar (1984) have recently reported the importance of elemental sulfur in the collectorless flotation of pyrite and pyrrhotite.

Research done by Furstenau and Sabacky (1981) demonstrated that most sulfide minerals, including sphalerite, were naturally floatable when ground in an essentially oxygen-free environment. Their work supported that of other researchers (Rao, 1969; Gaudin, 1957; Rogers, 1962; Gaudin et al, 1959; Sutherland and Wark, 1955; Yonezawa, 1960) demonstrating that copper activation improved the flotation of sphalerite. Recent work (Perry, Tsao, and Taylor, 1984) has shown that copper activation produces a copper-sulfide compound on the mineral surface.

Other researchers have given alternative explanations for collectorless flotation. Yoon (1981) postulated that the use of sodium sulfide produced a sulfur-enriched mineral surface which was able to be floated. This theory has been supported by research done by Luttrell and Yoon (1982), Luttrell (1982), and Luttrell and Yoon (1984a) on chalcopyrite and sphalerite ores. In addition, Luttrell (1982) pointed out that there is an optimum ratio of copper activation to sodium sulfide addition yielding maximum recoveries for sphalerite flotation, verifying an earlier suggestion by Yoon (1981). He also reiterated the importance of potential in chalcopyrite flotation.

Luttrell and Yoon (1983b, 1984 a and b) indicate the presence of

polysulfides on the surface of highly floatable chalcopyrite. Their results indicate the presence of an intermediate oxidation state between sulfide ion,  $S^{2-}$  and elemental sulfur,  $S^0$ . This species, which can be inferred from their X-ray photoelectron spectroscopy (XPS) data, had previously been identified as the sulfur peak from covellite,  $Cu_2S$ , by Buckley and Woods (1981). Since the formation of polysulfides occurs at oxidizing potentials, the theory that they are responsible for flotation is consistent with results obtained earlier by Heyes and Trahar (1977), Gardner and Woods (1979) and, Trahar (1983). Hamilton and Woods (1983) have examined the mechanism for formation of the polysulfides. The assertion that the oxidation of the mineral to form polysulfide responsible for flotation has been recently supported by Hodgson and Agar (1984).

Hamilton and Woods (1984) and Buckley and Woods (1984) have offered a seemingly different explanation for this oxidation product. Instead of polysulfide being responsible for the collectorless flotation, they considered the metal-deficient mineral surface to be responsible for the flotation. They have suggested that this species, though thermodynamically unstable, would have a sulfur lattice structure very similar to that of the unoxidized mineral. By using electrochemical techniques, Hamilton and Woods (1984) were able to identify the potentials involved in the production of these metal-deficient sulfides and relate these potentials directly to the flotation behavior of several sulfide minerals.

SCOPE OF WORK

This investigation has included microflotation of pure sphalerite for the purpose of establishing the effect of redox potential, pH, and copper activation on its flotation behavior without collector. Batch flotation has been carried out on samples of Elmwood Mine sphalerite ore to determine the role of dissolved oxygen, redox potential, and sodium sulfide and copper sulfate additions. Further, an attempt has been made to determine if there exists an optimum ratio between Cu and S additions. In an effort to explain the mechanisms involved in the collectorless flotation of sphalerite, X-ray photoelectron spectroscopy (XPS) was used to examine the samples obtained from microflotation tests. From this data, possible mechanisms for the collectorless flotation of sphalerite have been suggested.

## EXPERIMENTAL

### MATERIALS

#### Ore Samples

The ore samples used for this study were obtained from the Jersey Miniere Elmwood mine in Carthage, Tennessee. The company shipped coarse run-of-mine ore which was crushed upon receipt to -28 mesh and split into 1000 gram lots. These were stored at or below -20 C to minimize surface oxidation. Prior to flotation testing, a bag of sample was removed from the freezer and split into two 500-gram lots, and ground in a ceramic ball mill. The average analysis of the feed ore used in this work was 15.2% Zinc.

#### Pure Minerals

The pure sphalerite sample used for microflotation tests was also from the Elmwood mine as obtained from Ward Scientific Company. Specimens were visually checked for contamination and impure samples were discarded. The pure specimens were then crushed with a mortar and pestle, and the -65 + 100 mesh fraction was obtained by hand screening. The crushed samples were then further examined visually for foreign materials and obvious surface discoloration. All particles that were of a questionable nature were removed with tweezers and discarded.

The -65+100 mesh sample, thus prepared, was placed in a vacuum dessicator and stored under vacuum until use. The pure mineral assayed 65.9% zinc by weight. The complete composition of the sphalerite as

determined by electron microprobe analysis is in table 1.

### Reagents

Sodium sulfide flakes from Fisher Scientific were used as a surface cleaning agent in some batch flotation tests. These flakes were composed of 60 to 62%  $\text{Na}_2\text{S}$ , more than 35% water of crystallization, 1.5% sodium chloride and 2% unspecified sodium salts. For microflotation tests, where higher quality sodium sulfide was required, reagent grade sodium sulfide crystals ( $\text{Na}_2\text{S}\cdot 9\text{H}_2\text{O}$ ), also from Fisher Scientific, were employed. These were carefully scraped and washed with doubly distilled water as recommended by Chen and Morris (1972) to remove surface oxidation products before weighing and dissolution. This high-grade sodium sulfide solution was used to make the pulp potential more reducing. Fresh solution was prepared daily. The certified A.C.S. grade potassium permanganate obtained from Fisher Scientific Company was used to adjust the pulp to more oxidizing conditions in some batch experiments.

When sphalerite activation was desired, certified A.C.S. grade cupric sulfate ( $\text{CuSO}_4\cdot 5\text{H}_2\text{O}$ ) from Fisher Scientific was used.

Dowfroth 250 from Dow Chemical Company was used as frother in all batch flotation experiments. A 1% solution of this frother was prepared to more accurately control the dosage. Lime, sodium hydroxide and hydrochloric acid were used to adjust pH. For some microflotation experiments, buffer solutions were prepared. The pH 4 buffer was made from 0.5M potassium biphthalate. The pH 7 buffer made from sodium

Table 1. Summary of the composition of pure Elmwood sphalerite as determined by electron microprobe analysis.

| <u>ELEMENT</u> | <u>PER CENT BY WEIGHT</u> |
|----------------|---------------------------|
| Zinc(Zn)       | 65.86                     |
| Sulfur(S)      | 32.62                     |
| Gallium(Ga)    | 0.61                      |
| Germanium(Ge)  | 0.49                      |
| Iron(Fe)       | 0.24                      |
| Cadmium(Cd)    | 0.18                      |
| Total          | 100.0                     |

biphosphate ( $\text{KH}_2\text{PO}_4$ ). A solution of 0.05M sodium borate, sodium carbonate, and sodium hydroxide was used to make a pH 10 buffer solution. These chemicals were obtained from various suppliers.

Compressed ultra-pure nitrogen from Airco Industrial Gases was used as the carrier gas for microflotation experiments.

All water used in preparing solutions and in microflotation tests was doubly distilled. A Corning Megapure system was used for second-stage distillation. Batch flotation experiments were conducted in tap water drawn from the Virginia Tech-Blacksburg, Virginia, water system.

## EQUIPMENT

## Electrodes and Meters

Potential Electrodes --The potential across a bright platinum-saturated calomel electrode pair was monitored during flotation experiments. A standard porous calomel electrode was used for all microflotation work. For batch flotation, a reverse-sleeve calomel reference electrode was used to provide improved response time and to lessen the possibility of clogging. The electrodes were refilled weekly with saturated potassium chloride solution.

A pin-type platinum electrode was used for the microflotation potential measurements. In batch flotation, a disc-type platinum electrode was used to increase the surface area presented for slurry contact and to facilitate proper cleaning. These electrodes were developed by Orion Research and supplied by Fisher Scientific.

The platinum-calomel pair was connected, along with the sulfide ion electrodes and the pH electrode, to a Fisher Accumet model 750 digital pH/millivolt meter through a Fisher model 753 electrode switching box. The output from this meter was recorded as a function of time using a Pedersen model 27MR strip-chart recorder.

Oxygen Probe --Dissolved oxygen was monitored in the batch flotation work using a Lazar Research Labs model DO-166 dissolved oxygen probe. This was connected to an Altex model 3500 digital millivoltmeter. The dissolved oxygen levels were recorded as a function of time using a Hewlett-Packard strip-chart recorder.

The membrane and reference solution of the oxygen probe were changed weekly and replaced with manufacturers standard equipment supplied by Cole-Parmer Equipment Company. The probe was zeroed in a 5%  $\text{NaSO}_3$  solution and standardized to atmospheric conditions at the ambient temperature and pressure.

Sulfide Ion Electrode --In selected batch flotation tests, the sulfide ion concentration was monitored using an Orion Research silver/sulfide specific ion electrode coupled with an Orion Research double-junction reference electrode. The outer-chamber electrolyte of the reference electrode was changed daily and the inner-chamber electrolyte was changed weekly as suggested by the manufacturer.

This electrode pair was two-point calibrated using doubly distilled water as a blank and a sodium sulfide solution as a known concentration. The sulfide standard was prepared weekly and prevented from oxidizing using the SAOB II buffer described in the electrode instructions.

pH Electrode --Hydrogen ion concentration was monitored by use of an epoxy body, glass membrane pH electrode from Fisher Scientific. The electrode was calibrated in standard pH buffers obtained from American Scientific Products and Fisher Scientific. The electrode was cleaned between tests using 0.1 N HCl and rinsed thoroughly with doubly-distilled water.

### Microflotation Apparatus

Microflotation was carried out in a set-up identical to that used by Luttrell (1982). The diagram in figure 1 shows the arrangement. The pure mineral samples were floated in a Partridge and Smith type cell (1971) by sparging ultra-pure nitrogen through a medium porosity glass frit at the base of the cell. Gas flow was controlled and measured using a Gilmont micrometer capillary flowmeter. The particle suspension was gently stirred using a Teflon coated magnetic stir bar and a Sybron Nuova II magnetic stirrer.

### Batch Flotation Apparatus

The automated laboratory flotation cell developed by Luttrell and Yoon (1983) was used for all batch flotation experiments. A diagram is included as figure 2. This apparatus required no modifications for this work.

### X-ray Photoelectron Spectroscopy

The x-ray photoelectron spectroscopy (XPS) was done using an XSAM model 600 spectrophotometer manufactured by Kratos. The resolution limit of this machine was 0.46 eV. Additional information regarding this equipment may be obtained by contacting the manufacturer or the Polymer Lab, Department of Materials Engineering, Virginia Tech.

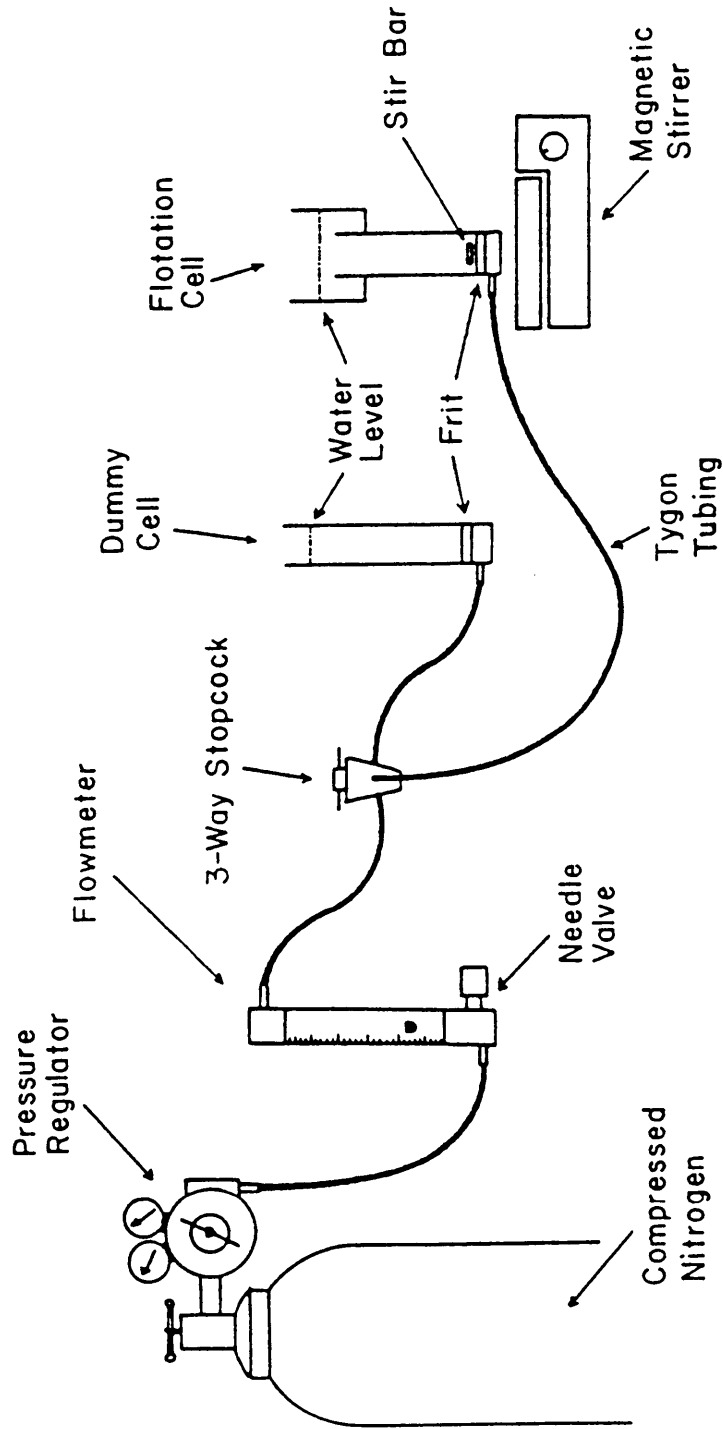


Figure 1. Schematic Diagram of the Microflotation Apparatus (Luttrell, 1982)

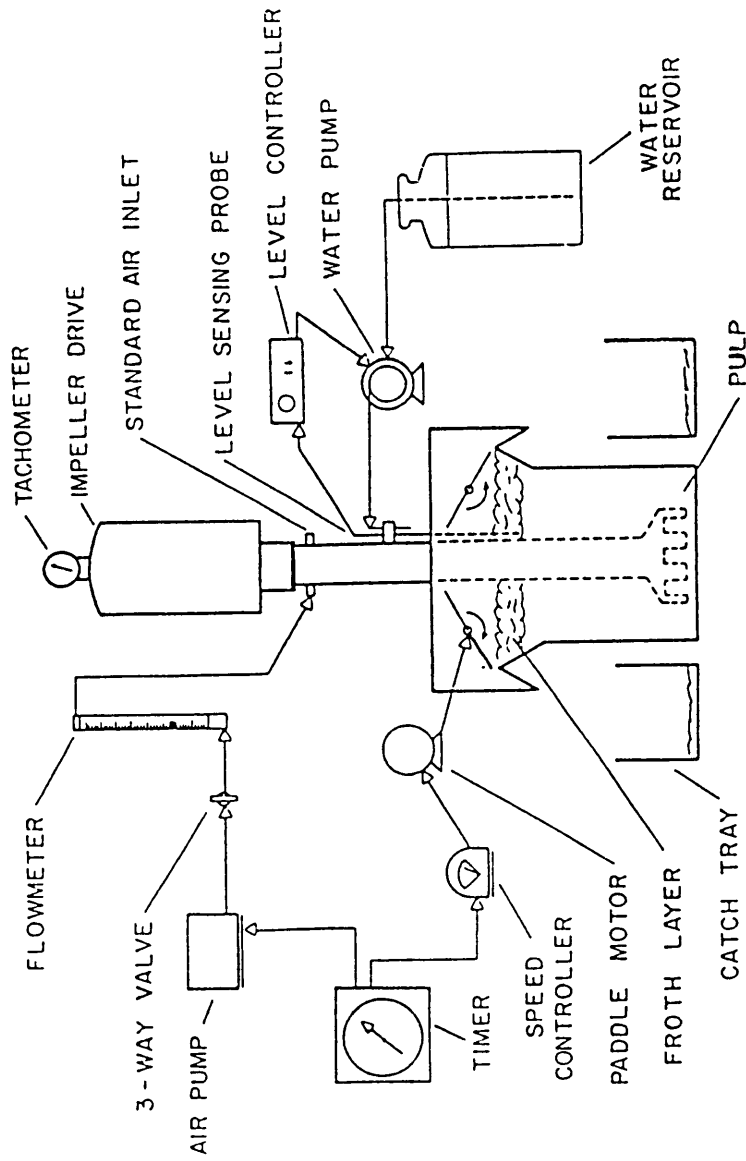


Figure 2. Schematic Diagram of the Batch Flotation Apparatus (Luttrell and Yoon, 1983a).

## PROCEDURE

## Measurement of Operating Variables

Potential --Potentials were measured by immersing a bright platinum-saturated calomel electrode pair into the suspension and reading a millivolt value from the voltmeter. This value was converted to the hydrogen scale by assuming the saturated calomel electrode had a potential of +0.242 volts, as determined by Bates (1964). This so-called "potential" is not intended to represent the reversible Nernst potential (Heyes and Trahar, 1977; Gardner and Woods, 1979). However, in general, positive potentials can be considered to be oxidizing and negative potentials, reducing.

The performance of the electrode pair was regularly checked using a ZoBell solution (Garrels and Christ, 1965). In addition, the surface of the platinum was mechanically cleaned with fine emery paper to ensure that any surface poisoning would be removed.

Dissolved Oxygen --The dissolved oxygen level was monitored by submerging the probe in the flotation pulp. The millivolt reading of the voltmeter was converted to dissolved oxygen content by a relationship where 0.1 volts equals 1 ppm oxygen. The probe was calibrated to atmospheric conditions and checked after each test.

Sulfide Ion --When sulfide ion concentration was measured, the specific ion- reference electrode pair was placed into the pulp

suspension. The meter was standardized to give the S level directly in ppm. This value was measured at the beginning of the experiment, at the onset of conditioning, at the beginning of flotation, and at the conclusion of the experiment in batch flotation tests.

pH --The pH was measured in an identical manner as were sulfide ion concentrations in batch flotation. In microflotation tests, pH was measured coincident with potential during conditioning.

#### Microflotation

The microflotation tests were carried out on pure sphalerite using the apparatus described earlier. In each test, 1.0 g of -65 +100 mesh particles was placed in an 150-ml beaker with approximately 70 ml of doubly distilled water. In the experiments conducted at constant pH, the buffers described earlier were used in place of doubly distilled water.

The potential measuring electrodes were placed in the suspension and the potential was adjusted to the desired level by reagent additions. Once the potential steadied at this level, the suspension was conditioned for 5 minutes. Except for those tests in which buffer solutions were used, no attempt was made to adjust the pH from the "natural pH" that resulted after reagent additions. After conditioning, the mixture was transferred to the flotation cell and flotation immediately begun. A one-minute flotation time was used. Forty ml/min ultra-pure nitrogen was passed through a glass frit for bubble

generation.

For tests where the sphalerite was desired to be copper-activated, the mineral sample was initially placed in a 250-ml Erlenmeyer flask with 50 ml  $\text{CuSO}_4$  solution. This mixture was conditioned for 15 minutes utilizing a wrist-action shaker. After this time, the  $\text{CuSO}_4$  was decanted off and the mineral was rinsed once with doubly-distilled  $\text{H}_2\text{O}$ . Then the sample was transferred to an 150-ml beaker, conditioned and floated as above. After flotation, both the floating and non-floating fractions were filtered, dried and weighed.

#### Batch Flotation

For batch flotation, approximately 500 g samples of the Elmwood ore, prepared as described earlier, were ground in a 10-inch ceramic ball mill with a 50% by volume (5 kg) charge of steel balls and 300 ml tap water. This pulp was transferred to the 2-liter plexiglas cell of the automated flotation machine. The necessary reagents were added, conditions adjusted, and conditioning begun. One minute prior to the starting of flotation, 1.5 ml of a 1% DF 250 solution (0.06 lb/ton) was added to the pulp.

At the appropriate time, the paddles of the machine were activated and the froth product collected for three minutes. This product was mixed with tap water to make up the volume of the cell, and floated to exhaustion as a cleaner stage.

The various pulp parameters were continuously monitored during conditioning and subsequent rougher flotation. In some tests, the level

of dissolved oxygen in the pulp was used to determine the starting point for flotation. In most tests, the pulp was conditioned six minutes after the addition of  $\text{Na}_2\text{S}$  and/or  $\text{CuSO}_4$  dosages.

For the series of tests which were designed to verify the ratio of  $\text{Cu}^{2+}$  to  $\text{S}^{2-}$  necessary for good flotation, the procedure described by Luttrell (1982), was used. The sodium sulfide was added and the pulp conditioned for 10 minutes. The copper sulfate was then added and the pulp conditioned an additional 5 minutes before floating to exhaustion.

In this series of tests, the pH was controlled at  $6.75 \pm 0.1$  by adding HCl as required. However, in the majority of batch tests, the pH was monitored but not controlled. A summary of reagent additions, pH, and other pertinent information for each test is included in Appendix A.

#### X-ray Photoelectron Spectroscopy

The samples for XPS analysis were prepared as for microflotation. More specifically, they were stirred in a beaker and reagents were added to bring potential to the desired level. Copper-activated specimens were in contact with  $5 \times 10^{-5}$  M  $\text{CuSO}_4$  for 15 minutes, then mixed with doubly-distilled water prior to potential adjustment. The treatment of these samples is summarized in table 2.

The samples were then dried in a vacuum dessicator and stored under vacuum until just prior to running the spectra. The powdered sample was then mounted on double-stick tape and placed on the probe for insertion into the spectrophotometer. A wide-scan spectrum was then collected, followed by narrow scans for elements of interest, such as sulfur.

Table 2. Summary of preparation conditions for samples for XPS analysis.

| <u>Sample No.</u> | <u>Weight</u> | <u>Product</u> | <u>Flotability</u> | <u>Potential</u> |
|-------------------|---------------|----------------|--------------------|------------------|
| 1C                | 0.12g         | Concentrate    | 11.65              | +476mV           |
| 2T                | 1.03          | Tailing        | 0.00               | +829             |
| 3T                | 1.05          | Tailing        | 0.00               | -276             |
| 4T*               | 0.97          | Tailing        | 4.90               | +829             |
| 5C*               | 1.08          | Concentrate    | 98.18              | +517             |
| 6T*               | 1.04          | Tailing        | 0.00               | -458             |

\*Copper-activated

## EXPERIMENTAL RESULTS

### MICROFLOTATION

The results of microflotation experiments were interpreted using flotability as the dependent variable. The outcomes were examined to determine the effect of potential,  $\text{Cu}^{2+}$  activation and pH. Flotability was defined as the percentage of the original 1-g sample of pure sphalerite collected in the head of the cell at the end of flotation. A summary of all experiments is included in Appendix B.

#### Effect of Potential

It was shown in this work that the flotation behavior of sphalerite is dependent on the potential of the solution prior to flotation. This agrees with the results obtained by other researchers for other sulfide mineral systems, such as Yoon and Luttrell (1984a) and Gardner and Woods (1979) for the chalcopyrite system.

Figure 3 shows these results for unactivated Elmwood sphalerite. This curve shows that flotation is negligible at potentials less than +200 mV, versus the standard hydrogen electrode (mv, SHE), and reaches a maximum of 35% at +432 mV. It drops off considerably at potentials greater than +600 mV. These results indicate that under certain potential conditions the surface of untreated sphalerite is at least partially hydrophobic.

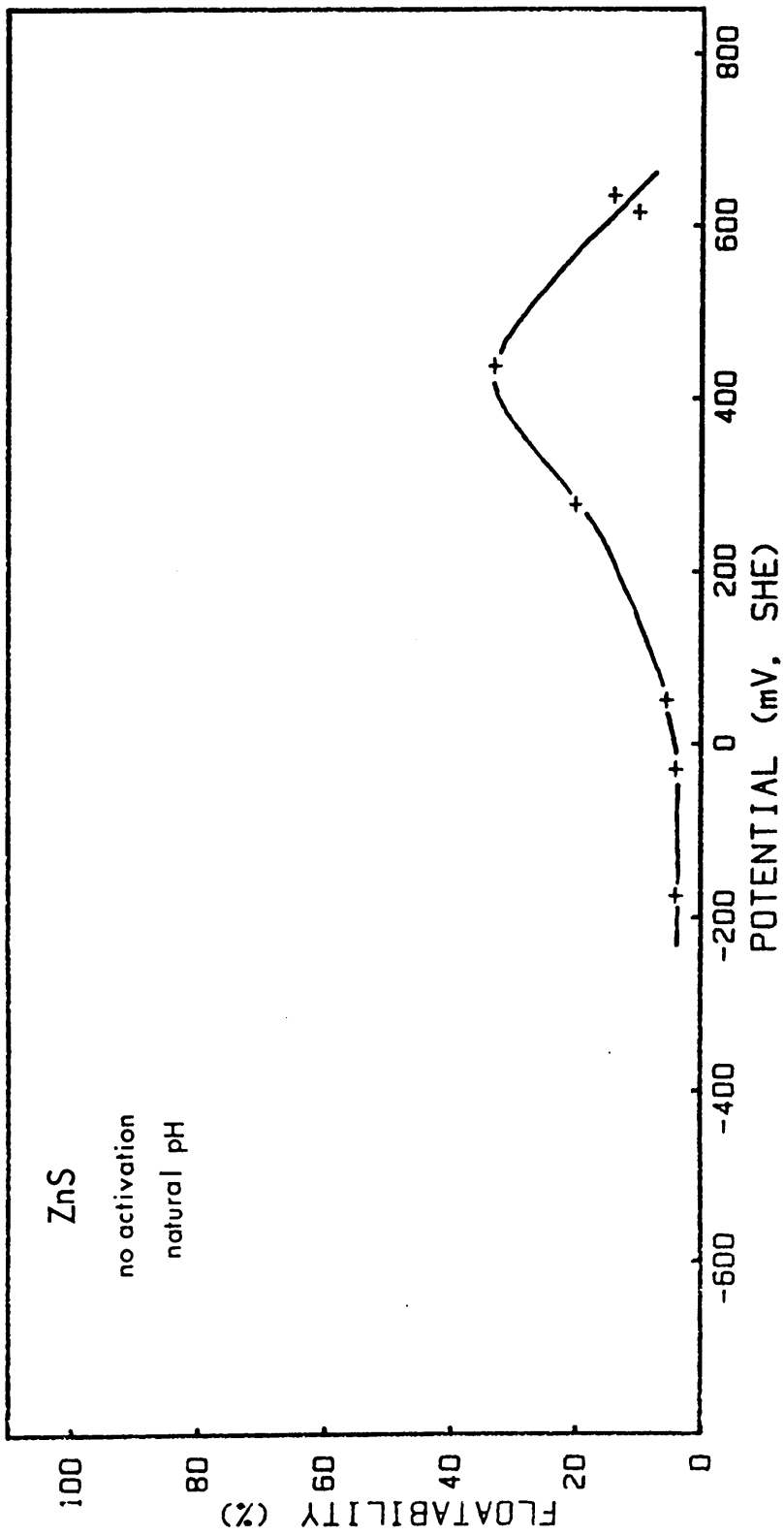


Figure 3. Flotation Behaviour of non-activated Elmwood Sphalerite.

### Effect of Copper Activation

In view of the observations of Furstenau and Sabacky (1981) and others, an attempt was made to improve the flotability of the sphalerite by activating the surface with cupric sulfate ( $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$ ). As was discussed in the procedure section, the mineral sample was treated with various concentrations of  $\text{CuSO}_4$  solutions to provide this activation. The samples thus treated were conditioned at numerous potentials and then floated. The results in figure 4 show that the behavior of activated sphalerite is also dependent on potential. The concentrations of copper used in the activation step determined the exact flotation characteristics of the sphalerite.

At a copper sulfate concentration of  $5 \times 10^{-5}$  M, the range and amount of flotation was drastically improved from the non-activated case. Recovery of 33% of the sphalerite was obtained at potentials as low as 0 mV, SHE. The flotability was above 80% between +200 and +600 mV, reaching a maximum of 91% between +250 and +425 mV.

Increasing the copper sulfate concentration by one-hundred fold to  $5 \times 10^{-3}$  M increased the range of good flotability greatly. More than 40% of the sphalerite floated at potentials down to -300 mV. The flotability was greater than 80% in the potential range of -100 to +450 mV with a maximum of 97% around +150 mV. The recovery decreased rapidly for potentials greater than +450 mV, becoming less than 2% at +800 mV.

Further increase in copper concentration did not improve the flotation, however. At a copper sulfate concentration of  $5 \times 10^{-2}$  M, flotation behavior became complex. After an initial rise in flotability

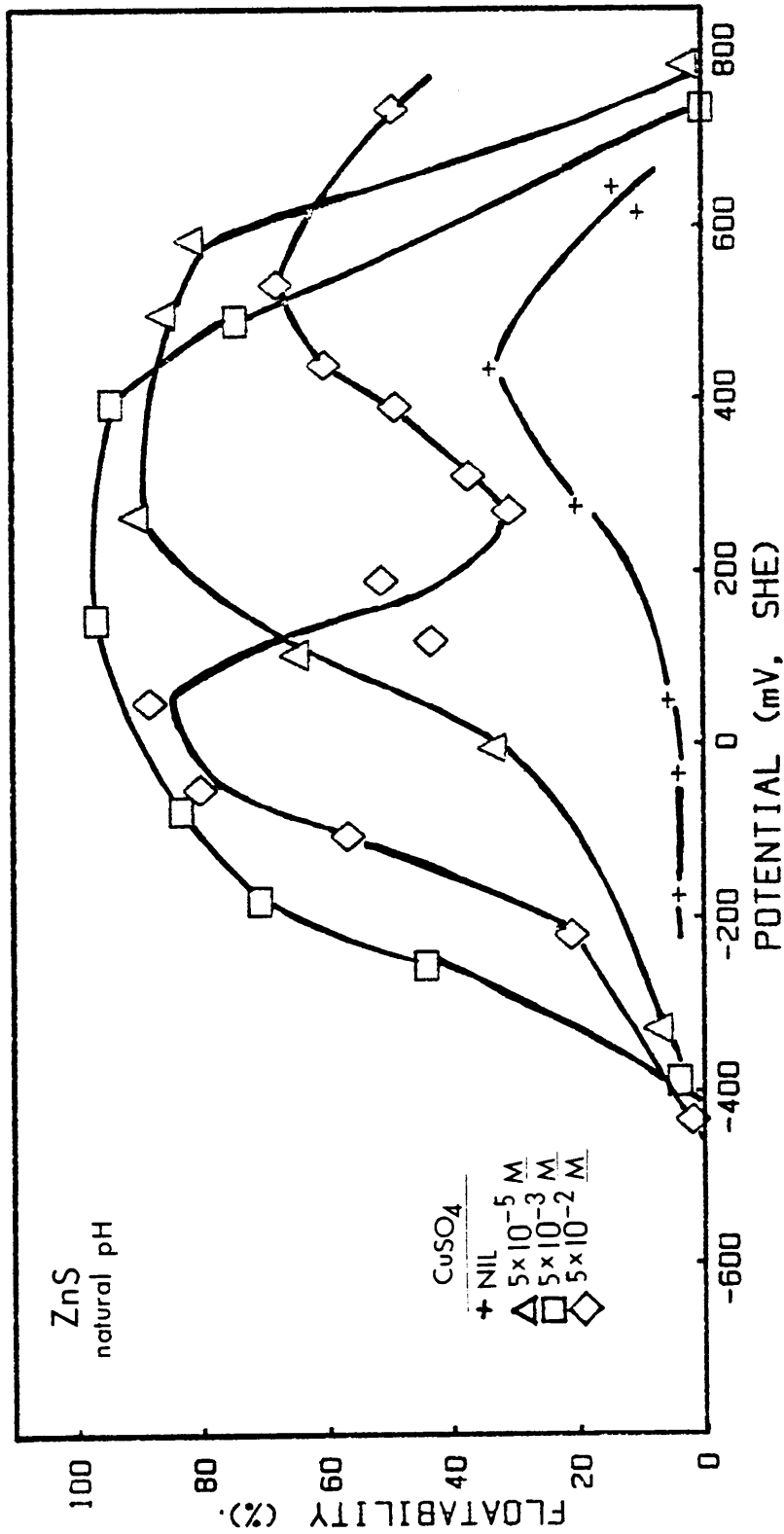


Figure 4. Effect of  $\text{CuSO}_4$  dosage on ZnS floatability at natural pH.  $\text{Na}_2\text{S}$  or  $\text{KMnO}_4$  added to adjust potential.

to a maximum of 88% at +50 mV, the recovery fell to 30% at +275 mV before rising to a second maximum of 67% at +535 mV. This bimodal response indicates that excess copper ions gave rise to two distinct flotation mechanisms, each having its own potential-dependent behavior.

#### Effect of pH

In the above tests, no attempt was made to control the solution pH. To examine the effect of this parameter on the hydrophobicity of pure sphalerite, microflotation experiments were carried out in pH 4, 7, and 10 buffer solutions. Both activated and non-activated series were conducted. The results of these experiments are shown in figure 5.

In both the activated and non-activated cases, the best results were obtained at pH 4. The non-activated sphalerite had a maximum flotability of 67% at approximately +200 mV at pH 4, compared with a 20% maximum for pH 7 and 7% for pH 10. Good flotability was exhibited for all potentials greater than 0 mV for pH4, with 25% of the sphalerite still floating at +1000 mV.

In the activated case, the flotability demonstrated even more variation with pH. At pH 4, the recovery was greater than 80% for all potentials between -100 and +900 mV, approaching 100% at potentials greater than 0 mV. Those tests conducted at pH 7 also showed flotabilities greater than 90% fo potentials between 0 and +700 mV. The recovery dropped drastically at potentials less than 0 mV and greater than +700 mV.

The experiments conducted at pH 10 showed a much narrower range of

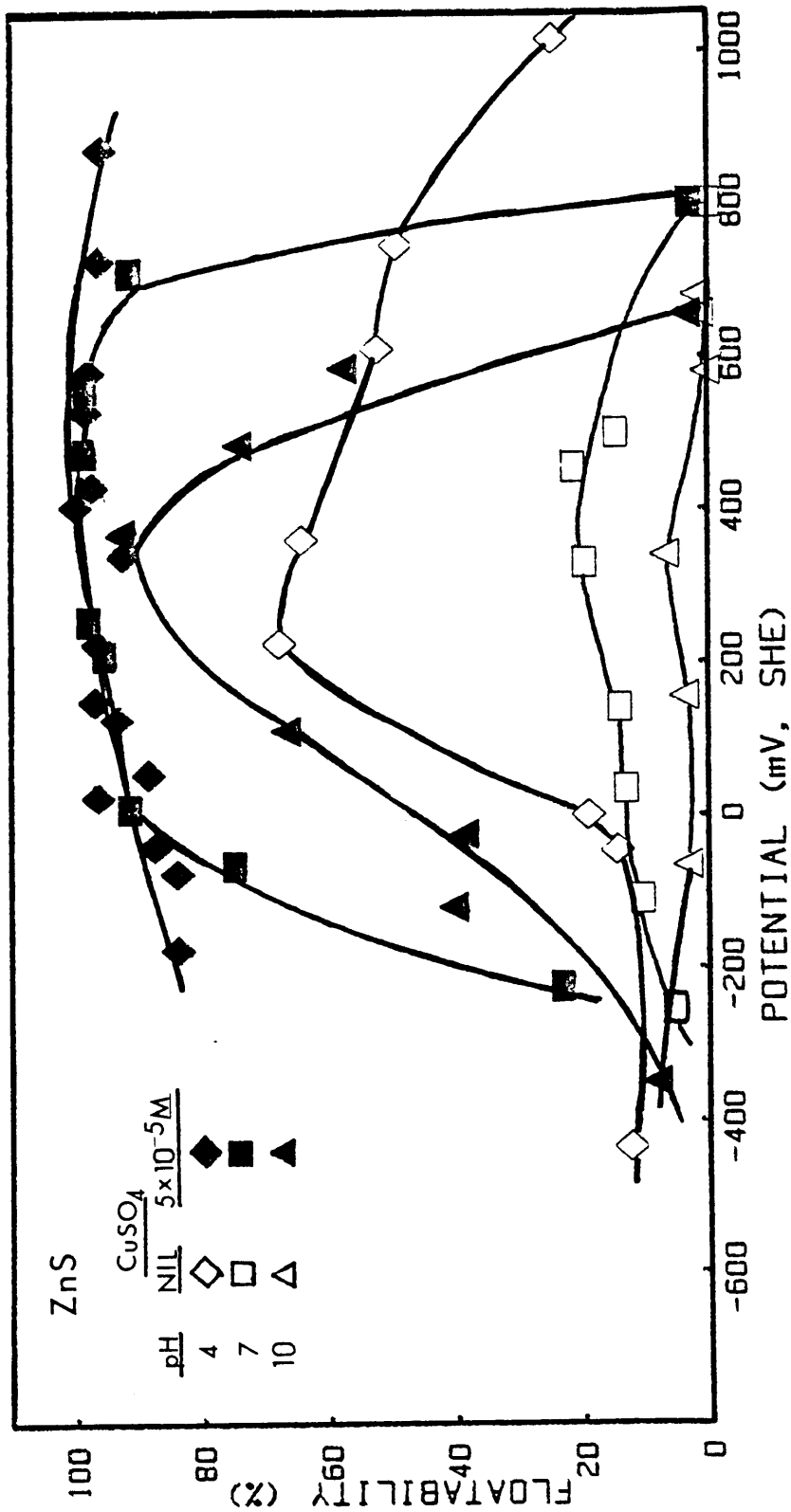


Figure 5. Effect of pH on the floatability of ZnS (non-activated and activated with  $5 \times 10^{-5} \text{ M CuSO}_4$ ).  $\text{Na}_2\text{S}$  or  $\text{KMnO}_4$  added to adjust potential.

good flotability. They exhibited a slow increase in recovery from about 7% at -350 mV to a maximum of about 92% at +350 mV. The flotability decreased rapidly for potentials above the maximum point.

The limit for reducing potentials obtainable in pH 4 and 7 buffers was due to limitations in the chemical system. It was found that addition of  $\text{Na}_2\text{S}$ , in an effort to reduce the potential below the minimum values shown in figure 5, resulted in a marked increase in pH.

## BATCH FLOTATION

Batch flotation experiments were conducted on 500-g lots of Elmwood mine sphalerite ore. Many pulp parameters, e.g. potential, dissolved oxygen, pH, and sulfide ion concentration, were monitored. Further, the effect of various reagent additions were examined. A complete summary of all batch experiments is in Appendix A.

### Effect of Sodium Sulfide Dosage

Figure 6 illustrates the influence of  $\text{Na}_2\text{S}$  and  $\text{CuSO}_4$  addition on flotation recovery of zinc from the Elmwood ore. It can be seen that any addition of  $\text{Na}_2\text{S}$  at  $\text{CuSO}_4$  levels of less than 1 lb/ton, depressed the sphalerite flotation compared to tests where no sodium sulfide was added. At cupric sulfate dosages greater than 1 lb/ton however, the recovery increased if 0.25 lb/ton  $\text{Na}_2\text{S}$  were added. The grade of the flotation concentrate was high for all of these tests. Increasing the Na S dosage to 0.50 lb/ton brought a decrease in flotability. Additional tests conducted at a sodium sulfide addition of 1.5 lb/ton produced zinc recoveries of less than 2.5% at  $\text{CuSO}_4$  dosages even up to 3.0 lb/ton, demonstrating that too much sodium sulfide is detrimental to flotation.

### Effect of Cupric Sulfate Dosage

The effect of cupric sulfate activation on sphalerite recovery is also shown in figure 6. Flotation is enhanced greatly from nil at 0.0 lb/ton addition, to a maximum of 65 to 85% at dosages between 0.5 and

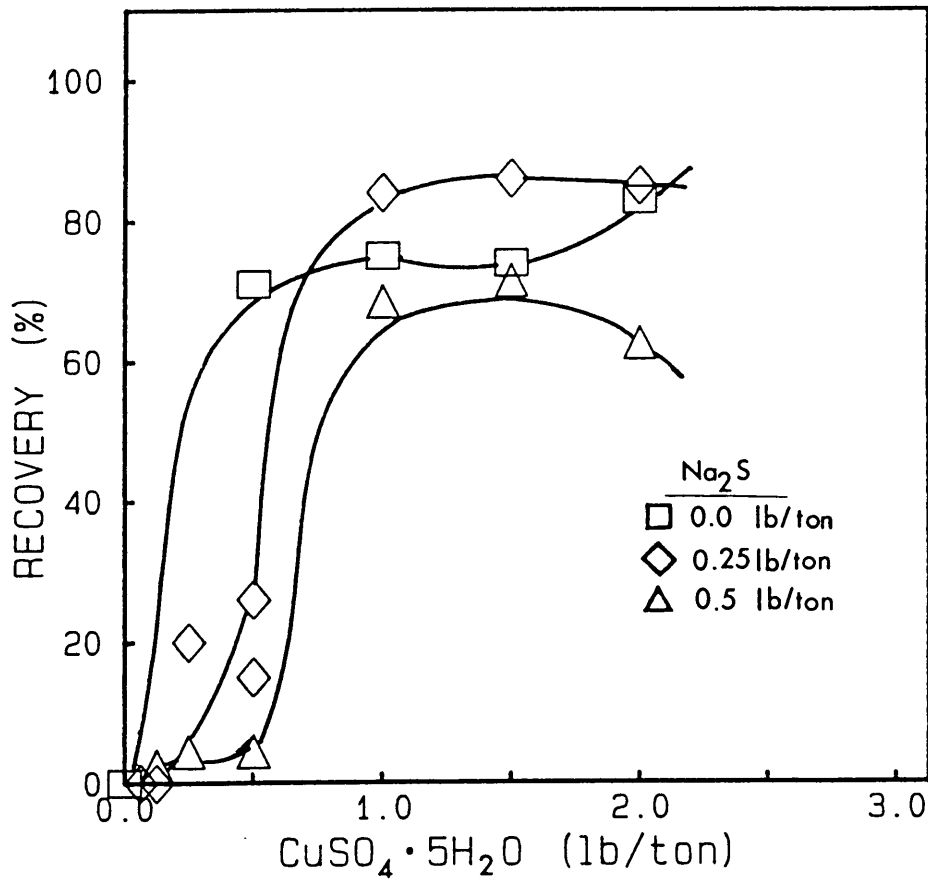


Figure 6. Effect of  $\text{CuSO}_4$  dosage on recovery of Elmwood sphalerite at constant  $\text{Na}_2\text{S}$  addition.

2.0 lb/ton. This general behavior is observed for 0.0, 0.25, and 0.5 lb/ton additions of  $\text{Na}_2\text{S}$ . In the case of 0.5 lb/ton sodium sulfide addition, the recovery reaches a maximum at a 1.5 lb/ton dosage of  $\text{CuSO}_4$ , then falls markedly at 2.0 lb/ton.

#### Effect of Cupric Ion/Sulfide Ion Ratio

As the results in figure 6 indicate, there is a relationship between both copper sulfate and sodium sulfide additions and zinc recovery in the Elmwood sphalerite system. This relationship is more clearly demonstrated in figure 7. Using an identical technique as Luttrell (1982), results showing the most favorable ratio of added copper ions to added sulfide ions for optimum recovery were obtained. The maximum recovery was obtained at atomic ratios of 0.31. The recovery curves were identical for Na S additions of both 0.25 and 0.50 lb/ton. The grade curves showed a slight difference, however.

#### Effect of Dissolved Oxygen

To examine the importance of dissolved oxygen in collectorless flotation, several experiments were conducted in which various methods were used to bring the pulp oxygen concentration to a level of 9 ppm. This level was chosen by examining the dissolved oxygen level at flotation for a test in which good recovery was obtained for a high-grade sphalerite product, as can be seen in figure 8. A summary of the most significant of these tests conducted at 9 ppm is included in table 3. The recoveries and grades for cleaner flotation show little

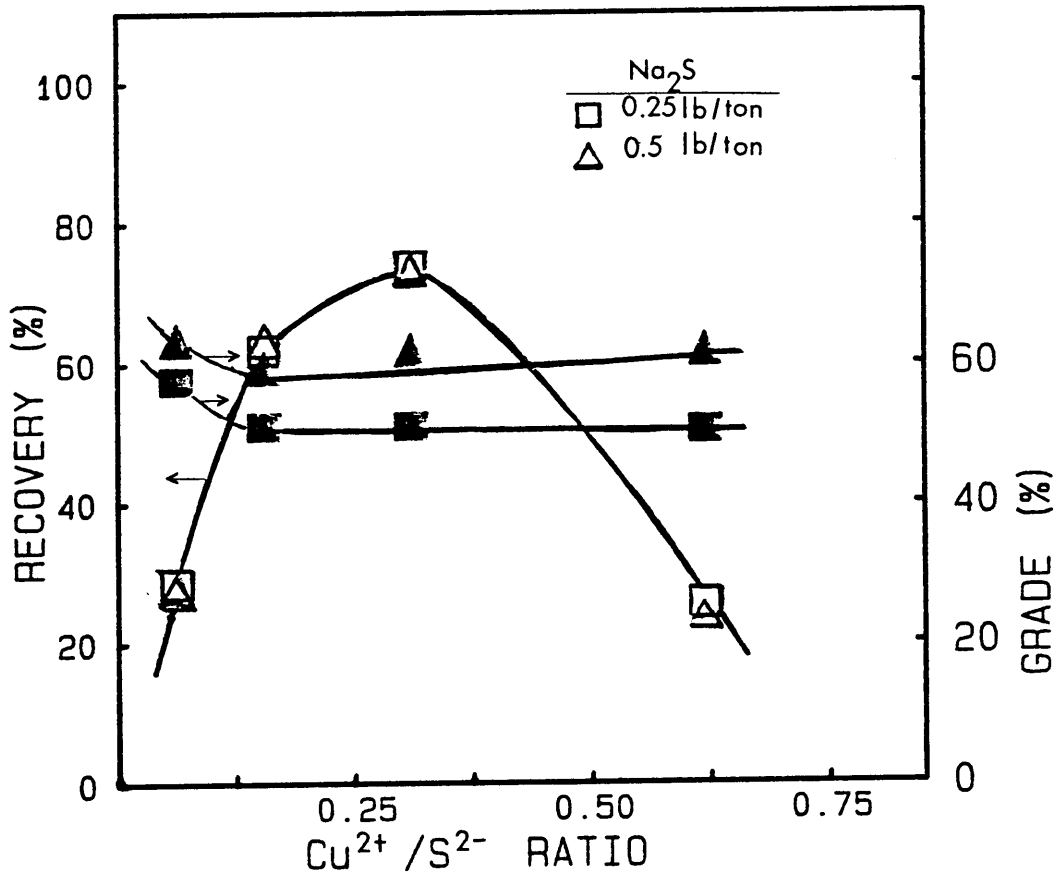


Figure 7. Effect of  $\text{Cu}^{2+}/\text{S}^{2-}$  ratio on recovery and grade of Elmwood sphalerite at pH 6.75.

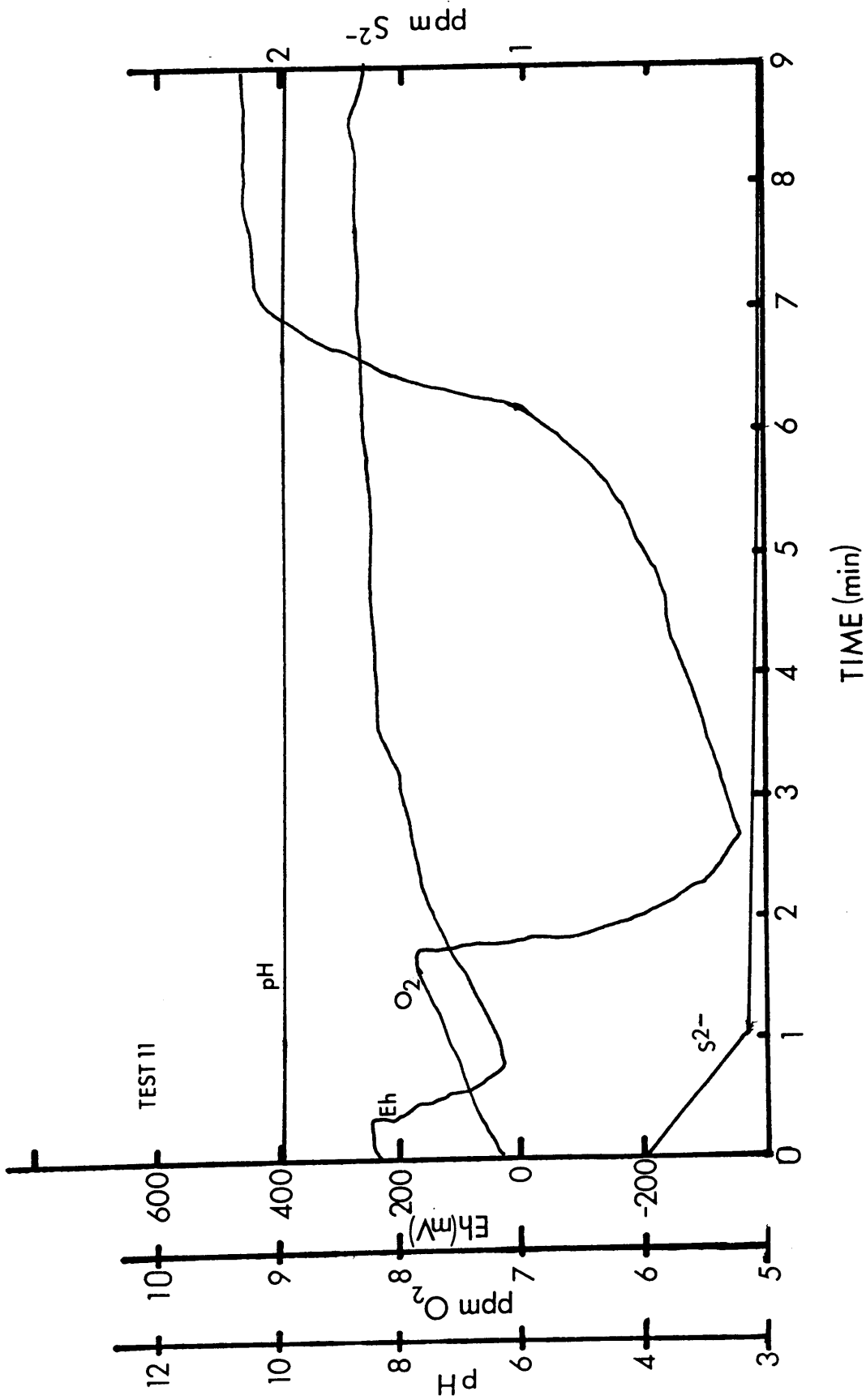


Figure 8. Record of operating variables for a test yielding high grade and recovery.

Table 3. Recovery and grade for flotation experiments conducted using 0.5 lb/ton  $\text{Na}_2\text{S}$  and 1.0 lb/ton  $\text{CuSO}_4$  under different conditions.

| <u>Test</u> | <u>Conditions</u>  | <u>Product</u> | <u>Grade</u> | <u>Recovery</u> |
|-------------|--|----------------|--------------|-----------------|
| 11          | Conditioned without air for 5 minutes  | Rougher        | 37.46        | 75.66           |
|             |  | Cleaner        | 54.09        | 69.20           |
| 30          | Conditioned without air until dissolved oxygen reached 9 ppm (12 min.)   | Rougher        | 38.37        | 73.79           |
|             |  | Cleaner        | 57.83        | 67.96           |
| 31          | Conditioned with air until dissolved oxygen reached 9 ppm (3 min.)   | Rougher        | 49.18        | 73.56           |
|             |  | Cleaner        | 55.06        | 67.76           |
| 32          | Conditioned without air 10 minutes, $\text{KMnO}_4$ added to bring dissolved oxygen to 9 ppm. (0.07g $\text{KMnO}_4$ ) | Rougher        | 42.29        | 90.57           |
|             |  | Cleaner        | 54.77        | 68.46           |

variation.

In test 32, the pulp was conditioned for 10 minutes and then 0.07 g of  $\text{KMnO}_4$  added. This produced an immediate increase in dissolved oxygen. The rougher concentrate in this test had a recovery of 90.57% at a grade of 42.29% Zn. This greatly exceeded the rougher recoveries obtained in the other tests and indicates the importance of dissolved oxygen, which is important in determining the pulp potential.

#### Effect of Permanganate Addition

In the series of tests conducted to examine the effect of potassium permanganate, the results indicate that there is an optimum level of permanganate addition. At a level of 0.2 lb/ton, the grade of the concentrate was 55.88% Zn at a recovery of 29.58%. When the dosage of  $\text{KMnO}_4$  was increased to 0.8 lb/ton, the grade decreased to 43.84% Zn, but the recovery was increased to 39.74%. Further increases in permanganate addition to 1.4 and 2.0 lb/ton gave very inferior results, with grades and recoveries indicating sphalerite depression.

#### Effect of Potential

As figure 8 and those in appendix D show, the pulp potential was monitored for all of the previously discussed batch flotation tests. The flotation data can be correlated to potential as well as the factors already examined by using this data. In general, the flotation of the ore was similar to that of the pure mineral observed in the microflotation tests. Specifically, the best flotation was obtained

when the pulp potential at the beginning of concentrate recovery was in the range of 0 to +600 mV SHE. Pulp potentials outside of this range greatly depressed the flotation of the sphalerite, although many other factors also affected the flotation behavior in a given test.

#### X-RAY PHOTOELECTRON SPECTROSCOPY

X-ray photoelectron spectroscopic (XPS) analysis was conducted on pure sphalerite samples after they had undergone microflotation testing. This surface analysis was done to identify the species that might have been responsible for flotation or the lack of flotation. In the cases where good flotability was observed, the microflotation concentrate was subjected to the analysis. In the other cases, the unfloated material was used. A standard of pure sulfur was also examined.

The raw spectra were deconvoluted using software developed by Kratos for use on their system. This provided detailed information regarding the oxidation states of sulfur present on the surface. Complete spectra and information for each sample is contained in appendix C. An example of raw and curve resolved Sulfur 2p spectra for sample 5C is included as figures 9 and 10.

Using accepted charge correction techniques, the binding energy of each peak was determined. This involved determining the binding energy of the carbon 1s peak and comparing it to the standard value of 284.9 eV. This difference was assumed to represent the shift in binding energies of all other peaks. When this shift was used to adjust the peak location of the curve-resolved S-2p peaks, the peaks could be

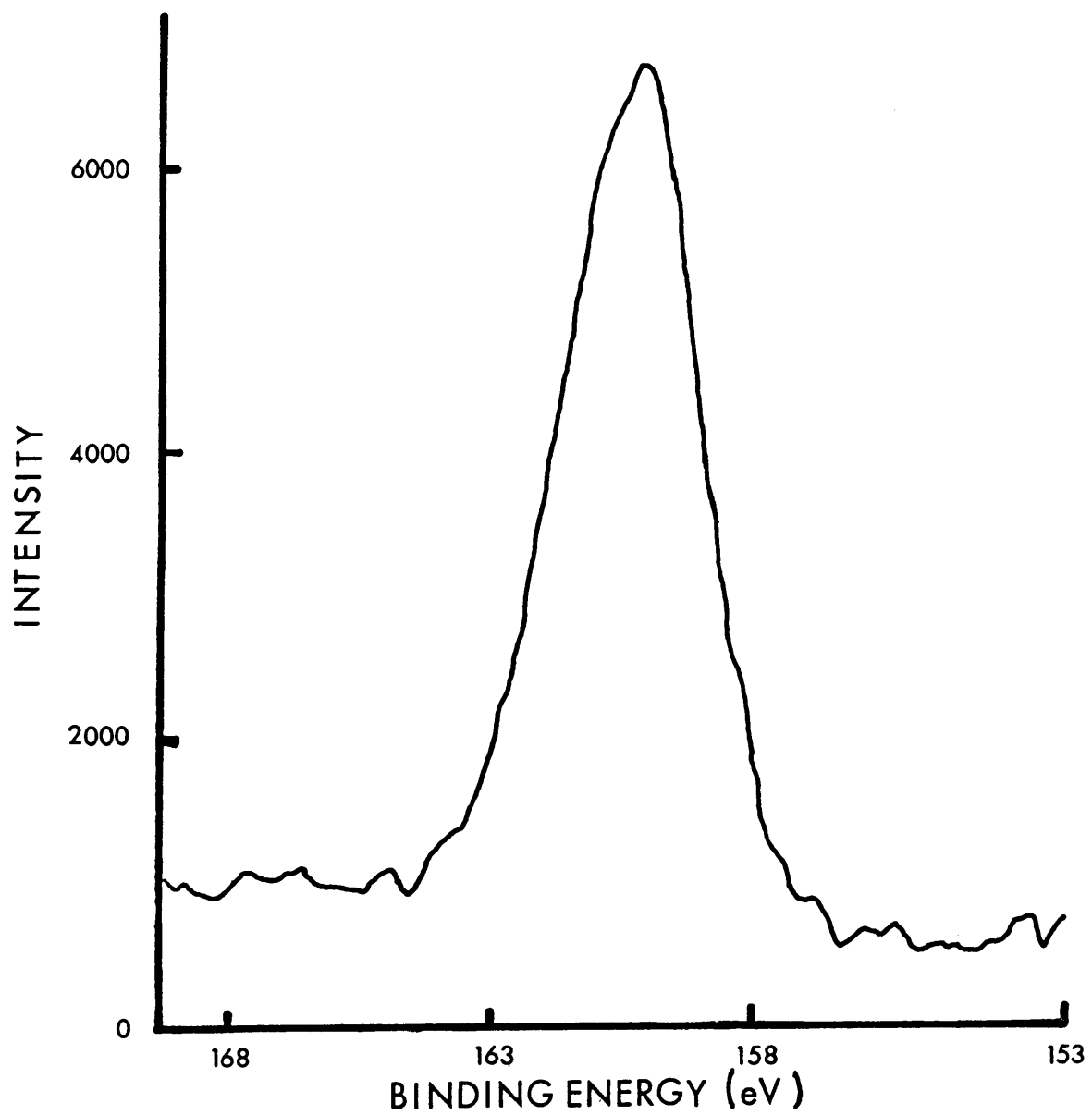


Figure 9. Raw Sulfur-2p spectra for sample 5C of pure sphalerite.

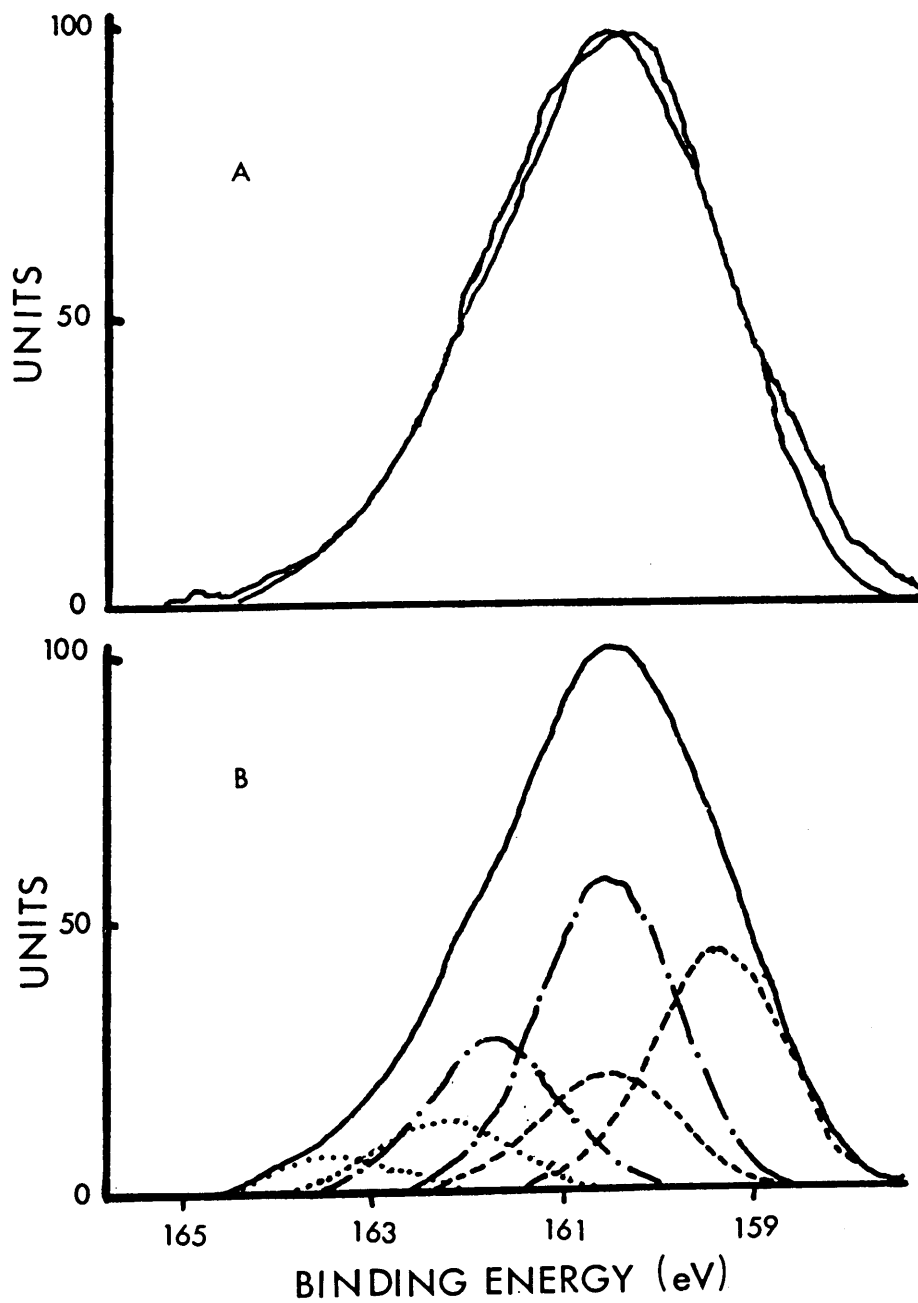


Figure 10. Sulfur-2p spectra for sample 5C of pure sphalerite. (A) Curve smoothed (B) Peak resolved.  $\cdots$   $S^0$ ,  $-\cdot-\cdot-$   $S_x^{2-}$ ,  $----$   $S^{2-}$ .

compared to literature values for identification of the species present. Table 4 gives those peak identifications.

In addition, the ratio of peak areas of the different sulfur species gives a measure of the relative amounts of each on that sample surface. These relative abundances are included in table 5. As it can be seen, the samples that were treated at high oxidizing potentials (2T and 4T) have a lot of oxidized surface species such as  $S^0$  and other more oxidized hydrophillic species, such as  $S_2O_3^{2-}$  and  $SO_4^{2-}$ , while samples 3T and 6T, which were treated at extremely reducing potentials, show a higher percentage of sulfur species in lower oxidation states. In addition, sample 5C, which alone showed good flotabilty, was the only sample to show the presence of  $S^-$  or  $S_x^{2-}$  on the surface. This seems to indicate a strong correlation of this species to flotability.

Table 4. Identification of surface sulfur species present on Elmwood sphalerite after various treatments as determined by XPS analysis.

| Sample | Corrected Peaks     |                     | Identification                | Reference |
|--------|---------------------|---------------------|-------------------------------|-----------|
|        | S-2p <sub>1/2</sub> | S-2p <sub>3/2</sub> |                               |           |
| Sulfur | 163.3               | 162.1               | S <sup>0</sup>                | 1,2       |
| 5C     | 163.3               | 162.1               | S <sup>0</sup>                | 1         |
|        | 161.6               | 160.4               | S <sub>x</sub> <sup>2-</sup>  |           |
|        | 160.4               | 159.2               | S <sup>2-</sup> (w/ Cu)       | 3         |
| 1C     | 163.5               | 162.3               | S <sup>0</sup>                | 4         |
|        | 162.4               | 161.2               | S <sup>2-</sup> (w/ Zn)       |           |
| 2T     | 164.2               | 163.0               | SO <sub>3</sub> <sup>2-</sup> | 5         |
|        | 163.3               | 162.1               | S <sup>0</sup>                |           |
|        | 162.0               | 160.8               | S <sup>2-</sup> (w/ Zn)       |           |
| 4T     | 164.7               | 163.5               | SO <sub>3</sub> <sup>2-</sup> |           |
|        | 163.3               | 162.1               | S <sup>0</sup>                |           |
|        | 160.9               | 159.7               | S <sup>2-</sup> (w/ Cu)       |           |
| 3T     | 163.4               | 162.2               | S <sup>0</sup>                |           |
|        | 162.4               | 161.2               | S <sup>2-</sup>               |           |
| 6T     | 163.5               | 162.3               | S <sup>0</sup>                |           |
|        | 162.4               | 161.2               | S <sup>2-</sup>               |           |

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References

- |                    |  |
|--------------------|--|
| 1. Luttrell (1982) | 3. Langer, Helmer, and Weichert (1969) |
| 2. Nordling (1972) | 4. Vesely and Langer (1971)            |
|                    | 5. Hercules (1970)                     |

Table 5. Relative surface abundance of various sulfur species present on Elmwood sphalerite after microflotation as determined by XPS analysis.

| Sample | Copper<br>Activ. | Abundance of Species<br>(% of Total S on Surface) |                              |                |                               | Pot. | Flot. |
|--------|------------------|---|------------------------------|----------------|-------------------------------|------|-------|
|        |                  | S <sup>2-</sup>                                   | S <sub>x</sub> <sup>2-</sup> | S <sup>0</sup> | SO <sub>3</sub> <sup>2-</sup> |      |       |
| 5C     | yes              | 38.52   | 49.53                        | 11.95          | -----                         | +517 | 98.18 |
| 1C     | no               | 74.81   | -----                        | 25.19          | -----                         | +476 | 11.65 |
| 2T     | no               | 18.10   | -----                        | 54.60          | 27.30                         | +829 | 0.00  |
| 4T     | yes              | 4.56  | -----                        | 67.92          | 27.52                         | +829 | 4.90  |
| 3T     | no               | 71.61   | -----                        | 28.39          | -----                         | -276 | 0.00  |
| 6T     | yes              | 72.14   | -----                        | 27.86          | -----                         | -458 | 0.00  |

## DISCUSSION

### FLOTATION

The results of microflotation experiments presented in previous sections demonstrate that pure sphalerite can be floated without use of collector, primarily at potentials above 0 mV, SHE. In order for this flotation behaviour to be significant, however, the sphalerite must be copper-activated. This activation, which can be represented by:



(Sutherland and Wark, 1955), changes the potential range where collectorless flotation is possible as well. The critical potential for flotation onset can be lowered to below 0 mV by increasing the cupric sulfate dosage added.

The pH is also a factor in determining the potential range where collectorless flotation is possible (see figure 5). In addition, the pH greatly influences the maximum flotability obtained. A wider range of good flotability was obtained at pH 4 where nearly 100% flotability was observed at potentials between -100 and +900 mV for activated sphalerite. The potential range for maximum flotation at pH 10, on the other hand, was extremely narrow, with the point of maximum recovery of about 92% occurring at +350 mV. This behavior seems to agree with Luttrell and Yoon's (1984a) reasoning that pH may be important in forming hydrophobic surface species such as elemental sulfur or polysulfides, and is consistent with the findings of Clifford, Purdy and Miller (1974).

Similar trends in behavior were noted for batch flotation of Elmwood sphalerite ore. The ore also required copper-activation to float without collector. The ore also required the addition of  $\text{Na}_2\text{S}$  as well to obtain maximum recovery and grade. This upholds observations by Yoon (1981) and Luttrell and Yoon (1984).

The effect of  $\text{Cu}^{2+}/\text{S}^{2-}$  ratio was shown in figure 7. The ratio, previously noted by Luttrell (1982) as producing the best results at a value of 0.17, was found in this case to have a different point yielding optimum recovery, 0.31. This difference may be attributed to the difference in the characteristics of the ores used in these two studies. It is important to note that just as Luttrell found, the flotation behavior was independent of reagent dosages if this ratio were maintained. The findings of this work generally concur with those of Luttrell's work, and thus, his argument relating the cupric ion/sulfide ion ratio to the semi-conducting properties and the resulting flotability of the sphalerite surface is reinforced. That is to say that, because of the insulating nature of sphalerite (band gap 3.67 eV)(Teichman, 1964), copper-activation may be necessary to create a semi-conducting surface which can become involved in electrochemical oxidation reactions. Ralston, Alabaster, and Healy's (1980) work indicating that surface formation of elemental sulfur is closely related to the semi-conducting properties of the mineral seems to support this contention.

The results of the present study also support the findings of Gaudin (1932) indicating that in general sodium sulfide and potassium

permanganate act as depressing agents in flotation. The results presented here provide further support for the evidence presented by others that this is due to the effect these reagents have on pulp potential.

## MECHANISMS

## Induced Hydrophobicity by Oxidation

Elemental Sulfur --Metal sulfides have long been thought to be thermodynamically unstable in the presence of oxygen. Because of this instability, their surfaces are readily oxidized under the prevailing conditions of flotation circuits. Sphalerite behaves slightly differently since, unlike most metal sulfides, it is an insulator rather than a semi-conductor. This electronic property is changed, however, when the surface of the ZnS is copper-activated. Because it is believed that the copper ions replace the zinc in the sphalerite lattice during activation, the surface essentially becomes CuS. The surface, which would now be semi-conducting, would be more easily oxidized to form elemental sulfur.

The XPS spectra show the presence of  $S^0$  on all samples, regardless of the treatment conditions used. The amount of elemental sulfur on the surface (based on peak intensities and distribution of each species) varied from sample to sample. The samples treated at high oxidizing potentials showed the most sulfur as expected. Unexpectedly, however, the samples treated under reducing potentials had the next highest amounts. This was probably due to the oxidation of sulfide ions present in a film of sodium sulfide solution adhering to the mineral surface during the drying of the sample. The samples which exhibited the best flotability appeared to have the least amount of  $S^0$  on the surface.

The possibility that some sample oxidation took place during the drying prior to XPS analysis, raises doubts about the validity of the

interpretation of the data to explain flotation behavior. The samples which floated well without  $\text{Na}_2\text{S}$  added may provide more reliable information. Further, the presence and relative amounts of other sulfur-containing species agrees well with expected behavior at the treatment potentials.

The apparent lack of correlation between elemental sulfur on the surface and flotation behavior would seem to negate the importance of elemental sulfur as a flotation inducer. However, since the large quantity of  $\text{S}^0$  on the surface of the reduced samples (3T and 6T) can be explained by the oxidation of extraneous  $\text{S}^{2-}$  ions upon drying, and the lack of flotability of the oxidized samples (2T and 4T) by the presence of higher-oxidation-state, strongly hydrophillic sulfur species such as  $\text{S}_2\text{O}_3^{2-}$ , the assertion that the observed flotability of samples 5C and 1C is due to the presence of elemental sulfur, is a valid one. It is also consistent with the proposal of Gardner and Woods (1979) that elemental sulfur is responsible for collectorless flotation in the chalcopyrite system. Recent work by Heyes and Trahar (1984) has demonstrated that the collectorless flotation of pyrite and pyrrhotite is related to the production of elemental sulfur on the surface, lending further support to this proposal.

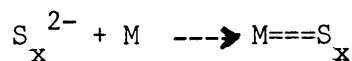
In addition, the results indicating that sphalerite floats better at acidic pH's is consistent with this theory. Many researchers have documented that the oxidation of sulfide minerals in acidic solutions results in the formation of elemental sulfur (Vizsolyi, Veltman, and Forward, 1963; Sato, 1966; Majima and Peters, 1966; Eadington and

Prosser, 1969; Bjorling, 1973). Thermodynamics also favors the increased stability of this elemental sulfur formed at pH's less than 7.5 (Garrels and Christ, 1965).

Polysulfides or Metal-Deficient Sulfides --As it can be seen in figure 10 and tables 3 and 4, the surface of sample 5C has a considerable amount of a sulfur species with a binding energy and oxidation state intermediate to  $S^{2-}$  and  $S^0$ . This species is best identifiable as a polysulfide. Polysulfides can form as a result of the interaction of sulfur with an aqueous solution of sulfide (Chen and Morris, 1972) or by the aging or oxidation of sulfides or hydrosulfides in solution (Karchmer, 1970). Chen and Gupta (1973) have also demonstrated that when sulfur is produced in the presence of sulfide, polysulfides are immediately formed in some cases.

The pH of a system is of extreme importance in determining the stability of polysulfides. Chen and Morris (1972) and Chen and Gupta (1973) discovered that the concentration of polysulfides in a pH 8 solution is several times greater than that of elemental sulfur. The situation is reversed at pH 6 however. The concentration of polysulfides should increase with increasing alkalinity, according to the mass balance calculations done by Chen and Morris.

It has been demonstrated (Allen and Hickling, 1957) that polysulfides can adsorb on a metal surface through the following



where  $S_x^{2-}$  represents the polysulfide and M the metal. Chen and Morris

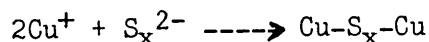
(1972) showed evidence that the oxygenation of mildly alkaline sulfide solutions is 100 times greater in the presence of transition metals. It is very possible that transition metal sulfides could behave in much the same way. This metal-polysulfide complex would probably be hydrophobic since the bonding of the sulfur atoms in the polysulfide chain is very much like that in elemental sulfur.

Thus polysulfides may also play important roles in the oxidation and flotation of copper-activated sphalerite as well as other sulfides. They may also be important components of electrochemical reaction systems involving sulfide minerals. Similar electrochemical behavior has been suggested for both production of elemental sulfur and polysulfides on galena surfaces (Ho and Conway, 1978). This would seem to indicate that it is difficult to differentiate between the two processes electrochemically.

The sample 5C was prepared in distilled water with a pH presumably of around 7. This corresponds to the optimum pH for formation of polysulfides from a sulfide solution (Chen and Morris, 1972). Since sphalerite is one of the most soluble sulfides, in the absence of oxygen, (Yoon, 1981) it is conceivable that enough  $S^{2-}$  ions are present to form these polysulfides.

Additional support is provided by the XPS data presented earlier (Table 3) which shows a peak at approximately 161.6 eV. This corresponds roughly to the peak assigned previously by Buckley and Woods (1981) and Luttrell (1982) as being  $Cu_2S$ . But as Luttrell points out, this peak could also be considered as an indication of the surface

presence of metal polysulfides. He suggests these may form by the reaction



where  $\text{Cu}^+$  represents the copper ions in the surface lattice and  $x$  represents the number of sulfur atoms in the polysulfide chain (usually 2-5). This is entirely possible since the formation of polysulfides from the  $\text{CuS}$  present on the surface would probably produce  $\text{Cu}^+$  ions on the surface by the reaction



where  $X$  and  $Y$  are probably equal to provide charge balance. Since the oxidation state of the sulfur in the middle of the polysulfide complex is nearly 0 and that on the end sulfur atoms is approximately  $-1$ , the XPS spectra may show these two peaks (Luttrell, 1982). The peaks at 161.6 and 163.3 eV for sample 5C correspond very closely to those indicated by Luttrell.

More recently, Hamilton and Woods (1983, 1984) and Buckley and Woods (1984) have attributed these intermediate peaks to the presence of a metal-deficient sulfide which they consider important to collectorless flotation. Their XPS data shows that the copper contained in this surface compound is present as copper (I), implying a formal oxidation state for the sulfur of  $-1/2$ , the same as is found in the polysulfide,  $\text{S}_4^{2-}$ .

From this data, an electronic structure of  $\text{Cu}_2\text{S}_4$  was theorized for the metal-deficient sulfide found on the chalcopyrite. They also assumed that all of the sulfur atoms would have the  $-1/2$  formal valence

state.

Other recent work (Perry, Tsao, and Taylor, 1984) suggests that the surface compound formed on copper-activated sphalerite is a copper (I) sulfide based on XPS and Auger parameter data. This compound, though identified as possibly being chalcocite ( $\text{Cu}_2\text{S}$ ) could actually be a metal-deficient sulfide. Thus, the copper-activation of sphalerite at appropriate oxidizing potentials could produce a metal-deficient sulfide which would cause flotation behavior such as was found for sample 5C.

Since, in general, polysulfide and metal-deficient sulfide formation is enhanced at higher pH, it is possible that these species are responsible for collectorless flotation in alkaline or nearly neutral solutions. The flotation behavior observed in acidic solutions may be caused by elemental sulfur, on the other hand.

### Inherent Hydrophobicity

The results of the microflotation tests conducted on pure and copper-activated sphalerite indicate that flotability decreases with increasing additions of sodium sulfide and thus with decreasing potential. To better understand the role that sodium sulfide addition plays in this system, it is important to consider the proportion of total sulfide present as  $S^{2-}$ ,  $HS^-$ , or  $H_2S$  in aqueous solutions as a function of pH. Jones and Woodcock (1978) demonstrated that  $HS^-$  ions make up the greatest portion of the total sulfide at pH values between 7 and 13. Therefore, it might be considered that hydrosulfide ions are depressing the natural flotation of sphalerite and copper activated sphalerite under the reducing conditions.

$HS^-$  ions have long been identified as depressants in xanthate flotation because of the competition of the hydrosulfide and xanthate ions for the mineral surface (Gaudin, 1957). Gardner and Woods (1979) and Trahar (1983) claimed that  $HS^-$  ions produce a reducing environment that prohibits the formation of elemental sulfur in collectorless flotation. Luttrell (1982) suggests alternatively that the adsorption of hydrosulfide ions on the mineral surface might render it hydrophilic. The S-H group can act as a proton donor and thus easily form hydrogen bonds (Vinogradov and Linnel, 1971) and, if the water molecules surrounding the mineral are proton acceptors, the adsorbed  $HS^-$  would give the mineral an hydrophilic character.

This argument implies that elemental sulfur may not be required for collectorless flotation. Failure to find a strong correlation between

elemental sulfur present on the mineral surface and flotability has been the case in much previous work (Finklestein et al, 1975; Heyes and Trahar, 1977; Pritzker, Yoon, and Dwight, 1980; Furstenau and Sabacky, 1981; and Luttrell, 1982). As was previously discussed, the present results cannot be easily interpreted using the elemental sulfur theory alone.

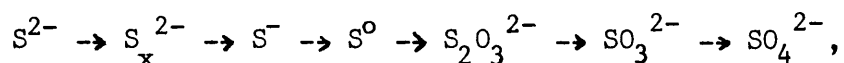
It has previously been indicated that the flotation response for both activated and non-activated sphalerite is best in acidic pH. If HS is responsible for hydrophilic depression of the mineral, this improved flotability may be due to the removal of  $\text{HS}^-$  from the system as  $\text{H}_2\text{S}$  gas. Of course, acidic conditions also favor the formation of elemental sulfur.

Under oxidizing conditions, the mineral may be depressed in a different way. At high oxidizing potentials, XPS shows large amounts of elemental sulfur, yet neither of these samples was floatable. This lack of flotability can be explained; at high oxidizing potentials, hydrophilic species such as  $\text{CuO}$ ,  $\text{CuSO}_4$ ,  $\text{CuS}_2\text{O}_3$ , and  $\text{MnO}_2$  are readily formed as shown by XPS spectra. These species adversely affect the hydrophobic nature of the sphalerite surface.

The assertion that some species (e.g.  $\text{HS}^-$ ,  $\text{CuSO}_4$ ) is rendering the mineral surface hydrophilic is based on the assumption that clean sulfides are inherently hydrophobic. The increased flotability of copper-activated sphalerite could be explained on the basis that the  $\text{CuS}$  formed is less soluble than the  $\text{ZnS}$  (Yoon, 1981). The inherent flotability of sulfides is due to the inability of sulfide ions to form

H-bonds with the surrounding water molecules (Finkelstein et al, 1975; Furstenau and Sabacky, 1981). The weakness of this natural flotability theory is that sulfide minerals are extremely susceptible to oxidation. At even  $1 \times 10^{-10}$  M levels of dissolved oxygen, oxidation is likely to occur (Gaudin, 1972).

The present study has verified that collectorless flotation of sphalerite is related to the oxidation mechanism of the sphalerite or the CuS on the surface. Since oxidation of sulfide proceeds in an orderly progression from



the flotability of a mineral will be most influenced by the degree of oxidation of the mineral at the time of flotation. The results of the flotation experiments and the XPS experiments indicate that the chemical environment can cause this oxidative progression to be limited or to progress fully, producing the desired flotation behavior. In addition, these results favor the conclusion that polysulfides or metal-deficient sulfides are the species responsible for collectorless flotation of sphalerite in nearly neutral and slightly alkaline solutions. Elemental sulfur is probably responsible for flotation in acidic solutions, however.

## SUMMARY AND CONCLUSIONS

The results of the present investigation may be summarized as follows:

1. Microflotation experiments conducted on non-activated and copper-activated samples of pure sphalerite in the absence of collector demonstrated that flotation was related to the potential of the chemical system. In general, better flotability was obtained in the range of potentials from 0 to +600 mV versus SHE.

2. The use of copper sulfate was essential in obtaining good flotation in both the microflotation and batch flotation tests. The concentration of the cupric sulfate used for activation was extremely important in determining the exact potential range where good flotability occurred.

3. In batch flotation tests conducted with Elmwood mine sphalerite ore, the use of both cupric sulfate and sodium sulfide was required to obtain maximum grade and recovery in collectorless flotation practice. Good flotability was obtained for a wide range of reagent additions as long as the atomic ratio of  $\text{Cu}^{2+}/\text{S}^{2-}$  was maintained at 0.31.

4. The atomic ratio of  $\text{Cu}^{2+}/\text{S}^{2-}$  required for optimum flotability of the Elmwood sphalerite differed from that obtained by earlier research on a different ore. This seems to indicate that the ratio may be a function of each particular ore.

5. The sphalerite recovery in batch flotation tests was found to improve in neutral to acidic solutions. The increased flotability was

postulated to be caused by the increased stability of  $S^0$  in acidic solution or to the removal of hydrophillic  $HS^-$  by protonization to  $H_2S$  gas.

6. The X-ray photoelectron spectroscopic analyses of the flotation products indicated the possibility of polysulfide ions or metal-deficient sulfides on the surface of a highly-flotable copper-activated sample. The chemical reactions necessary to produce this species could have taken place under the conditions of this test. This supports Luttrell and Yoon's work (1984b) as to the role of polysulfides in collectorless flotation.

7. Sodium sulfide and potassium permanganate were found in general to depress flotation. This may support the theory that sulfide surfaces are inherently hydrophobic. The depressant effect of  $Na_2S$  may be caused by the adsorption of  $HS^-$  which may hydrogen bond to surrounding water molecules.  $KMnO_4$  may cause the formation of hydrophillic oxidation products, such as manganese dioxide, on the mineral surface.

8. Collectorless flotation of sphalerite may be viewed as being the result of superficial oxidation of the mineral surface. Insufficient or over oxidation can result in inferior flotation behavior. The use of specific reagents, such as potassium permanganate or sodium sulfide, can speed up or limit the oxidation.

## INDUSTRIAL APPLICATION

The results of the present study have demonstrated that collectorless flotation of sphalerite is possible using only a frother after treatment with copper sulfate and sodium sulfide. Since the cost of sodium sulfide is substantially less than that of collectors used in industry today, the collectorless flotation of sphalerite may result in savings in operation costs. In addition, the need for pH regulators such as lime is eliminated in collectorless flotation.

The current economic crisis in the mining industry makes it essential that cost cutting measures be implemented. With the price of zinc at only 34.5 cents per pound, many mining companies have found it impossible to make a reasonable profit and have ceased to operate. More efficient and cost-effective methods of mining and beneficiation are required to rejuvenate the industry.

In addition, the results of this work underscore the importance of the potential of the flotation pulp as a major variable in achieving good flotation behavior, as has been pointed out by other investigators (Luttrell and Yoon, 1984a; Heyes and Trahar, 1977; Walker, Stout, and Richardson, 1982). Industrial utilization of potential electrodes in flotation control may lead to more efficient metal processing and recovery.

## RECOMMENDATIONS FOR FURTHER WORK

Based on the results of the present investigation, further study is recommended in the following areas:

1. The physical and chemical significance of the cupric ion/sulfide ion ratio should be investigated. Special attention should be paid to the crystal structure and electronic properties of the sphalerite that are important in determining this ratio.
2. Photo-conductivity and other semi-conducting properties of copper-activated sphalerite should be investigated and quantified.
3. Additional XPS studies should be conducted to determine the surface species present on sphalerite after treatment at various pH's and cupric sulfate concentrations.
4. Research to determine the collectorless flotation characteristics of covellite and chalcocite should be conducted and the results compared to those obtained for copper-activated sphalerite.
5. Analysis of the surface of highly-flotable sphalerite should be conducted while the mineral is still in the flotation pulp at the prevailing test conditions using a technique such as Fourier Transform Infrared Spectroscopy (FTIR). This would prevent the problems associated with the possible oxidation of the mineral surface as a result of the drying required prior to using analytical techniques such as XPS.

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APPENDIX A: BATCH FLOTATION CONDITIONS AND  
METALLURGICAL BALANCE SHEETS

This appendix contains a summary of the conditions under which each batch flotation test was conducted, as well as the metallurgical balance sheets from these tests. The charts of the continuously monitored operating parameters, such as potential, are contained in appendix D. The procedure for determining the assays reported in the metallurgical balance sheets is given in appendix E.

Table A-1. Summary of the Conditions for Batch Flotation

| Test | Pot. SHE<br>mV | pH    | [S <sup>2-</sup> ]<br>ppm | [O <sub>2</sub> ]<br>ppm | CuSO <sub>4</sub><br>lb/ton | Na <sub>2</sub> S<br>lb/ton |
|------|----------------|-------|---------------------------|--------------------------|-----------------------------|-----------------------------|
| 1    | +380           | 8.50  | 0.000                     | -----                    | 0.00                        | 0.00                        |
| 2    | +354           | 8.20  | 0.000                     | 8.10                     | 0.00                        | 0.00                        |
| 3    | +340           | 8.30  | 0.000                     | 7.78                     | 1.50                        | 0.00                        |
| 5    | -058           | 10.50 | 2.800                     | 9.50                     | 0.00                        | 1.50                        |
| 6    | +217           | 9.90  | 0.022                     | 8.61                     | 3.00                        | 1.50                        |
| 7    | +270           | 8.90  | 0.004                     | 8.60                     | 1.50                        | 1.50                        |
| 8    | -053           | 10.72 | -----                     | 8.00                     | 0.25                        | 1.50                        |
| 9    | -058           | 10.30 | 1.700                     | 9.70                     | 0.15                        | 1.50                        |
| 10   | -023           | 9.94  | 0.980                     | 10.50                    | 0.00                        | 0.50                        |
| 11   | +243           | 9.90  | 0.002                     | 9.02                     | 1.00                        | 0.50                        |
| 13   | +062           | 7.40  | 0.390                     | 7.20                     | 0.50                        | 0.50                        |
| 14   | +119           | 6.40  | 0.340                     | 7.70                     | 0.10                        | 0.50                        |
| 15   | +003           | 7.60  | 0.920                     | 0.90                     | 0.25                        | 0.50                        |
| 16   | +330           | 8.30  | 0.000                     | 8.90                     | 1.00                        | 0.25                        |
| 17   | +180           | 8.90  | 0.000                     | 8.35                     | 0.50                        | 0.25                        |
| 18   | +230           | 8.38  | 0.004                     | 7.75                     | 0.25                        | 0.25                        |
| 19   | +090           | 7.60  | 0.099                     | 7.00                     | 0.10                        | 0.25                        |
| 20   | +030           | 6.80  | 0.070                     | 7.95                     | 0.05                        | 0.25                        |
| 21   | +325           | 8.74  | 0.000                     | 7.90                     | 0.50                        | 0.00                        |
| 22   | +350           | 8.50  | 0.000                     | 6.85                     | 1.00                        | 0.00                        |
| 23   | +310           | 8.50  | 0.000                     | 7.10                     | 2.00                        | 0.00                        |
| 25   | +220           | ----- | -----                     | 8.50                     | 0.50                        | 0.25                        |
| 26   | +535           | ----- | -----                     | 8.80                     | 1.50                        | 0.25                        |
| 27   | +390           | ----- | -----                     | 8.62                     | 2.00                        | 0.25                        |
| 28   | +320           | ----- | -----                     | 8.85                     | 1.50                        | 0.50                        |
| 29   | +330           | ----- | -----                     | 8.65                     | 2.00                        | 0.50                        |
| 30   | +260           | ----- | -----                     | 9.00                     | 1.00                        | 0.50                        |
| 31   | +310           | ----- | -----                     | 9.00                     | 1.00                        | 0.50                        |
| 32   | +400           | ----- | -----                     | 9.20                     | 1.00                        | 0.50                        |

Table A-2. Metallurgical Balance Sheets for Batch Flotation on Elmwood Sphalerite.

## Test 1

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 1.28     | 11.03  | 14.12   | 0.89         |
| Cl. Tail | 5.15     | 10.80  | 55.62   | 3.49         |
| Tailing  | 93.57    | 16.30  | 1525.19 | 95.62        |
| Feed     | 100.00   | 15.95  | 1594.93 | 100.00       |

## Test 2

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 0.48     | 15.84  | 7.60    | 0.58         |
| Cl. Tail | 7.89     | 13.29  | 104.86  | 7.97         |
| Tailing  | 91.63    | 13.14  | 1204.02 | 91.45        |
| Feed     | 100.00   | 13.16  | 1316.48 | 100.00       |

## Test 3

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 19.54    | 60.00  | 1172.40 | 75.00        |
| Cl. Tail | 11.10    | 19.08  | 211.76  | 13.55        |
| Tailing  | 69.36    | 2.58   | 178.95  | 11.45        |
| Feed     | 100.00   | 15.63  | 1563.11 | 100.00       |

## Test 5

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 0.83     | 6.24   | 5.18    | 0.29         |
| Cl. Tail | 10.31    | 12.50  | 128.85  | 7.24         |
| Tailing  | 88.86    | 18.52  | 1646.05 | 92.47        |
| Feed     | 100.00   | 17.80  | 1780.08 | 100.00       |

Table A-2 (cont.)

## Test 6

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 1.28     | 28.91  | 37.00   | 2.24         |
| Cl. Tail | 8.65     | 25.70  | 222.28  | 13.45        |
| Tailing  | 90.07    | 15.43  | 1389.78 | 84.28        |
| Feed     | 100.00   | 16.49  | 1649.06 | 100.00       |

## Test 7

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 1.72     | 17.13  | 29.46   | 1.76         |
| Cl. Tail | 10.68    | 14.35  | 153.26  | 9.13         |
| Tailing  | 87.60    | 17.07  | 1495.33 | 89.11        |
| Feed     | 100.00   | 16.78  | 1678.05 | 100.00       |

## Test 8

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 1.02     | 9.84   | 10.04   | 0.59         |
| Cl. Tail | 4.64     | 10.57  | 49.04   | 2.89         |
| Tailing  | 94.34    | 17.36  | 1637.74 | 96.52        |
| Feed     | 100.00   | 16.97  | 1696.82 | 100.00       |

## Test 9

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 0.70     | 8.36   | 5.85    | 0.40         |
| Cl. Tail | 6.72     | 11.17  | 75.05   | 5.11         |
| Tailing  | 92.58    | 14.98  | 1387.07 | 94.49        |
| Feed     | 100.00   | 14.68  | 1467.97 | 100.00       |

## Test 10

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 0.61     | 7.79   | 4.75    | 0.33         |
| Cl. Tail | 3.58     | 9.98   | 35.73   | 2.49         |
| Tailing  | 95.81    | 14.57  | 1396.02 | 97.16        |
| Feed     | 100.00   | 14.37  | 1436.50 | 100.00       |

## Test 11

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 18.24    | 54.09  | 986.54  | 69.20        |
| Cl. Tail | 10.55    | 8.71   | 91.87   | 6.44         |
| Tailing  | 71.21    | 4.88   | 347.37  | 24.36        |
| Feed     | 100.00   | 14.26  | 1425.78 | 100.00       |

## Test 13

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 1.51     | 46.15  | 53.07   | 4.55         |
| Cl. Tail | 1.81     | 12.41  | 22.46   | 1.92         |
| Tailing  | 96.68    | 11.29  | 1091.36 | 93.53        |
| Feed     | 100.00   | 11.67  | 1166.89 | 100.00       |

## Test 14

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 0.54     | 26.83  | 14.49   | 1.25         |
| Cl. Tail | 3.69     | 5.96   | 21.99   | 1.90         |
| Tailing  | 95.77    | 11.73  | 1123.82 | 96.85        |
| Feed     | 100.00   | 11.60  | 1160.30 | 100.00       |

## Test 15

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 1.16     | 53.34  | 61.88   | 5.07         |
| Cl. Tail | 0.72     | 35.91  | 97.68   | 5.00         |
| Tailing  | 98.12    | 11.04  | 1081.57 | 88.93        |
| Feed     | 100.00   | 12.21  | 1221.13 | 100.00       |

## Test 16

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 21.27    | 56.17  | 1194.77 | 84.38        |
| Cl. Tail | 3.14     | 2.18   | 6.83    | 0.48         |
| Tailing  | 75.59    | 2.83   | 214.41  | 15.14        |
| Feed     | 100.00   | 14.16  | 1416.02 | 100.00       |

## Test 17

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 4.54     | 47.11  | 213.88  | 15.44        |
| Cl. Tail | 4.97     | 18.30  | 90.95   | 6.57         |
| Tailing  | 90.49    | 11.94  | 1080.45 | 77.99        |
| Feed     | 100.00   | 13.85  | 1385.28 | 100.00       |

## Test 18

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 5.89     | 49.96  | 294.29  | 20.82        |
| Cl. Tail | 6.29     | 12.39  | 77.94   | 5.52         |
| Tailing  | 87.82    | 11.65  | 1041.00 | 73.66        |
| Feed     | 100.00   | 14.13  | 1413.23 | 100.00       |

## Test 19

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 0.26     | 1.39   | 0.36    | 0.02         |
| Cl. Tail | 3.31     | 10.85  | 35.90   | 2.22         |
| Tailing  | 96.43    | 16.37  | 1579.45 | 97.76        |
| Feed     | 100.00   | 16.14  | 1614.71 | 100.00       |

## Test 20

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 0.76     | 9.63   | 7.32    | 0.50         |
| Cl. Tail | 5.17     | 11.20  | 57.93   | 3.99         |
| Tailing  | 94.07    | 14.76  | 1388.07 | 95.51        |
| Feed     | 100.00   | 14.53  | 1453.32 | 100.00       |

## Test 21

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 18.17    | 55.51  | 1002.62 | 71.23        |
| Cl. Tail | 8.86     | 14.22  | 125.00  | 8.90         |
| Tailing  | 72.97    | 3.85   | 281.27  | 19.87        |
| Feed     | 100.00   | 14.16  | 1415.89 | 100.00       |

## Test 22

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 19.17    | 49.77  | 954.13  | 75.75        |
| Cl. Tail | 7.49     | 21.56  | 161.50  | 12.62        |
| Tailing  | 73.34    | 1.96   | 143.99  | 11.43        |
| Feed     | 100.00   | 12.60  | 1259.62 | 100.00       |

## Test 23

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 19.76    | 58.25  | 1111.50 | 83.68        |
| Cl. Tail | 5.79     | 28.57  | 165.40  | 12.45        |
| Tailing  | 74.45    | 0.69   | 51.31   | 3.87         |
| Feed     | 100.00   | 13.28  | 1328.21 | 100.00       |

## Test 25

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 7.24     | 57.78  | 418.32  | 28.54        |
| Cl. Tail | 10.07    | 15.29  | 153.96  | 9.77         |
| Tailing  | 82.69    | 12.14  | 1003.98 | 63.69        |
| Feed     | 100.00   | 15.76  | 1576.26 | 100.00       |

## Test 26

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 20.97    | 61.35  | 1280.59 | 86.00        |
| Cl. Tail | 1.25     | 27.81  | 34.76   | 2.33         |
| Tailing  | 77.88    | 2.23   | 173.67  | 11.67        |
| Feed     | 100.00   | 14.89  | 1489.02 | 100.00       |

## Test 27

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 20.15    | 55.65  | 1121.35 | 85.15        |
| Cl. Tail | 1.65     | 23.76  | 40.04   | 3.04         |
| Tailing  | 78.16    | 1.99   | 155.54  | 11.81        |
| Feed     | 100.00   | 13.17  | 1316.93 | 100.00       |

## Test 28

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 20.19    | 56.95  | 1144.82 | 70.98        |
| Cl. Tail | 1.79     | 28.51  | 51.03   | 3.15         |
| Tailing  | 75.02    | 5.37   | 418.97  | 25.87        |
| Feed     | 100.00   | 16.20  | 1619.82 | 100.00       |

## Test 29

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 18.67    | 53.70  | 1002.58 | 62.34        |
| Cl. Tail | 5.83     | 36.82  | 214.66  | 13.35        |
| Tailing  | 75.50    | 5.18   | 391.09  | 24.31        |
| Feed     | 100.00   | 16.08  | 1608.33 | 100.00       |

## Test 30

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 16.97    | 57.83  | 981.38  | 67.96        |
| Cl. Tail | 10.80    | 7.79   | 84.13   | 5.83         |
| Tailing  | 72.23    | 5.24   | 378.49  | 26.48        |
| Feed     | 100.00   | 14.44  | 1444.00 | 100.00       |

## Test 31

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 18.06    | 55.06  | 994.38  | 67.76        |
| Cl. Tail | 3.89     | 21.88  | 85.11   | 5.80         |
| Tailing  | 78.05    | 4.97   | 387.92  | 26.44        |
| Feed     | 100.00   | 14.67  | 1467.41 | 100.00       |

## Test 32

| Product | Weight % | % Zinc | Unit Zn | Distribution |
|---------|----------|--------|---------|--------------|
| Conc.   | 18.54    | 54.77  | 1015.43 | 68.46        |
| Cl.Tail | 13.23    | 24.79  | 327.97  | 22.11        |
| Tailing | 68.23    | 2.05   | 139.87  | 9.43         |
| Feed    | 100.00   | 14.83  | 1483.27 | 100.00       |

## Test R1A

| Product | Weight % | % Zinc | Unit Zn | Distribution |
|---------|----------|--------|---------|--------------|
| Conc.   | 5.74     | 61.91  | 355.36  | 24.32        |
| Cl.Tail | 7.11     | 49.53  | 352.16  | 24.10        |
| Tailing | 87.15    | 8.65   | 753.95  | 51.58        |
| Feed    | 100.00   | 14.61  | 1461.37 | 100.00       |

## Test R1B

| Product | Weight % | % Zinc | Unit Zn | Distribution |
|---------|----------|--------|---------|--------------|
| Conc.   | 7.44     | 63.09  | 469.39  | 27.41        |
| Cl.Tail | 4.44     | 45.37  | 201.40  | 11.76        |
| Tailing | 88.12    | 11.82  | 1041.55 | 60.83        |
| Feed    | 100.00   | 17.12  | 1712.34 | 100.00       |

## Test R1C

| Product | Weight % | % Zinc | Unit Zn | Distribution |
|---------|----------|--------|---------|--------------|
| Conc.   | 16.20    | 61.57  | 1078.22 | 72.92        |
| Cl.Tail | 18.53    | 12.43  | 230.33  | 16.33        |
| Tailing | 64.77    | 2.34   | 151.56  | 10.75        |
| Feed    | 100.00   | 14.10  | 1410.11 | 100.00       |

## Test R10

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 15.88    | 60.24  | 955.41  | 63.25        |
| Cl. Tail | 27.08    | 9.65   | 261.13  | 17.29        |
| Tailing  | 57.08    | 5.15   | 283.98  | 19.46        |
| Feed     | 100.00   | 15.11  | 1510.50 | 100.00       |

## Test R2A

| Product     | Weight % | % Zinc | Unit Zn | Distribution |
|-------------|----------|--------|---------|--------------|
| Conc.       | 8.16     | 50.87  | 413.47  | 25.39        |
| Cl. Tailing | 10.51    | 36.52  | 383.83  | 23.57        |
| Tailing     | 81.33    | 10.22  | 831.19  | 51.04        |
| Feed        | 100.00   | 16.28  | 1628.49 | 100.00       |

## Test R2B

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 7.35     | 58.17  | 427.55  | 27.64        |
| Cl. Tail | 5.24     | 32.61  | 170.88  | 11.05        |
| Tailing  | 87.41    | 10.85  | 948.40  | 61.31        |
| Feed     | 100.00   | 15.47  | 1548.83 | 100.00       |

## Test R2C

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 22.90    | 51.14  | 1171.11 | 73.63        |
| Cl. Tail | 8.25     | 27.30  | 225.23  | 14.16        |
| Tailing  | 68.85    | 2.82   | 194.16  | 12.21        |
| Feed     | 100.00   | 15.91  | 1590.50 | 100.00       |

## Test R2D

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 17.34    | 52.40  | 908.62  | 62.01        |
| Cl. Tail | 27.47    | 8.65   | 237.62  | 16.22        |
| Tailing  | 55.19    | 5.78   | 315.00  | 21.77        |
| Feed     | 100.00   | 14.65  | 1465.24 | 100.00       |

## Test P1

| Product  | Weight % | % Zinc | Unit Zn | Distribution |
|----------|----------|--------|---------|--------------|
| Conc.    | 16.57    | 43.84  | 726.43  | 39.74        |
| Cl. Tail | 5.04     | 10.64  | 53.63   | 2.93         |
| Tailing  | 78.39    | 13.37  | 1048.07 | 57.33        |
| Feed     | 100.00   | 18.28  | 1828.13 | 100.00       |

## Test P2

| Product | Weight % | % Zinc | Unit Zn | Distribution |
|---------|----------|--------|---------|--------------|
| Conc.   | 4.28     | 17.64  | 75.50   | 5.16         |
| Tailing | 95.72    | 14.50  | 1387.94 | 94.84        |
| Feed    | 100.00   | 14.63  | 1463.44 | 100.00       |

## Test P3

| Product | Weight % | % Zinc | Unit Zn | Distribution |
|---------|----------|--------|---------|--------------|
| Conc.   | 10.50    | 44.22  | 464.32  | 30.27        |
| Tailing | 89.49    | 11.95  | 1069.41 | 69.73        |
| Feed    | 100.00   | 15.34  | 1533.73 | 100.00       |

## Test P4

| Product | Weight % | % Zinc | Unit Zn | Distribution |
|---------|----------|--------|---------|--------------|
| Conc.   | 2.78     | 11.11  | 30.89   | 2.22         |
| Tailing | 97.22    | 14.01  | 1362.05 | 97.78        |
| Feed    | 100.00   | 13.93  | 1392.94 | 100.00       |

APPENDIX B: MICROFLOTATION CONDITIONS

| <u>Test</u> | <u>pH</u> | <u>Pat. (SHE)</u> | <u>Recovery</u> | <u>CuSO<sub>4</sub></u> |
|-------------|-----------|-------------------|-----------------|-------------------------|
| 30          | 5.70      | +432              | 35.40           | 0                       |
| 32          | 11.25     | -173              | 6.90            | 0                       |
| 33          | 10.20     | -028              | 5.70            | 0                       |
| 34          | 9.60      | +047              | 7.80            | 0                       |
| 35          | 6.67      | +272              | 22.70           | 0                       |
| 36          | 6.90      | +644              | 16.30           | 0                       |
| 38          | 6.50      | +615              | 12.10           | 0                       |
| 39          | 5.46      | +590              | 81.42           | 5x10 <sup>-5</sup>      |
| 40          | 7.62      | +266              | 90.99           | "                       |
| 41          | 9.55      | +102              | 64.75           | "                       |
| 42          | 10.32     | -006              | 33.33           | "                       |
| 43          | 11.62     | -332              | 7.63            | "                       |
| 44          | 7.30      | +732              | 0.00            | "                       |
| 46          | 5.60      | +502              | 85.35           | "                       |
| 47          | 6.70      | +412              | 91.24           | "                       |
| 48          | 5.42      | +400              | 94.44           | 5x10 <sup>-3</sup>      |
| 49          | 10.94     | -248              | 44.04           | "                       |
| 50          | 11.24     | -383              | 2.50            | "                       |
| 51          | 9.81      | -076              | 83.33           | "                       |
| 52          | 8.96      | +150              | 97.12           | "                       |

| test | pH    | Pot. (SHE) | Recovery | CuSD               |
|------|-------|------------|----------|--------------------|
| 53   | 8.07  | +794       | 1.85     | $5 \times 10^{-3}$ |
| 54   | 9.67  | +489       | 74.19    | "                  |
| 55   | 10.97 | -177       | 76.85    | "                  |
| 56   | 5.53  | +374       | 95.54    | $5 \times 10^{-2}$ |
| 57   | 10.91 | -212       | 21.10    | "                  |
| 58   | 11.68 | -437       | 1.94     | "                  |
| 59   | 10.07 | -103       | 55.76    | "                  |
| 60   | 10.10 | -044       | 80.51    | "                  |
| 61   | 10.03 | +118       | 43.22    | "                  |
| 62   | 8.76  | +198       | 51.72    | "                  |
| 63   | 9.04  | +272       | 29.62    | "                  |
| 64   | 5.99  | +747       | 54.31    | "                  |
| 65   | 6.14  | +445       | 60.00    | "                  |
| 66   | 8.90  | +301       | 36.36    | "                  |
| 67   | 9.03  | +055       | 88.89    | "                  |
| 68   | 6.08  | +534       | 67.18    | "                  |
| 69   | 4.66  | +395       | 48.72    | "                  |
| 94   | 4.00  | +356       | 63.39    | 0                  |
| 95   | 4.00  | -002       | 18.10    | 0                  |
| 96   | 4.00  | +227       | 66.36    | 0                  |

| Test | pH   | Pot. (SHE) | Recovery | CuSO <sub>4</sub>  |
|------|------|------------|----------|--------------------|
| 97   | 4.00 | -437       | 12.28    | 0                  |
| 98   | 4.00 | -047       | 14.18    | 0                  |
| 99   | 4.00 | +1015      | 23.58    | 0                  |
| 100  | 4.00 | +817       | 51.40    | 0                  |
| 101  | 4.00 | +748       | 49.49    | 0                  |
| 103  | 4.00 | +410       | 98.94    | $5 \times 10^{-5}$ |
| 104  | 4.00 | +054       | 88.68    | "                  |
| 105  | 4.00 | +153       | 95.15    | "                  |
| 106  | 4.00 | +227       | 95.15    | "                  |
| 107  | 4.00 | +343       | 92.66    | "                  |
| 108  | 4.00 | +439       | 95.33    | "                  |
| 109  | 4.00 | +864       | 95.50    | "                  |
| 110  | 4.00 | +740       | 95.37    | 0                  |
| 111  | 4.00 | +468       | 95.28    | 0                  |
| 112  | 4.00 | +033       | 95.41    | 0                  |
| 113  | 4.00 | -034       | 43.64    | 0                  |
| 114  | 4.00 | +038       | 93.04    | $5 \times 10^{-5}$ |
| 115  | 7.55 | -075       | 83.64    | 0                  |
| 116  | 4.60 | +068       | 86.99    | 0                  |
| 117  | 4.00 | +532       | 96.12    | $5 \times 10^{-5}$ |
| 118  | 4.00 | -040       | 86.27    | $5 \times 10^{-5}$ |

| Test | pH    | PoL. (SHE) | Recovery | CuSO <sub>4</sub>  |
|------|-------|------------|----------|--------------------|
| 119  | 7.00  | +397       | 96.40    | 0                  |
| 120  | 7.00  | -170       | 73.79    | 0                  |
| 121  | 7.00  | -270       | 23.21    | 0                  |
| 122  | 7.00  | -073       | 93.52    | 0                  |
| 123  | 7.00  | +013       | 94.55    | 0                  |
| 124  | 7.00  | +166       | 95.50    | 0                  |
| 125  | 7.00  | +577       | 96.23    | 0                  |
| 126  | 7.00  | +703       | 43.81    | 0                  |
| 127  | 7.00  | +342       | 96.12    | 0                  |
| 128  | 7.00  | +450       | 95.73    | 0                  |
| 129  | 7.00  | +491       | 94.64    | $5 \times 10^{-5}$ |
| 130  | 7.00  | -273       | 32.73    | "                  |
| 131  | 7.00  | -151       | 94.39    | "                  |
| 132  | 7.00  | -070       | 93.52    | "                  |
| 133  | 7.00  | +098       | 97.03    | "                  |
| 134  | 7.00  | +687       | 91.26    | "                  |
| 135  | 7.00  | +387       | 95.10    | "                  |
| 136  | 7.00  | +569       | 95.24    | "                  |
| 137  | 7.00  | +277       | 96.00    | "                  |
| 138  | 10.00 | +341       | 6.19     | 0                  |
| 139  | 10.00 | -425       | 7.41     | 0                  |
| 140  | 10.00 | -066       | 2.86     | 0                  |
| 141  | 10.00 | +149       | 3.00     | 0                  |
| 142  | 10.00 | +580       | 0.00     | 0                  |
| 143  | 10.00 | +649       | 0.00     | 0                  |
| 144  | 10.00 | +680       | 0.00     | 0                  |
| 145  | 10.00 | +347       | 93.01    | $5 \times 10^{-5}$ |
| 146  | 10.00 | -353       | 7.14     | "                  |
| 147  | 10.00 | -121       | 39.58    | "                  |

| Test | pH    | Pot. (SHE) | Recovery | CuSO <sub>4</sub>  |
|------|-------|------------|----------|--------------------|
| 148  | 10.00 | +355       | 91.67    | $5 \times 10^{-5}$ |
| 149  | 10.00 | -100       | 51.55    | "                  |
| 150  | 10.00 | -066       | 21.65    | "                  |
| 151  | 10.00 | -030       | 38.14    | "                  |
| 152  | 10.00 | +104       | 65.66    | "                  |
| 153  | 10.00 | +239       | 67.01    | "                  |
| 154  | 10.00 | +645       | 1.03     | "                  |
| 155  | 10.00 | +476       | 73.96    | "                  |
| 156  | 10.00 | +578       | 56.44    | "                  |
| 157  | 10.00 | +329       | 98.97    | $5 \times 10^{-3}$ |
| 158  | 10.00 | -115       | 30.30    | "                  |
| 159  | 10.00 | -046       | 44.44    | "                  |
| 160  | 10.00 | +265       | 10.20    | "                  |
| 161  | 10.00 | +071       | 61.05    | "                  |
| 162  | 10.00 | +640       | 0.00     | "                  |
| 163  | 10.00 | +569       | 22.22    | "                  |
| 164  | 10.00 | +510       | 62.47    | "                  |
| 165  | 10.00 | +445       | 67.31    | "                  |
| 166  | 10.00 | +222       | 51.35    | "                  |

| test | pH    | Pot. (SHE) | Recovery | CuSO <sub>4</sub>  |
|------|-------|------------|----------|--------------------|
| 167  | 7.00  | +551       | 14.43    | 0                  |
| 168  | 7.00  | -111       | 10.42    | 0                  |
| 169  | 7.00  | -251       | 5.21     | 0                  |
| 170  | 7.00  | +022       | 13.54    | 0                  |
| 171  | 7.00  | +252       | 10.42    | 0                  |
| 172  | 7.00  | +142       | 14.29    | 0                  |
| 173  | 7.00  | +803       | 0.99     | 0                  |
| 174  | 7.00  | +324       | 19.39    | 0                  |
| 175  | 7.00  | +452       | 21.34    | 0                  |
| 176  | 7.00  | +554       | 97.94    | $5 \times 10^{-5}$ |
| 177  | 7.00  | -227       | 23.30    | "                  |
| 178  | 7.00  | -069       | 75.00    | "                  |
| 179  | 7.00  | +346       | 76.53    | "                  |
| 180  | 7.00  | +007       | 91.18    | "                  |
| 181  | 7.00  | +202       | 94.90    | "                  |
| 182  | 7.00  | +252       | 97.98    | "                  |
| 183  | 7.00  | +472       | 98.10    | "                  |
| 184  | 7.00  | +800       | 2.65     | "                  |
| 185  | 7.00  | +703       | 96.40    | "                  |
| 186  | 10.00 | +312       | 91.75    | "                  |

| test | pH    | Pot. (SHE) | Recovery | CuSO <sub>4</sub>  |
|------|-------|------------|----------|--------------------|
| 187  | 10.00 | +194       | 73.74    | $5 \times 10^{-5}$ |
| 188  | 10.00 | +140       | 84.21    | $5 \times 10^{-3}$ |
| 189  | 10.00 | +257       | 91.67    | "                  |
| 190  | 10.00 | +307       | 87.50    | "                  |
| 191  | 10.00 | -257       | 16.35    | "                  |

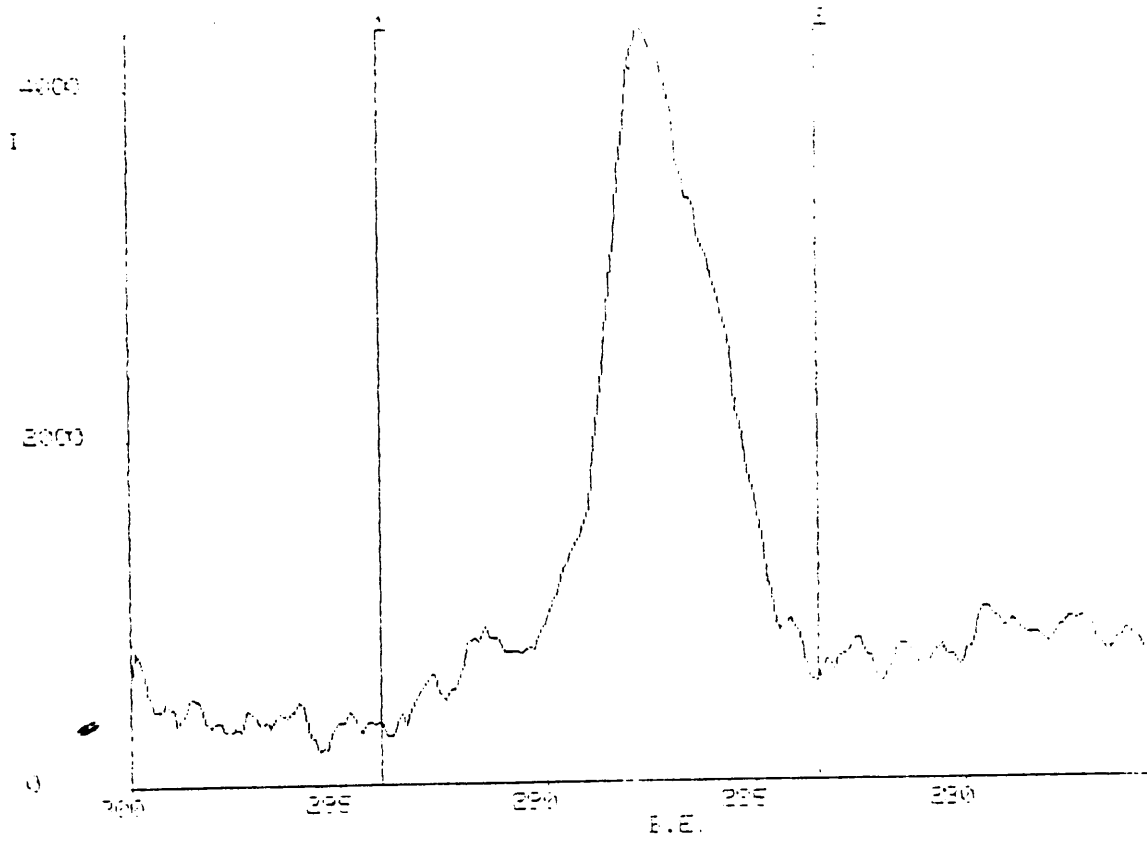
APPENDIX C: XPS SPECTRA AND CALCULATIONS

Since most specimens exhibited complex carbon-1s peaks, the assignment of peak location for calculation of sample charging was not trivial. The assignment of particular peak locations was based on an analysis of the geometry of the carbon-1s peaks, an assumption that sample charging would be relatively consistent between samples, and a practical examination of the identity of the various sulfur species indicated using different charge correction factors. Peak location and the difference in binding energy between peaks in the Sulfur-2p region as given in the literature were used to correlate the data obtained in the present work and to define the charge correction factor for each sample. The assignments of the binding energy for the carbon-1s peak thus calculated result in reasonable identities for the sulfur species found on each sample.

Table C-1. The binding energy of the carbon-1s peak and the charge correction factor (arrived by assigning the standard carbon peak to 284.9 eV) for the samples of Elmwood sphalerite and a sulfur standard.

| Sample      | Carbon-1s Peak | Charge Correction |
|-------------|----------------|-------------------|
| Sulfur std. | 287.7 eV       | 2.8 eV            |
| 1C          | 291.9          | 7.0               |
| 2T          | 291.1          | 6.2               |
| 3T          | 292.0          | 7.1               |
| 4T          | 290.8          | 5.9               |
| 5C          | 291.3          | 6.4               |
| 6T          | 290.0          | 5.1               |

ANAL: SULFUR STAN C18  
RUN: 1040 29-HPF-84 FEG: 3.0KV ST:0.10 #1 13. #SM 3 SWELL: 0.200



RUN : Y040 / 23-APR-84  
 SAMPLE: 1  
 NAME : SULFUR STD  
 TITLE : DR 100N  
 15KV 20MA SLIT=2 CUR=9.0

EXCITATION : MG ANALYSER MODE: PRR  
 SCAN MODE : T FACTOR : 0

MAGNIFICATION: L RESOLUTION: M

BASELINE THRESHOLD: 0 MIN. PEAK WIDTH: 1.00

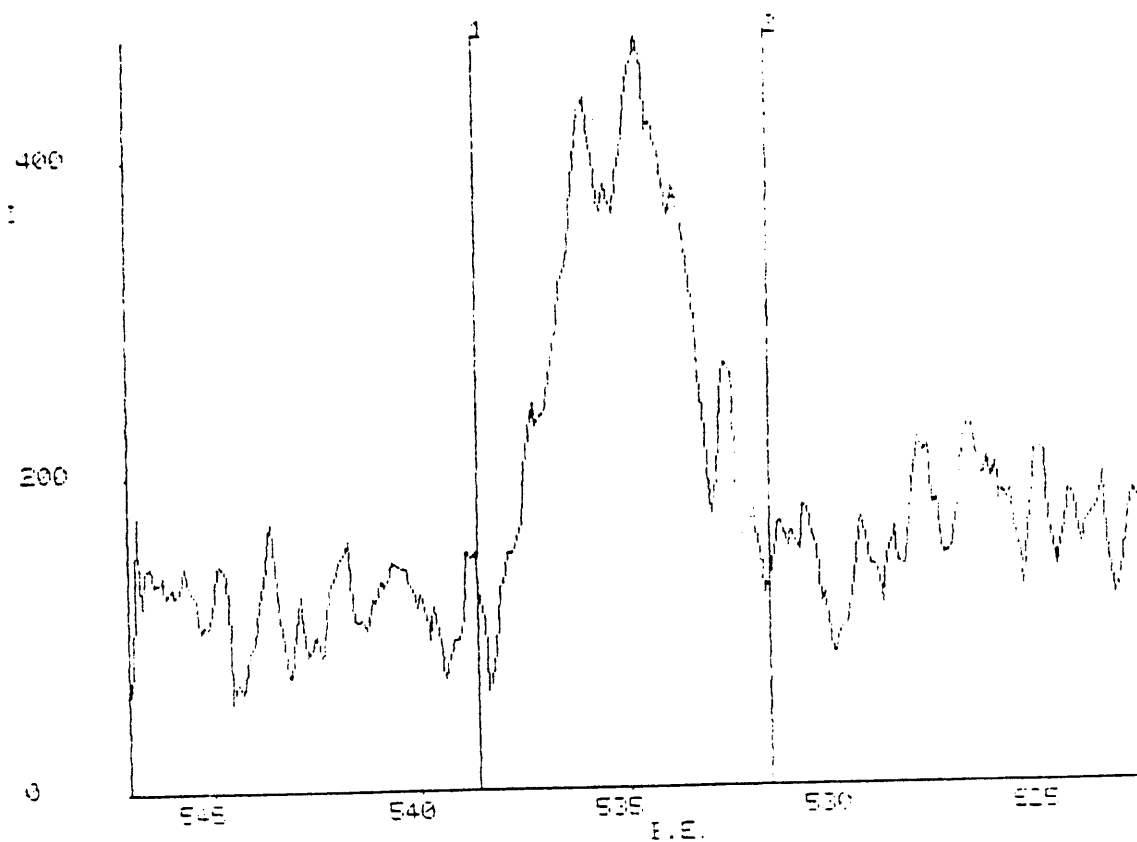
REGION: 3 - WITH MARKERS  

| ELMT | K.E.   | STEP | DWELL | #CHAN | #AVG |
|------|--------|------|-------|-------|------|
| XC1  | 988.80 | 0.10 | 0.200 | 250   | 3    |

SCAN: 1

| PEAK | START E.V.<br>K.E. | END E.V.<br>E.E. | MAX  | AREA   | FWHM |
|------|--------------------|------------------|------|--------|------|
| 1    | 985.90             | 987.70           | 3890 | 134816 | 3.00 |

REE SULFUR STAN 013  
END  
RUN: -1040 X 22-APP-84 PEG: 4.001 ST: 0.10 #1. 250 #SW: 1 DWELL: 0.200



RUN : Y040 / 29-APP-84  
 SAMPLE: 1  
 NAME : SULFUR STD  
 TITLE : IR Y00N  
 13KV 20MA SLIT=2 CUR=9.0

EXCITATION : MG ANALYSER MODE: FRP  
 SCAN MODE : T FACTOR : 0

MAGNIFICATION: L RESOLUTION: M

BASELINE THRESHOLD: 0 MIN. PEAK WIDTH: 1.00

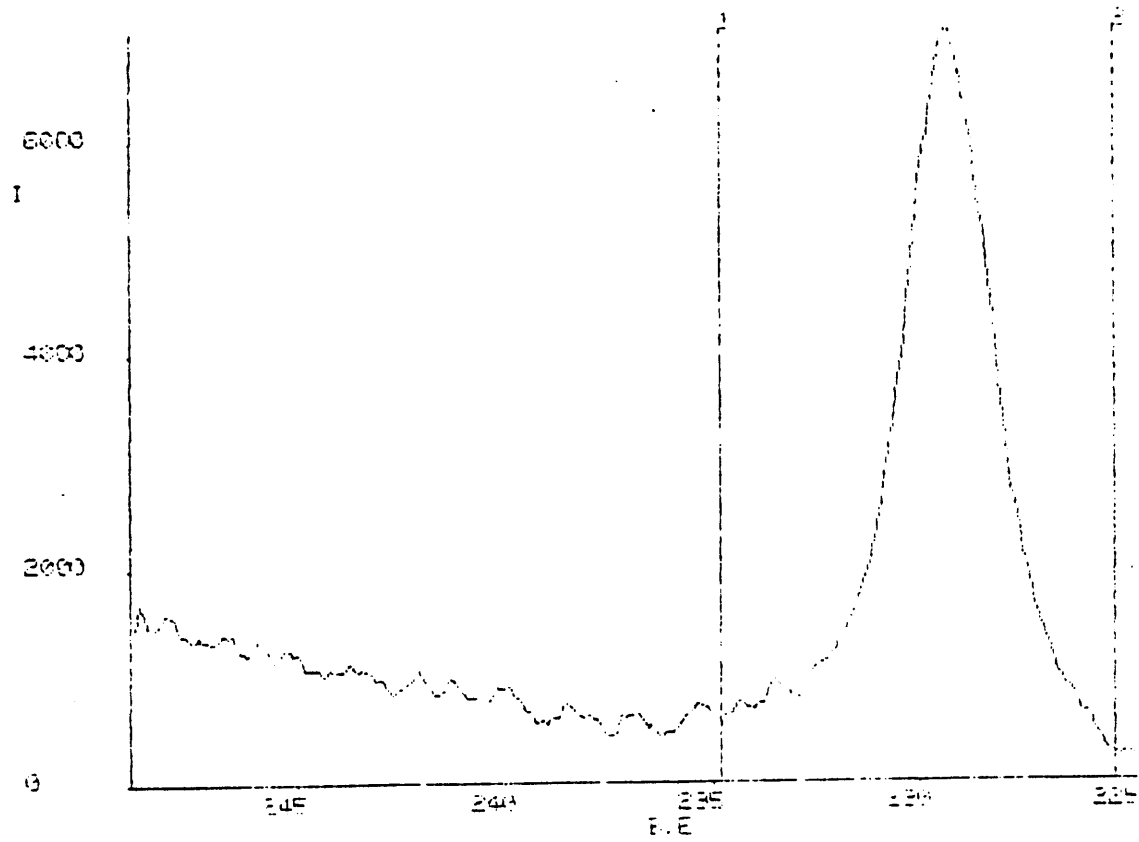
REGION: 4 - WITH MARKERS

| ELMT | K.E.   | STEP | DNELL | #CHAN | #AVG |
|------|--------|------|-------|-------|------|
| X01  | 721.60 | 0.10 | 0.200 | 250   | 1    |

SCAN: 1

| PEAK | K.E.   | START E.V. | E.E.   | END E.V. | MAX | ARE-  | FUHH |
|------|--------|------------|--------|----------|-----|-------|------|
| 1    | 718.80 | 708.80     | 534.80 | 731.80   | 357 | 12417 | 8.40 |

DATA SHEET 1741 881  
END  
RUN: 1040 29-FEB-84 RES: 2.000 ST: 0.10 BI SP: 450 S: 0.000 0.000



RUN : 1044 03-APR-82  
 SAMPLE :  
 NAME : SULFUR STAIN  
 TITLE : DR. 100V  
 15KV 20MA SPLIT=8 CUR=9.0

EXCITATION : MG ANALYSER MODE: FRP  
 SCAN MODE : T FACTOR : 0

MAGNIFICATION: 1 RESOLUTION: M

BASELINE THRESHOLD: 0 MIN PEAK WIDTH: 1.00

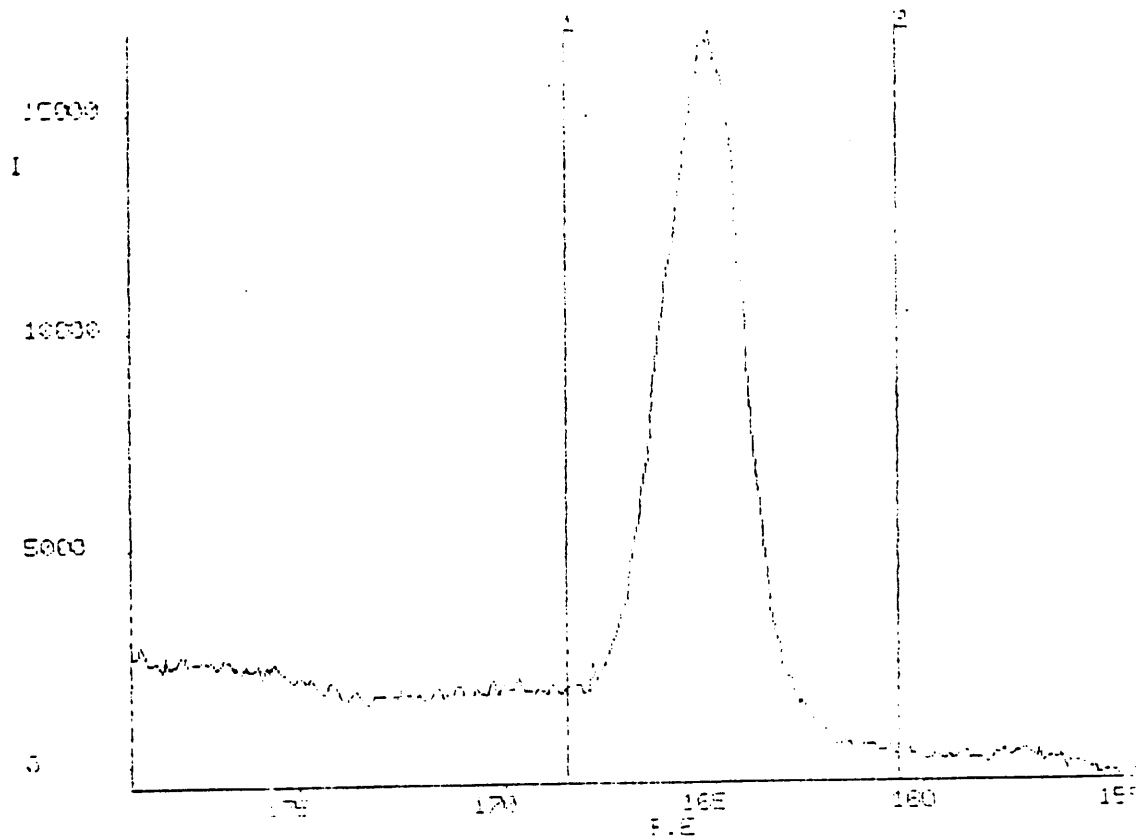
REGION: 2 - WITH MARKERS

| ELMT | K.E.    | STEP | DWELL | #CHAN | #AVG |
|------|---------|------|-------|-------|------|
| XS2S | 1019.60 | 0.10 | 0.200 | 250   | 3    |

SCAN: 1

| PEAK | START E.V. : 1024.60 END E.V. : 1028.50 |        | MAX  | AREA   | FWHM |
|------|---|--------|------|--------|------|
|      | K.E.                                    | E.E.   |      |        |      |
| 1    | 1024.60                                 | 228.90 | 6522 | 150734 | 2.40 |

XSU SULFUR STAN 92F  
END  
RUN: Y040 / 29-APR-64 REG: 1.001 ST: 0.10 NO. 150 PSW: 3 SWELL: 0.200



RUN : Y244 / 23-APR-84  
 SAMPLE: 1  
 NAME : SULFUR STD  
 TITLE : DR YOUN  
 15.7 204A SPLIT=2 CUR=5.0

EXCITATION : MG ANALYSER MODE: FRP  
 SCAN MODE : T FACTOR : 0

MAGNIFICATION: L RESOLUTION: 6

BASELINE THRESHOLD: 0 MIN. PEAK WIDTH: 1.00

REGION: 1 - WITH MARKERS  

| ELMT | K.E.    | STEP | INELL | #CHAN | #AVG |
|------|---------|------|-------|-------|------|
| XS   | 1039.50 | 0.10 | 0.200 | 250   | 3    |

SCAN: 1

| PEAK | START E.V. : 1074.50 |        | END E.V. : 1099.50 |        | FWHM |
|------|----------------------|--------|--------------------|--------|------|
|      | R.E.                 | E.E.   | MAX                | AREA   |      |
| 1    | 1039.50              | 105.10 | 15756              | 373120 | 2.20 |

RUN REG SCAN STEP #CH  
 Y040 1 1 .10 32

ELMT K.E. DWELL  
 NO 1039.00 .200

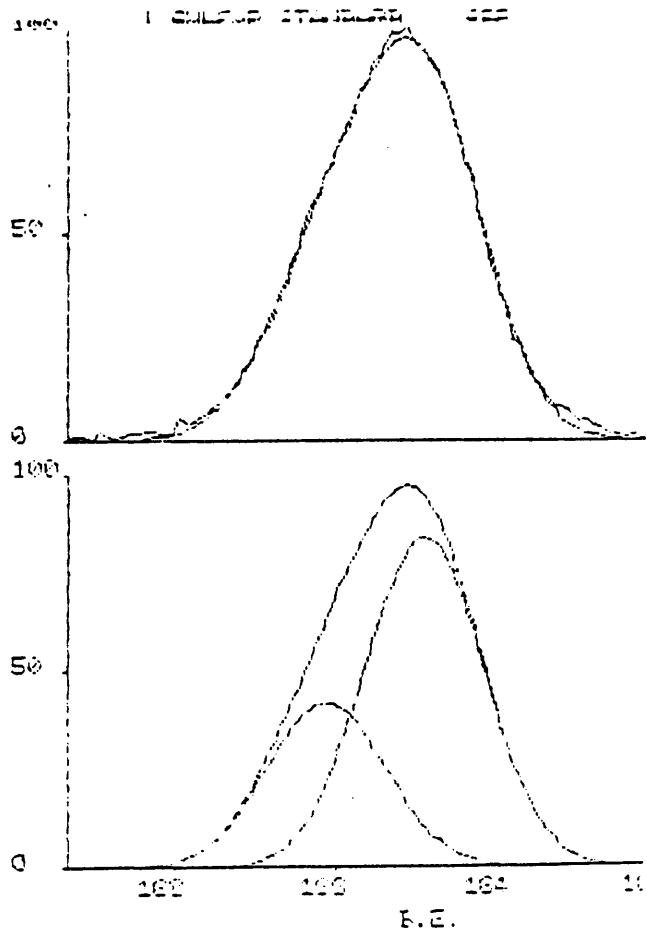
START EV: 123.00  
 END EV: 121.20

100% AREA = 070832

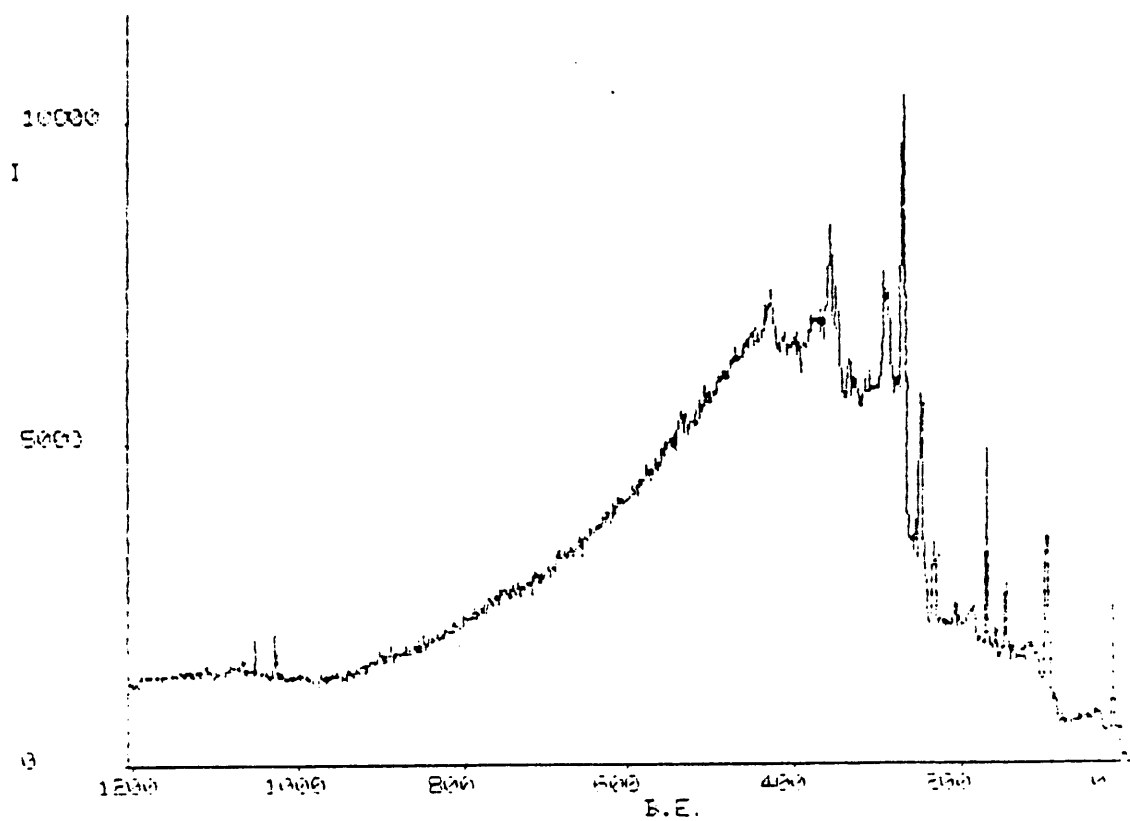
100% INTENSITY = 15200

FIT=4

| PK | ENERGY | MAX   | FWHM | AREA |
|----|--------|-------|------|------|
| 1G | 123.10 | 42.00 | 1.70 | 32.7 |
| 2G | 124.90 | 34.00 | 1.70 | 65.4 |



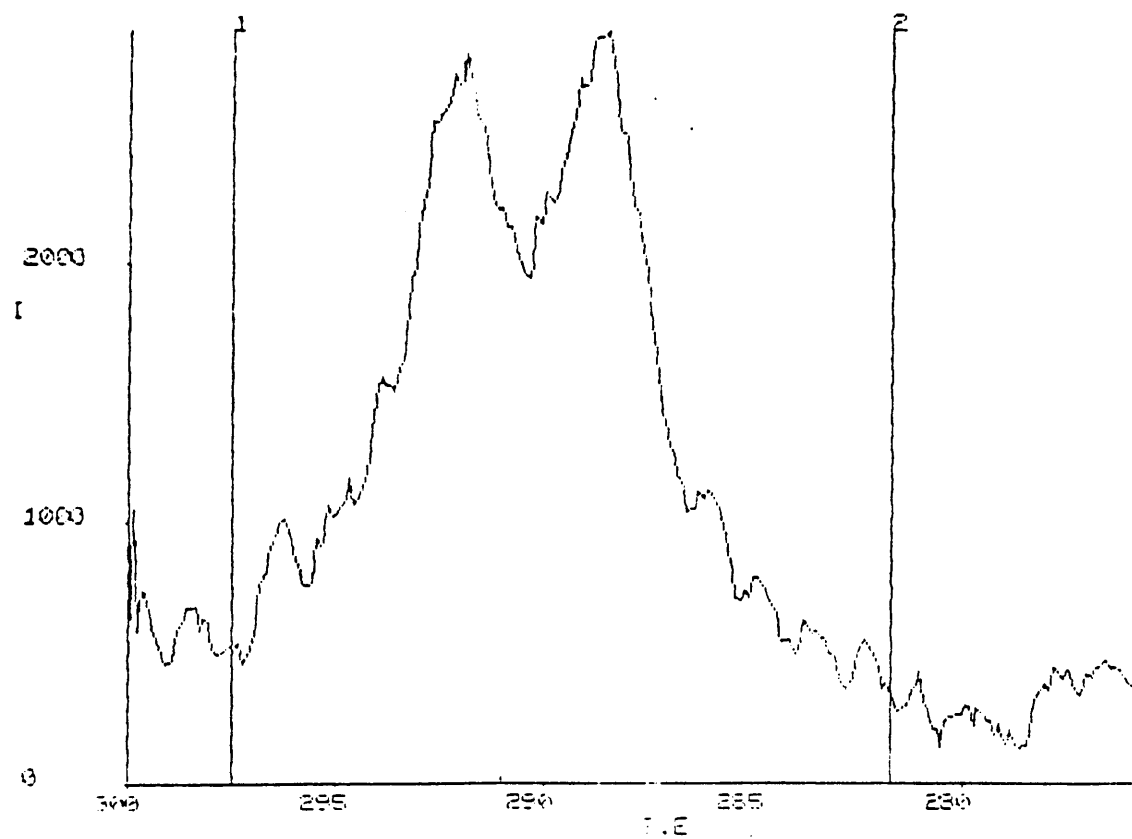
\*\*\*E 10  
END  
RUN: -Y020 / 29-APR-84 REG: 1.001 ST: 1.00 #C: 1250 #SW: 2 DWELL: 0.100



M1 10 013

SIC

RUN: Y010 / 29-APR-34 REG: 2.001 ST: 0.10 #C: 250 #SW: 2 DWELL: 0.200



RUN : Y010 / 29-APR-84  
 SAMPLE: 1  
 NAME : IC  
 TITLE : IR YOON  
 13KV 20MA SLIT=2 CUR=9.0

EXCITATION : MG ANALYSER MODE: FRR  
 SCAN MODE : T FACTOR : 0

MAGNIFICATION: L RESOLUTION: M

BASELINE THRESHOLD: 0 MIN. PEAK WIDTH: 1.00

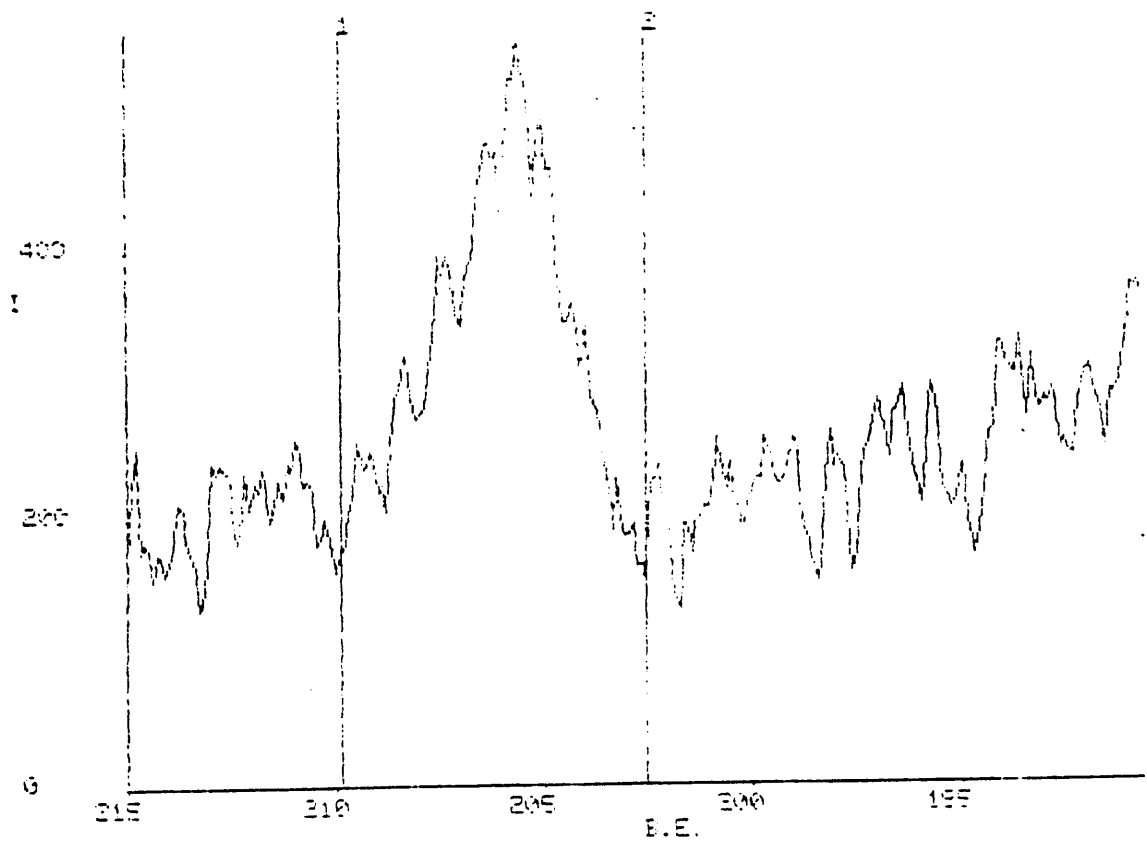
REGION: 2 - WITH MARKERS

| ELMT | K.E.   | STEP | DWELL | #CHAN | #AVG |
|------|--------|------|-------|-------|------|
| XC1  | 968.60 | 0.10 | 0.200 | 250   | 2    |

SCAN: 1

| PEAK | K.E.   | START E.V. :<br>B.E. | END E.V. :<br>MAX | AREA   | FWHH |
|------|--------|----------------------|-------------------|--------|------|
| 1    | 968.00 | 208.60               | 2507              | 164405 | 6.00 |

WAC 10 010  
E10  
RUN: 1000 / 28-11-7-34 REV: 2.001 ST: 0.10 SC: 250 POW: 1 SWEET: 0.200



RUN : Y030 / 23-APR-84  
 SAMPLE: 1  
 NAME : IC  
 TITLE : DR YOON  
 13KV 20MA SLIT=2 CUR=8.0

EXCITATION : MG ANALYSER MODE: FRR  
 SCAN MODE : T FACTOR : 0

MAGNIFICATION: L RESOLUTION: M

BASELINE THRESHOLD: 0 MIN. PEAK WIDTH: 1.00

REGION: 2 - WITH MARKERS  

| ELMT | K.E.    | STEP | DWELL | #CHAN | #AVG |
|------|---------|------|-------|-------|------|
| XCL  | 1053.60 | 0.10 | 0.200 | 250   | 1    |

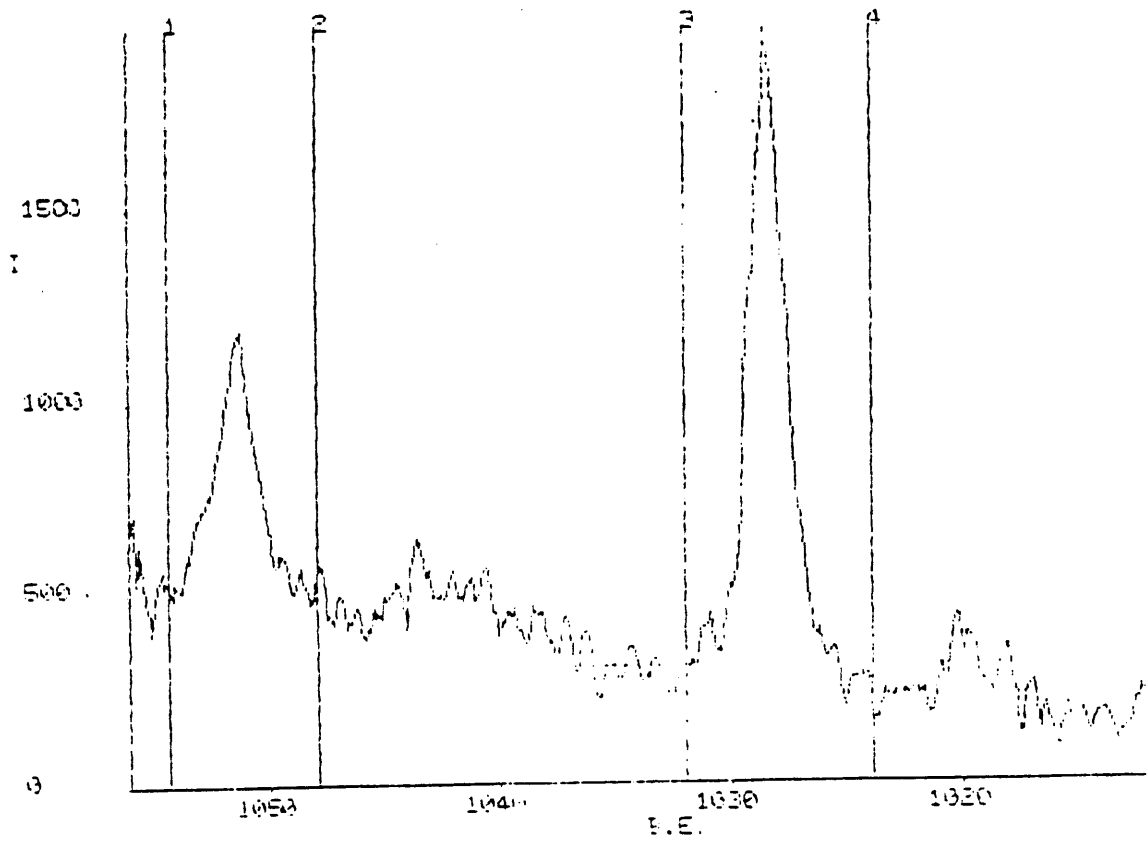
SCAN: 1

| PEAK | START E.V. : 1038.60 |        | END E.V. : 1063.50 |       | FWHM |
|------|----------------------|--------|--------------------|-------|------|
|      | K.E.                 | B.E.   | MAX                | AREA  |      |
| 1    | 1048.00              | 205.60 | 376                | 12202 | 3.10 |

XMS 10 2N2P

END

Run: -Y030 / 23-APR-84 REG: 1.001 ST: 0.10 #1: 450 #SW: 2 DWELL: 0.200



RUN : Y030 / 23-APR-84  
 SAMPLE: 1  
 NAME : 10  
 TITLE : IR YOON  
 15KV 2004 SLIT=2 CUR=9.0

EXCITATION : MG                    ANALYSER MODE: PPA  
 SCAN MODE : T                    FACTOR : 0

MAGNIFICATION: L                    RESOLUTION: 6

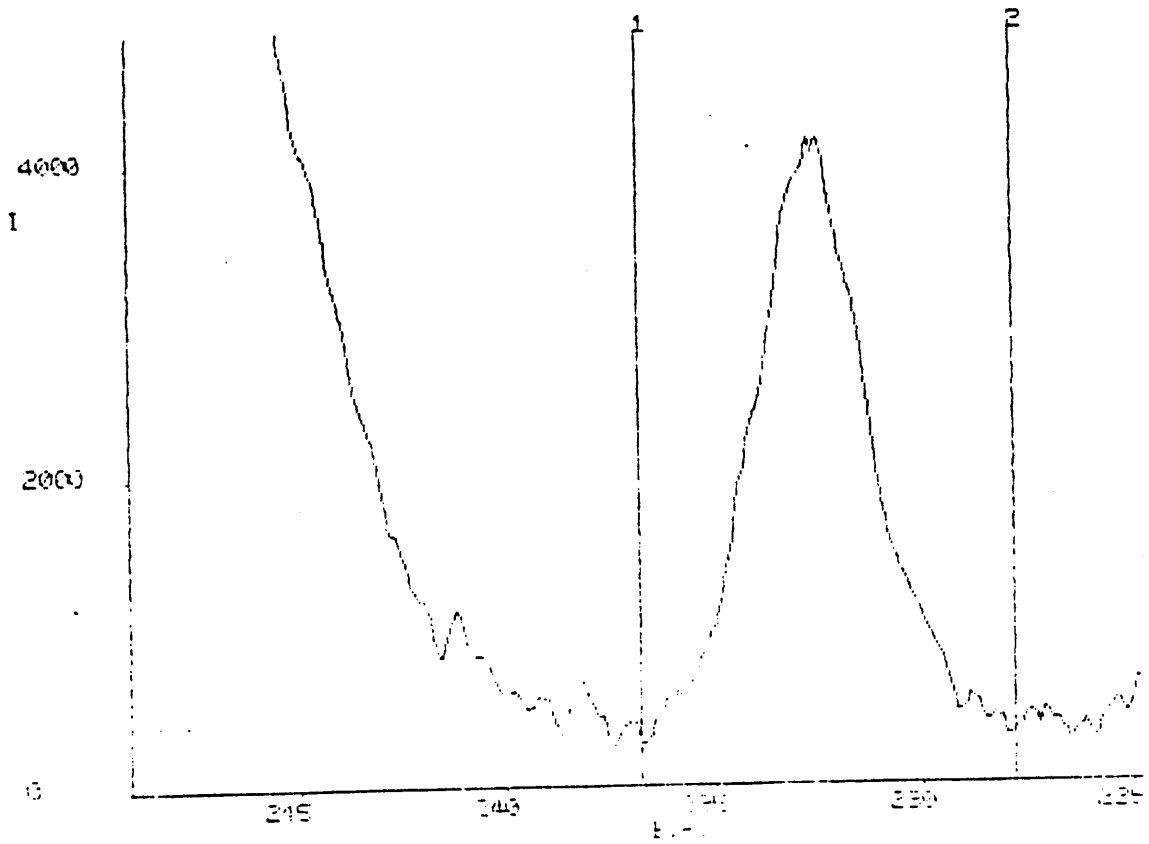
BASELINE THRESHOLD: 0            MIN. PEAK WIDTH: 1.00

REGION: 1 - WITH MARKERS  
 ELMT    K.E.        STEP        DWELL        #CHAN        #AVG  
 XZN     232.60      0.10        0.200        450          2

SCAN: 1

| PEAK | START E.V. : 197.80 |         | END E.V. : 242.50 |       | FWHM |
|------|---------------------|---------|-------------------|-------|------|
|      | K.E.                | B.E.    | HAN               | AREA  |      |
| 1    | 202.30              | 1051.30 | 1881              | 14524 | 1.00 |
| 2    | 225.30              | 1028.30 | 1721              | 33143 | 1.00 |

1141 1C S2S  
END  
RUN: -Y030 / 29-APR-84 REG: 3.001 ST: 0.10 #C: 250 #SW: 4 DWELL: 0.200



RUN: 1039 - 20777-34  
 SAMPLE: 1  
 COUNT: 10  
 TITLE: IN 10014  
 10KV XMM SLIT 22 CUR 9.0

EXCITATION: 15 MA ANALYSER MODE: FFF  
 SCAN MODE: 1 FACTOR: 0

MAGNIFICATION: 10 RESOLUTION: 14

BASELINE THRESHOLD: 0 MIN. PEAK WIDTH: 1.00

REGION: 3 - WITH MARKERS  
 ELIMT K.E. STEP DWELL #CHN #AVG  
 XS25 1019.60 0.10 0.200 250 4

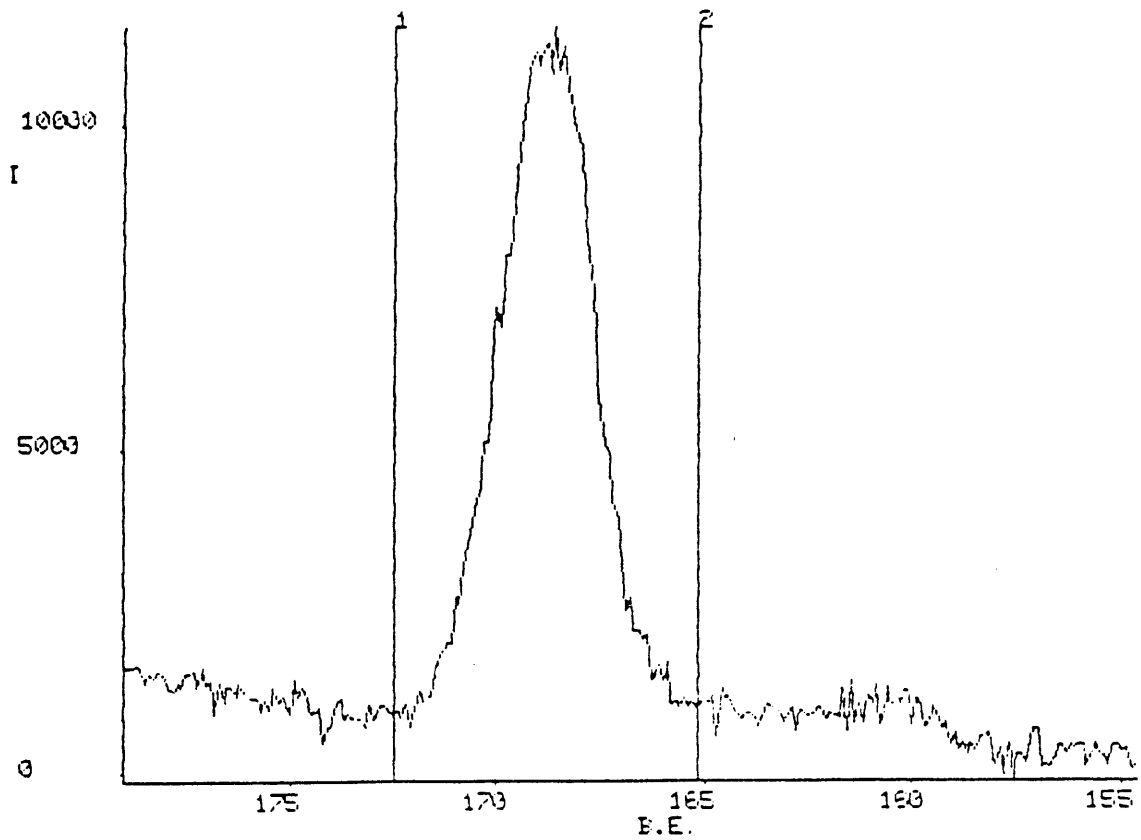
SCAN: 1

| PEAK | START E.V. : 1004.60 |        | END E.V. : 1029.50 |        | FWHM |
|------|----------------------|--------|--------------------|--------|------|
|      | K.E.                 | E.E.   | MAX                | AREA   |      |
| 1    | 1021.00              | 230.20 | 3095               | 125198 | 2.94 |

\*SU 1C 32P

END

RUN: -Y010 / 29-APR-84 REG: 1.001 ST: 0.10 #C: 250 #SW: 5 DWELL: 0.200



RUN : Y010 / 29-APR-84  
 SAMPLE: 1  
 NAME : 1C  
 TITLE : DR YOON  
 13KV 20MA SLIT=2 CUR=9.0

EXCITATION : MG ANALYSER MODE: FRR  
 SCAN MODE : T FACTOR : 0

MAGNIFICATION: L RESOLUTION: M

BASELINE THRESHOLD: 0 MIN. PEAK WIDTH: 1.00

REGION: 1 - WITH MARKERS

| ELMT | K.E.    | STEP | DWELL | #CHAN | #AVG |
|------|---------|------|-------|-------|------|
| XS   | 1029.60 | 0.10 | 0.200 | 250   | 5    |

SCAN: 1

| PEAK | START E.V. : 1074.60 |        | END E.V. : 1099.50 |        | FWHM |
|------|----------------------|--------|--------------------|--------|------|
|      | K.E.                 | E.E.   | MAX                | AREA   |      |
| 1    | 1085.00              | 108.60 | 10302              | 284414 | 2.30 |

RUN REG SCAN STEP #CH  
 Y010 1 1 .10 88

ELMT K.E. DWELL  
 XS 1029.60 .200

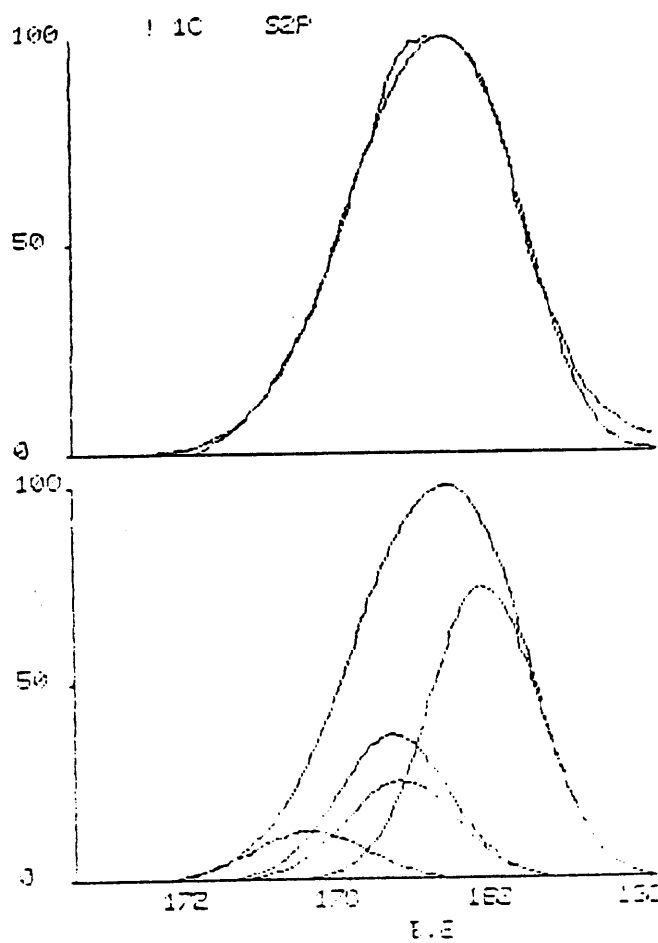
START EV: 173.60  
 END EV: 184.90

100% AREA =232932

100% INTENSITY =10200

FIT=10

| PK | ENERGY | MAX   | FWHM | AREA |
|----|--------|-------|------|------|
| 1G | 170.50 | 12.00 | 1.70 | 9.1  |
| 2G | 169.30 | 25.00 | 1.70 | 16.3 |
| 3G | 168.40 | 37.00 | 1.70 | 24.2 |
| 4G | 168.20 | 74.00 | 1.70 | 48.4 |



RUN REG SCAN STEP #CH  
 1010 1 1 .10 00

ELM7 K.E. DWELL  
 75 1000.60 .200

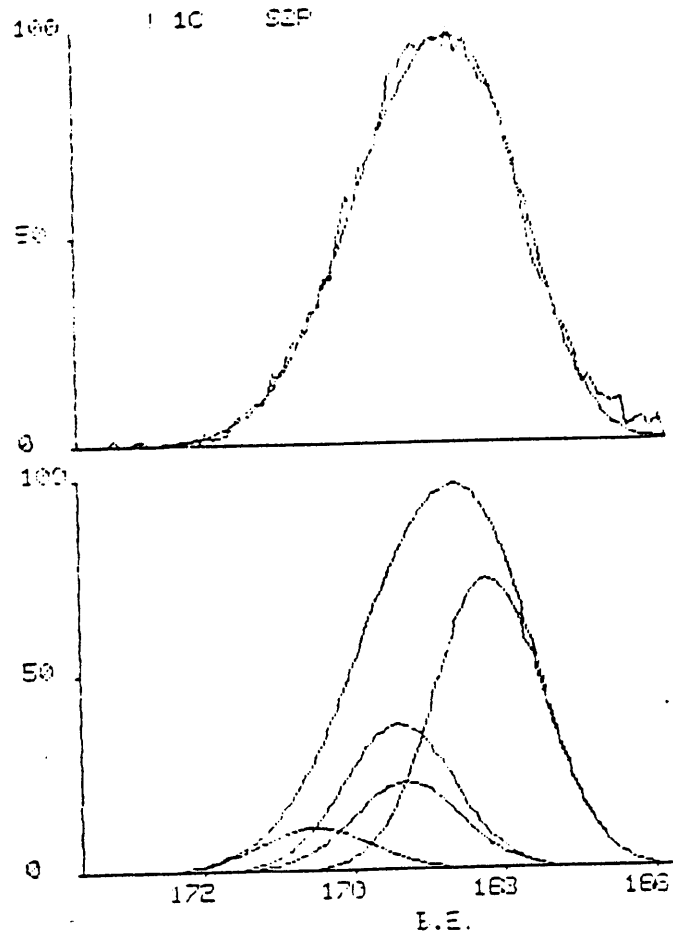
START EV: 173.60  
 END EV: 164.90

100% AREA = 231604

100% INTENSITY = 10510

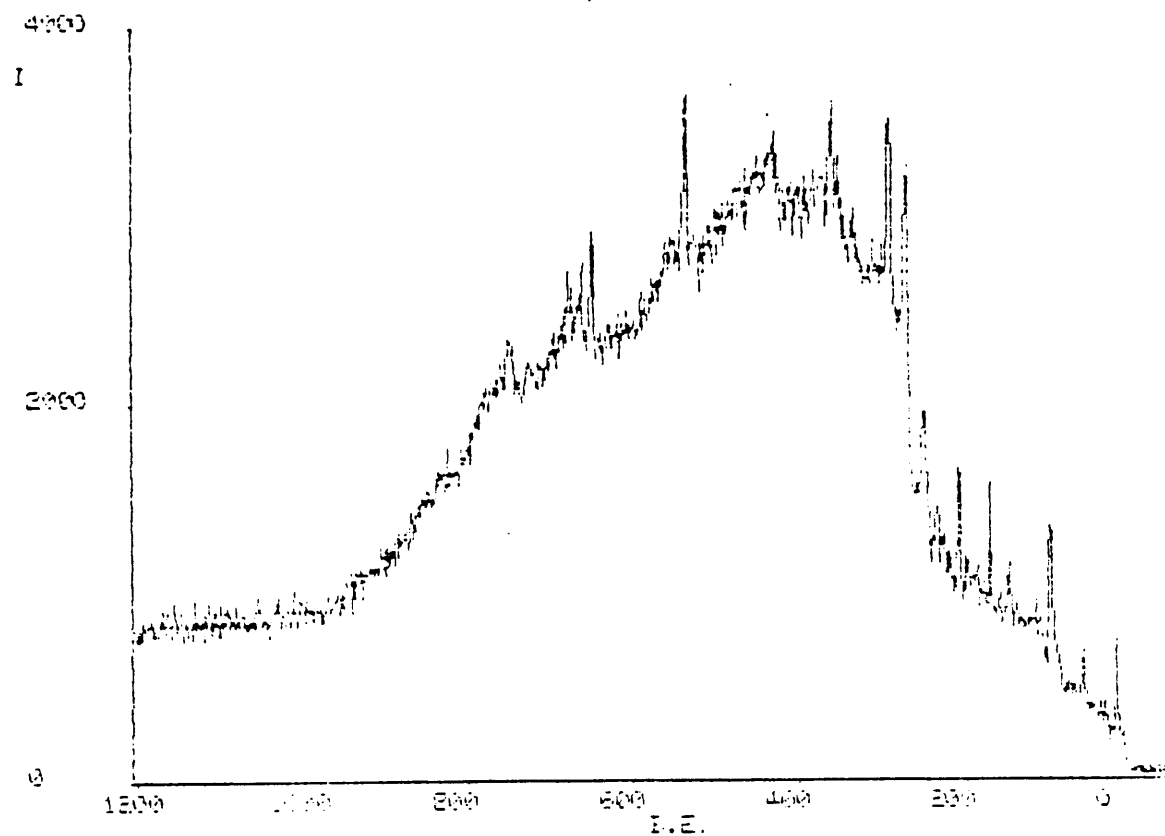
FIT=18

| PK | ENERGY | MAX   | FWHM | AREA |
|----|--------|-------|------|------|
| 10 | 170.50 | 11.00 | 1.70 | 7.4  |
| 20 | 169.30 | 22.00 | 1.70 | 14.9 |
| 30 | 169.40 | 37.00 | 1.70 | 25.0 |
| 40 | 168.20 | 74.00 | 1.70 | 50.1 |



VPE 2T  
C10

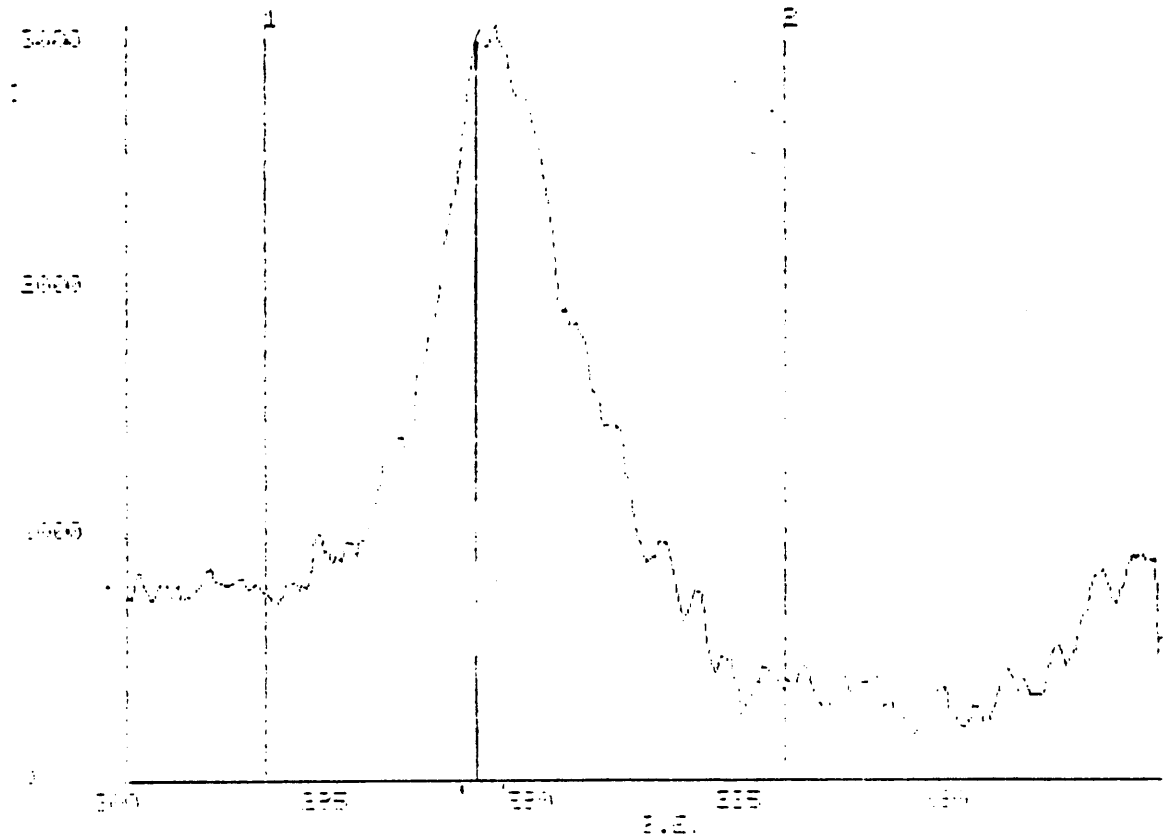
RUN: -V100 / 29-APR-84 REG: 1.001 ST: 1.00 #0: 1250 #GM: 1 DWELL: 0.100



RM1 ET C15

END

VOL. 0180 / EB-PR-84 REA: 3.001 ET:0.10 #0. LEO 430: 2 DWELL: 0.200



RUN : Y120 / 29-APR-84  
 SAMPLE: 1  
 LANE : 27  
 TITLE : DR YOON  
 13KV 20MA SLIT=2 CUR=9.0

EXCITATION : MG ANALYSER MODE: FRR  
 SCAN MODE : T FACTOR : 0

MAGNIFICATION: L RESOLUTION: M

BASLINE THRESHOLD: 0 MIN. PEAK WIDTH: 1.00

REGION: 3 - WITH MARKERS

| SCAN | K.E.   | STEP | DWELL | #CHAN | #AVG |
|------|--------|------|-------|-------|------|
| NO1  | 953.50 | 0.10 | 0.200 | 350   | 2    |

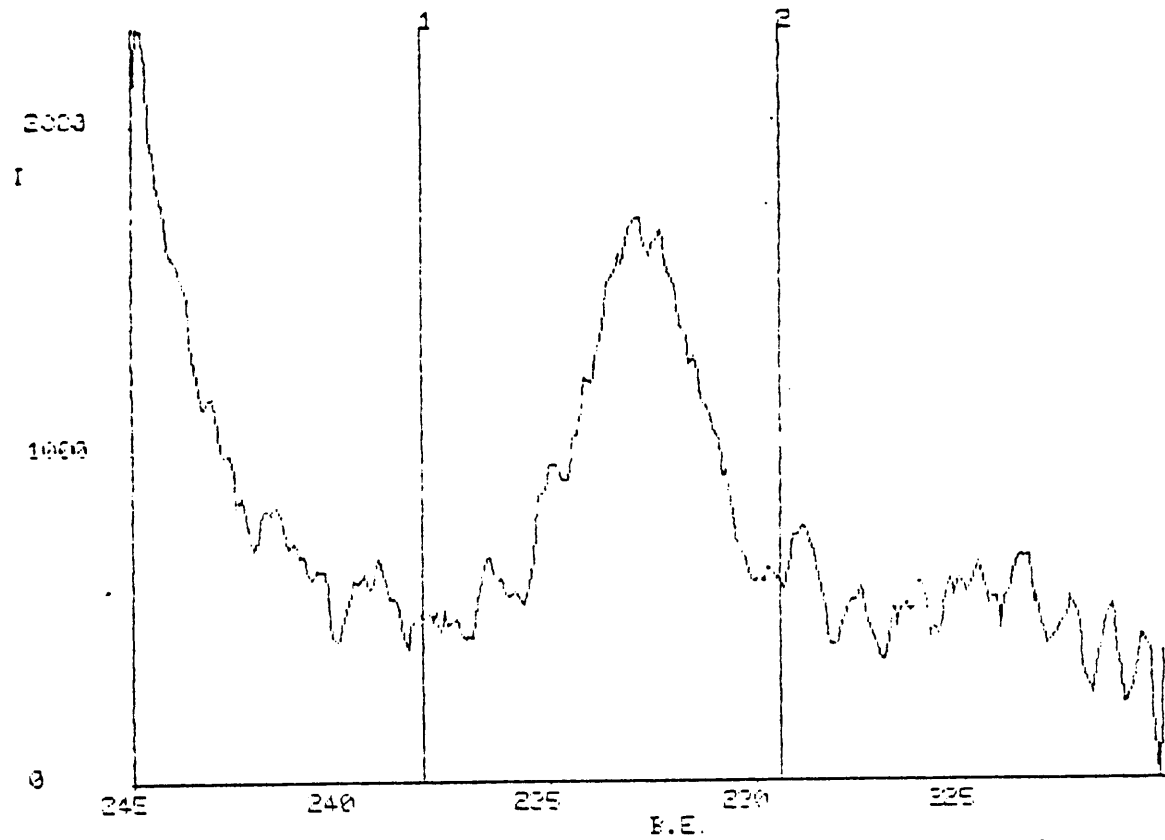
SCAN: 1

| PEAK | START E.V. :<br>K.E. | END E.V. :<br>B.E. | MAX  | AREA   | FWHM |
|------|----------------------|--------------------|------|--------|------|
| 1    | 962.50               | 991.10             | 2513 | 111379 | 3.50 |

M41 2T 925

E.V.C.

P.M.: -1.120 / 29-APR-84 REG: 2.001 ST: 0.10 #C: 250 #SW: 5 DWELL: 0.200



RUN : Y120 / 29-APR-84  
 SAMPLE: 1  
 NAME : ZT  
 TITLE : DR YOON  
 13KV 20MA SLIT=2 CUR=9.0

EXCITATION : MG ANALYSER MODE: FRR  
 SCAN MODE : T FACTOR : 0

MAGNIFICATION: L RESOLUTION: M

BASELINE THRESHOLD: 0 MIN. PEAK WIDTH: 1.00

REGION: 2 - WITH MARKERS

| EVENT | K.E.    | STEP | DWELL | #CHAN | #AVG |
|-------|---------|------|-------|-------|------|
| K523  | 1023.80 | 0.10 | 0.200 | 250   | 5    |

SCAN: 1

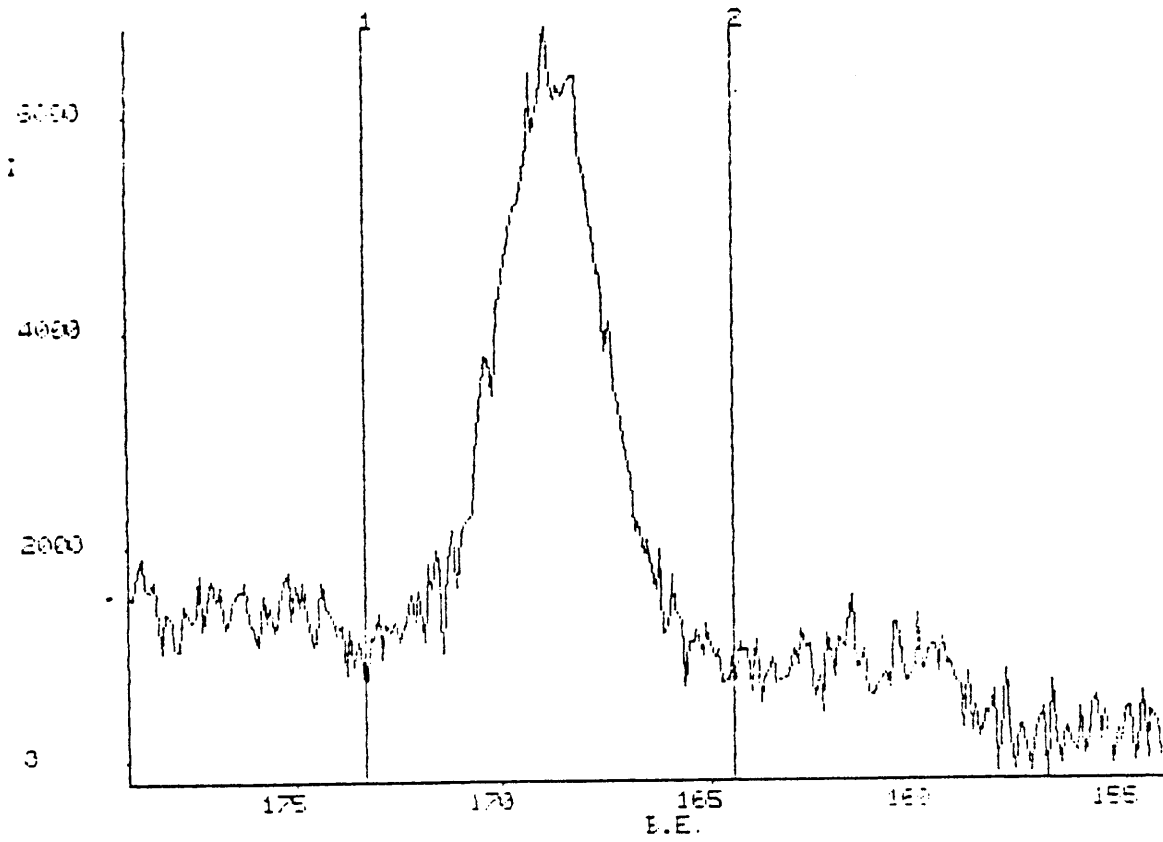
| PEAK | K.E.    | B.E.   | MAX  | AREA  | FWHM |
|------|---------|--------|------|-------|------|
| 1    | 1020.70 | 332.90 | 1178 | 30373 | 3.10 |

START E.V. : 1000.60 END E.V. : 1033.50

END ET 32F

END

REQ: -Y120 / 29-APR-64 REG: 1.001 ST: 0.10 #C: 250 #SW: 10 DWELL: 0.200



RUN : Y120 / 29-APR-84  
 SAMPLE: 1  
 NAME : 2T  
 TITLE : DR YOON  
 13KV 20MA SLIT=2 CUR=9.0

EXCITATION : MG ANALYSER MODE: FRR  
 SCHM MODE : T FACTOR : 0

MAGNIFICATION: L RESOLUTION: M

BASELINE THRESHOLD: 0 MIN. PEAK WIDTH: 1.00

REGION: 1 - WITH MARKERS

| EURT | K.E.    | STEP | DWELL | #CHAN | #AVG |
|------|---------|------|-------|-------|------|
| 25   | 1033.80 | 0.10 | 0.200 | 250   | 10   |

SCAN: 1

| PEAK | K.E.    | B.E.   | MAX  | AREA   | FWHH |
|------|---------|--------|------|--------|------|
| 1    | 1034.70 | 163.30 | 5627 | 181164 | 3.10 |

START E.V. : 1074.80 END E.V. : 1099.50

RUN REG SCAN STEP #CH  
Y120 1 1 .10 88

ELMT K.E. DWELL  
XS 1089.60 .200

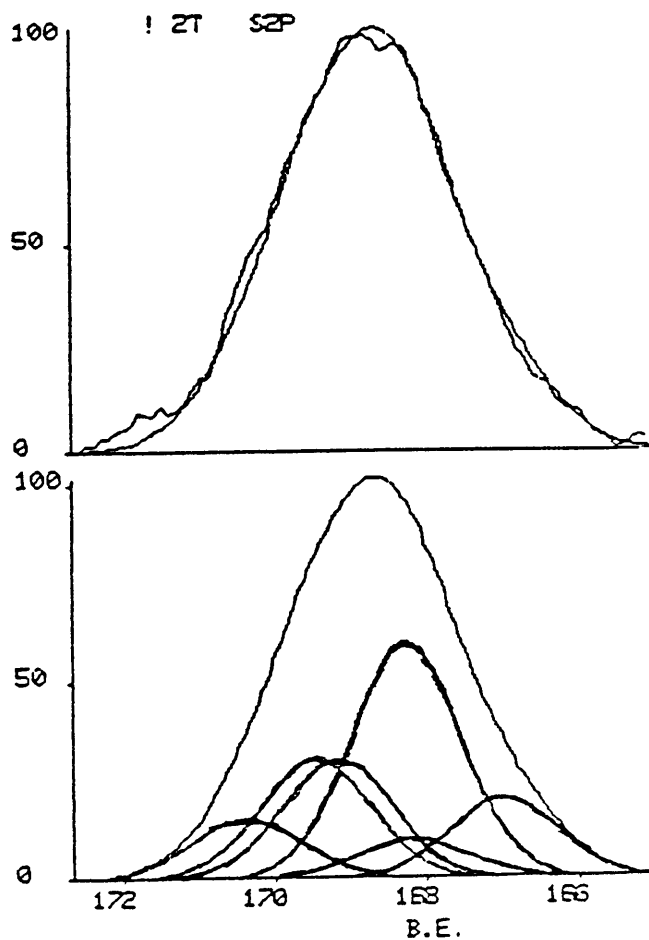
START EV: 172.70  
END EV: 164.00

100% AREA =164644

100% INTENSITY =5387

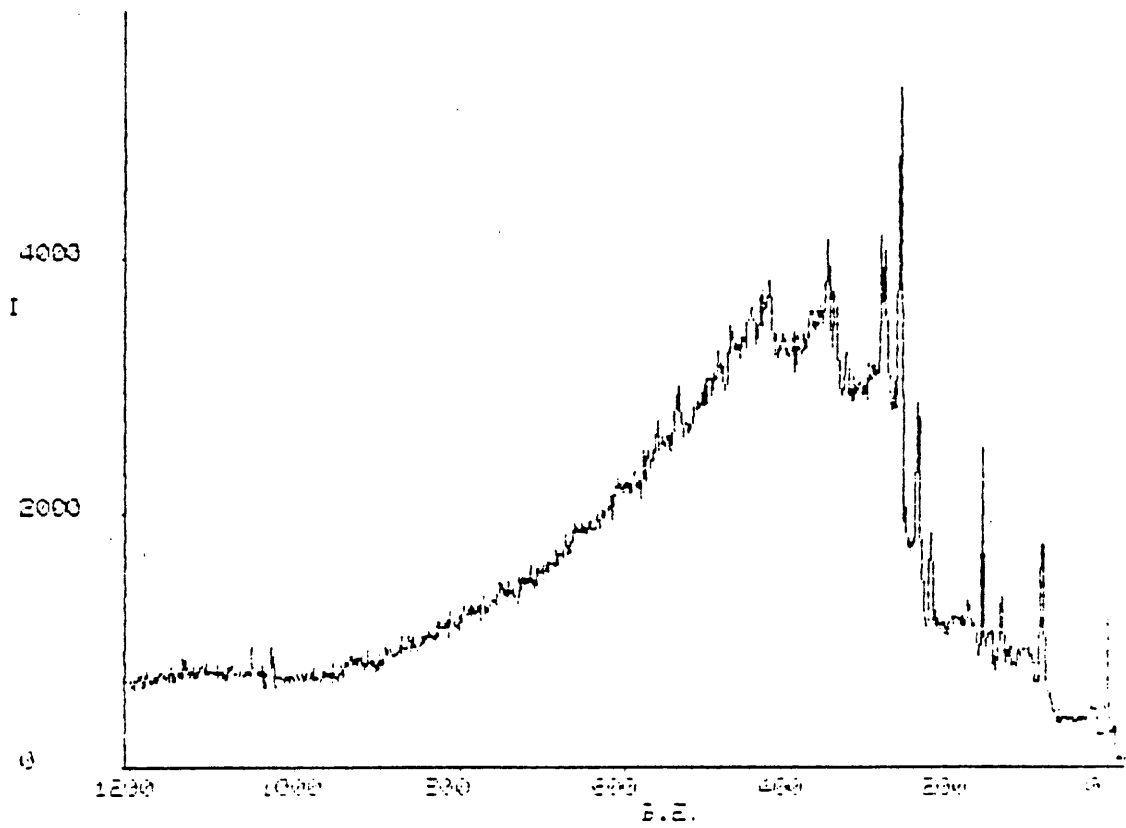
FIT=15

| PK | ENERGY | MAX   | FWHM | AREA |
|----|--------|-------|------|------|
| 1G | 170.40 | 15.00 | 1.70 | 8.9  |
| 2G | 169.20 | 30.00 | 1.70 | 17.8 |
| 3G | 169.50 | 30.00 | 1.70 | 17.8 |
| 4G | 168.30 | 60.00 | 1.70 | 35.6 |
| 5G | 168.20 | 10.00 | 1.70 | 5.9  |
| 6G | 167.00 | 20.00 | 1.70 | 11.8 |



\*EE ST  
END

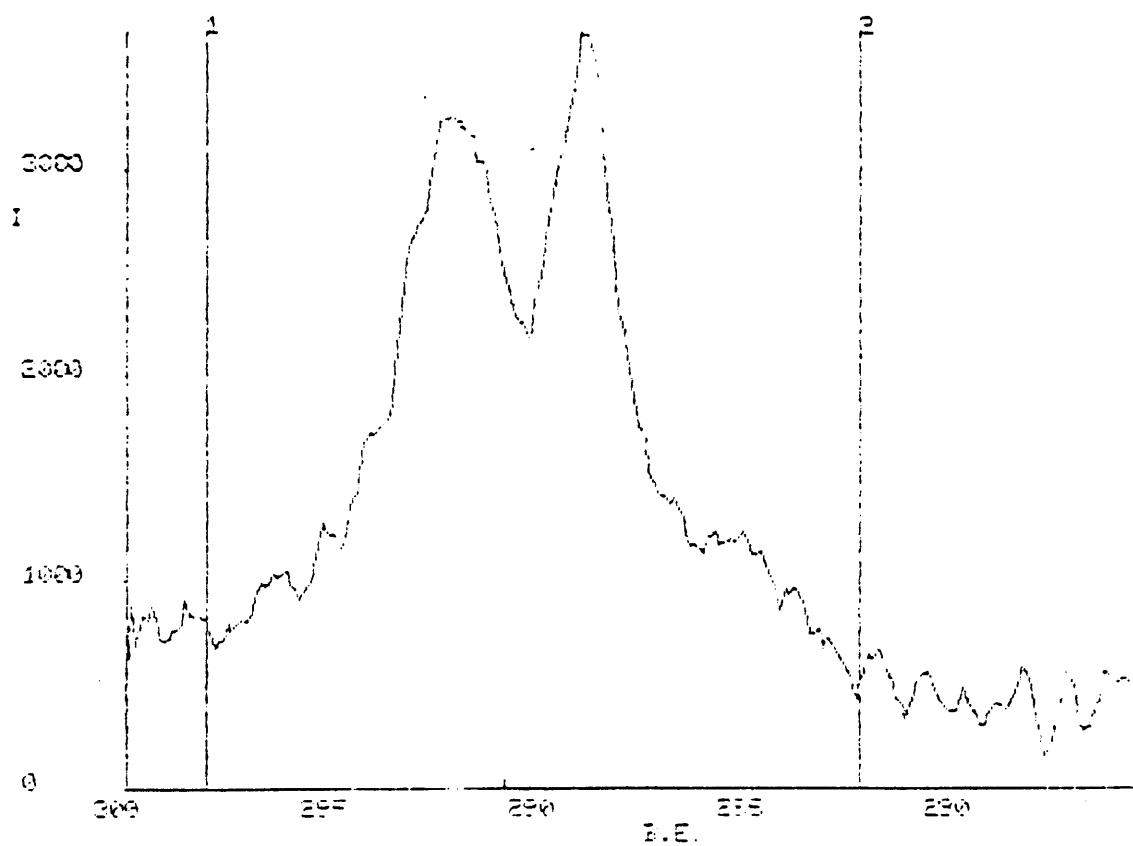
RUN:-Y110 / 29-APR-84 REG: 1.001 ST: 1.00 #C: 1253 #SW: 1 DWELL: 0.100



M41 37 C18

EIE

RUN: -Y100 / 29-APR-84 REG: 5.001 ST: 0.10 #C: 250 #S4: 2 DWELL: 0.200



RUN : Y100 / 29-APR-84  
 SAMPLE: 1  
 NAME : ST  
 TITLE : DR YOON  
 13KV 20MA SLIT=2 CUR=9.0

EXCITATION : MG ANALYSER MODE: FRR  
 SCAN MODE : T FACTOR : 0

MAGNIFICATION: L RESOLUTION: M

BASELINE THRESHOLD: 0 MIN. PEAK WIDTH: 1.00

REGION: 3 - WITH MARKERS  

| ELMT | K.E.   | STEP | DWELL | #CHAN | #AVG |
|------|--------|------|-------|-------|------|
| XC1  | 963.60 | 0.10 | 0.200 | 250   | 2    |

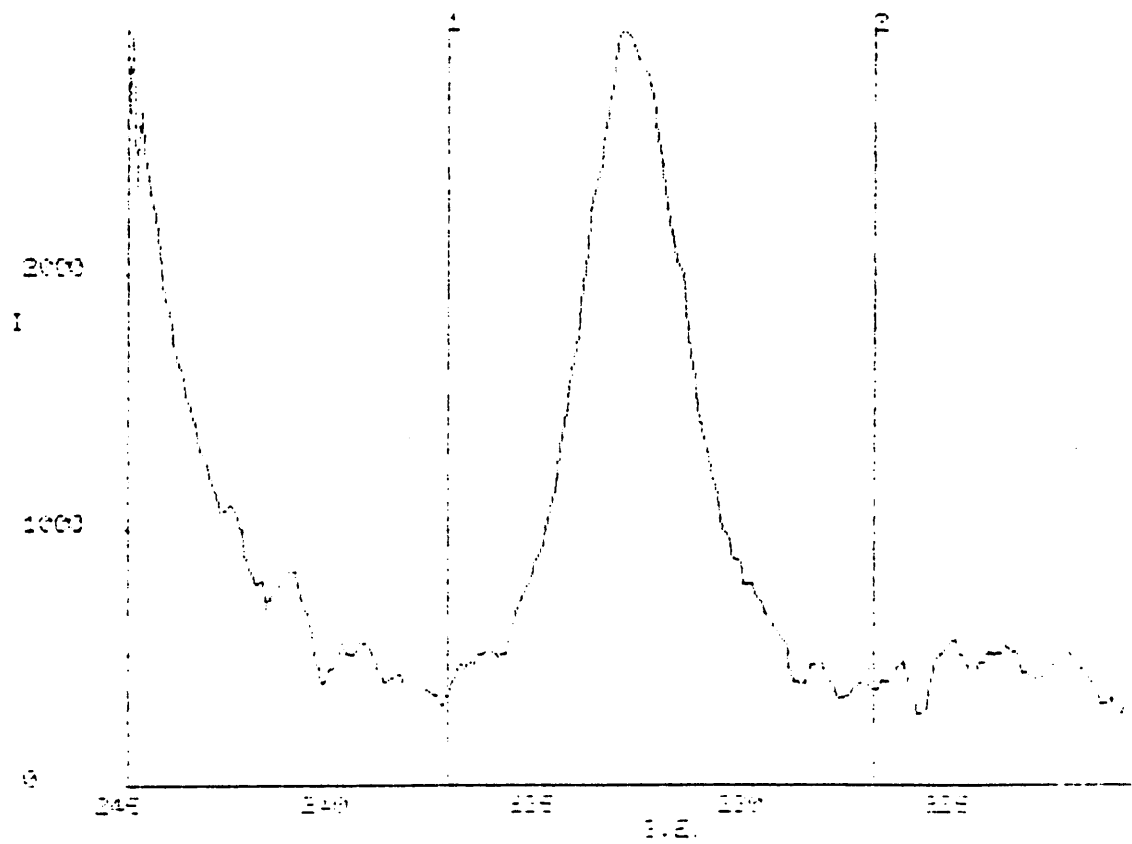
SCAN: 1

| PEAK | START E.V. : 553.60 |        | END E.V. : 978.50 |        | FWHM |
|------|---------------------|--------|-------------------|--------|------|
|      | K.E.                | B.E.   | MAX               | AREA   |      |
| 1    | 564.60              | 225.00 | 3111              | 180310 | 5.40 |

M41 ST 123

END

RUN: -Y100 / 23-HPF-34 RES: 2.001 ST: 0.10 #C: 250 #SM: 3 SWELL: 0.107



RUN : Y100 / 23-APR-84  
 SAMPLE: 1  
 NAME : ST  
 TITLE : DR YOON  
 15KV 30MA SLIT=2 CUR=3.0

EXCITATION : MG ANALYSER MODE: PEP  
 SCHM MODE : T FACTOR : 0

MAGNIFICATION: L RESOLUTION: M

BASELINE THRESHOLD: 0 MIN. PEAK WIDTH: 1.00

REGION: 2 - WITH PAR-ERS

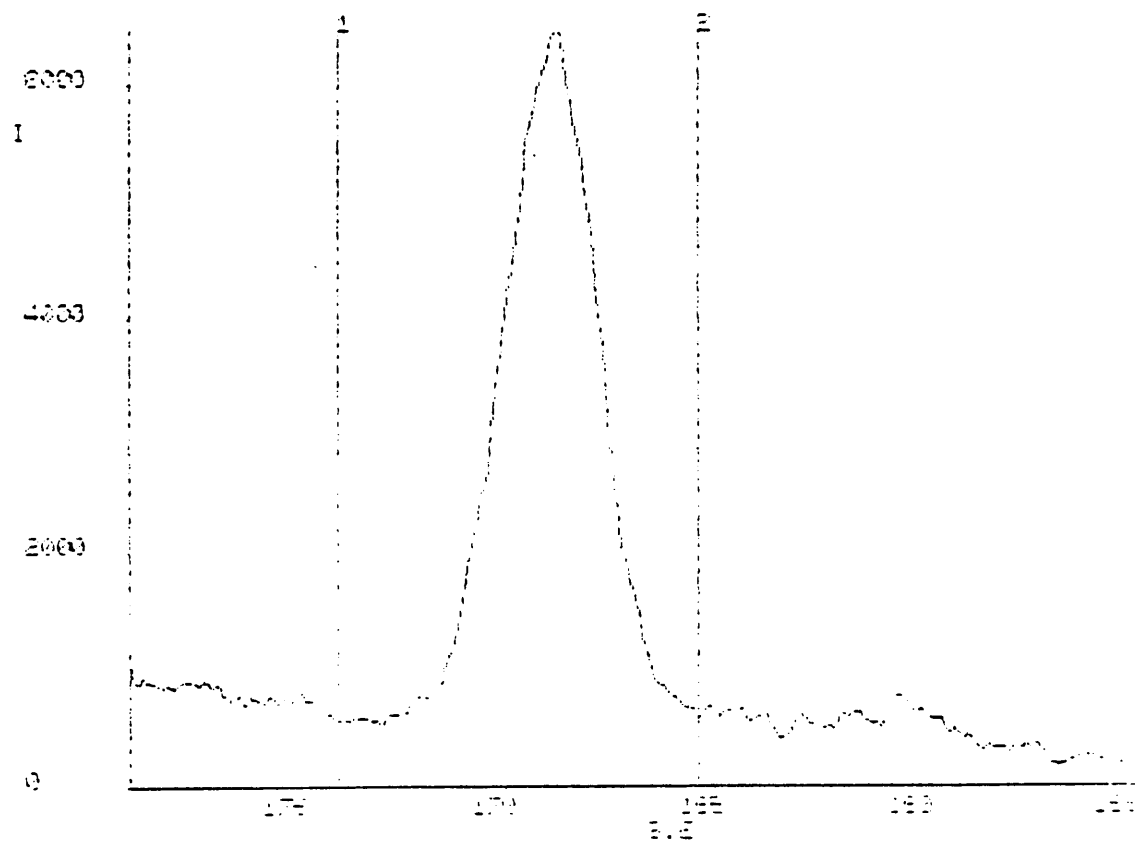
| ELMT | K.E.    | STEP | DWELL | #CHAN | #AVG |
|------|---------|------|-------|-------|------|
| X326 | 1025.50 | 0.10 | 0.200 | 250   | 3    |

SCAN: 1

| PEAK | START E.V.<br>K.E. | END E.V.<br>E.V. | AREA   | AREA | AREA   |
|------|--------------------|------------------|--------|------|--------|
| 1    | 1020.50            | 1035.50          | 233.00 | 2333 | 233.00 |

ASU ST SEP  
END

RUN: -1100 / 23-APR-84 REG: 1.001 ST: 0.10 AC: 250 #84 3 DWELL: 0.000



RUN : 1100 / 28-APR-84  
 SAMPLE: 1  
 NAME : BT  
 TITLE : DR YOON  
 15KV 20MA SLIT=2 CUR=3.0

EXCITATION : MG ANALYSER MODE: PRR  
 SCAN MODE : T FACTOR : 0

MAGNIFICATION: L RESOLUTION: M

BASLINE THRESHOLD: 0 MIN. PEAK WIDTH: 1.00

REGION: 1 - WITH MARKERS

| EUNT | K.E.    | STEP | DWELL | EDWIN | #AVG |
|------|---------|------|-------|-------|------|
| XS   | 1089.00 | 0.10 | 0.200 | 250   | 3    |

SCAN: 1

| PEAK | START E.V. : 1074.00 END E.V. : 1099.50 |        | COUNT | AREA   | FWHM |
|------|---|--------|-------|--------|------|
|      | K.E.                                    | E.V.   |       |        |      |
| 1    | 1084.00                                 | 125.00 | 8337  | 153913 | 2.00 |

RUN REG SCAN STEP #CH  
Y100 1 1 .10 86

ELMT K.E. DWELL  
XS 1089.60 .200

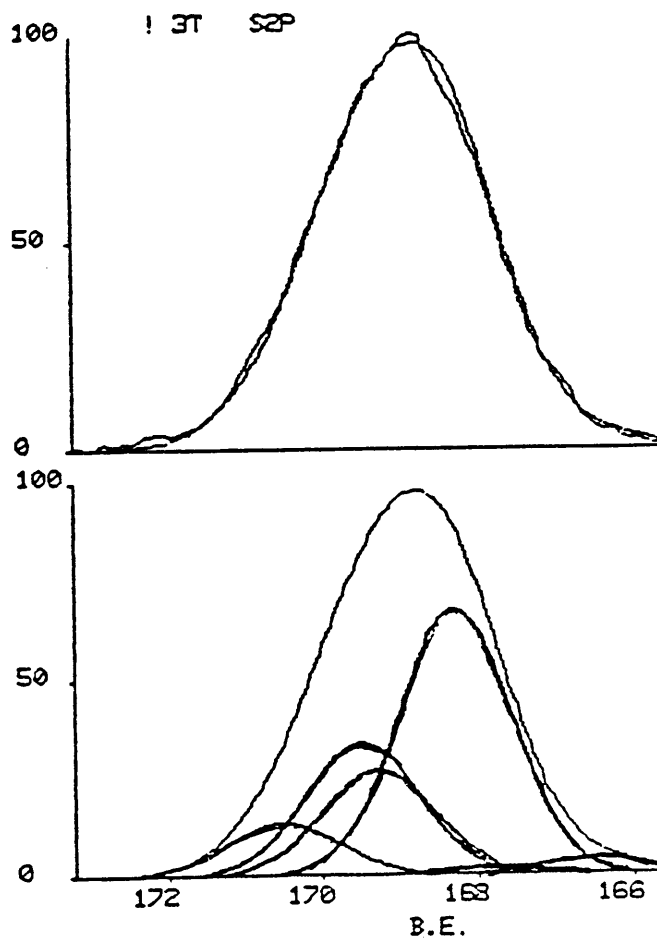
START EV: 173.20  
END EV: 164.70

100% AREA =157937

100% INTENSITY =5878

FIT=7

| PK | ENERGY | MAX   | FWHM | AREA |
|----|--------|-------|------|------|
| 1G | 170.50 | 13.50 | 1.70 | 9.1  |
| 2G | 169.30 | 27.00 | 1.70 | 18.2 |
| 3G | 169.50 | 34.00 | 1.70 | 22.9 |
| 4G | 168.30 | 68.00 | 1.70 | 45.9 |
| 5G | 167.70 | 2.00  | 1.70 | 1.3  |
| 6G | 166.50 | 4.00  | 1.70 | 2.6  |



RUN REG SCAN STEP #CH  
Y100 1 1 .10 86

ELMT K.E. DWELL  
XS 1089.60 .200

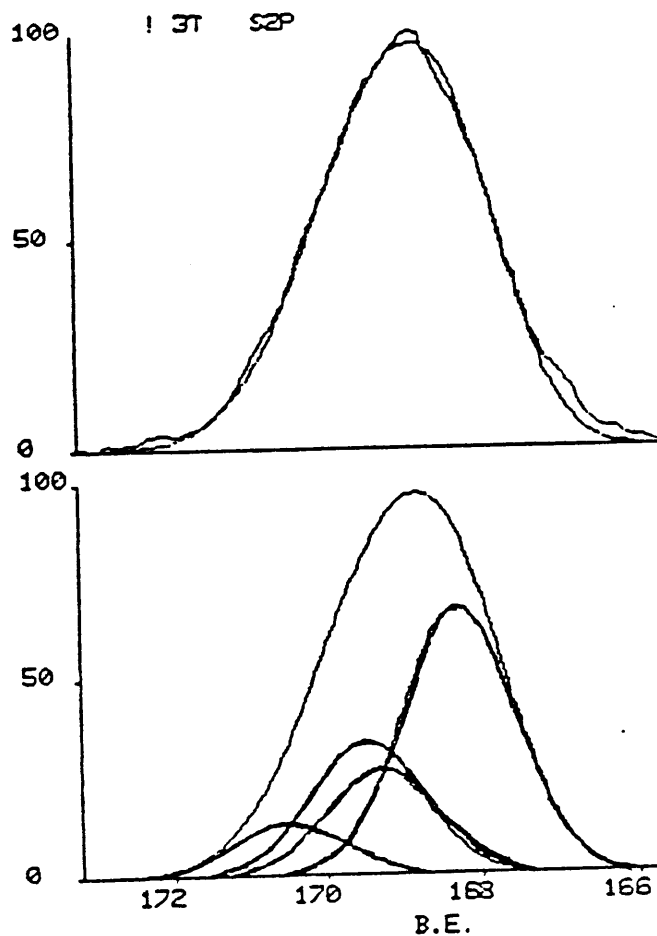
START EV: 173.20  
END EV: 164.70

100% AREA =157937

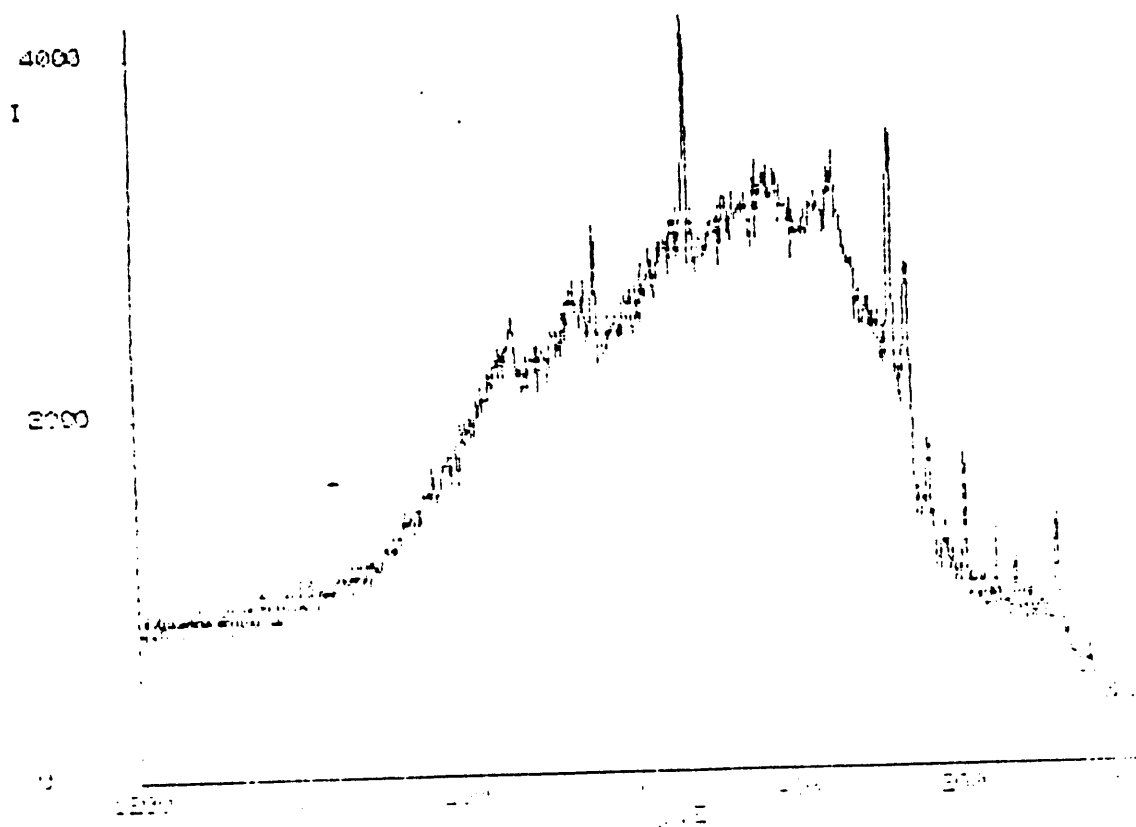
100% INTENSITY =5878

FIT=11

| PK | ENERGY | MAX   | FWHM | AREA |
|----|--------|-------|------|------|
| 1G | 170.50 | 13.50 | 1.70 | 9.1  |
| 2G | 169.30 | 27.00 | 1.70 | 18.2 |
| 3G | 169.50 | 34.00 | 1.70 | 22.9 |
| 4G | 168.30 | 68.00 | 1.70 | 45.9 |



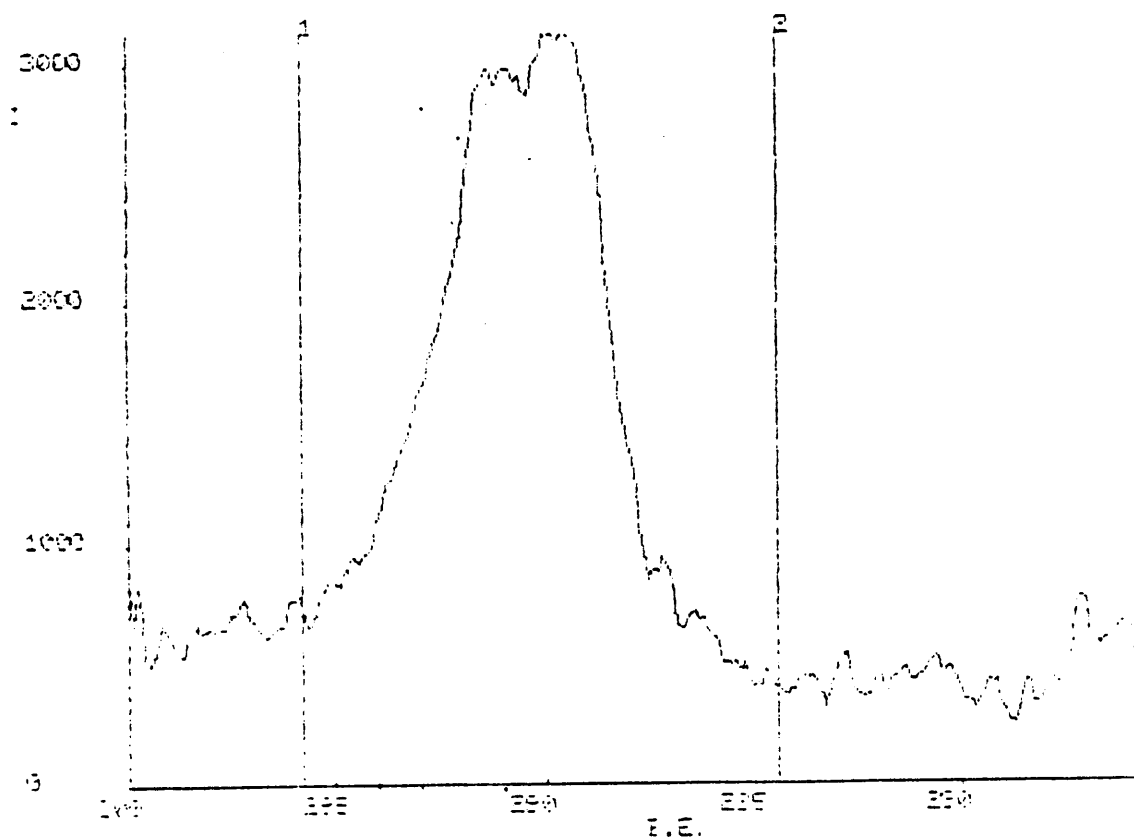
\*BE 47  
END  
RUN: -Y090 / 25-PP-34 REG: 1.001 ST: 1.00 #C: 1250 #SW: 1 DWELL: 0.100



M41 47 C15

END

RUN: -1050 / 28-PP-34 REG: 3.001 ST: 0.10 RD: 250 #SW: 2 DWELL: 0.200



RUN : Y000 23-APR-84  
 SAMPLE: 1  
 NAME : CT  
 TITLE : DR Y000  
 15KV 20MA SLIT=2 CUR=3.0

EXCITATION : MA ANALYSER MODE: PRR  
 SCAN MODE : T FACTOR : 0

MAGNIFICATION: L RESOLUTION: M

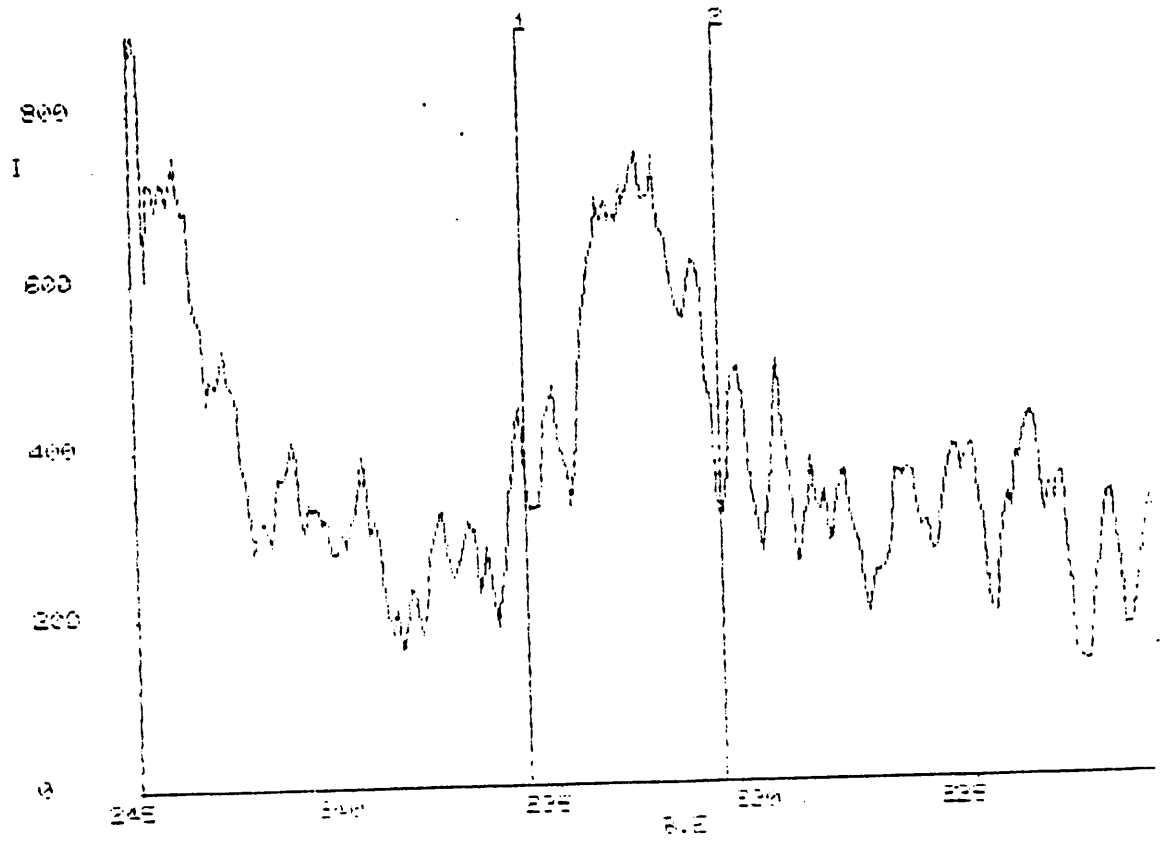
BASELINE THRESHOLD: 0 MIN. PEAK WIDTH: 1.00

| REGION: | 3 - WITH MARKERS |      |       |       |      |
|---------|------------------|------|-------|-------|------|
| ELVT    | K.E.             | STEP | DWELL | #CHAN | #AVG |
| XC1     | 553.80           | 0.10 | 0.200 | 250   | 2    |

SCAN: 1

| PEAK | START E.V. :<br>K.E. | END E.V. :<br>B.E. | MAX  | AREA   | FWHM |
|------|----------------------|--------------------|------|--------|------|
| 1    | 554.20               | 559.40             | 2627 | 121540 | 4.10 |

M41 4T 823  
END  
RUN: -Y030 / 23-APR-84 REG: 2.001 ST: 0.10 #C: 250 #SW. 3 DWELL: 0.200



RUN : Y050 / 23-APR-84  
 SAMPLE: 1  
 NAME : 4T  
 TITLE : IR Y00H  
 13KV 20MA SLIT=2 CUR=9.0

EXCITATION : MG ANALYSER MODE: FRR  
 SCAN MODE : T FACTOR : 0

MAGNIFICATION: L RESOLUTION: 0

BASELINE THRESHOLD: 0 MIN. PEAK WIDTH: 1.00

REGION: 2 - WITH MARKERS  
 ELMT K.E. STEP DWELL #CHAN #AVG  
 X028 1028.60 0.10 0.200 250 3

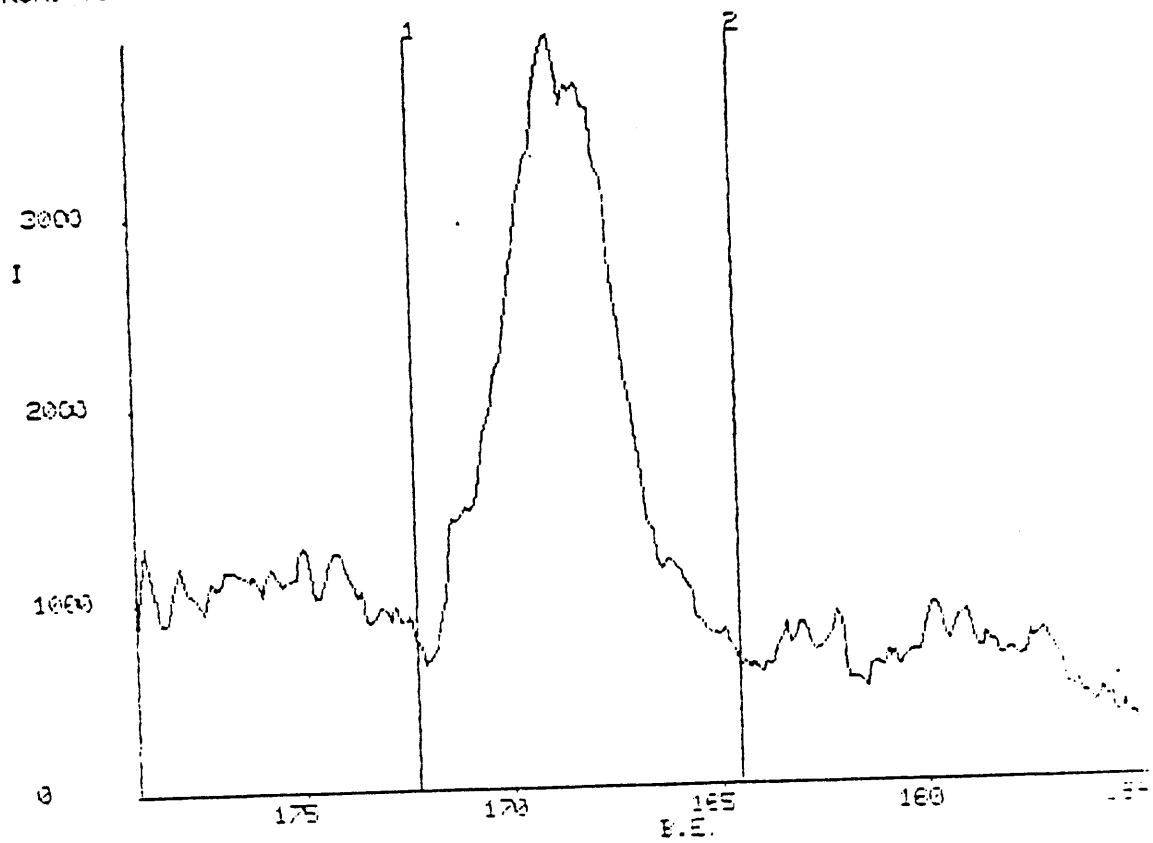
SCAN: 1

| PEAK | START E.V.<br>K.E. | END E.V.<br>E.V. | AREA | AREA  | FWHM |
|------|--------------------|------------------|------|-------|------|
| 1    | 1028.60            | 1033.50          | 033  | 10337 | 3.00 |

XSU 4T SZP

END

RUN: -Y060 / 29-APR-84 REG: 1.001 ST: 0.10 #C: 250 #SW: 10 DWELL: 0.200



RUN : YDS0 / 29-APR-84  
 SAMPLE: 1  
 NAME : 4T  
 TITLE : IR YOON  
 13KV 20MA SLIT=2 CUR=0.0

EXCITATION : MG ANALYSER MODE: FRR  
 SCAN MODE : T FACTOR : 0

MAGNIFICATION: L RESOLUTION: M

BASELINE THRESHOLD: 0 MIN. PEAK WIDTH: 1.00

| REGION: | 1       | WITH MARKERS |       |       |      |  |
|---------|---------|--------------|-------|-------|------|--|
| ELMT    | K.E.    | STEP         | DWELL | #CHAN | #AVG |  |
| X5      | 1039.60 | 0.10         | 0.200 | 250   | 10   |  |

SCAN: 1

| * PEAK | START E.V.<br>K.E. | END E.V.<br>B.E. | MAX  | AREA   | FWHM |
|--------|--------------------|------------------|------|--------|------|
| 1      | 1034.60            | 1039.00          | 3218 | 102764 | 2.30 |

RUN REG SCAN STEP #CH  
Y020 1 1 .10 85

ELMT K.E. DWELL  
XS 1089.60 .200

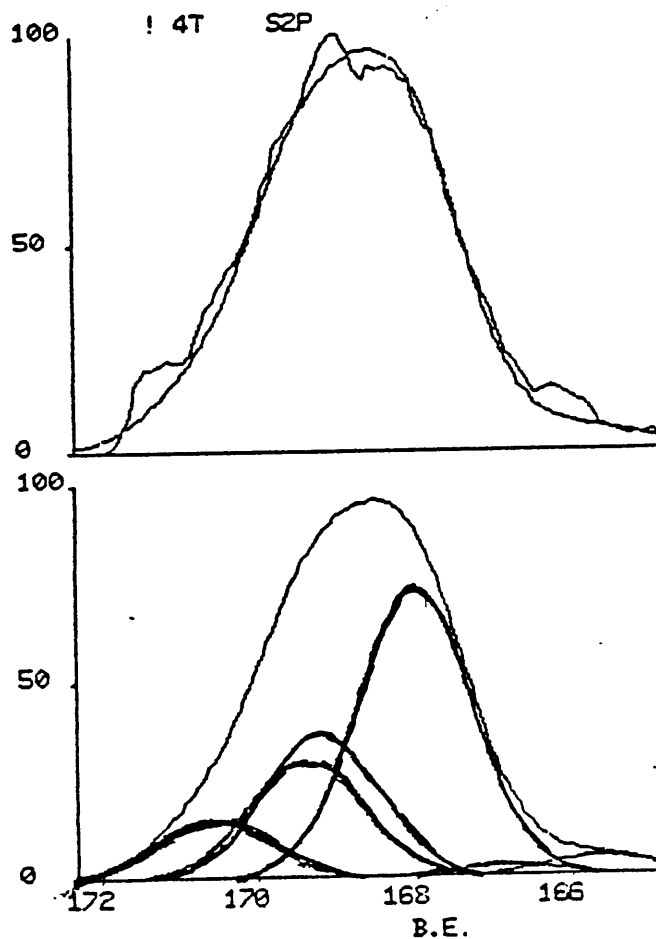
START EV: 172.30  
END EV: 163.90

100% AREA =99424

100% INTENSITY =3201

FIT=39

| PK | ENERGY | MAX   | FWHM | AREA |
|----|--------|-------|------|------|
| 1G | 170.60 | 15.00 | 1.70 | 8.6  |
| 2G | 169.40 | 30.00 | 1.70 | 17.5 |
| 3G | 169.20 | 37.00 | 1.70 | 21.6 |
| 4G | 168.00 | 74.00 | 1.70 | 43.2 |
| 5G | 165.60 | 5.00  | 1.70 | 2.9  |
| 6G | 166.80 | 2.50  | 1.70 | 1.4  |



RUN REG SCAN STEP #CH  
Y080 1 1 .10 94

ELMT K.E. DWELL  
XS 1089.60 .200

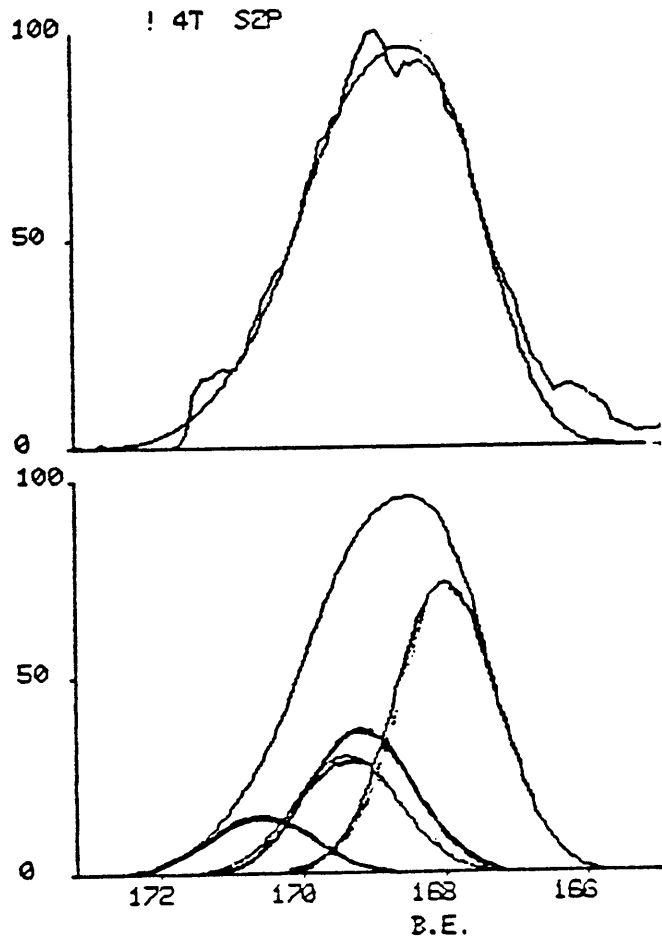
START EV: 173.20  
END EV: 163.90

100% AREA =94822

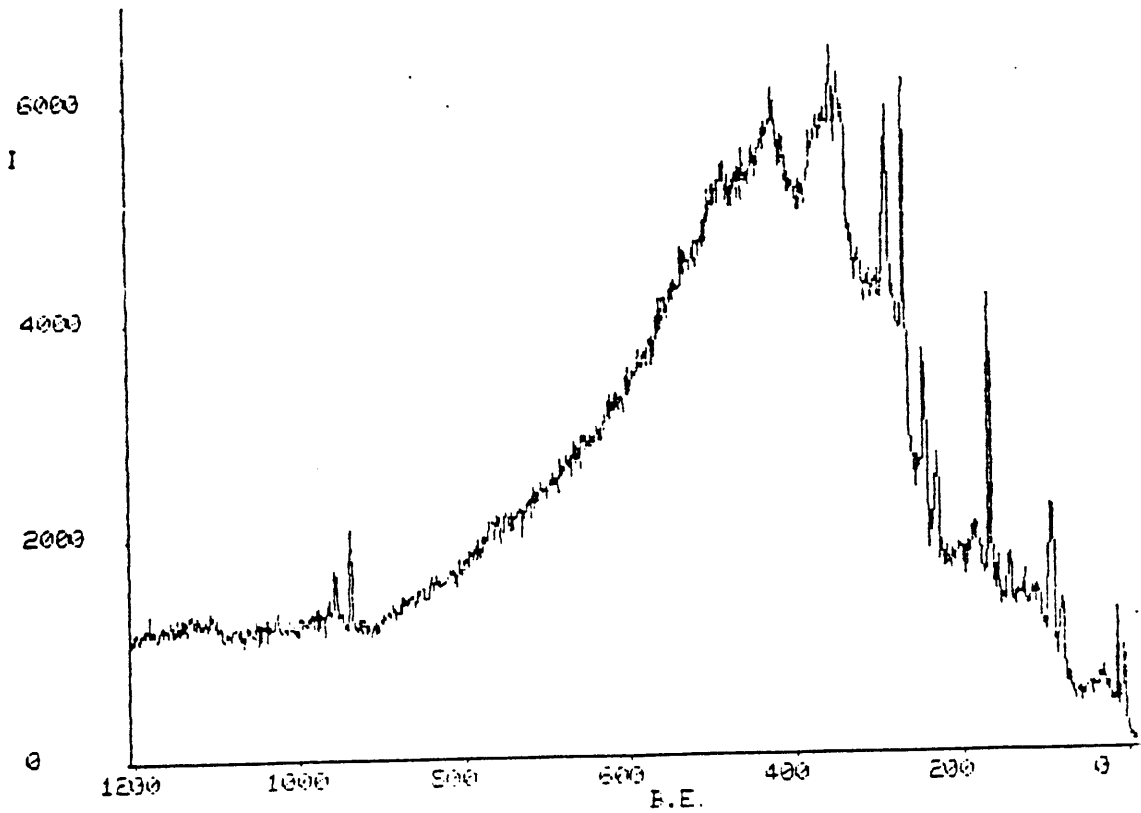
100% INTENSITY =3124

FIT=4S

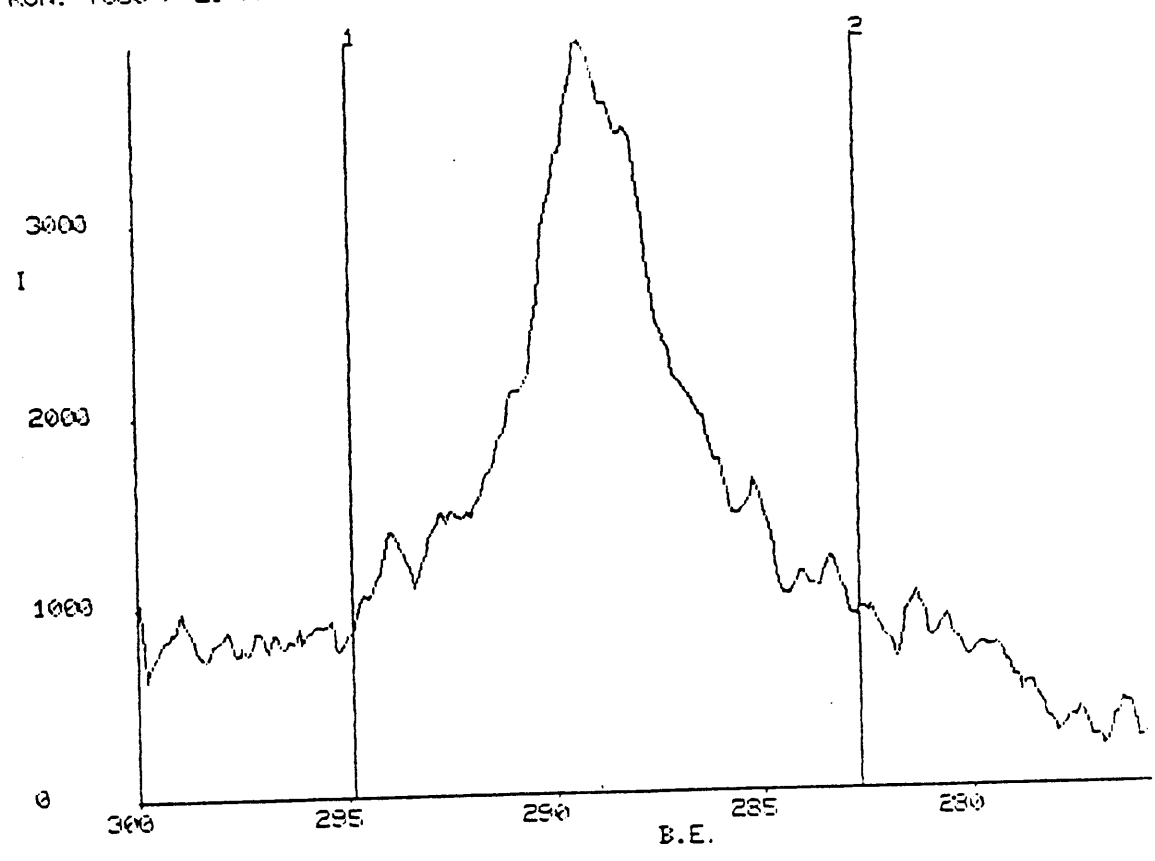
| PK | ENERGY | MAX   | FWHM | AREA |
|----|--------|-------|------|------|
| 1G | 170.60 | 15.00 | 1.70 | 8.9  |
| 2G | 169.40 | 30.00 | 1.70 | 17.9 |
| 3G | 168.20 | 37.00 | 1.70 | 22.1 |
| 4G | 168.00 | 74.00 | 1.70 | 44.2 |



\*SE SC  
END  
RUN: -Y070 / 29-APR-84 REG: 1.001 ST: 1.00 #C: 1250 #SM: 1 DWELL: 0.101



#41 50 C15  
EID  
RUN: -1060 / 23-APR-84 REG: 3.001 ST: 0.10 #C: 250 #SW: 2 DWELL: 0.200



RUN : Y060 / 29-APR-84  
SAMPLE: 1  
NAME : SC  
TITLE : DR YOON  
13KV 20MA SLIT=2 CUR=9.0

EXCITATION : MG ANALYSER MODE: FRR  
SCAN MODE : T FACTOR : 0

MAGNIFICATION: L RESOLUTION: M

BASELINE THRESHOLD: 0 MIN. PEAK WIDTH: 1.00

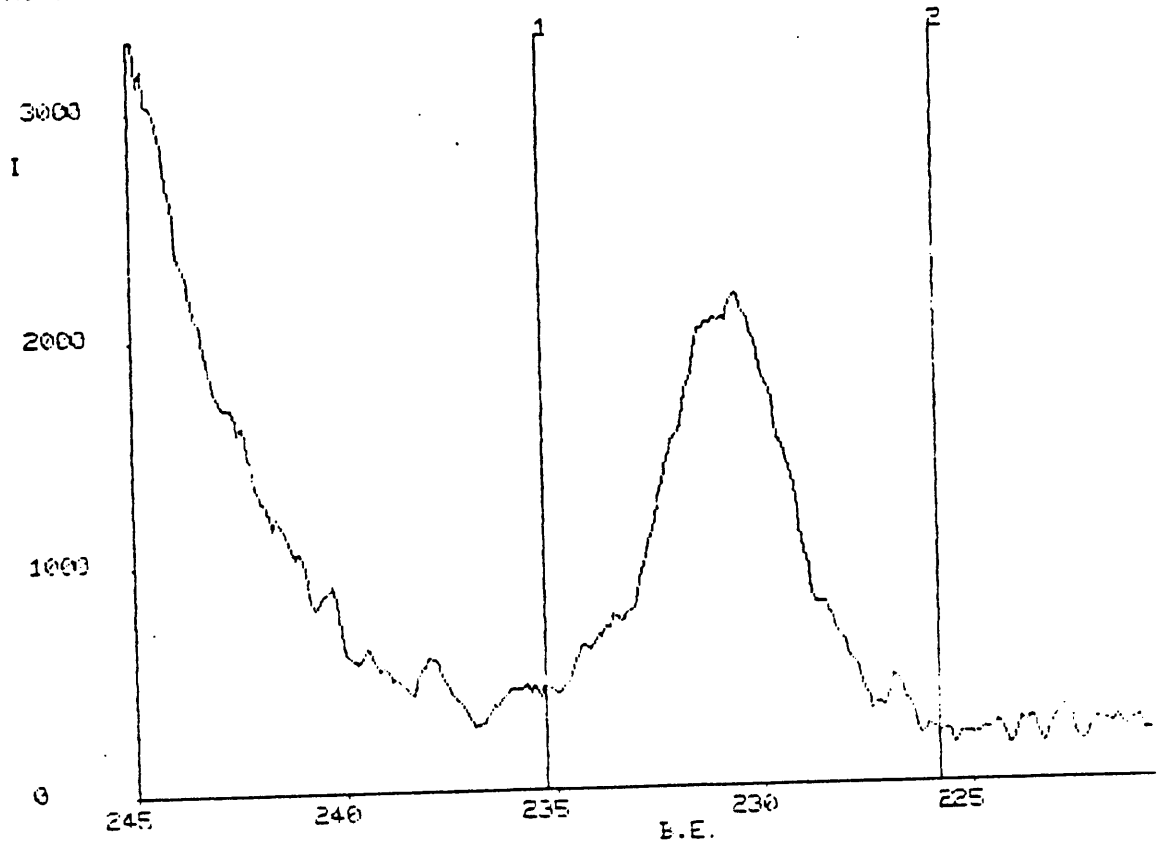
REGION: 3 - WITH MARKERS

| ELMT | K.E.   | STEP | DWELL | #CHAN | #AVG |
|------|--------|------|-------|-------|------|
| XC1  | 968.60 | 0.10 | 0.200 | 250   | 2    |

SCAN: 1

| PEAK | START E.V. :<br>K.E. | END E.V. :<br>B.E. | MAX  | AREA   | FWHM |
|------|----------------------|--------------------|------|--------|------|
| 1    | 964.30               | 289.30             | 3019 | 127276 | 3.00 |

#41 5C 325  
END  
RUN: -Y060 / 29-APR-84 REG: 2.001 ST: 0.10 #C: 250 #SW: 2 DWELL: 0.200



RUN : Y060 / 29-APR-84  
 SAMPLE: 1  
 NAME : SC  
 TITLE : DR YOON  
 13KV 20MA SLIT=2 CUR=9.0

EXCITATION : MG ANALYSER MODE: FRR  
 SCAN MODE : T FACTOR : 0

MAGNIFICATION: L RESOLUTION: M

BASELINE THRESHOLD: 0 MIN. PEAK WIDTH: 1.00

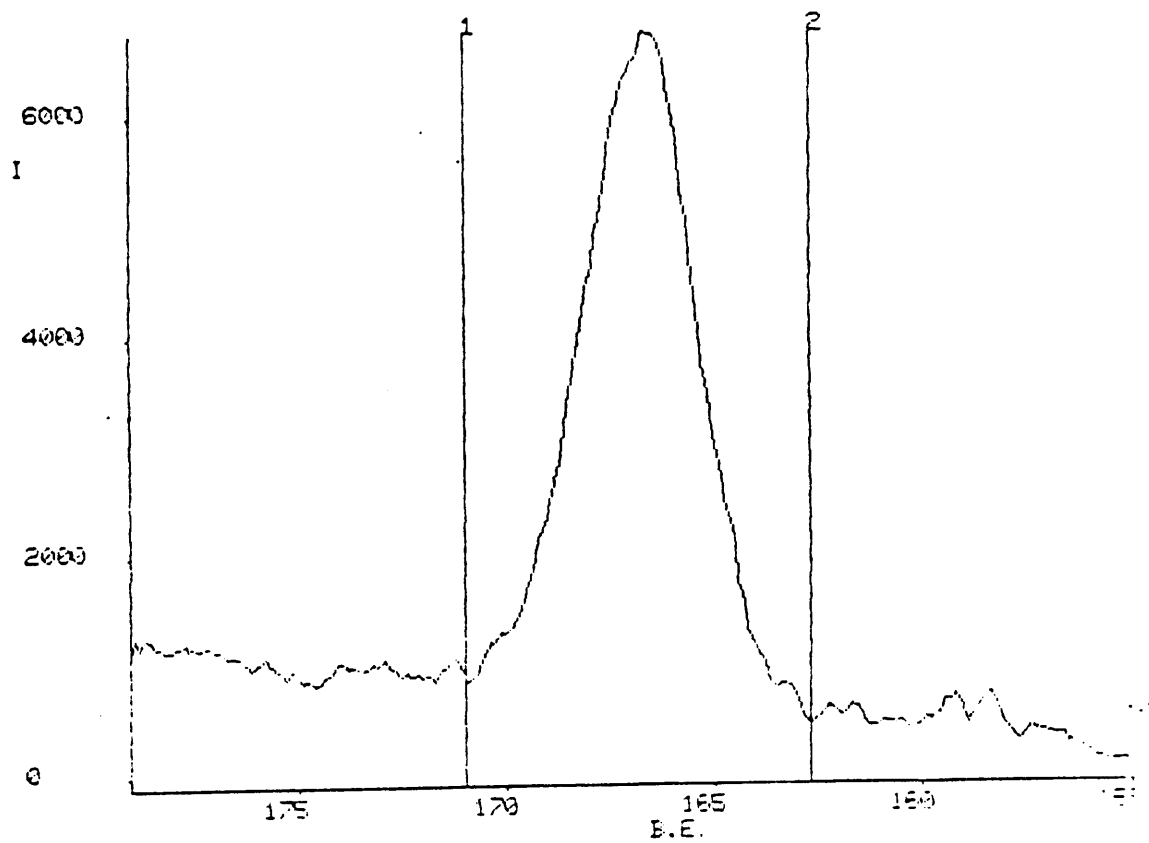
REGION: 2 - WITH MARKERS

| ELMT | K.E.    | STEP | DWELL | #CHAN | #AVG |
|------|---------|------|-------|-------|------|
| XS2S | 1023.60 | 0.10 | 0.200 | 250   | 2    |

SCAN: 1

| PEAK | START E.V. : 1008.60 |        | END E.V. : 1033.50 |       | FWHM |
|------|----------------------|--------|--------------------|-------|------|
|      | K.E.                 | E.E.   | MAX                | AREA  |      |
| 1    | 1023.10              | 230.50 | 1823               | 65178 | 3.30 |

\*SU 50 SEP  
END  
RUN: -Y060 / 29-APR-84 REG: 1.001 ST: 0.10 #C: 250 #SW: 3 DWELL: 0.20



RUN : Y060 / 29-APP-34  
 SAMPLE: 1  
 NAME : SC  
 TITLE : DR YOON  
 13KV 20MA SLIT=2 CUR=9.0

EXCITATION : MG ANALYSER MODE: FRR  
 SCAN MODE : T FACTOR : 0

MAGNIFICATION: L RESOLUTION: M

BASELINE THRESHOLD: 0 MIN. PEAK WIDTH: 1.00

REGION: 1 - WITH MARKERS

| ELMT | K.E.    | STEP | DWELL | #CHAN | #AVG |
|------|---------|------|-------|-------|------|
| XS   | 1089.60 | 0.10 | 0.200 | 250   | 3    |

SCAN: 1

| PEAK | START E.V. : 1074.60 |        | END E.V. : 1099.50 |        | FWHM |
|------|----------------------|--------|--------------------|--------|------|
|      | K.E.                 | B.E.   | MAX                | AREA   |      |
| 1    | 1037.10              | 165.50 | 6106               | 204353 | 3.10 |

ROW REG SCH. STEP #Ch  
 0000 1 1 .10 39

EDIT W.E. DWELL  
 NS 1089.60 1200

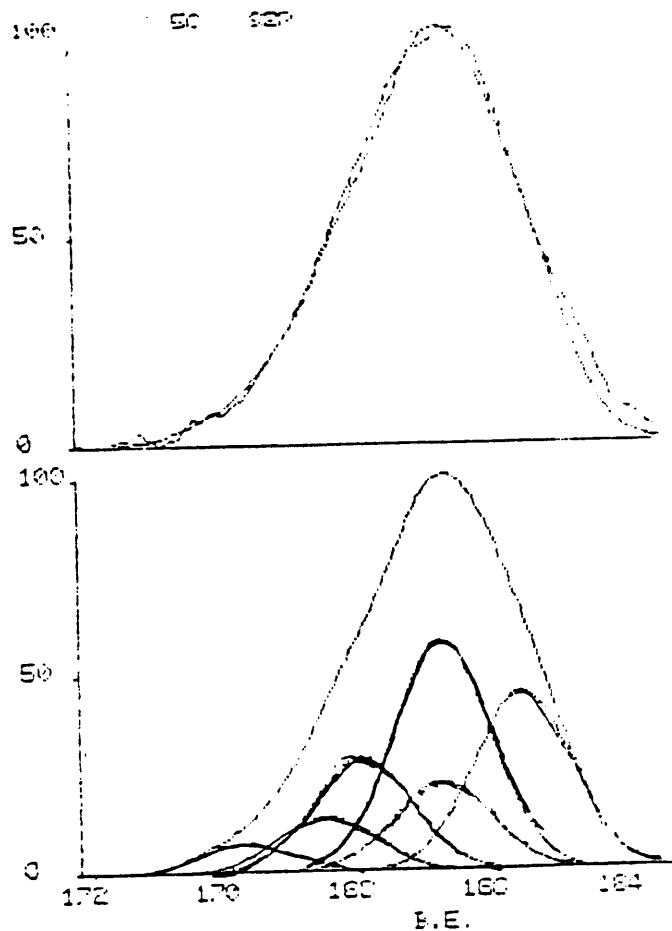
START EV: 172.20  
 END EV: 182.40

100% AREA =201051

100% INTENSITY =6042

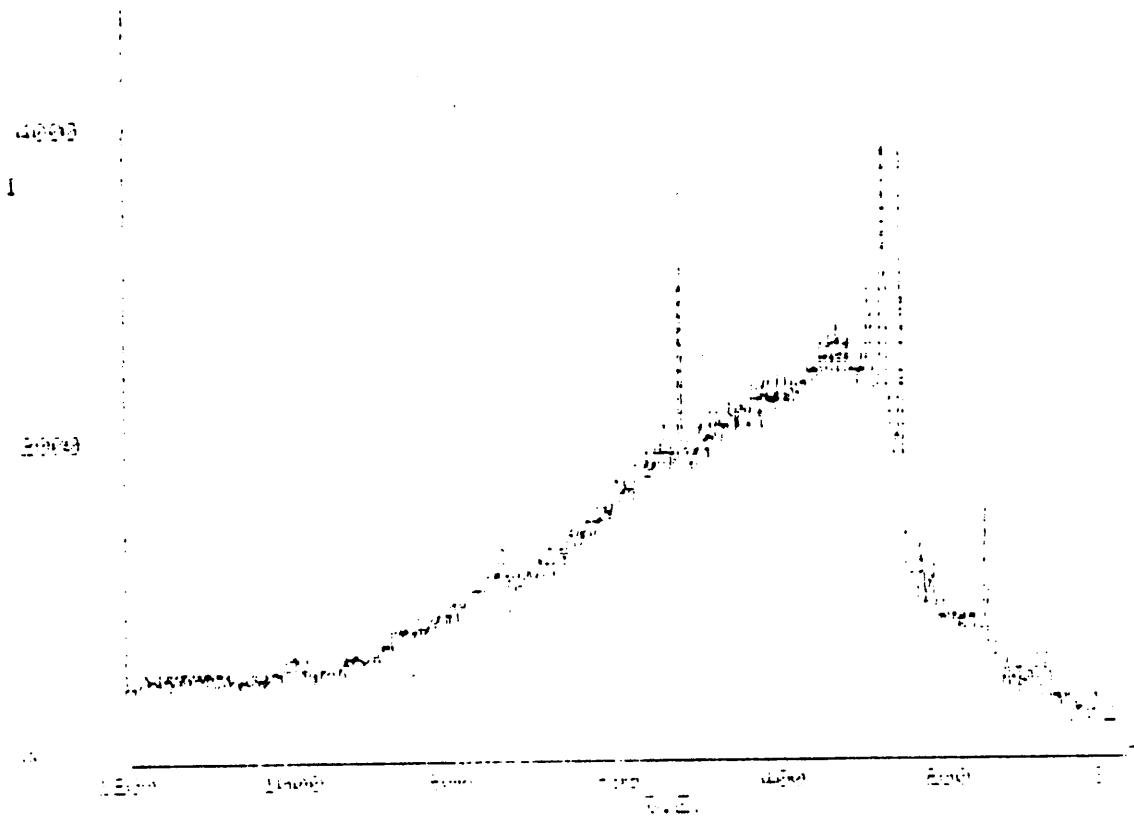
FIT=18

| PK | ENERGY | MAX   | FWHM | AREA |
|----|--------|-------|------|------|
| 18 | 185.70 | 7.00  | 1.70 | 3.6  |
| 20 | 185.50 | 14.00 | 1.70 | 7.6  |
| 30 | 185.40 | 29.00 | 1.70 | 15.7 |
| 40 | 185.30 | 52.00 | 1.70 | 31.5 |
| 50 | 185.20 | 82.00 | 1.70 | 12.2 |
| 60 | 185.10 | 45.00 | 1.70 | 24.5 |

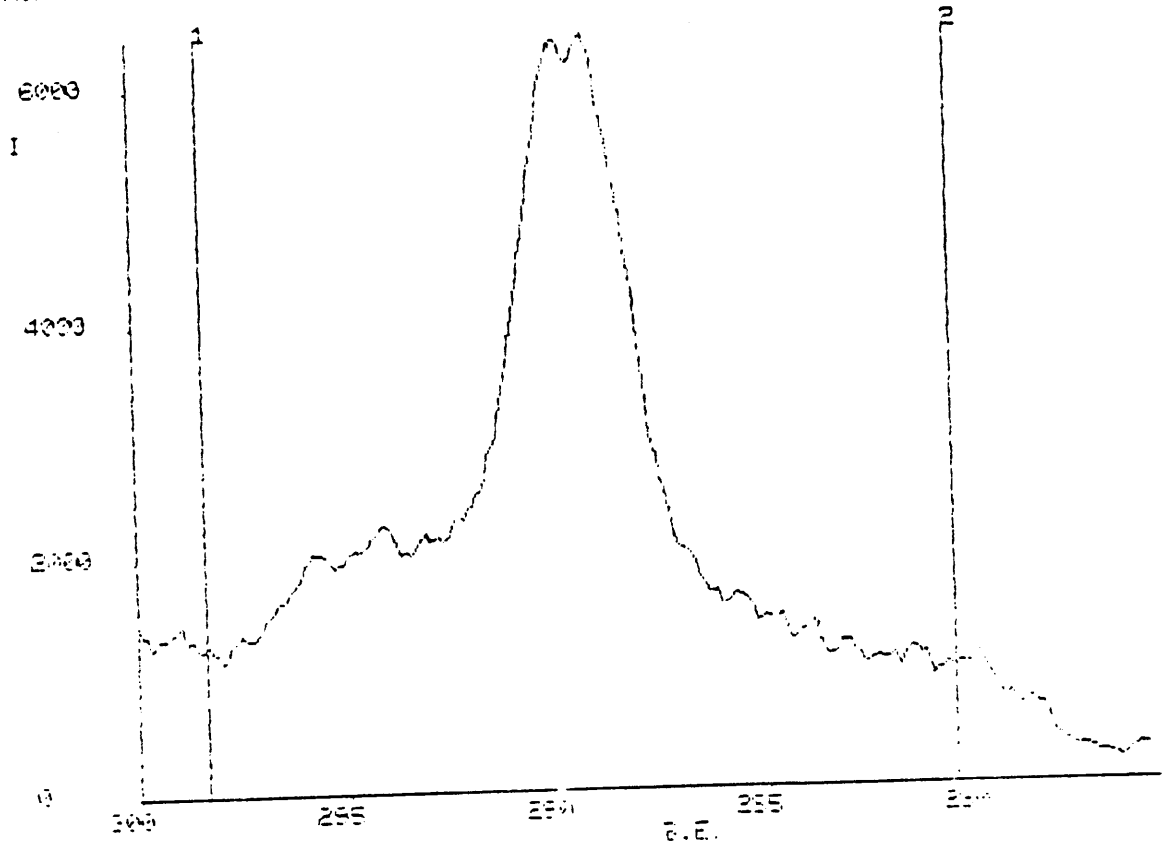


AGE 57  
END

RDN: 1150 29-APR-84 REG: 1.001 ST: 1.00 SO: 1250 #30 1 UNELL: 0.10



NAME: ST 018  
END  
RUN: Y1140 / 29-APR-84 REG: 3.001 ST: 0.10 #0: 250 #SW: 3 DWELL: 0.200



RUN : Y140 / 29-APR-84  
 SAMPLE: 1  
 NAME : 6T  
 TITLE : IS 100N  
 15KV 20MA SLIT=2 CUR=0.0

EXCITATION : MG ANALYSER MODE: TRR  
 SCAN MODE : T FACTOR : 0

MAGNIFICATION: L RESOLUTION: M

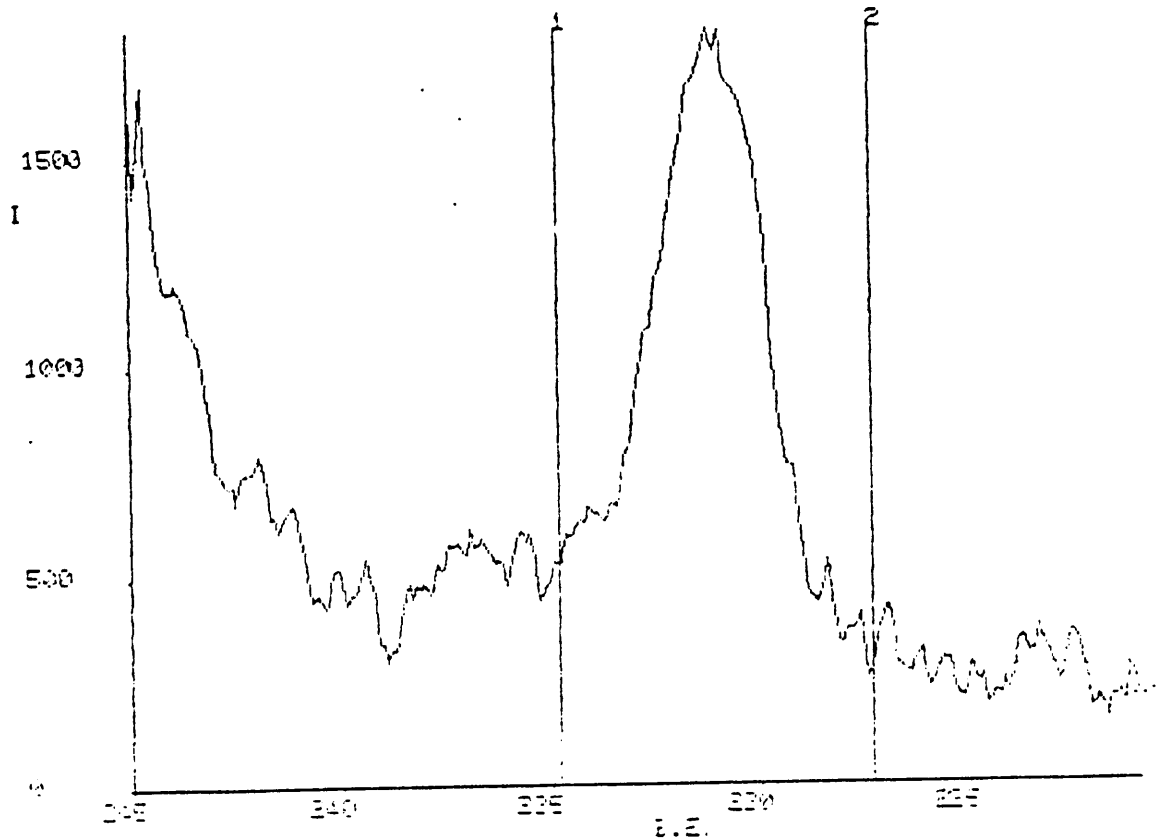
BASELINE THRESHOLD: 0 MIN. PEAK WIDTH: 1.00

REGION: 3 - WITH MARKERS  
 ELMT K.E. STEP DWELL #CHAN #AVG  
 X01 888.80 0.10 0.200 350 3

SCAN: 1

| PEAK | START E.V.<br>K.E. | END E.V.<br>E.E. | AREA | AREA   | FWHM |
|------|--------------------|------------------|------|--------|------|
| 1    | 884.20             | 885.20           | 8884 | 240844 | 3.10 |

\*\*\*1 6T 525  
END  
RUN: -Y140 / 29-APR-84 REG: 2.001 ST: 0.10 #C: 250 #SW: 4 DWELL: 0.200



RUN : Y140 / 23-APR-84  
 SAMPLE: 1  
 NAME : BT  
 TITLE : IR YOON  
 13KV 30MA SLIT=2 CUR=9.0

EXCITATION : MG ANALYSER MODE: FRR  
 SCAN MODE : T FACTOR : 0

MAGNIFICATION: L RESOLUTION: M

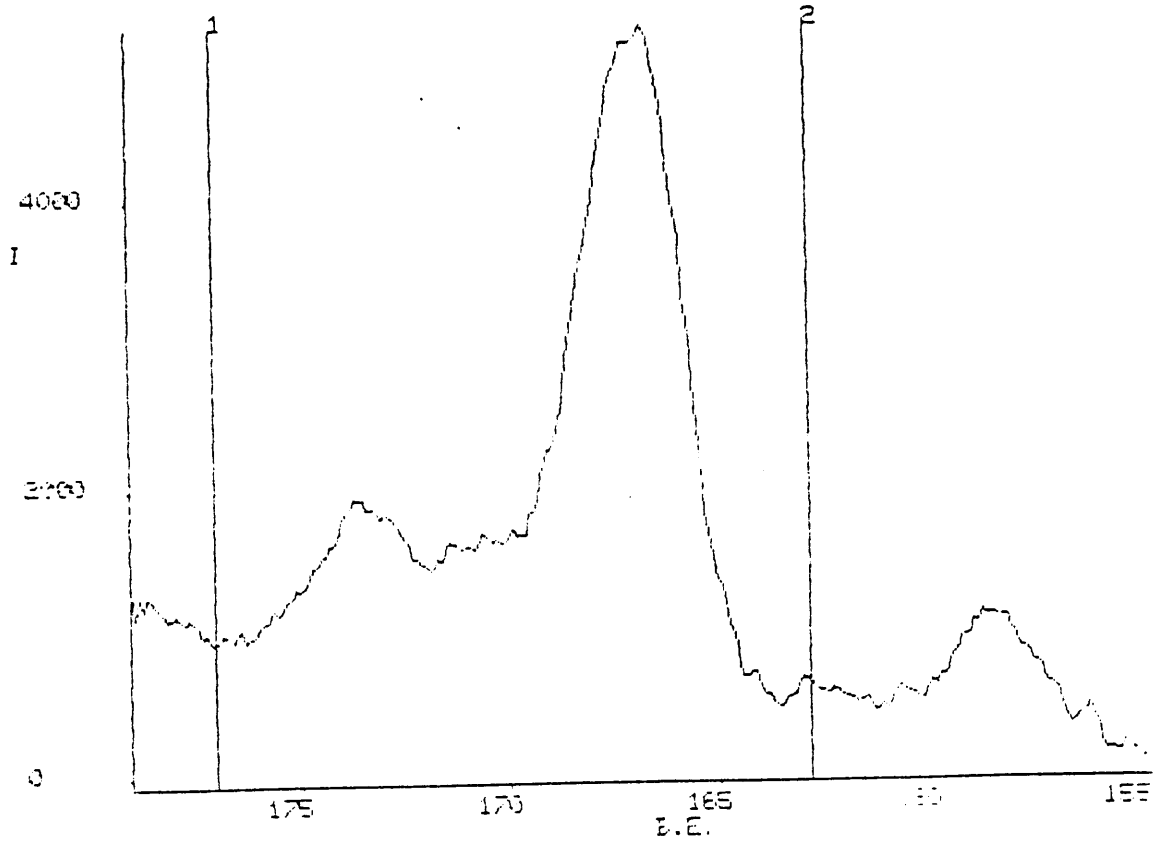
BASELINE THRESHOLD: 0 MIN. PEAK WIDTH: 1.00

REGION: 2 - WITH MARKERS  
 ELMT K.E. STEP DWELL #CHAN #AVG  
 XSE3 1033.60 0.10 0.200 250 4

SCAN: 1

| PEAK | START E.V.<br>K.E. | END E.V.<br>E.E. | MAX  | AREA  | FLHH |
|------|--------------------|------------------|------|-------|------|
| 1    | 1032.60            | 231.10           | 1374 | 42332 | 2.30 |

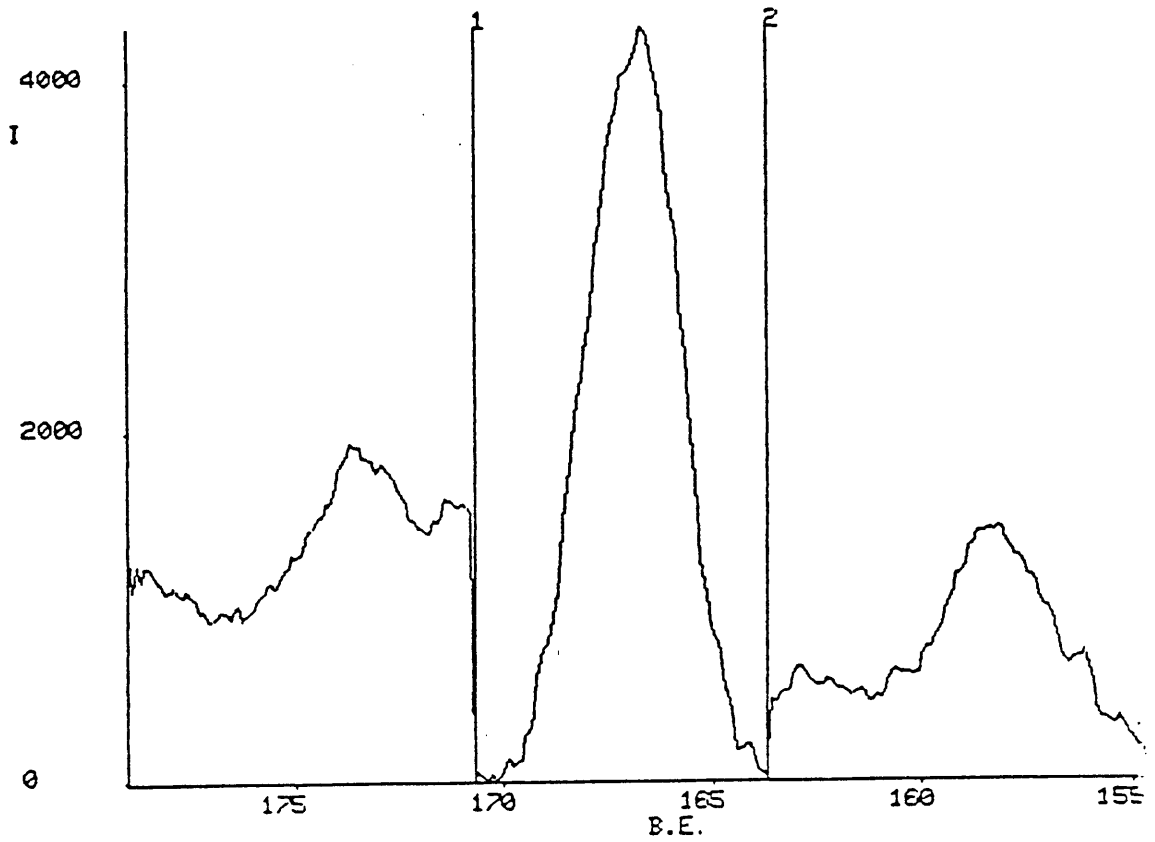
\*SU 6T S2P  
END  
RUN: -Y140 / 29-APR-84 REG: 1.001 ST: 0.10 #C: 250 #SW: 5 DWELL: 0.200



\*RE 6T S2P

END

RUN:-Y140 / 29-APR-84 REG: 1.001 ST:0.10 #C: 250 #SW: 5 DWELL: 0.200



RUN : Y140 / 23-APR-84  
 SAMPLE: 1  
 NAME : 6T  
 TITLE : DR YOON  
 13KV 20MA SLIT=2 CUR=9.0

EXCITATION : MG ANALYSER MODE: FRR  
 SCAN MODE : T FACTOR : 0

MAGNIFICATION: L RESOLUTION: M

BASELINE THRESHOLD: 0 MIN. PEAK WIDTH: 1.00

REGION: 1 - WITH MARKERS  
 ELMT K.E. STEP DWELL #CHAN #AVG  
 X3 1038.60 0.10 0.200 250 5

SCAN: 1

| PEAK | START E.V.<br>K.E. | END E.V.<br>E.E. | MAX  | AREA   | FWHM |
|------|--------------------|------------------|------|--------|------|
| 1    | 1038.90            | 163.70           | 4525 | 182431 | 2.20 |

RUN REG SCAN STEP #CH  
Y140 1 1 .10 71

ELMT K.E. DWELL  
XS 1089.60 .200

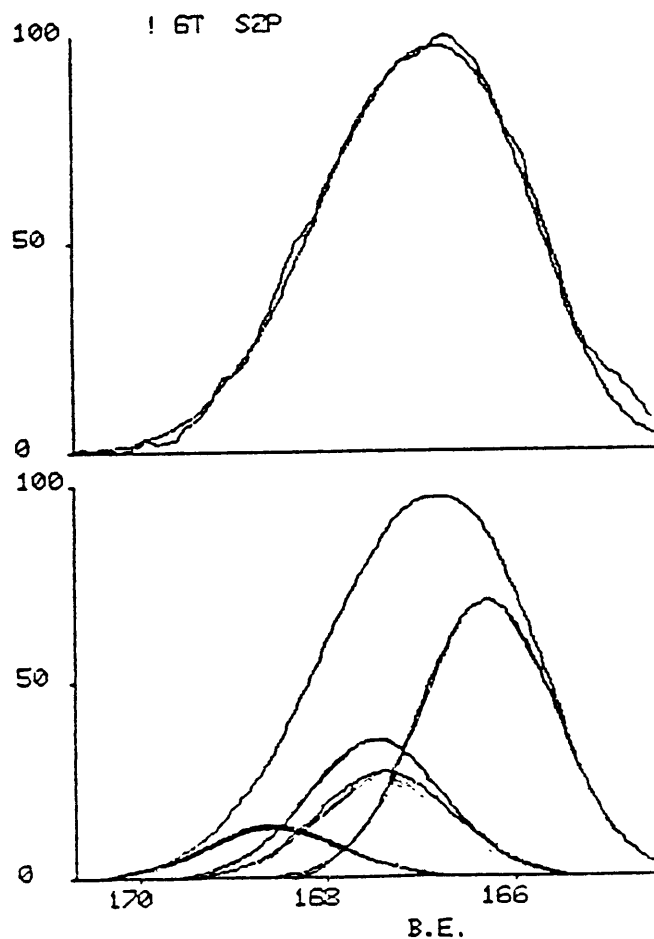
START EV: 170.70  
END EV: 163.70

100% AREA =118503

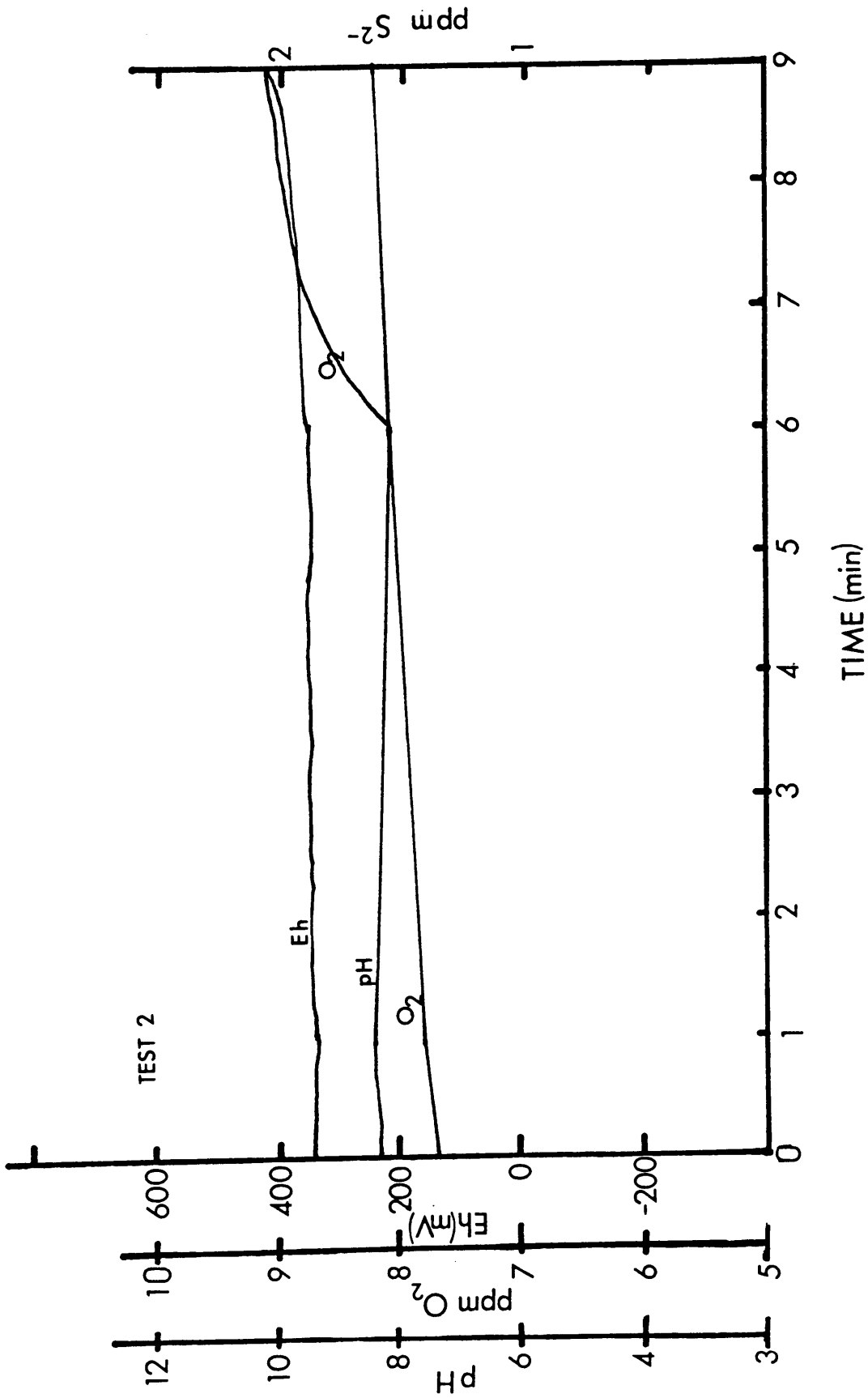
100% INTENSITY =4306

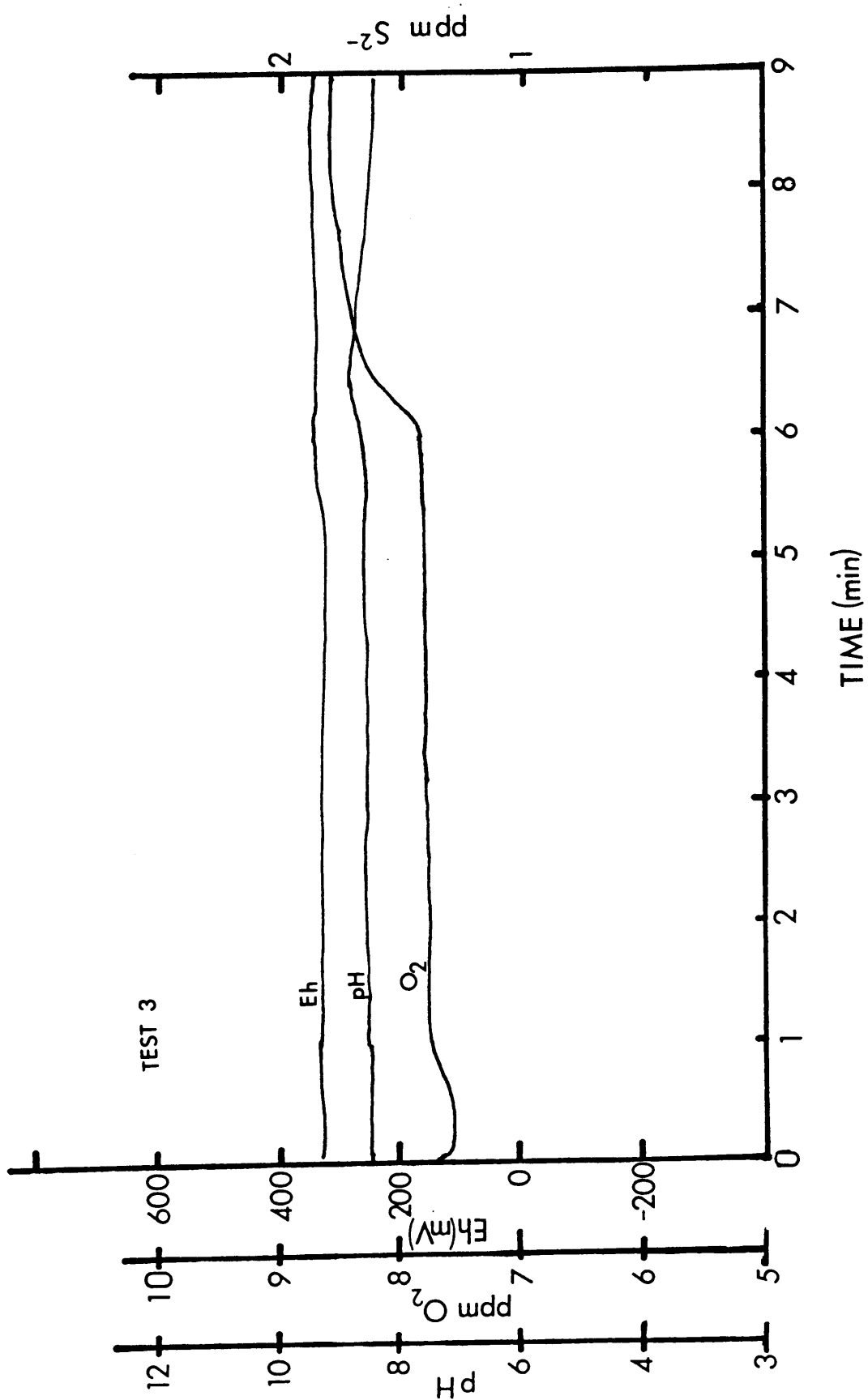
FIT=L7

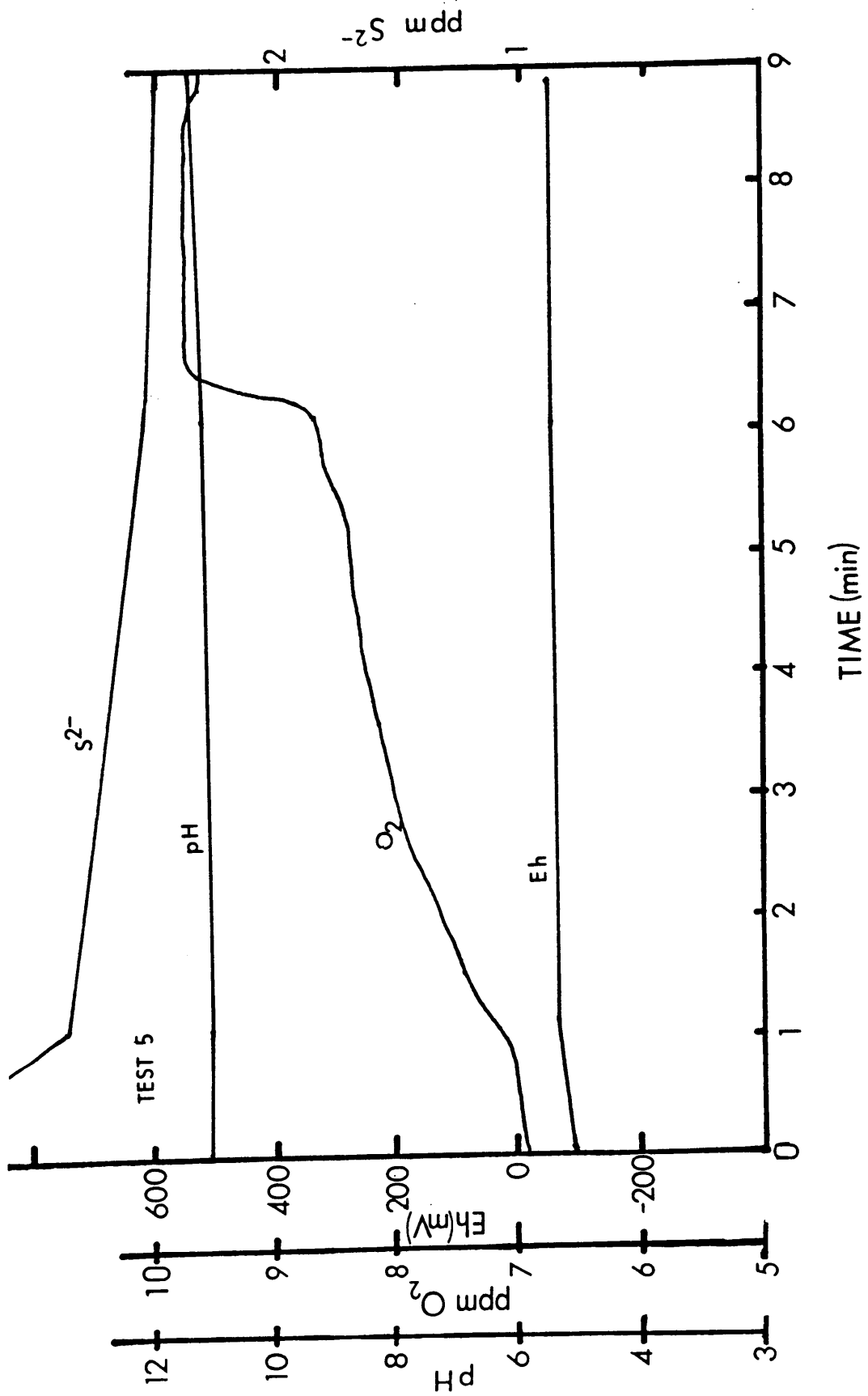
| PK | ENERGY | MAX   | FWHM | AREA |
|----|--------|-------|------|------|
| 1G | 168.60 | 13.50 | 1.70 | 8.8  |
| 2G | 167.40 | 27.00 | 1.70 | 17.8 |
| 3G | 167.50 | 35.00 | 1.70 | 23.0 |
| 4G | 166.30 | 70.00 | 1.70 | 46.1 |



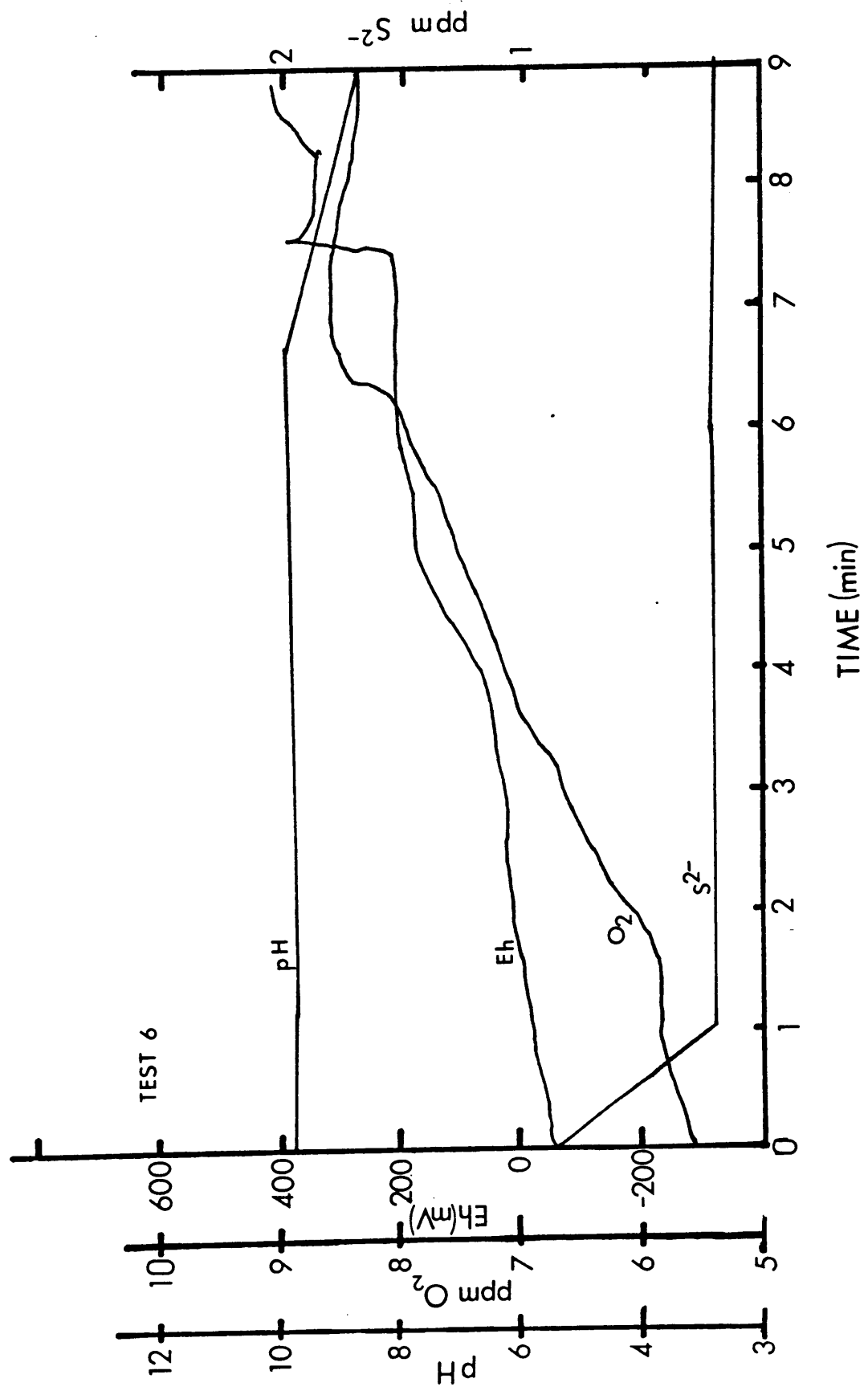
APPENDIX D: RECORDS OF OPERATING ELECTRODES IN  
BATCH FLOTATION

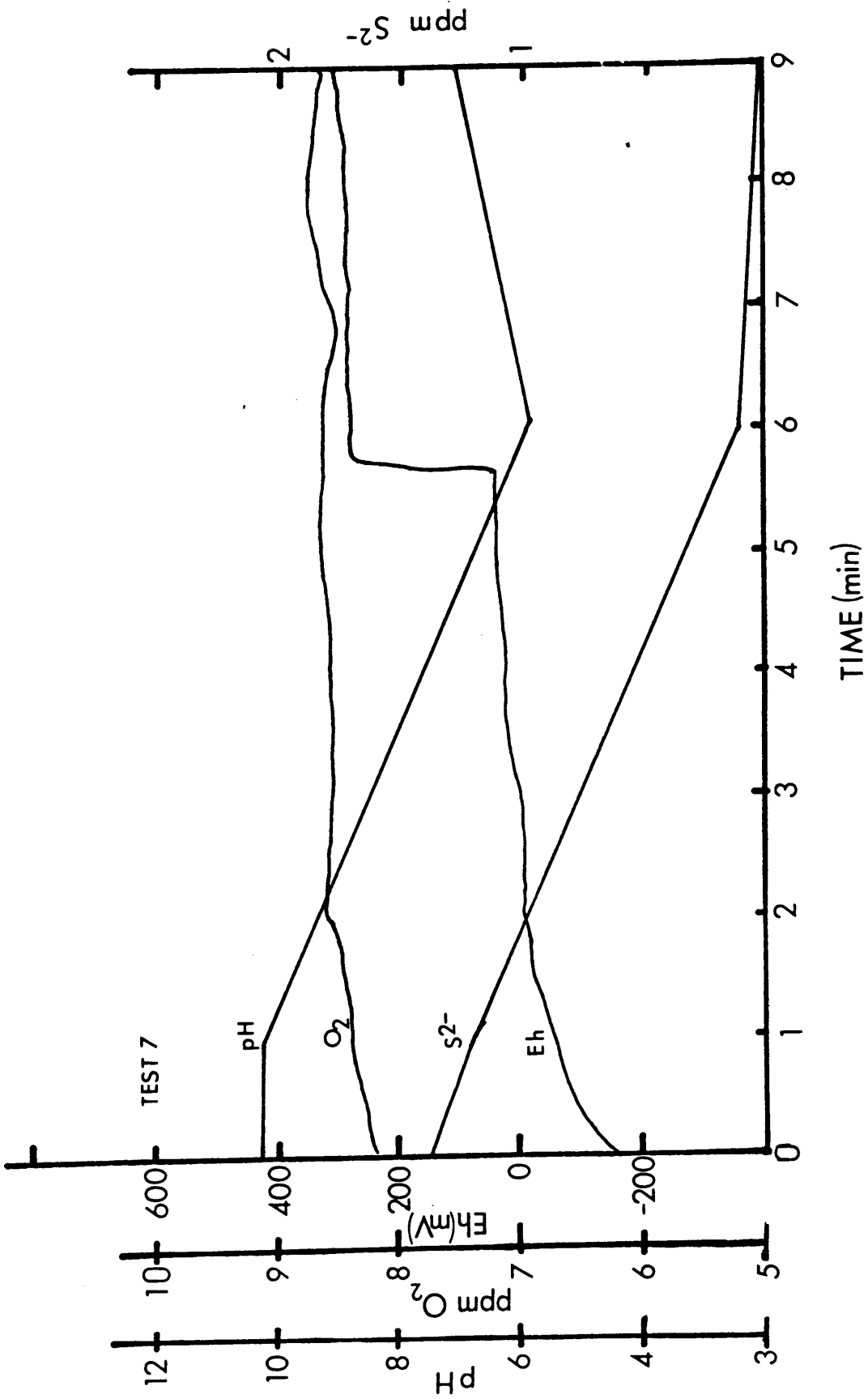


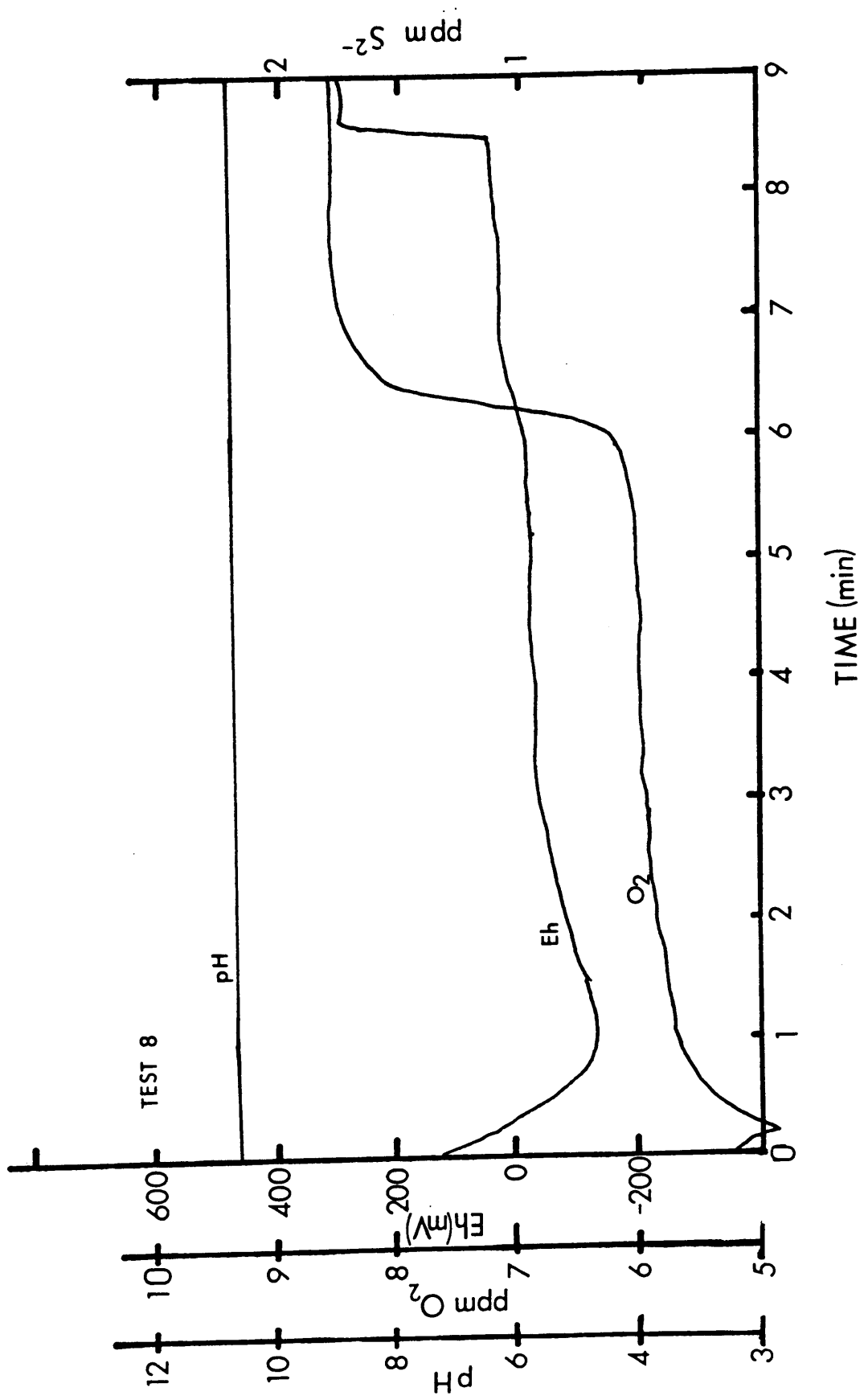


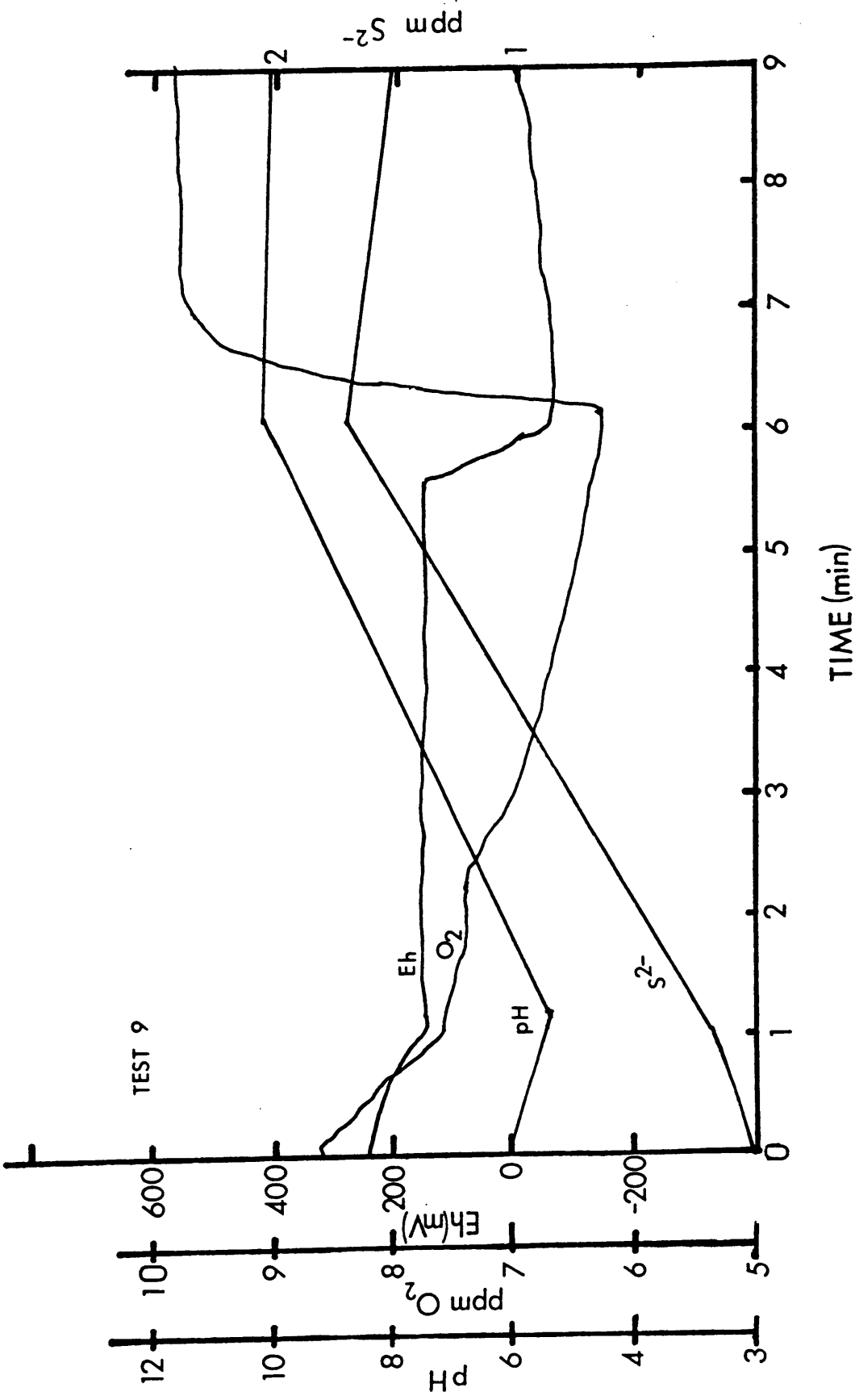


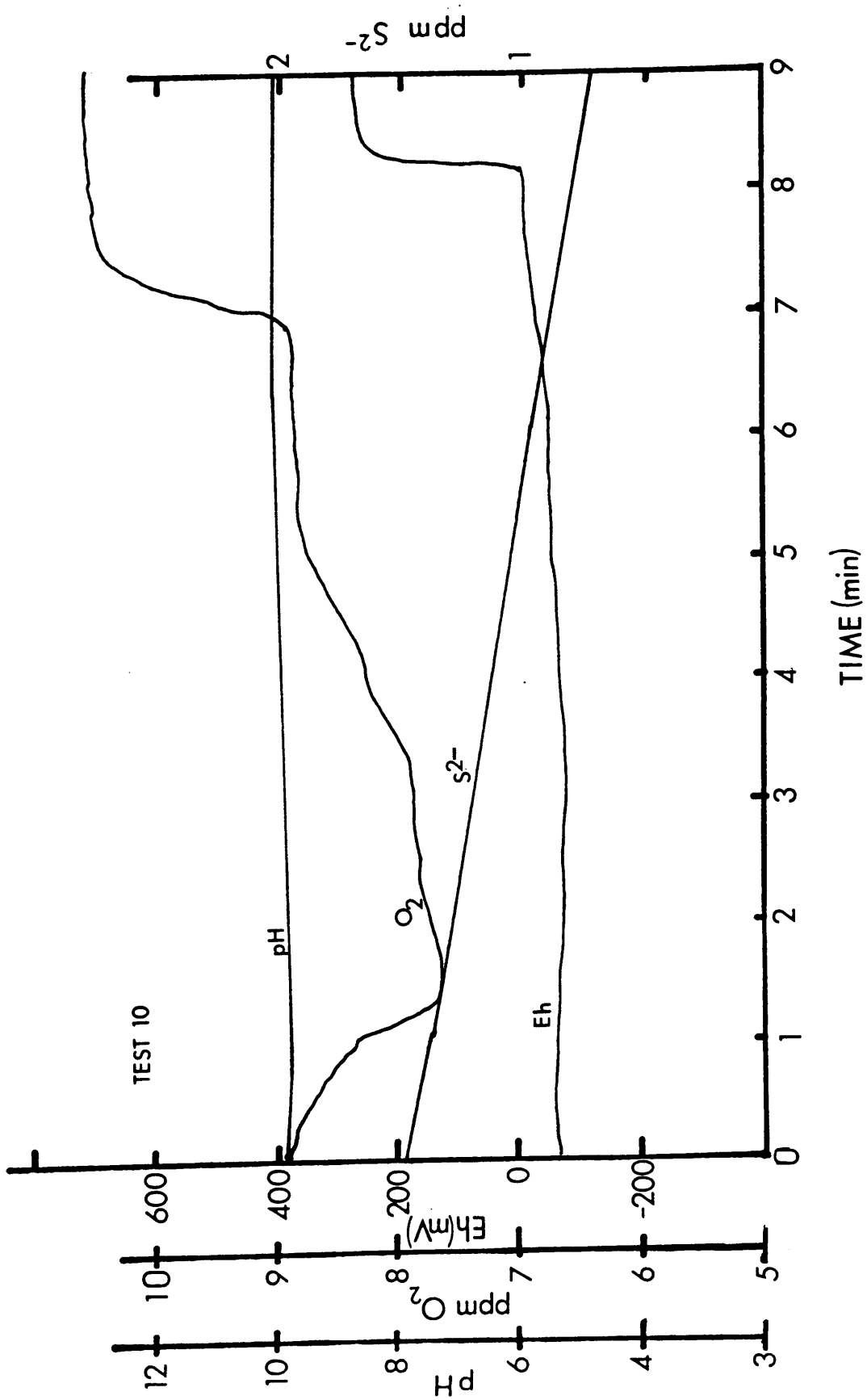
S<sup>2-</sup> ppm

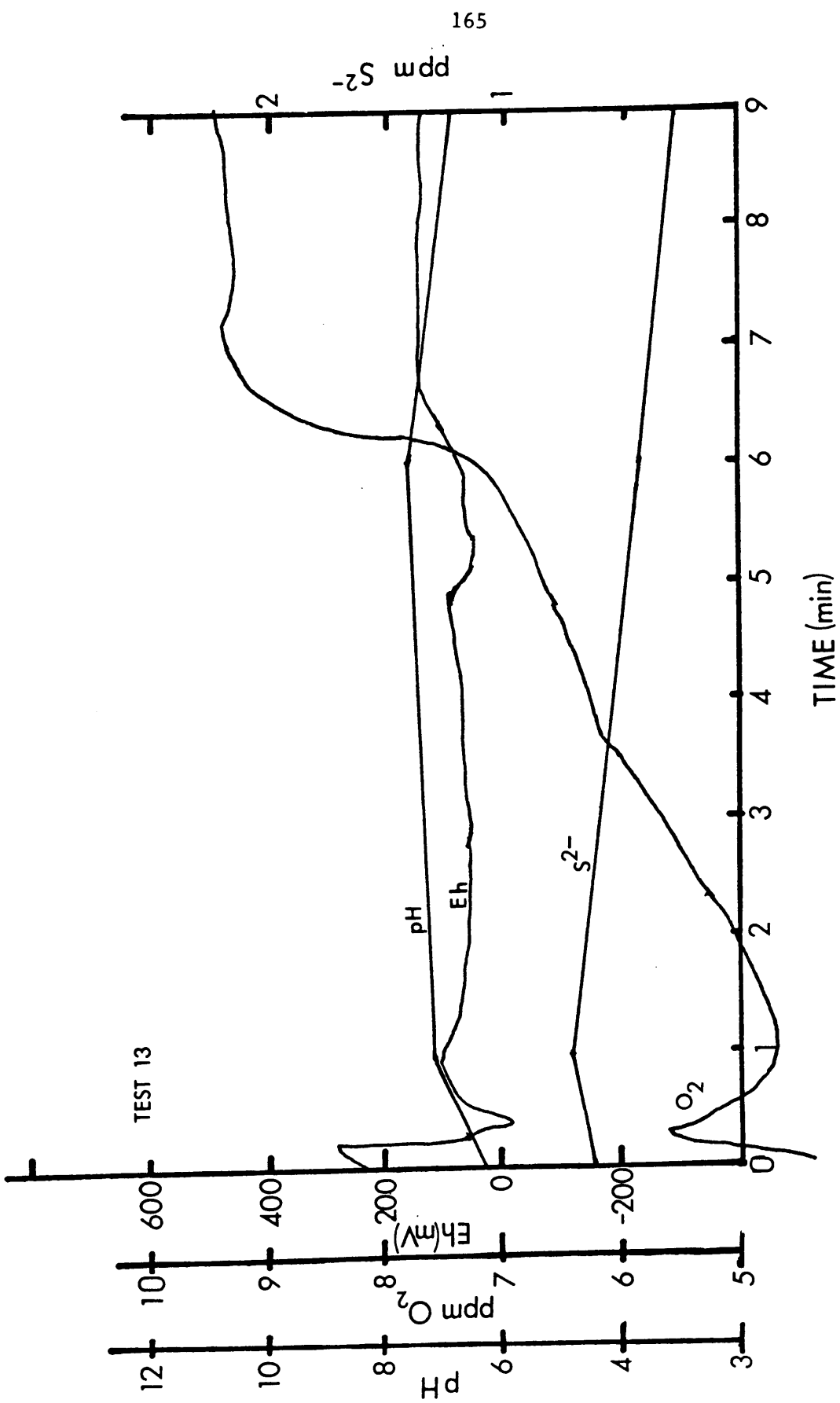


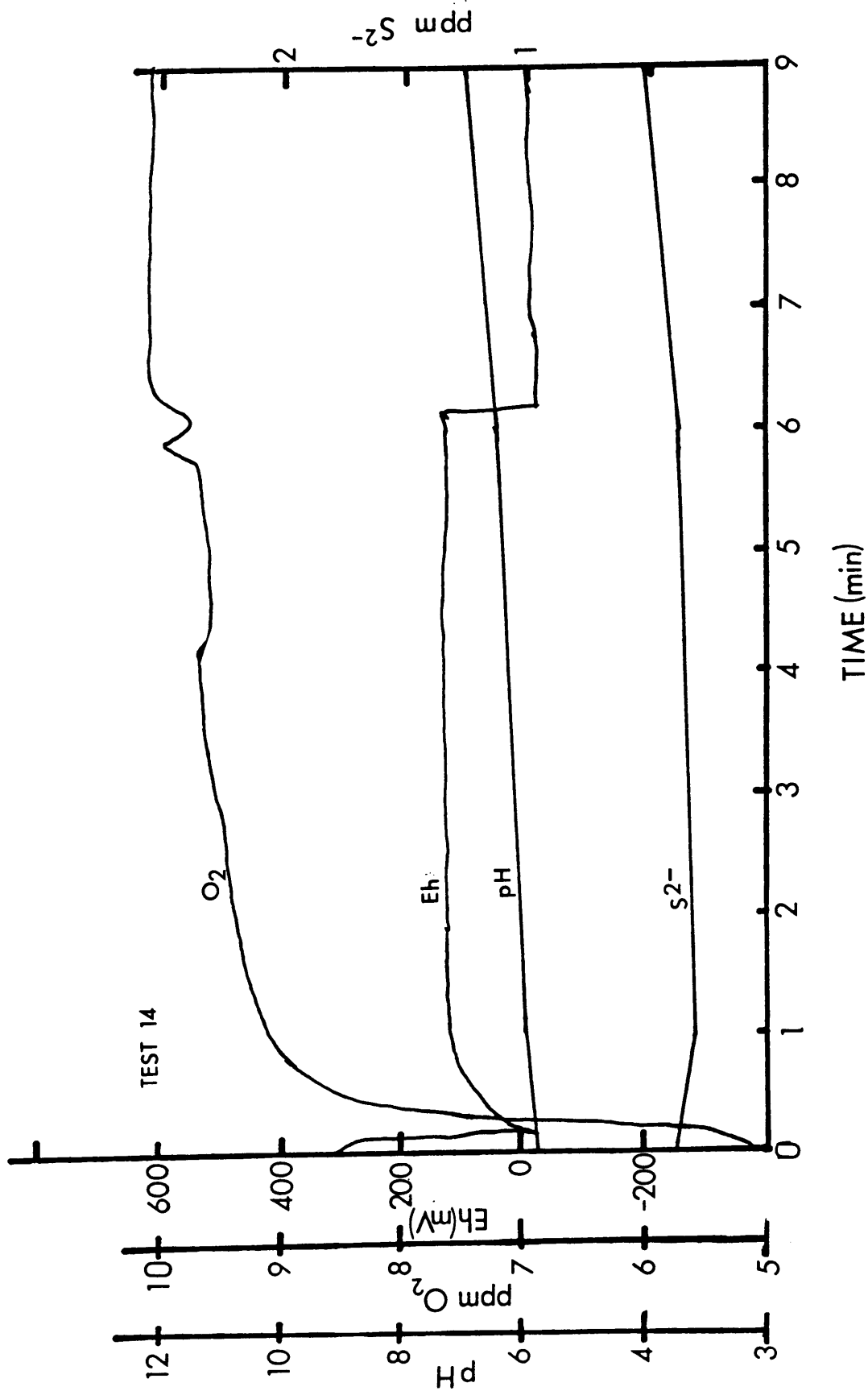


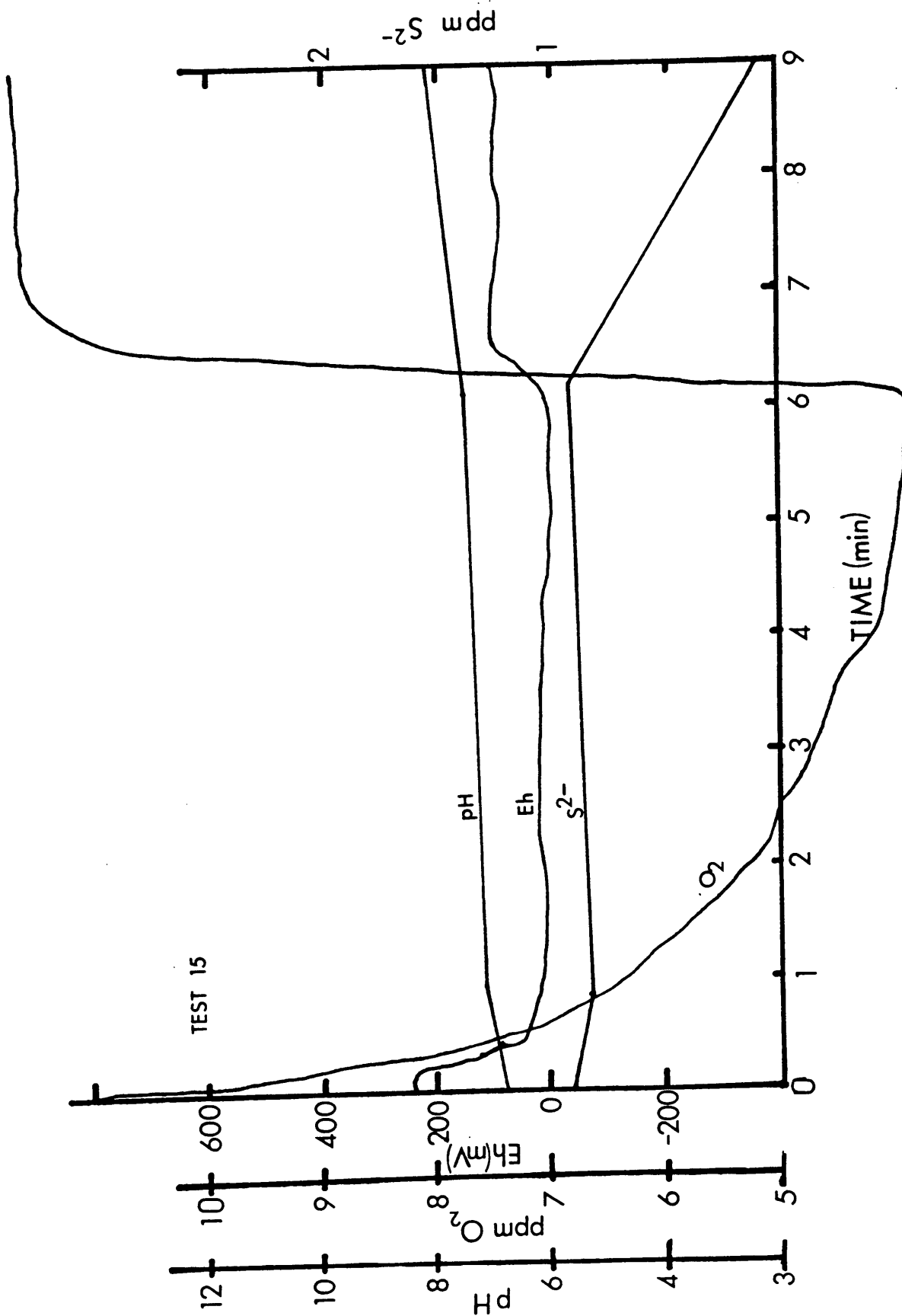


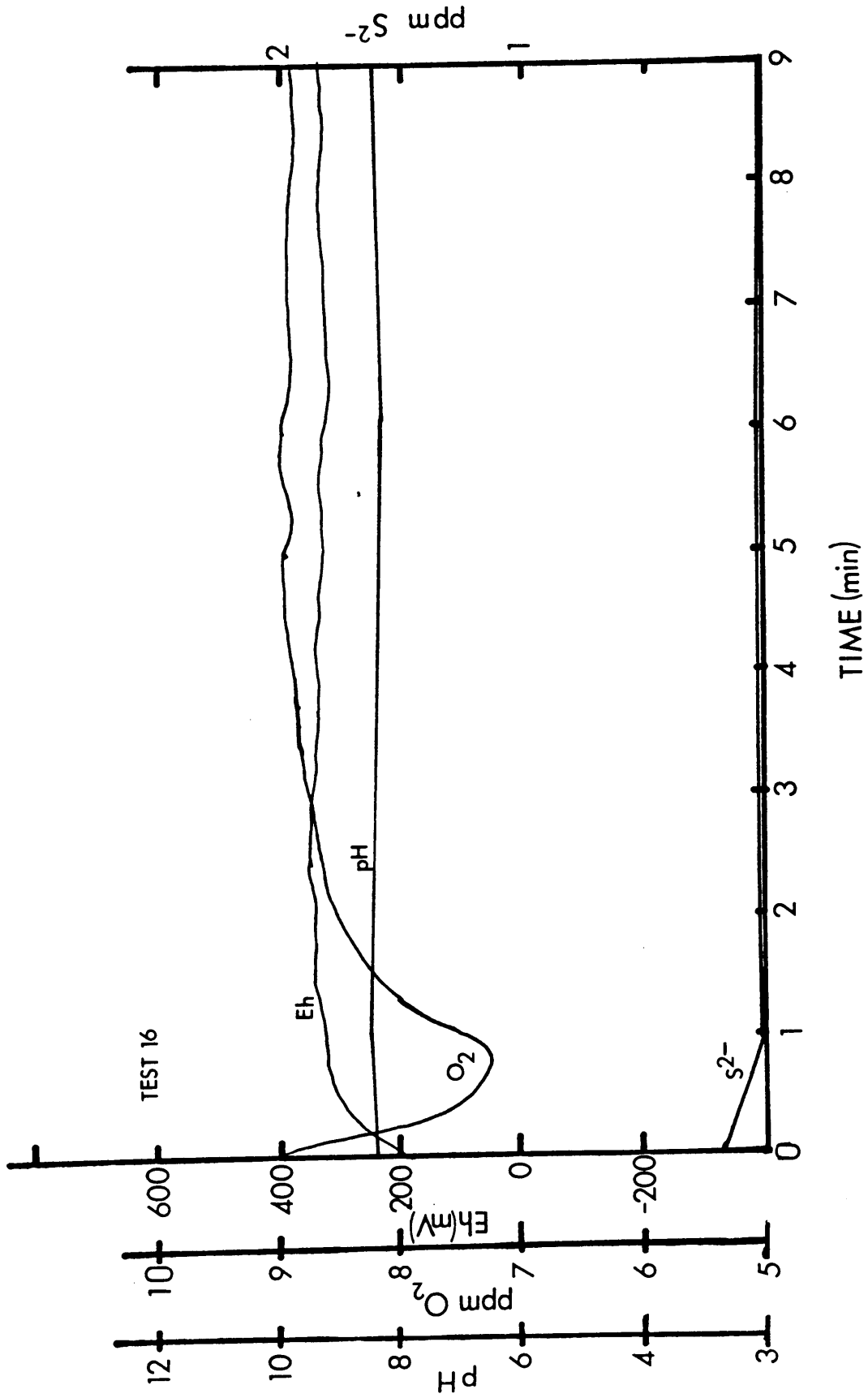


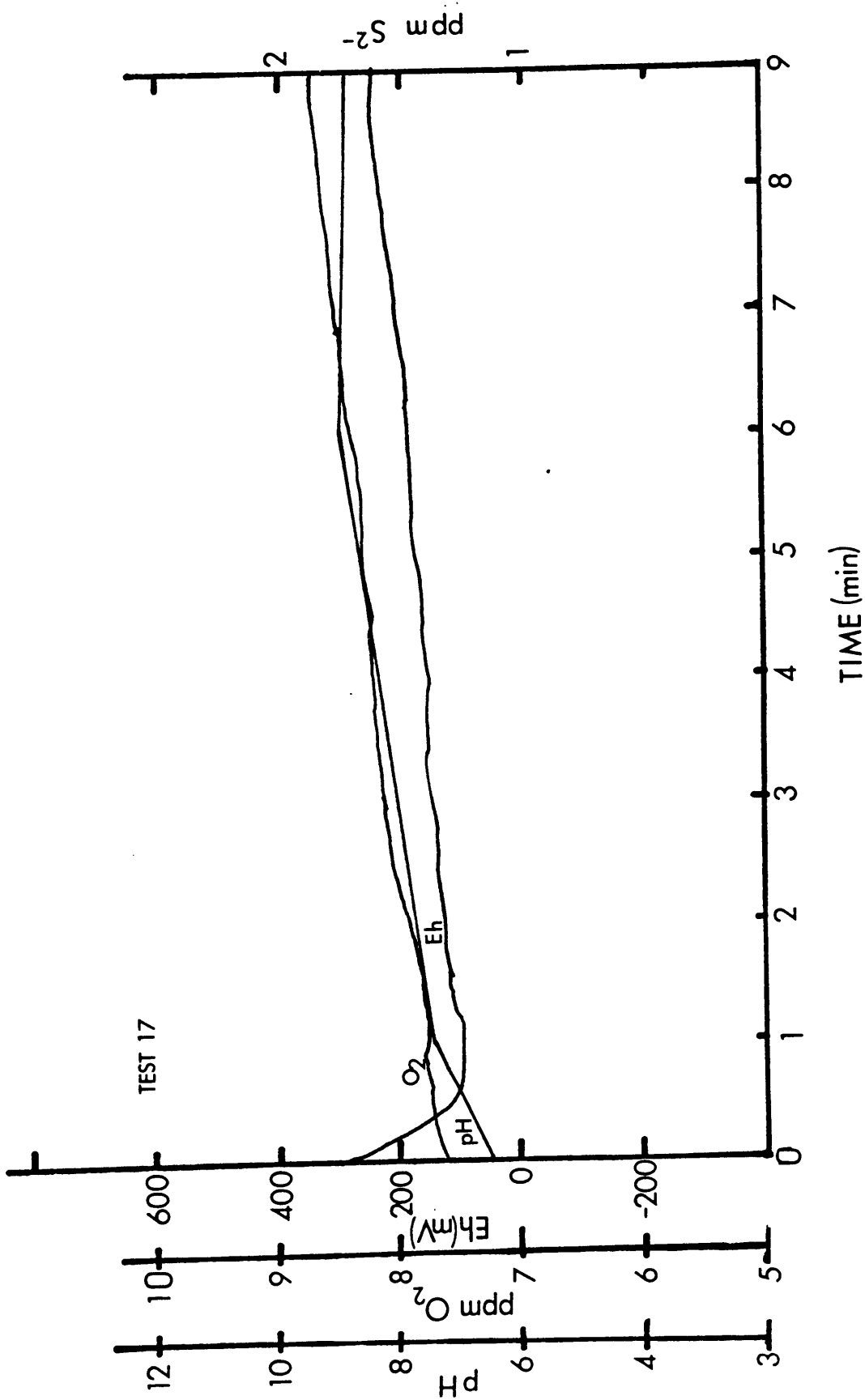


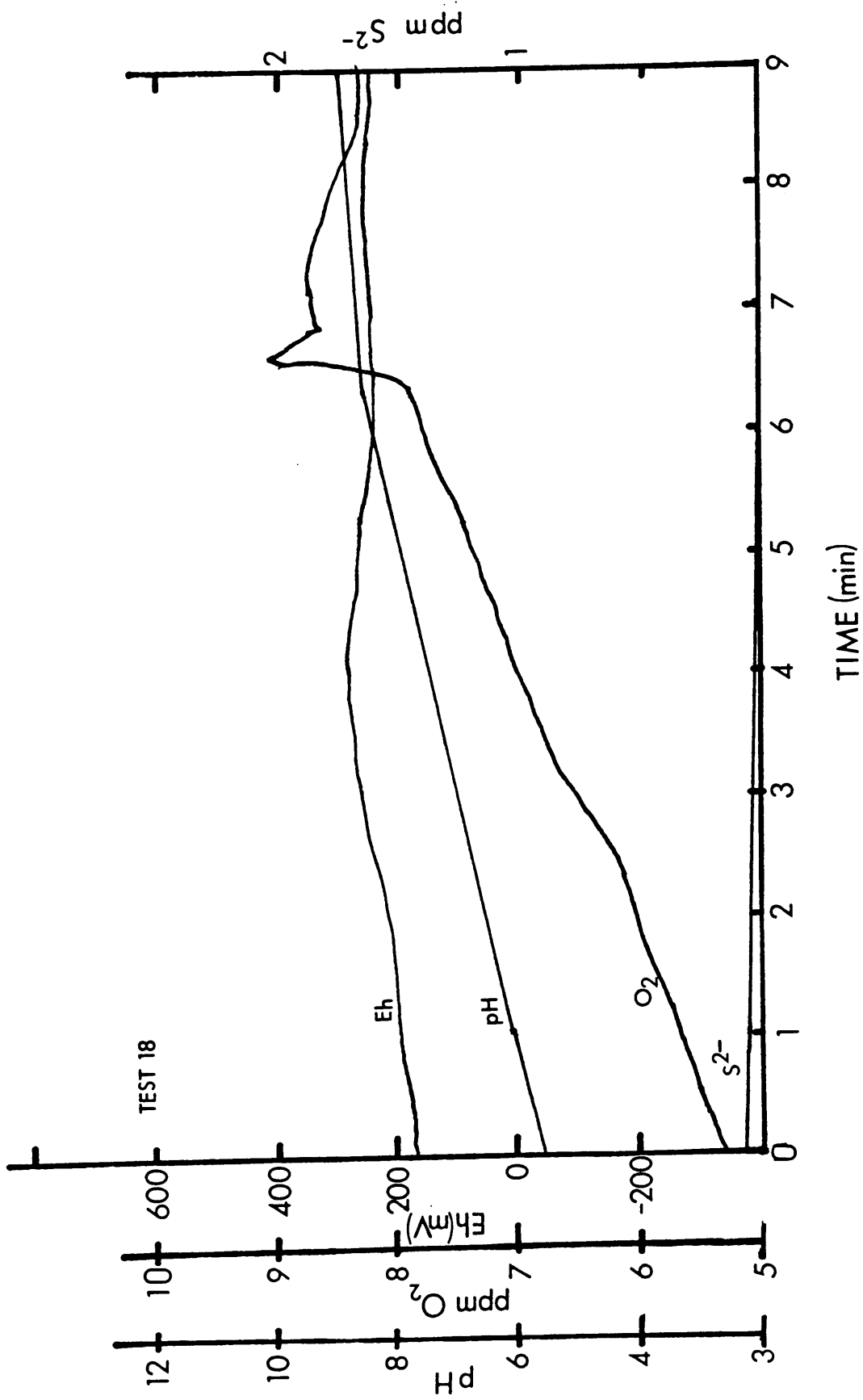


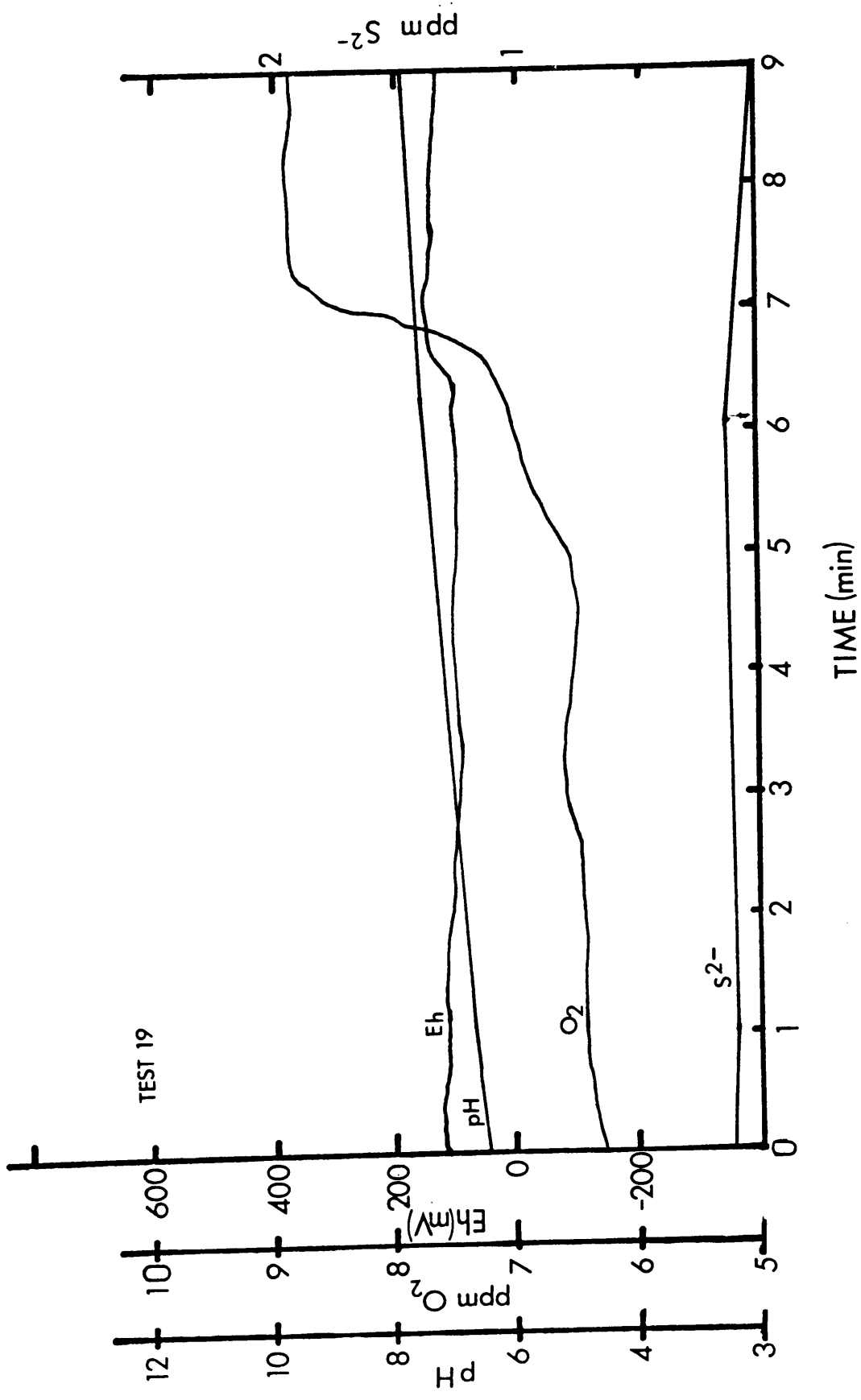


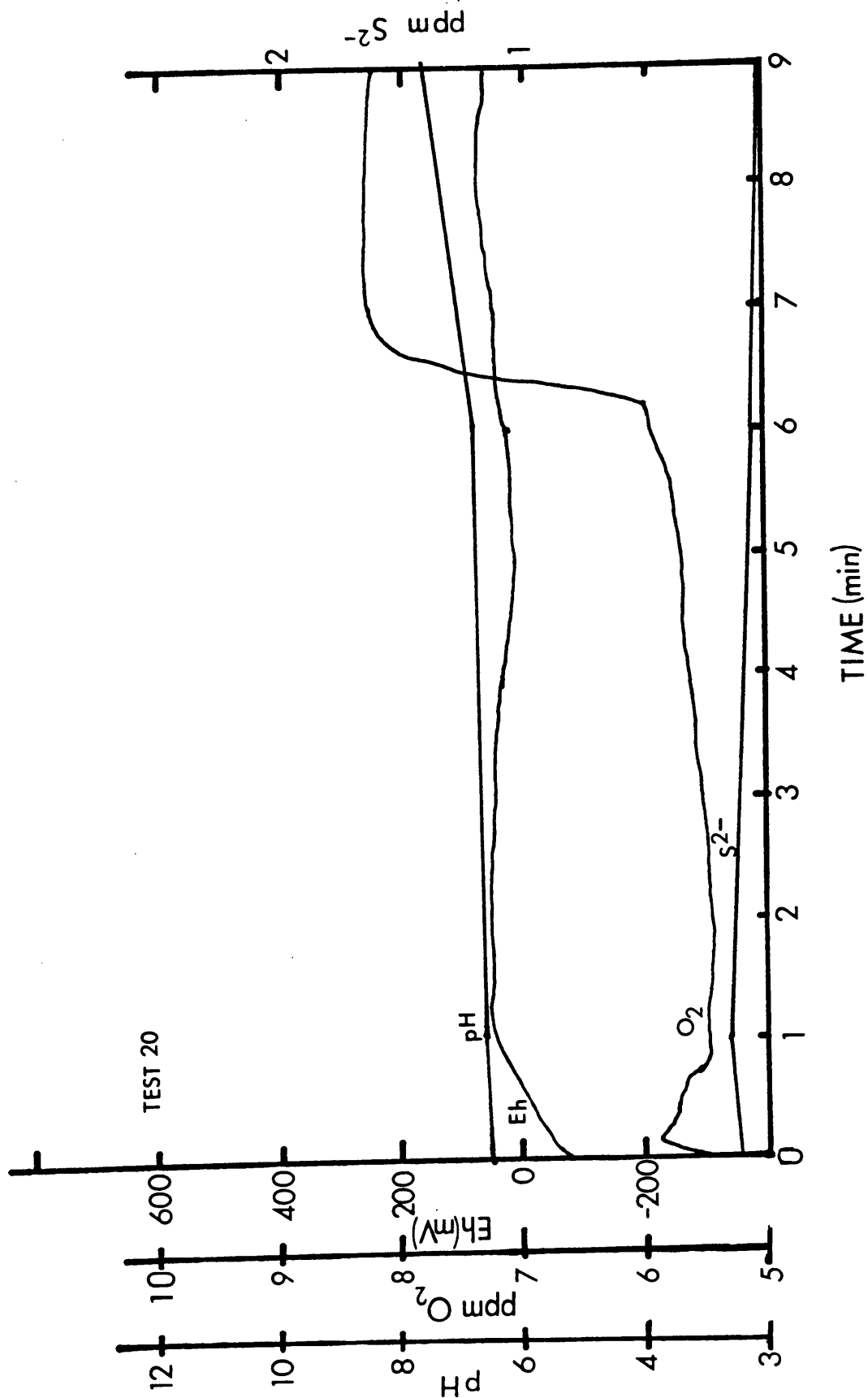


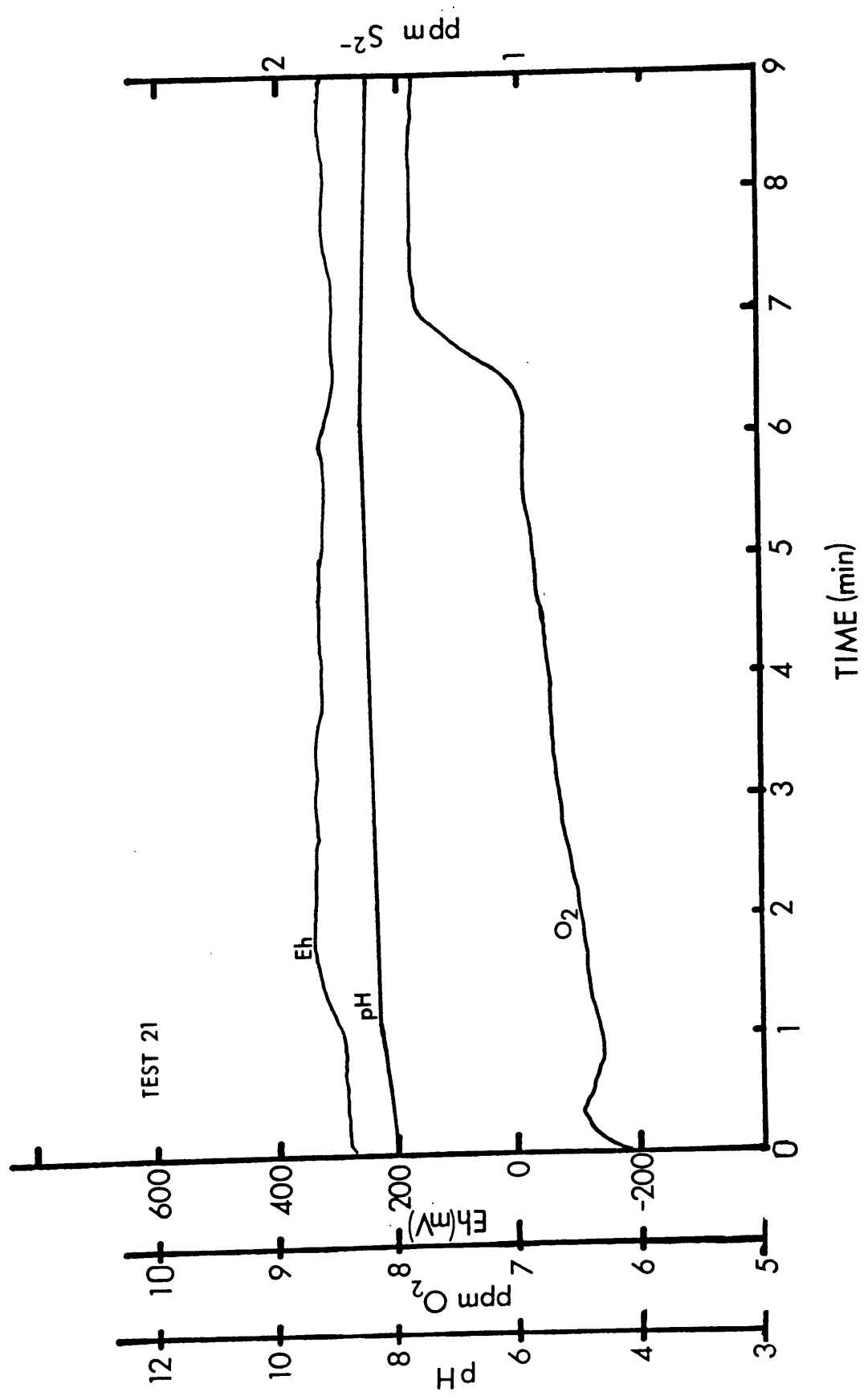


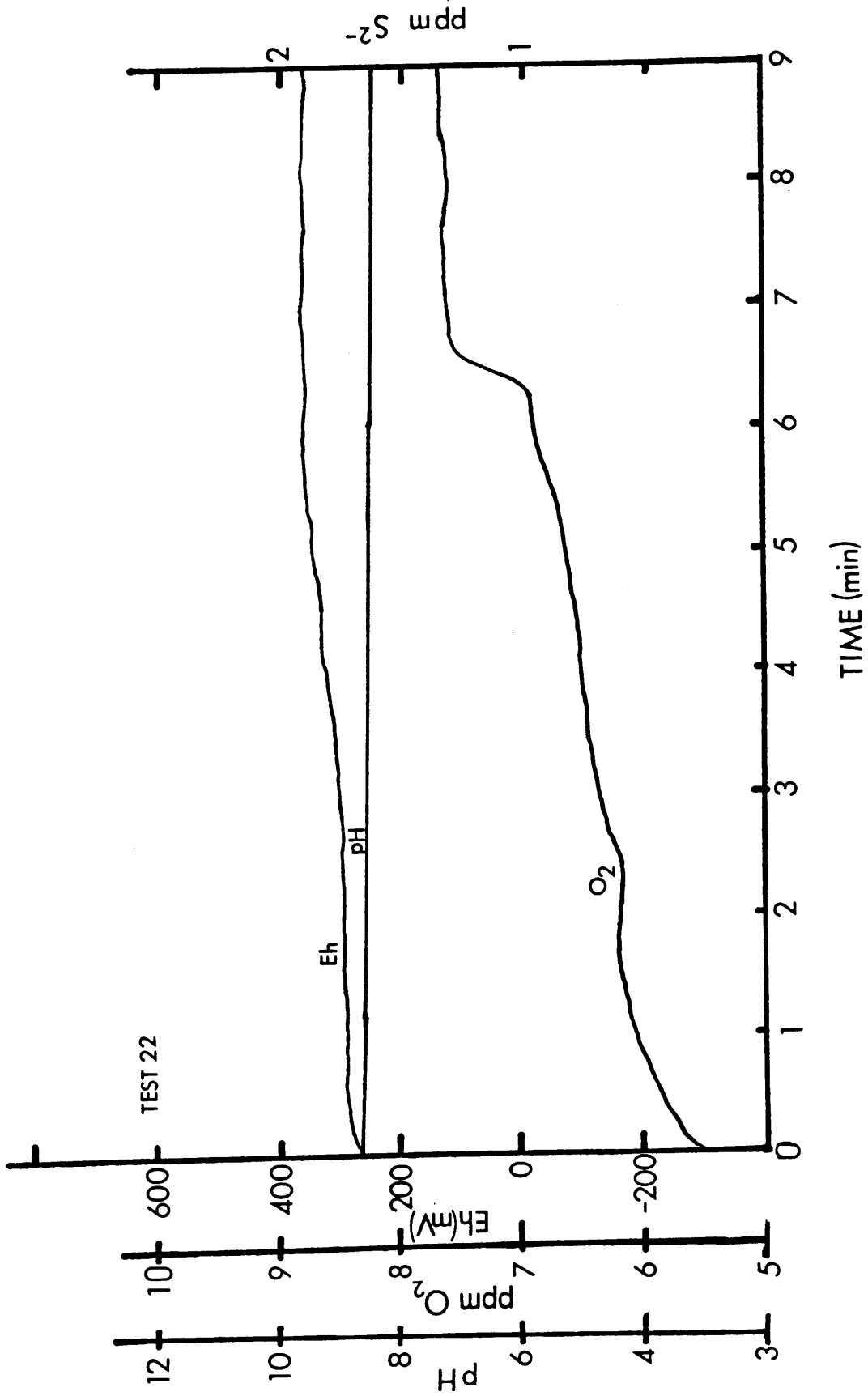


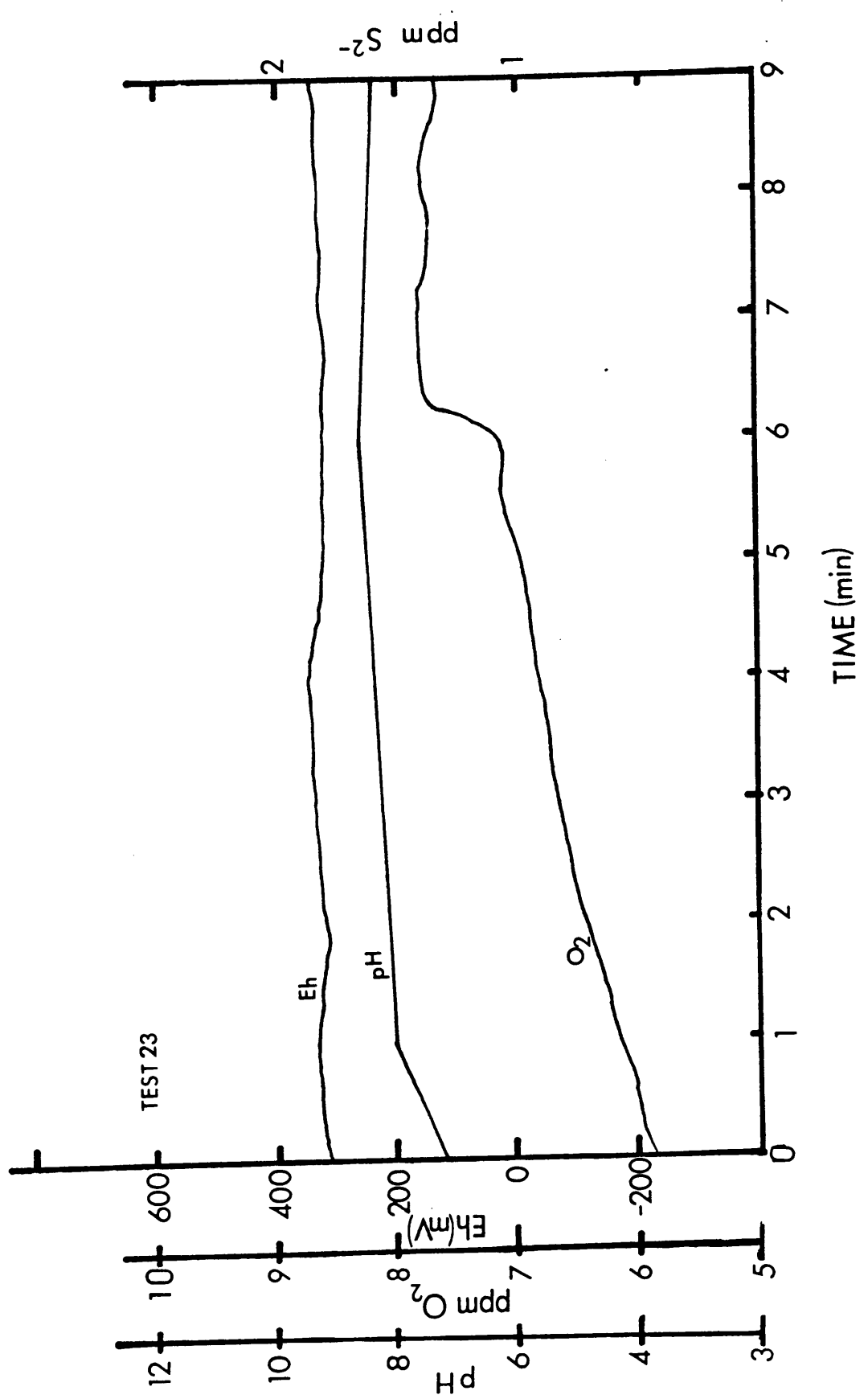


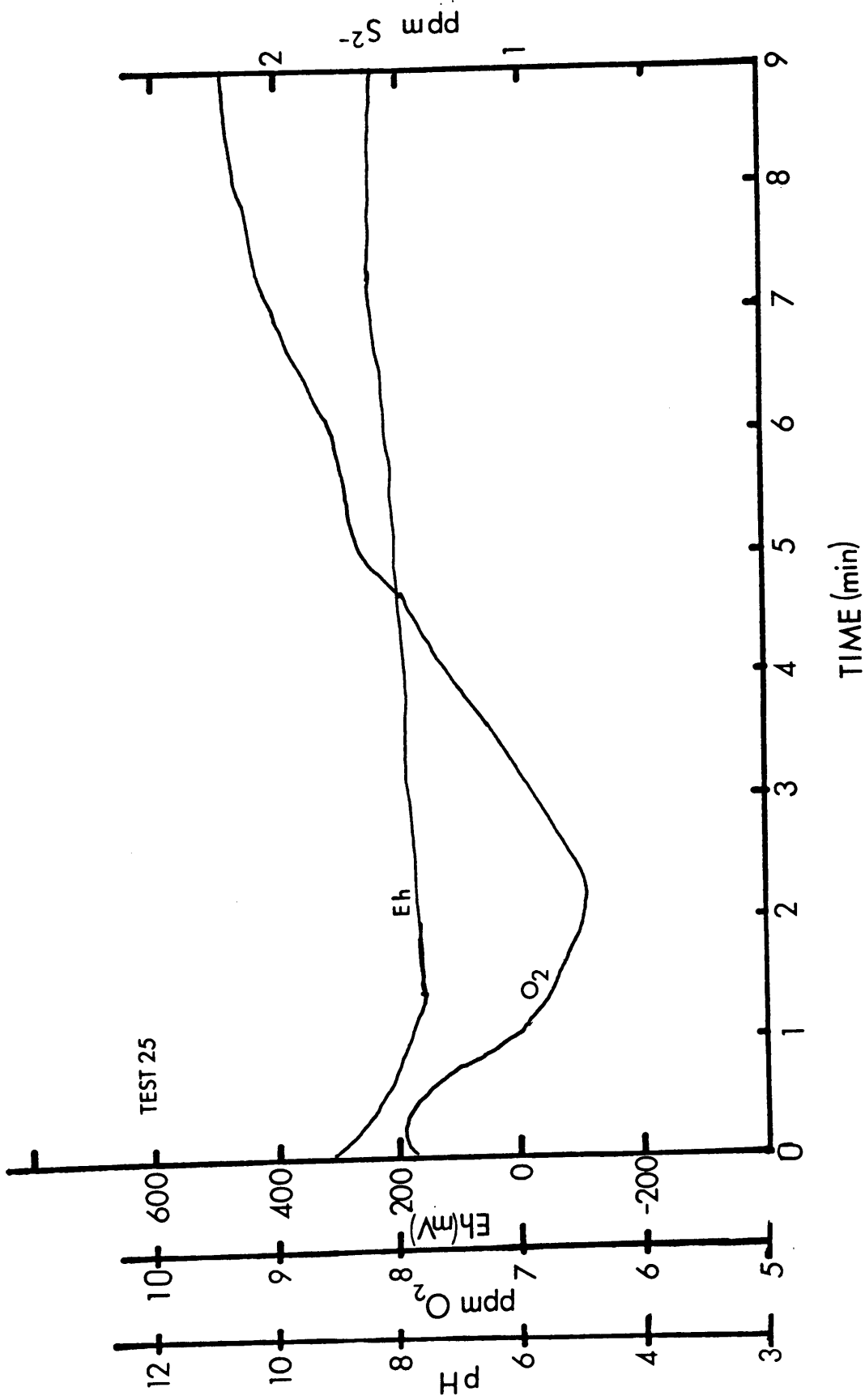


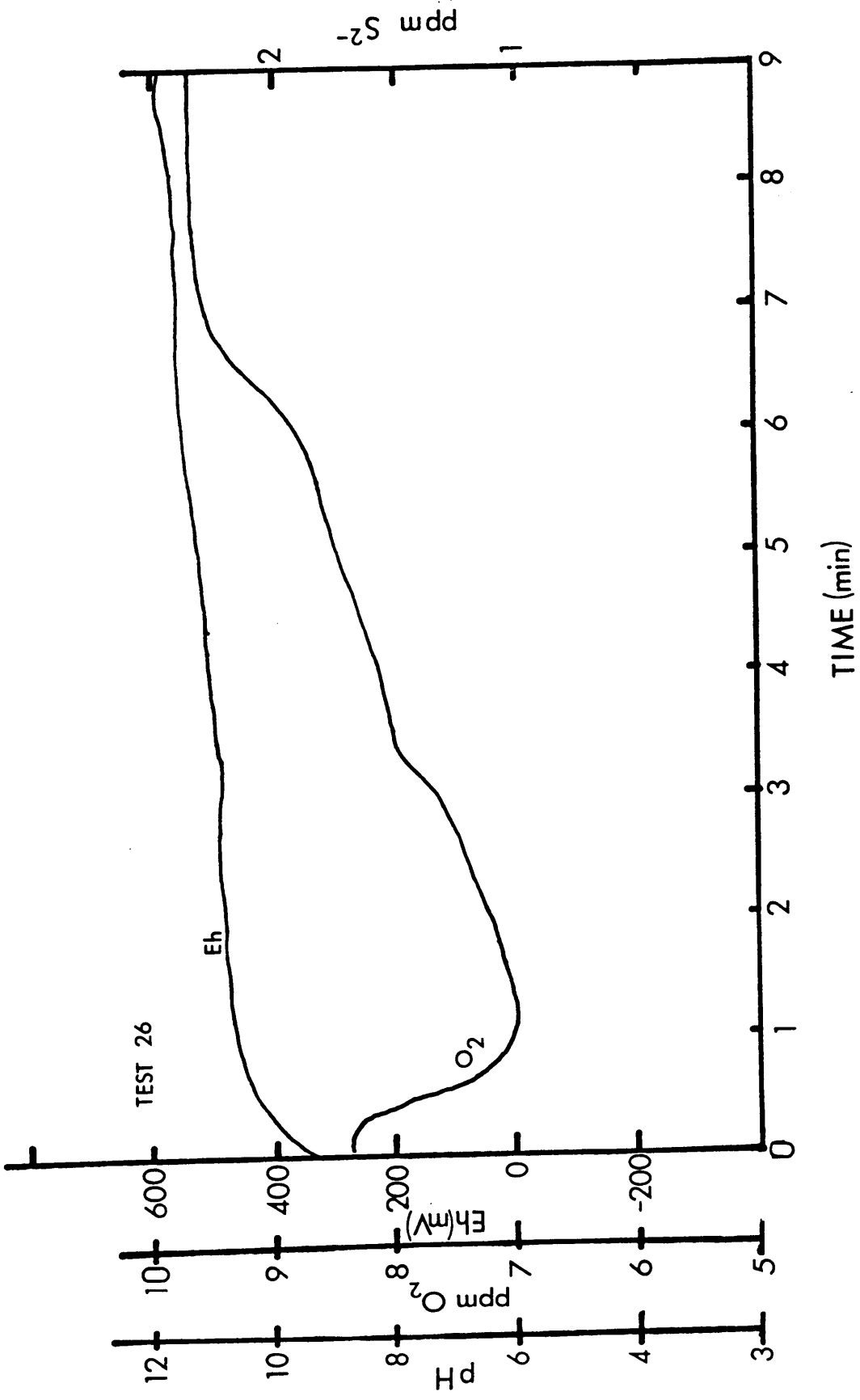


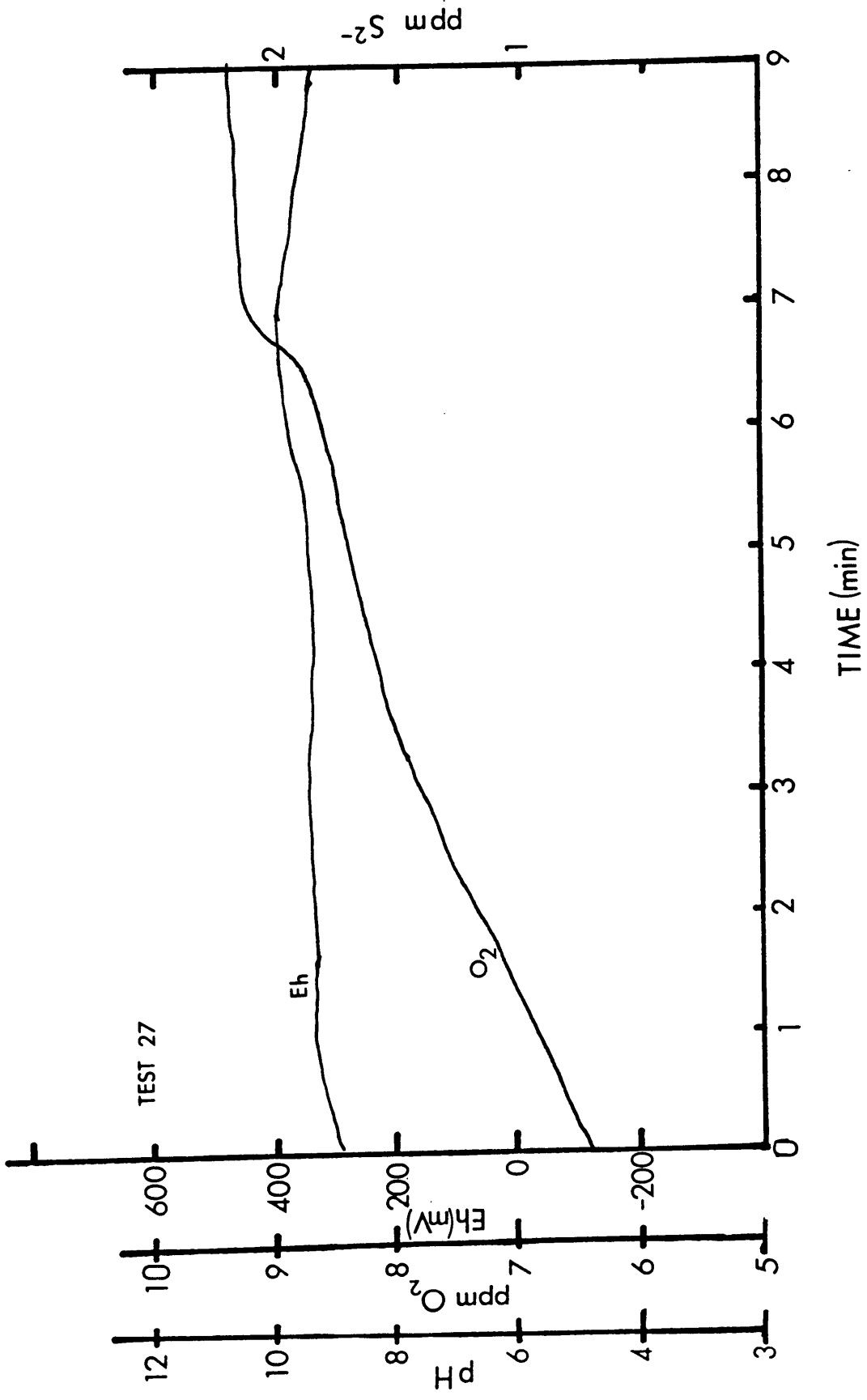


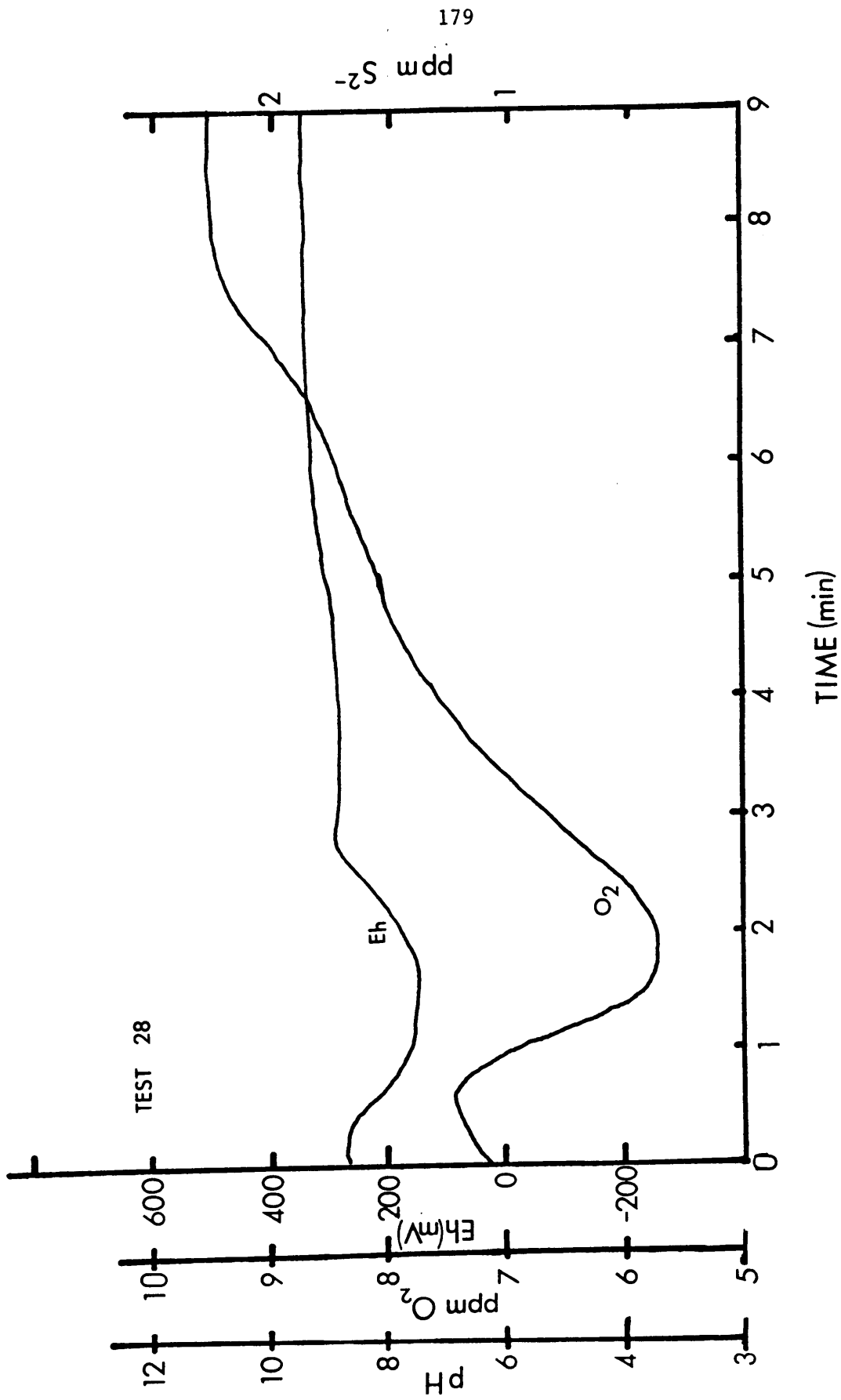


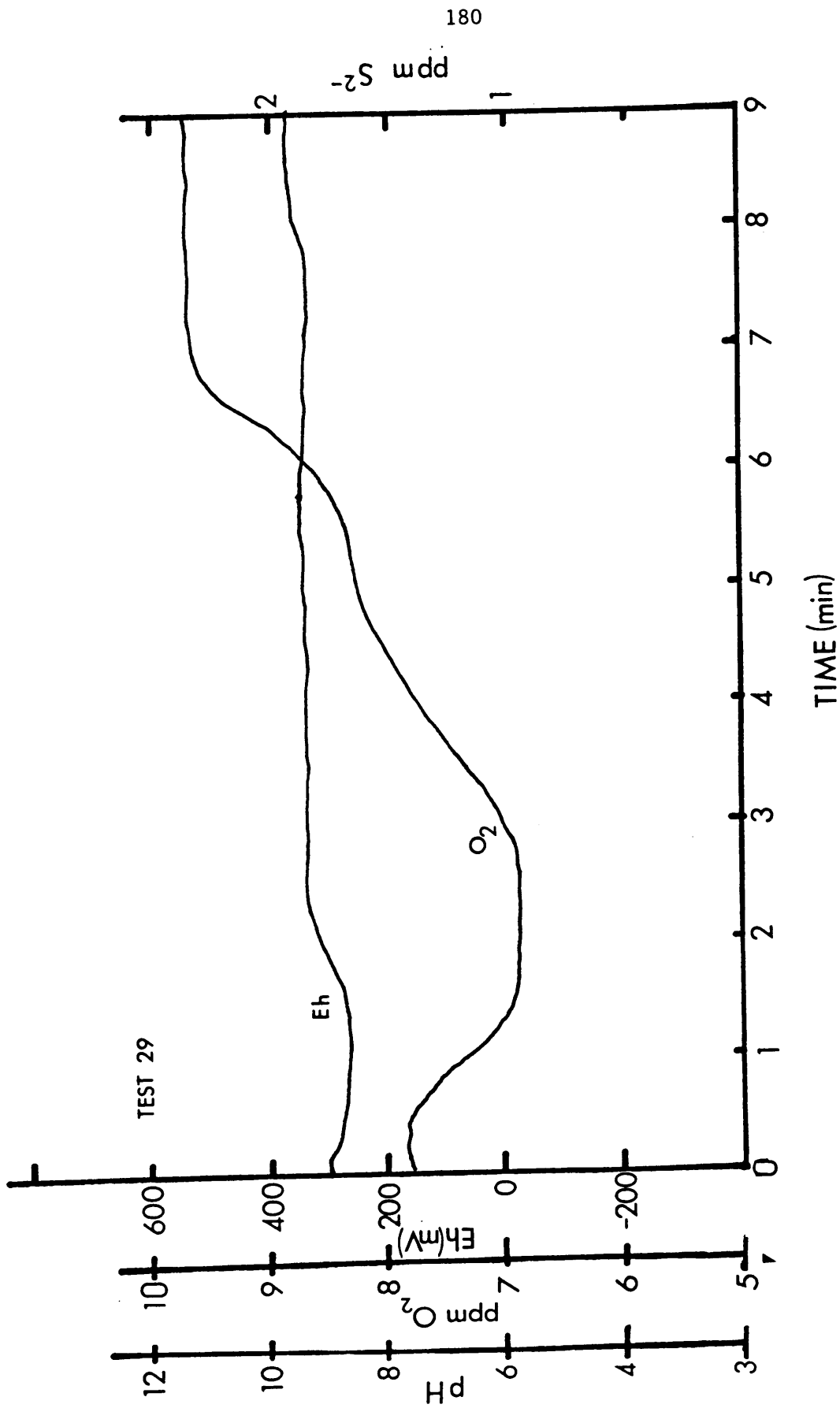


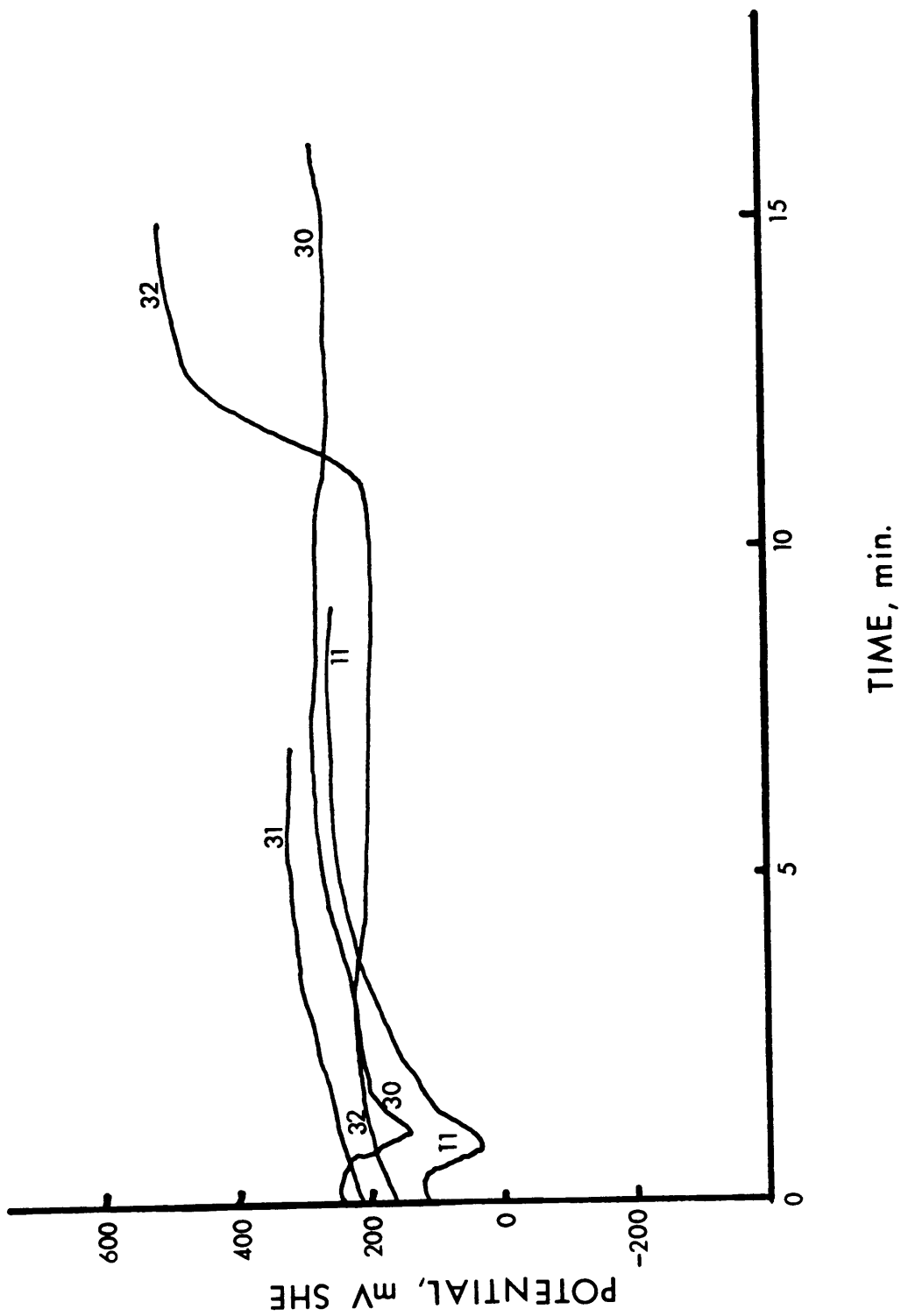


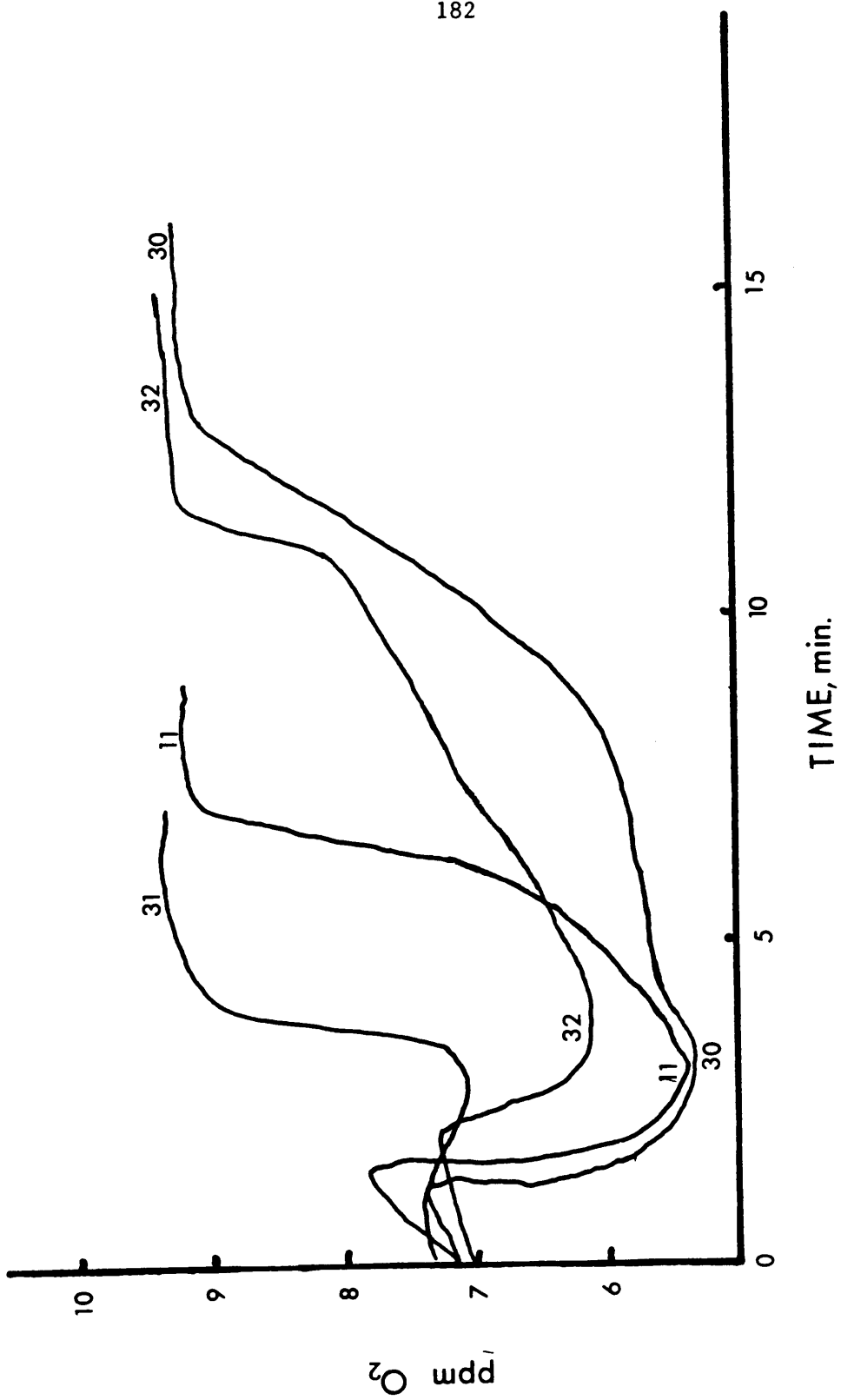












APPENDIX E: ASSAY PROCEDURE

## ASSAY PROCEDURE

An amount of flotation product, between one-half and two grams, depending on the expected concentration, was accurately weighed into a 125-ml Erlenmeyer flask. Twenty ml of aqua regia (50% HCl-50% HNO<sub>3</sub>) was added and the flask placed on a hotplate and heated to boiling. A reflux boiling funnel was placed in the mouth of the flask to prevent excessive splashing of the solution and to wash the sides of the flask. The sample was slowly boiled until the solution had nearly all evaporated. The sides of the flask were then rinsed with enough water to dissolve any metal ions.

The samples, thus digested, were allowed to cool and filtered through Whatman number 2 filter paper to remove any undissolved solids. The filtrate was then transferred to a volumetric flask and diluted with doubly distilled water. Further dilution was often necessary to bring the zinc concentration in the unknown sample to within the range of the calibration standards.

The zinc content of each sample was then determined using a Spectraspan IV Plasma Emission Spectro-

meter, set to detect zinc in the wavelength region around 2025.51 Angstroms. All glassware used in these procedures was cleaned in Micro cleaning solution, rinsed with tap water, soaked for a minimum of 2 hours in 20%  $\text{HNO}_3$ , then rinsed with distilled and doubly distilled water. This procedure was carried out to ensure that no extraneous metal ions would interfere with the analysis.

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