

The Quest for a Simple Bioactive Analog of Paclitaxel as a Potential Anticancer Agent

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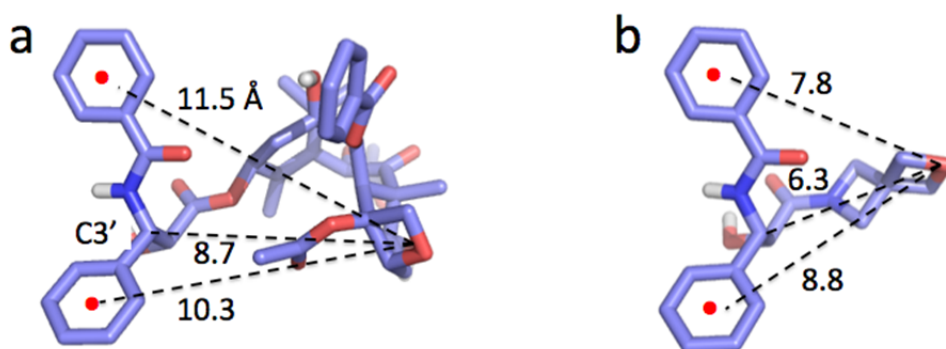
CONSPECTUS

Paclitaxel (PTX), introduced into the clinic in 1991, has revealed itself as an effective anti-microtubule drug for treatment of a range of otherwise intractable cancers.

Along with docetaxel (DTX)

and in combination with other agents such as cisplatin, it has proven to be a first-line therapy.

Unfortunately, PTX and DTX carry severe liabilities such as debilitating side effects, rapid onset of resistance and rather complex molecular structures offering substantial challenges to ease of synthetic manipulation. Consequently, the past 15 years has witnessed many efforts to synthesize and test highly modified analogs based on intuitive structural similarity relationships with the PTX molecular skeleton as well as efforts to mimic the conformational profile of the ligand observed in the macromolecular tubulin-PTX complex. Highly successful improvements in potency, up to 50-fold increases in IC_{50} , have been achieved by constructing bridges between distal centers in PTX that imitate the conformer of the electron crystallographic binding pose. Much less successful have been numerous attempts to truncate PTX by replacing the baccatin core with simpler moieties to achieve PTX-like potencies and application of a wide range of flexible synthesis-based chemistries. Reported efforts, characterized by a fascinating array of baccatin substitutes, have failed to surpass the bioactivities of PTX in both microtubule disassembly assays and cytotoxicity against a range of cell types. Most of the structures retain the main elements of the PTX C13 side chain, while seeking a smaller rigid bicycle as a baccatin replacement adorned with substituents to mimic the C2 benzoyl moiety and the oxetane ring. We surmise that past studies have been handicapped by solubility and membrane permeability issues, but primarily by the existence of an expansive taxane binding pocket



and the discrepancy in molecular size between PTX and the pruned analogs. A number of these molecules offer molecular volumes 50-60% that of PTX, fewer contacts with the tubulin protein, severe mismatches with the PTX pharmacophore, lessened capacity to dispel binding site waters contributing to ΔG_{bind} and unanticipated binding poses. The latter is a critical drawback, if molecular designs of simpler PTX structures are based on a perceived or known PTX binding conformation. We conclude that design and synthesis of a highly cytotoxic tubulin-assembly agent based on the paclitaxel pharmacophore remains an unsolved challenge, but one that can be overcome by focus on the architecture of the taxane binding site independent of the effective, but not unique, hand-in-glove match represented by the PTX-tubulin complex.

1. Introduction

The decade of the 1970s was bookended by two significant discoveries in the area of naturally occurring anticancer agents: the isolation and structure elucidation of the diterpenoid taxol **1** (as it was then called) in 1971,¹ and its subsequent identification as a promoter of microtubule assembly in 1979.² A personal account of these two events has recently been published.³ These discoveries led ultimately to the first clinical demonstration of the activity of taxol against ovarian cancer in 1989,⁴ its approval for treatment of ovarian and breast cancers by the FDA in 1991 and 1993, respectively, and its controversial name change to paclitaxel.⁵ Docetaxel (**2**)⁶ and cabazitaxel (**3**)⁷ are the only other taxanes currently in clinical use (Figure 1).

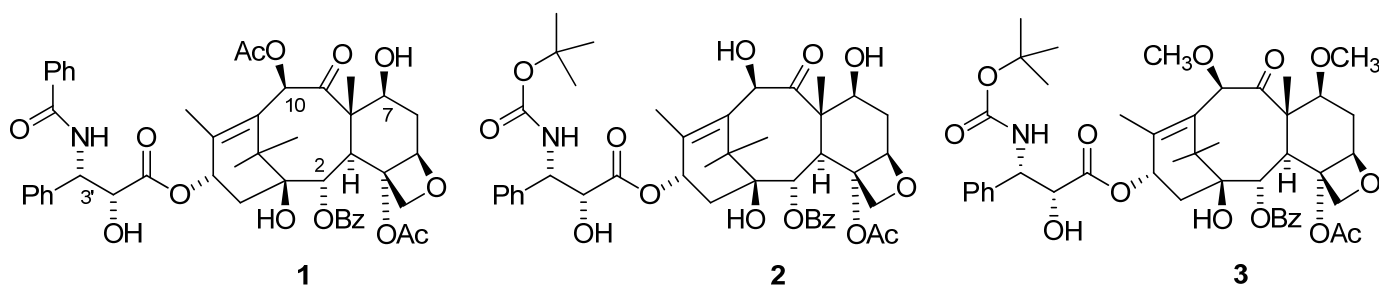


FIGURE 1. The structures of paclitaxel (Taxol) (**1**) docetaxel (Taxotere) (**2**), and Cabazitaxel (**3**).

The mechanism of action of paclitaxel is intimately associated with its activity as a potent promoter of microtubule assembly and, thus, as an antimetabolic agent,^{8,9} but it has also shown activity as a neuroprotective agent.¹⁰ Under physiological conditions the major therapeutic effect of taxol and taxol-like compounds is the slowing of tubulin dynamics rather than tubulin polymerization,⁹ and a recent publication has argued that its interference with intracellular trafficking on microtubules is probably its most important function.¹¹ Since both of these effects depend on or are at least closely related to the

ability of paclitaxel to bind to microtubules and to promote the polymerization of tubulin to microtubules, this activity remains the key to its overall effectiveness as an anticancer agent.

The numerous paclitaxel analogs in clinical trial¹² and the vast majority of the analogs that have been evaluated retain the basic paclitaxel skeleton and differ in substituents on the ring system and on the side chain. This approach, while successful, begs the question “Is the paclitaxel skeleton essential for bioactivity? Is it possible that a simplified structure could be constructed with equal or even enhanced bioactivity as compared to paclitaxel?” This account lays out the options for such a structure, and describes the progress made towards its synthesis. The selection of compounds for discussion has been limited to those prepared by total synthesis, or those compounds prepared from paclitaxel or 10-deacetyl baccatin III designed to test the microtubule-bound conformation of the drug. This definition necessarily excludes the many thousands of analogs prepared by chemical modification of paclitaxel or 10-deacetyl baccatin III.

The structure-activity relationships of paclitaxel have been investigated extensively and reviewed on several occasions.¹³⁻¹⁶ For the purposes of this review, the major structural features necessary for activity are the C13 *N*-acyl- β -phenylisoserine side chain or some variation of it, an acyl group at C4, and the C2 benzoate group or some variation thereof. While the oxetane ring was originally thought to be essential for activity based on early results that ring-opened analogs were inactive,¹⁷ the cyclopropane analog **4** and the D-seco analog **5** reveal potency similar to paclitaxel in the stabilization of microtubules (Figure 2).^{18,19} Thus, the oxetane ring is no longer regarded as essential.

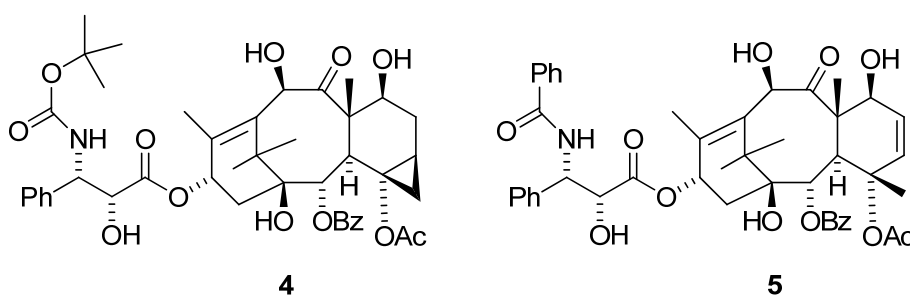


FIGURE 2. Active oxetane ring-opened structures **4** and **5**.

2. Simplified analogs designed without the assistance of the structure of the ligand-MT complex

One of the first attempts to design a simplified analog was made by Fuji and coworkers, who used molecular mechanics to calculate the distance between the C4 and C13 carbons of paclitaxel as 4.5 Å.²⁰ This information was used to design **6a** – **6f** (Figure 3), where the distance *D*, corresponding to

best tubulin-assembly activity, 13-fold more potent than paclitaxel at stabilizing microtubules. Its close analog **9b** was evaluated in the NCI 60-cell line screen, but it proved much less active than paclitaxel, and the compounds were abandoned as cancer therapeutics.²³ The reason for the discrepancy between microtubule stabilization and antiproliferative activities was not investigated, but it could be due in part to the lack of water-solubility or poor cell membrane penetration.

Frejd et al. designed **10** (Figure 5) based on comparative modeling of its spiro[6-hydroxybicyclo[2.2.2]octan-2-one-3,1'-cyclohex-3'-en]-2'-one core with the baccatin core of one of the X-ray structures of paclitaxel using MacroModel. The compound was inactive in an assay to determine microtubule stabilization. The authors concluded this was likely due to the lack of a 2-benzoate group.²⁴

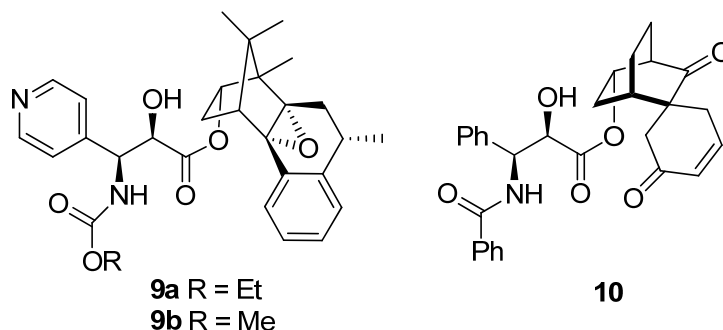


FIGURE 5. Klar's and Frejd's simplified structures.

Analogs based on the adamantane core were prepared by Zefirova and coworkers. The synthesis of **11a**²⁵ was followed by the synthesis and biological evaluation of **11b** – **13** (Figure 6). All four compounds were over a thousand-fold less cytotoxic than paclitaxel, but all likewise showed some level of tubulin polymerization activity as judged by the relative amount of tubulin pelleted after incubation with each substance.^{26,27}

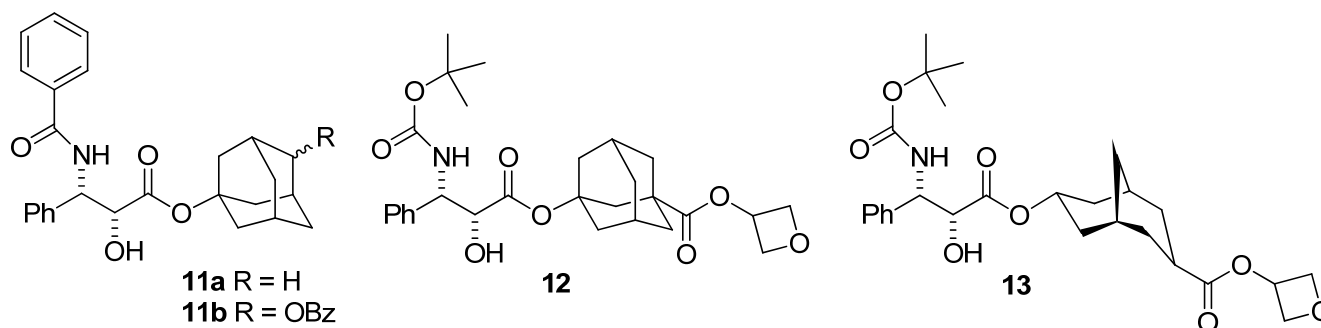


FIGURE 6. Zefirova's simplified structures.

Vauzeilles and Beau prepared simplified compounds with the general structure **14** (Figure 7), where R represents one of seven substituted benzenes or one of three substituted benzyl groups. The

structures were designed based on an overlay of the MM3* conformation of the β -L-glucurono- γ -lactone core of **14** with the taxane core of paclitaxel. The only analog to show any tubulin polymerization activity was **14d**, but this was only very weakly active (IC_{50} 90 μ M) compared with paclitaxel's 0.5 μ M response in the same assay.²⁸

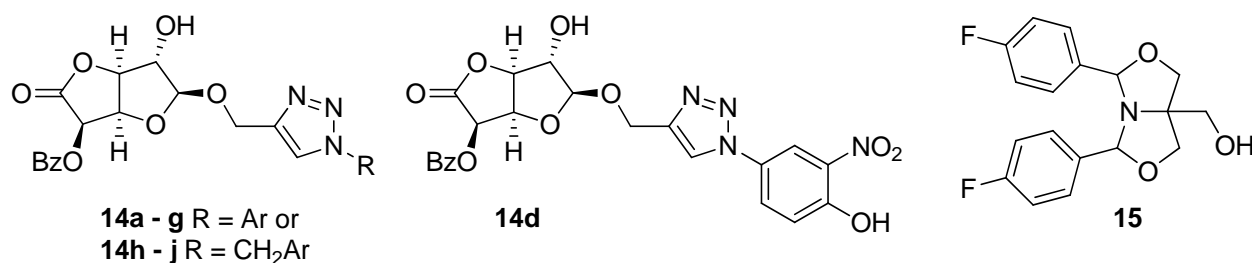


FIGURE 7. Vauzeilles, Beau's, and Shintani's simplified structures.

Another analog with two fused five-membered rings was evaluated by Shintani et al., who found that the small molecule designated GS-164 (**15**) (Figure 7) had a similar tubulin-assembly effect at 40 μ M to that of paclitaxel at 5 μ M. GS-164 was however had at least 1000-fold less growth inhibitory potency than paclitaxel in a series of cell lines.²⁹ It was also found to have activity as a neuroprotective agent.³⁰

Roussi et al. prepared analogs based on a cholic acid precursor (Figure 8). The key *cis* A-B ring junction provides a U-shaped conformation similar to that of baccatin III. None of the analogs showed inhibitory activity against microtubule disassembly. Compounds **16a** and **16c** did reveal weak cytotoxicity to KB cells (IC_{50} 's 2.7 and 7.2 μ M, respectively), while **16b** was not cytotoxic. It showed modest inhibition of microtubule assembly; the opposite of that from paclitaxel!³¹

Howarth et al. synthesized protected **17a – 17d** (Figure 8) based on the hypothesis that paclitaxel behaves as a GTP mimic, with the baccatin III core acting as the guanosine part of GTP and the side chain representing a triphosphate. The deprotected derivative was not prepared, but the protected analogs were weakly cytotoxic against the colon cancer cell line SW480.³² No results of microtubule disassembly assays were reported.

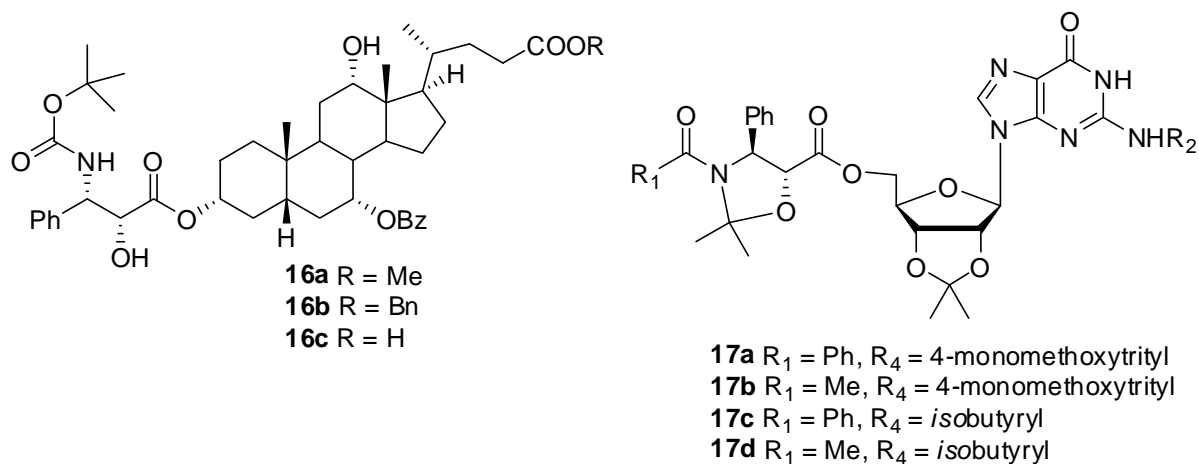


FIGURE 8. Roussi's and Haworth's simplified structures.

3. The solution and microtubule-bound conformations of paclitaxel

As noted earlier, the bioactivity of paclitaxel is closely linked to its ability to bind to microtubules and stabilize them, leading to mitotic arrest.³³⁻³⁵ It has been presumed that the design of a simple bioactive analog of paclitaxel is likely to be successful if the resulting structure possesses a 3D shape closely matching the microtubule-bound conformation of paclitaxel. In this context, the determination of conformation becomes a matter of prime importance. The major conformational variations among candidate tubulin-binding structures are in the C13 side chains as illustrated by Figure 9. For additional details, see the Supporting Information.

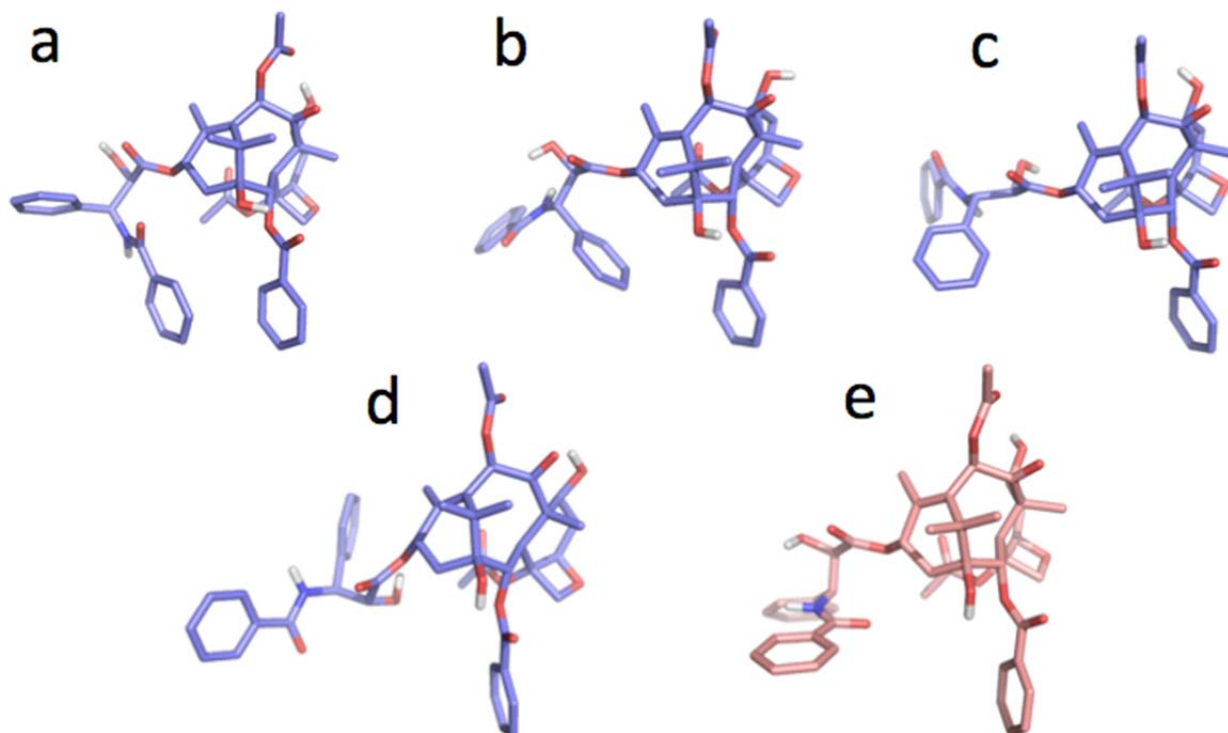


FIGURE 9. Conformations of paclitaxel, a) non-polar,^{36,37,38} b) polar collapsed,^{37,39} c) extended,^{37,38} d) REDOR Taxol,⁴⁰ e) T-Taxol.⁴¹

Various conformationally constrained taxoids have challenged the proposed polar hydrophobic collapse and non-polar poses. Georg prepared bridged analogs **18a** and **18b** (Figure 10) as a test of the “hydrophobic collapse” conformation, but neither demonstrated tubulin-assembly activity, providing indirect evidence that the bound state is unlike this conformer.⁴²

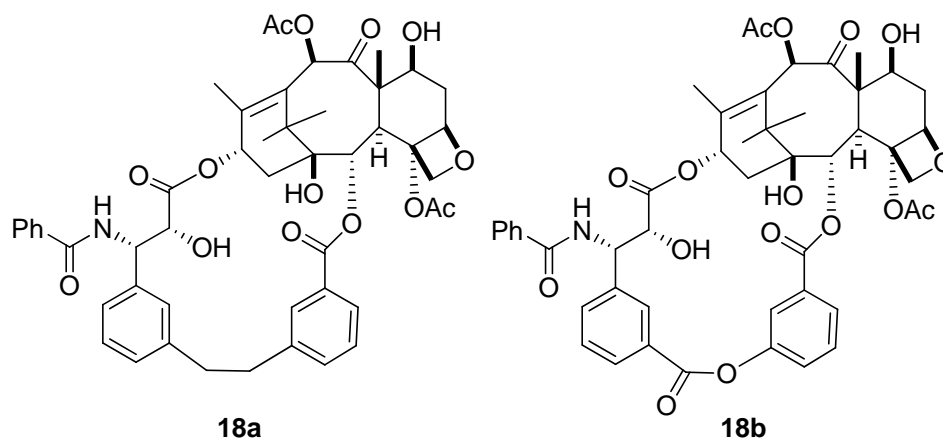


FIGURE 10. Bridged analogs presumed to mimic the “hydrophobic collapse” conformation of paclitaxel.

A series of macrocyclic taxanes was prepared by Ojima to test his proposed common pharmacophore for paclitaxel and the epothilones.⁴³ Bridged compound **19** (Figure 11) gave an IC_{50} value of $0.39 \mu\text{M}$ against human breast cancer cells (MDA-435yLCC6-WT) and 37% tubulin polymerization vs. paclitaxel. This is a reversal of the pattern found in most of the other compounds discussed herein; with good tubulin polymerization activity but weaker activity against cells. This could mean that **19** operates by a different mechanism against cells than paclitaxel. A later paper reported the synthesis of **19** and a large number of similar congeners with differing ring sizes, but **19** was the most active in the series.⁴⁴

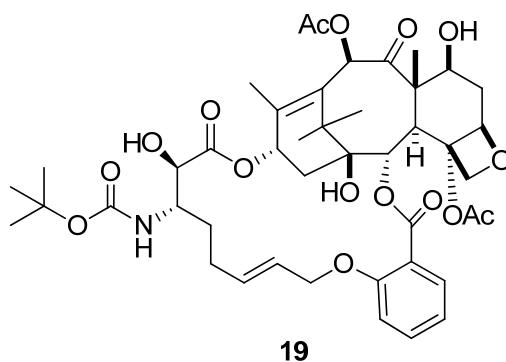


FIGURE 11. Bridged paclitaxel analog mimicking the “common pharmacophore” conformation of paclitaxel.

Two non-taxoids, **20** and **21** (Figure 12) were also prepared by Ojima to test the common pharmacophore concept.⁴⁵ Both compounds showed only micromolar cytotoxicities against four different cell lines and no tubulin assembly activity. It would appear that the limited cytotoxic activities arise by a mechanism of action different from that of paclitaxel.

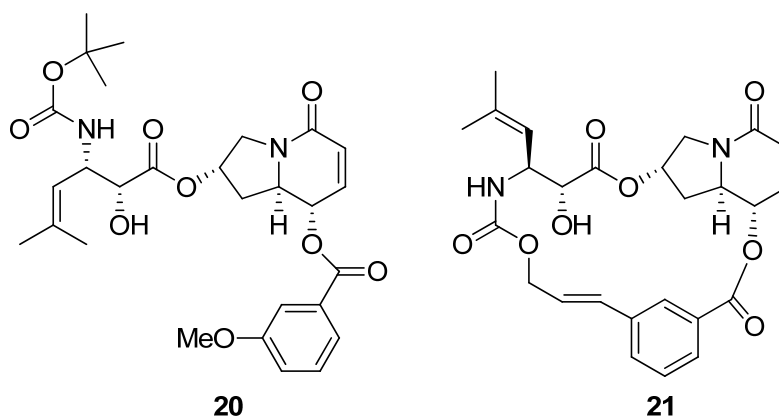


FIGURE 12. Non-taxoid analogs to mimic the "common pharmacophore" conformation of paclitaxel.

Ojima also prepared bridged **22** (Figure 13) and some related compounds based on the x-ray crystallographic structure of paclitaxel. Compound **22** was the most cytotoxic (IC_{50} 0.067 μ M, LCC6-WT human breast cancer cells) compared with 0.004 μ M for paclitaxel, but no tubulin assembly data were reported. Thus, the mechanism of action was not established.⁴⁶

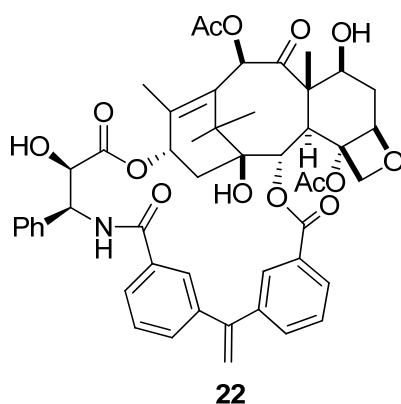


FIGURE 13. Bridged analog that mimics the x-ray structure of paclitaxel (**22**).

These results, taken together, indicate that the activities of bridged paclitaxels vary widely with the nature of the bridging linker, but that none of the proposed models are capable of guiding the synthesis of constrained analogs with activity superior to paclitaxel itself.

Two other conformations for the tubulin-bound structure of paclitaxel have been proposed. The T-taxol structure was initially identified by one of us by mapping to the electron crystallographic density.⁴¹ The related REDOR-taxol structure was proposed by Ojima in 2005⁴⁰ based on initial REDOR experiments reported in 2000 by Bane et al.⁴⁷ The structure was modified⁴⁸ by a second set of REDOR NMR data⁴⁹ and by modeling the structure in a reshaped 1JFF tubulin structure.⁵⁰ T-taxol and REDOR-taxol differ considerably in the C13 side chain conformations leading to alternative orientations of the C2' hydroxyl group and the two terminal phenyl moieties emanating from C3'. These deviations furnish a molecular volume for the REDOR conformer that is $\sim 15 \text{ \AA}^3$ larger than the T-Taxol conformer. The two conformations have been compared by Ojima⁴⁸ and by Snyder^{51,52,53} with significantly different conclusions. High level density functional calculations suggest the T-taxol conformer to be 5-7 kcal/mol more stable than the REDOR form.⁵³ In spite of the differences, the general outline of the microtubule-bound conformation is clear enough to allow the design of simplified analogs with a somewhat common overall shape.

Experimental validation of the taxol binding conformation, or a closely related form, was provided by a series of REDOR NMR experiments on labeled paclitaxels. The first analysis in 2000 employed the labeled paclitaxel **23**,⁴⁷ while a second set of REDOR measurements was performed on labeled **24** and **25** (Figure 14).⁴⁹ These studies led to the assignment of internuclear distances shown in **26** and **27** (Figure 15).⁴⁹

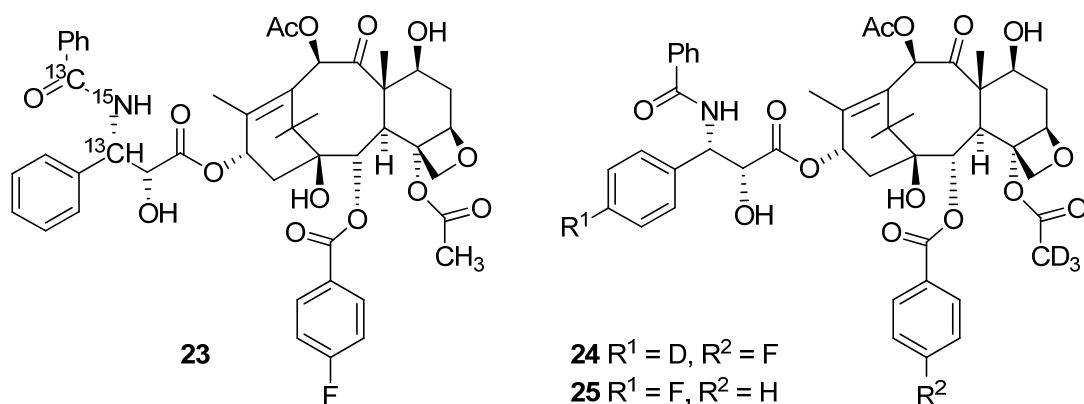


FIGURE 14. Labeled paclitaxels used for REDOR internuclear distance determinations.

Internuclear distances determined by REDOR NMR were compared with those predicted by the major conformers depicted in Figure 9, with the results shown in Table 1.⁴⁹ Both the T-taxol and the REDOR-taxol conformations are consistent with the REDOR NMR data, although neither set of

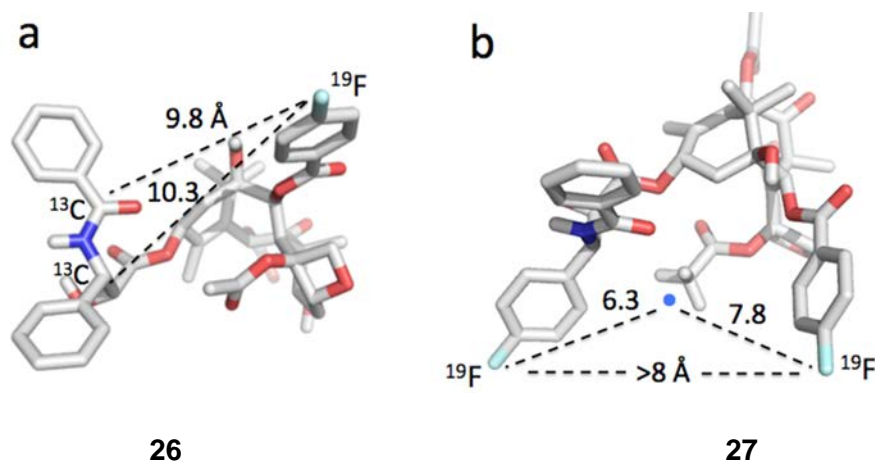
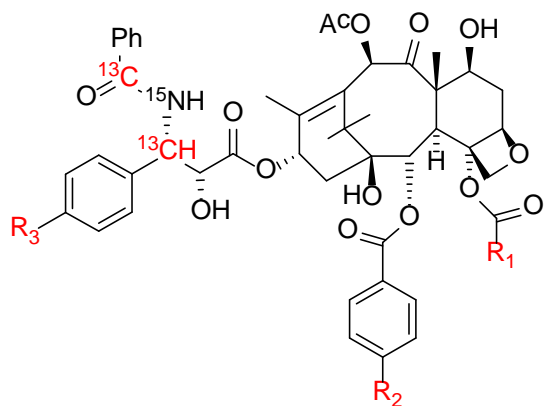


FIGURE 15. Internuclear distances among centers in microtubule-bound paclitaxel from REDOR NMR. a) C2-benzoyl (*p*-F) to the C3' carbon and the C(=O) carbon of the benzamide; b) C2-benzoyl (*p*-F), C4-CD₃ acetate and C3'-phenyl (*p*-F).

distances in **26** and **27** are able to resolve the C13/C2'OH conformational issue.⁵¹

TABLE 1. Comparison of predicted and observed internuclear distances for microtubule-bound paclitaxel^a



Distances (Å)	Polar	Non-polar	REDOR PTX	T-Taxol	Expt
R ¹ -R ²	7.9	8.0	7.6	7.9	7.8
R ¹ -R ³	5.9	7.2	6.1	6.6	6.3
R ² -R ³	4.6	12.5	13.1	12.2	>8
R ² - ¹³ CH	9.6	8.5	9.5	9.9	10.3
R ² - ¹³ CO	10.4	6.2	9.9	9.1	9.8

^aNumbers in bold agree with the REDOR data within ± 0.8 Å

Additional powerful support for the taxol binding conformation was provided by the synthesis of bridged paclitaxels locked into conformations designed to mimic those of T-taxol (Figure 16).⁵⁴ Two of these bridged analogs, **28** and **29**, which best matched the T-taxol conformation showed enhanced activity compared with paclitaxel. Compound **28** (IC₅₀ 0.30 nM, A2780 ovarian cancer cells) was 22-fold more cytotoxic than paclitaxel (IC₅₀ 6.6 nM) and also approximately double the tubulin-assembly

activity of paclitaxel. The related compound **29** with a saturated linker was slightly less potent to cells but slightly better at promoting tubulin assembly. Many other related compounds were prepared, but **28** proved to be the most potent analog.⁵⁵

Compound **30** was synthesized as a test of the REDOR-taxol conformer.⁵⁰ It was approximately equipotent with paclitaxel against a panel of six cell lines, and it promoted tubulin assembly as well as paclitaxel.

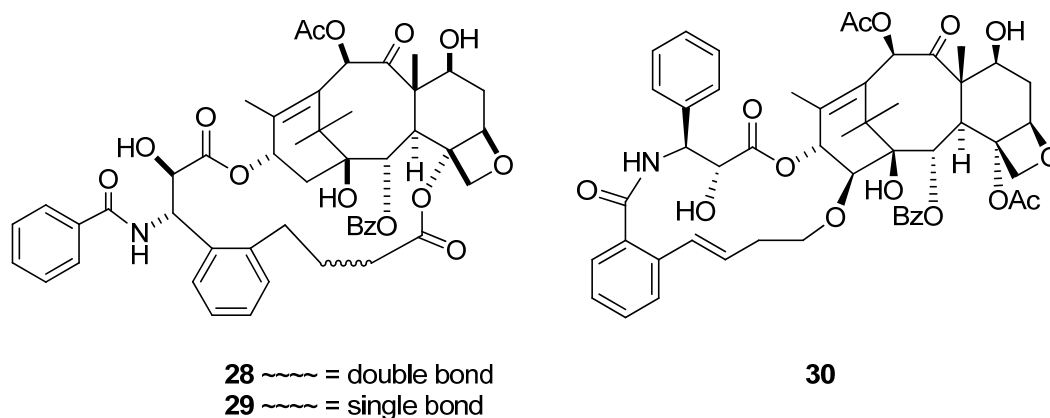


FIGURE 16. Constrained paclitaxel analogs with equal or better potency than paclitaxel.

Taken together, the combined evidence from REDOR NMR studies and from the synthesis of conformationally constrained paclitaxel analogs provides strong support for the electron crystallographic T-taxol/tubulin structures^{41,56} as being the best models of the tubulin-bound conformation of paclitaxel. The question remains, is it possible to construct compounds which retain paclitaxel's bioactivity with the same basic shape but with much less structural complexity?

4. Simple compounds based on the microtubule-bound conformation of paclitaxel

Bridged bicycles **31** – **34** (Figure 17) were prepared by Ganesh et al. as an attempt to design bioactive compounds based on the T-taxol conformation.⁵⁷ All had similar antiproliferative activities, with IC_{50} values in the 10 – 18 μ M range against the A2780 cell line, as opposed to 0.02 μ M for paclitaxel. All were able to promote tubulin assembly and stabilize the resulting microtubules to cold-induced disassembly at a dose of 30 μ M, compared with paclitaxel's IC_{50} of 0.4 μ M. Thus, these compounds exhibit definite but weak tubulin-assembly activity as well as antiproliferative activity.

Compound **32** was modeled into the β -tubulin paclitaxel-binding site and shown to be able to adopt a conformation similar to T-taxol with the C-4 and C-13 side chains matching closely. The C-2 benzoyl

phenyl ring overlapped that of paclitaxel, but not exactly. Being forced deeper into the hydrophobic pocket, the phenyl experienced steric congestion with Leu230 and Leu275 tubulin side chains. The lack of activity of these compounds may be due to this observation or to insolubility, or both.

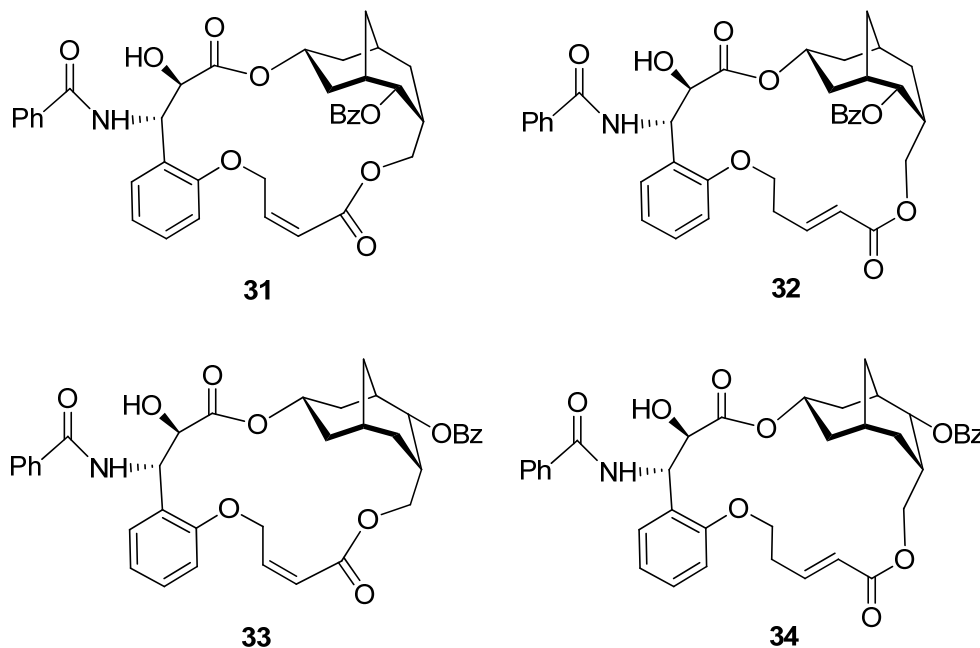


FIGURE 17. Bicyclononane mimics.

Tricyclic bridged **36** and the related unbridged **35** (Figure 18) were prepared by Sun et al. in a test of the REDOR-taxol conformation.⁵⁸ Unbridged **35** was the best mimic of the REDOR-taxol conformation, with IC_{50} values of 3.8 – 8.3 μ M against a panel of four non drug-resistant cell lines; paclitaxel had IC_{50} values of 0.002 – 0.05 μ M when tested against the same cell lines. Compound **35** showed weak binding to tubulin, as determined by displacement of the fluorescent paclitaxel analog Flutax-2 with an estimated binding constant of 50 – 100 μ M. Bridged **36** was inactive to all cell lines except the A2780 ovarian cancer line, where it furnished an IC_{50} of 15 μ M. Molecular dynamics simulations for **36** showed significant differences between the conformational stabilities of the tubulin-docked structures of **36** and REDOR-taxol, suggesting it to be a reason for the differences in potency.

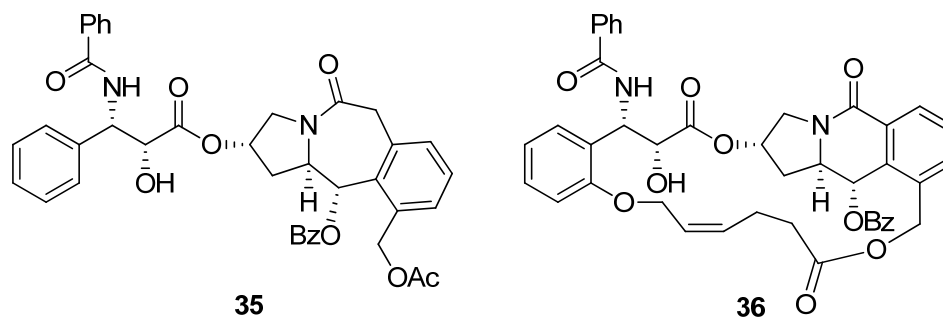


FIGURE 18. Ojima's tricyclic alkaloid mimics.

The final example of an alkaloid mimic was reported by Zhao et al. in 2011. Bridged alkaloid **37** (Figure 19) related to Sun's alkaloid **36**,⁵⁸ and two related compounds were prepared from *cis*-4-hydroxyproline, based on computational analysis to determine which core structure best served as the optimal mimic of the baccatin structure.⁵⁹ The design also incorporated a basic tertiary nitrogen in an attempt to increase water solubility through salt formation. Of the resulting compounds, **37** and its open chain precursor **38** had IC₅₀ values of 4.5 and 5.8 μM respectively against the A2780 ovarian cancer cell line. These values compare with 0.015 μM for paclitaxel in the same assay. Interestingly the open-chain alkaloid **38** was almost as potent as the bridged alkaloid **37**, indicating that bridging in this molecular system has no significant effect on activity.

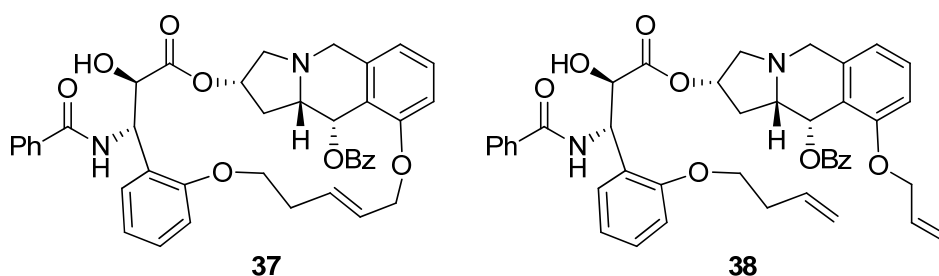


FIGURE 19. Zhao's tricyclic alkaloid mimics.

The compounds proved to be relatively insoluble in water even as salts, leading to an important discovery for their tubulin assembly activity. The initial good tubulin assembly activity was subsequently reinterpreted to be a false positive due to light scattering by insoluble substances. Determination of tubulin assembly activity by the alternative method of observing DAPI fluorescence (4',6-diamidino-2-phenylindole) failed to detect significant tubulin polymerization.

The failure of bridged **37** to show significantly increased activity compared with open chain **38** is in telling contrast with the case of the bridged *A-nor*-taxol **40** with at least 20-fold improved antiproliferative activity compared with its unbridged precursor **39** (Figure 20).⁶⁰ The reasons for these differences are not well understood at present.

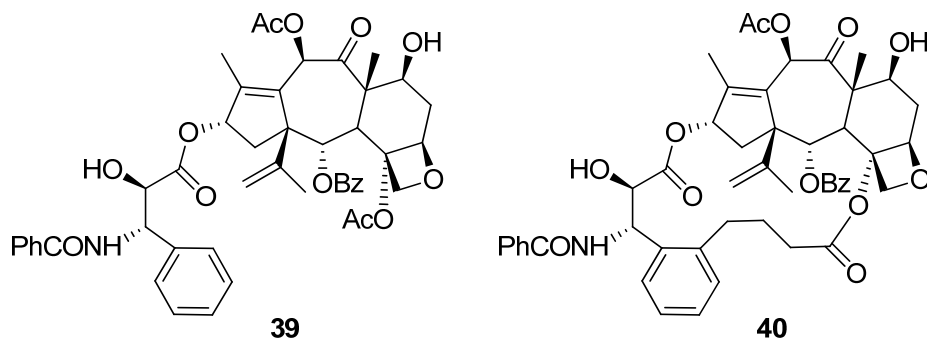


FIGURE 20. *A-nor-taxol* and its bridged analog.

6. Summary and conclusions

A summary of the structures and bioactivities of the compounds discussed is provided in the Appendix, Table S1. The attempts made to date to design simplified mimics of paclitaxel with bioactivities similar to those of paclitaxel have so far not proved to be completely successful. None of the mimics combines significant activity in cells with significant tubulin-assembly activity. In terms of cell-based activity, the simple analogs **7a** and **7b** are the most potent, with activities ranging from 3.5 to 7.7-fold less potent than paclitaxel in three different cell lines. This substantial activity is unlikely due to tubulin-assembly, however, since these compounds are 120-200 fold less potent than paclitaxel, and most of the remaining compounds are at least 100-fold less potent. Compounds **20** and **35** are exceptions with potencies only 1-2 orders of magnitude less than paclitaxel against drug-resistant cell lines.

The situation for tubulin-polymerization activity is more complex, since different papers report the results in different ways. In sum, the only really active analogs are Klar's pentacyclic compounds,²³ of which **9a** and **9b** are good examples. These compounds appear to be more potent tubulin polymerization agents than paclitaxel, although the data were determined in an unusual way, and paclitaxel appears to be less active under these conditions than might have been expected.

The final point to note is that efforts to lock some of the simple compounds into the electron crystallographic taxol-tubulin conformation do not appear to improve bioactivity. In fact, it can be reduced. Bridged **36** is significantly less active than the similar but unbridged **35**,⁵⁸ while bridged **37** and its uncyclized precursor **38** have essentially the same activities in a cell-based assay.⁵⁹

The reasons for the failure to develop simple bioactive tubulin-assembly promoters based on the T-taxol-tubulin structure and the related REDOR pharmacophore are not completely clear, since the structural target was highly successful in the design of bridged paclitaxel derivatives with improved

activity.^{54,60} There may be several reasons for this. One impediment for the simplified mimics is likely to be their high insolubility and possibly poor membrane penetration, which presumably reduces their cellular uptake and their ability to encounter tubulin. Second, several important factors most certainly center around differences in molecular size, in particular, as it relates to the large taxane binding pocket and the discrepancy between the 3D space occupied by Taxol and the simplified ligands. The latter not only access fewer contact anchor points, but it is likely they only poorly match key pharmacophore elements as indicated in Fig. 21. There is little chance that **7a** and **7b**, the most active truncated paclitaxel analogs to date, can employ the same set of contacts with tubulin, implying that if they bind to the taxane binding pocket, they must certainly adopt a significantly different binding pose. Alternatively, as argued above, these compounds may likely find a binding nest somewhere else on the protein. Consequently, using the paclitaxel binding conformation or pharmacophore in the design phase for such significantly pruned agents would be misleading. This contrasts with bridged analogs like **28 - 30**, which retain the full baccatin core and are constrained closely to the paclitaxel binding conformation.

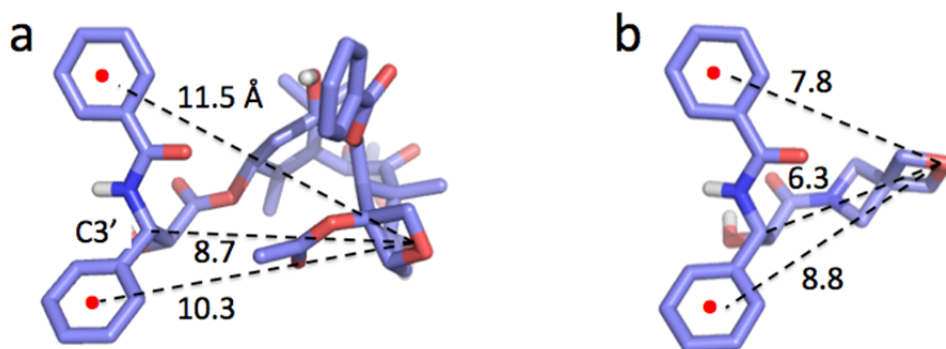


FIGURE 21. Comparison of interatomic distances (Å) for Taxol and **7a** with the same C13-side chain conformation. a) Taxol (pdf code 1JFF) with distances from the C13 phenyl ring centroids and C3' to the oxetane oxygen, b) Compound **7a** with similar distances to the spirocyclic oxetane oxygen.

A third barrier to promoting tubulin assembly is likewise related to molecular size. The volume of T-Taxol solved in 1JFF is 831 Å³,⁶¹ while those for the most active truncated taxanes discussed herein (**7a** and **7b**) are only 376 and 391 Å³ respectively, a little under half the volume. Compounds **9a** and **9b**, which promote potent tubulin polymerization but fail to kill cells, likewise occupy only 60-65% of the volume presented by paclitaxel. Ligand occupation of tubulin protein clefts displaces water molecules into the microtubule lumen contributing to the free energy of binding (ΔG) via $-T\Delta S$. This term can be significant for many taxanes.⁶² Small molecules, however, carry a reduced capacity to

dislodge water molecules from the same pockets. Thus, even if favorable geometries might be achieved by truncated taxanes, a combination of fewer ligand protein contacts, alternative binding poses and an entropy disadvantage for reduced scaffolds may well limit the extent of potency equivalency relative to the taxanes, epothilones and other agents known to bind the taxane site. The design and synthesis of a highly cytotoxic tubulin-assembly agent based on the paclitaxel pharmacophore thus remains an, as yet, unsolved challenge.

With the knowledge surveyed here, we project that a potentially more fruitful approach would focus on exploitation of the properties of the taxane binding pocket alone and ignore perceived or observed paclitaxel conformations. It is worth noting that the taxoid site is home to not only PTX, but also the epothilones,⁶³ discodermolide,⁶⁴ dictyostatin,⁶⁵ and eleutherobin and sarcodictyin A.⁶⁶ While fully-confirmed structures of the corresponding tubulin complexes are not known, a reasonable binding hypothesis recognizes that a subset of pharmacophore elements might be shared by these molecules as proposed for PTX and epothilone,⁶⁷ but that each structure likewise utilizes a separate set of pharmacophore points dictated by a combination of individual structure and available conformations. In addition, the large tubulin binding cavity most likely accommodates different ligands in slightly different subsites. This implies that the remaining unoccupied space can house both highly ordered and less tightly bound waters that contribute differently to each ligand binding profile. Consequently, a potentially effective strategy for designing novel and easily prepared scaffolds to mimic the biological effects of PTX might employ two complementary sub-strategies. First, a fragment-based approach has the capacity to identify and exploit novel sets of pharmacophore anchor points associated with unique binding poses.^{68,69} Second, exploration of the water network surrounding a bound ligand with tools such as Watermap⁷⁰ or SZMAP and GAMEPLAN⁷¹ offers opportunities to take advantage of enhanced binding free energy contributions by solvent manipulation. This one-two structure-based ligand optimization process, and others like it, may well overcome the limitations of the more intuitive PTX-based approaches of the past.

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SUPPORTING INFORMATION AVAILABLE

A discussion of the C13 side chain conformations of paclitaxel and Table S1 summarizing the structures and bioactivities of all the compounds discussed is provided as Supporting Information. This information is available free of charge via the Internet at <http://pubs.acs.org/>.

BIOGRAPHICAL INFORMATION

David G. I. Kingston was born in London, England, and received his B.A and Ph.D. degrees (Lord Todd and D. W. Cameron) from Cambridge University in 1960 and 1964, respectively. After postdoctoral appointments at M.I.T. and Cambridge, he moved to SUNY Albany in 1966, and then to Virginia Tech in 1971, where he is currently a University Distinguished Professor of Chemistry and Director of the Virginia Tech Center for Drug Discovery. He has worked extensively on the chemistry, microtubule binding, and nanoparticle delivery of paclitaxel. He has carried out conservation and drug discovery in Suriname and Madagascar, and he is currently using ethnobotany to search for antimalarial agents from plants. He is a recipient of the Research Achievement Award of the American Society of Pharmacognosy and the ACS Guenther Award for Natural Products, and is a member of the National Advisory Council for Complementary and Alternative Medicine. He also serves as one of several lay pastors in his local church.

James P. Snyder was born in Seattle, USA, and received his B.S. and Ph.D. degrees from St. Martin's College and Cornell Universities, respectively, followed by postdoctoral fellowships at the Friedrich-Alexander and Columbia Universities. The subsequent road passed through university appointments, industrial research and back to the university again: Yeshiva University (New York City), University of Copenhagen (Denmark), Merck Sharp and Dohme (New Jersey), Searle Pharmaceuticals (Chicago), IRBM (Institute for Research in Molecular Biology, Rome, Italy) and Emory University (Atlanta, Georgia), where he is currently Director of Biostructural Research. The same path has crossed chemical, structural, theoretical, biological and biochemical boundaries that have been projected into a palette of multi-disciplinary parallel investigations and a course for undergraduates that links our daily lives to the root fundamentals of science: "Chemistry Biology and Molecular Modeling". He still rides his yellow scooter to work daily through the hills and dales of Atlanta.

FOOTNOTES

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