

Techniques for Processing Airborne Imagery for Multimodal Crop Health Monitoring and Early Insect Detection

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ABSTRACT

During their growth, crops may experience a variety of health issues, which often lead to a reduction in crop yield. In order to avoid financial loss and sustain crop survival, it is imperative for farmers to detect and treat crop health issues. Interest in the use of unmanned aerial vehicles (UAVs) for precision agriculture has continued to grow as the cost of these platforms and sensing payloads has decreased. The increase in availability of this technology may enable farmers to scout their fields and react to issues more quickly and inexpensively than current satellite and other airborne methods. In the work of this thesis, methods have been developed for applications of UAV remote sensing using visible spectrum and multispectral imagery. An algorithm has been developed to work on a server for the remote processing of images acquired of a crop field with a UAV. This algorithm first enhances the images to adjust the contrast and then classifies areas of the image based upon the vigor and greenness of the crop. The classification is performed using a support vector machine with a Gaussian kernel, which achieved a classification accuracy of 86.4%. Additionally, an analysis of multispectral imagery was performed to determine indices which correlate with the health of corn crops. Through this process, a method for correcting hyperspectral images for lighting issues was developed. The Normalized Difference Vegetation Index values did not show a significant correlation with the health, but several indices were created from the hyperspectral data. Optimal correlation was achieved by using the reflectance values for 740 nm and 760 nm wavelengths, which produced a correlation coefficient of 0.84 with the yield of corn. In addition to this, two algorithms were created to detect stink bugs on crops with aerial visible spectrum images. The first method used a superpixel segmentation approach and achieved a recognition rate of 93.9%, although the processing time was high. The second method used an approach based upon texture and color and achieved a recognition rate of 95.2% while improving upon the processing speed of the first method. While both methods achieved similar accuracy, the superpixel approach allows for detection from higher altitudes, but this comes at the cost of extra processing time.

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Daniel Scott Whitehurst

GENERAL AUDIENCE ABSTRACT

Crops can experience a variety of issues as they grow, which can reduce the amount of resulting crop. In order to avoid losing their crops and money, it is critical for farmers to detect and treat these issues. The current methods for detecting the issues can be expensive and have slow turnaround time to find the results. Unmanned aerial vehicles (UAVs) have emerged as a potential to improve upon the current methods and reduce the cost and turnaround time for determining issues. The UAVs can use a wide array of sensors to quickly and easily acquire information about the crop field. Using a variety of cameras, data can be gathered from the wavelengths which can be seen by humans as well as many other wavelengths outside of our visible spectrum. The work in this thesis uses images acquired from visible spectrum cameras as well as multispectral data, which uses a different range of wavelengths. A method was created to process the visible spectrum images to classify areas of the field based upon the health of the crop. This method was implemented on a server to allow a farmer to upload their images through the internet and have the data processed remotely. In addition to this, multispectral images were used to analyze the health of corn crops. The multispectral data can be used to create index values based upon various wavelengths of data. Many index values were analyzed and created to find relationships between these values and the health of the crops and strong relationships were found between some of the indices and the crop health. The final portion of this work uses standard visible spectrum images to detect the presence of stink bugs on crops. Two separate methods were created for this detection and both of these methods were able to accurately find stink bugs with a high success rate. The first method was able to detect the stink bugs from farther away than the second method, however the second method was able to perform the detection much faster.

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Nomenclature

ac	Acre
lbs	Pounds (mass)
NDVI	Normalized Difference Vegetation Index
NIR	Near-infrared
nm	Nanometer
SVM	Support Vector Machine
UAV	Unmanned Aerial Vehicle

Chapter 1 Introduction

With increased access to unmanned aircraft that are capable of high-resolution overhead imagery, farmers are now empowered to scout fields with much higher accuracy and coverage area. This image acquisition capability is complemented by the ability to interpret the data so that an appropriate response is taken.

Crops can regularly suffer from a variety of issues, which a farmer will want to address. The issues suffered can include both biotic and abiotic stresses. These may involve diseases, improper treatment with chemicals, misapplication of water, as well as insect problems. The current methods of detecting and resolving these issues are lacking. These methods include manual inspection, whole field treatment with pesticides or fungicides, and photographic inspection. The current manual inspection methods involve the farmers looking through the fields at ground level or from a slightly elevated height. These methods of inspection do not provide proper scope to truly locate issues and also can be manually intensive to see the scope of the field. A current method of photographic inspection involves aerial imagery, generally acquired from a manned aircraft, which can be used for the crop inspection. This method is also manually intensive and the cost for this inspection method is very high, which limits its applicability for most farmers. The other current method is treating large portions of the field based on field-average thresholds instead of targeting treatments to the specific problem areas. Knowing the problem areas to target can help reduce the amount of treatment and save money for the farmer.

The rise in availability and use of unmanned aircrafts can allow a farmer to improve upon the current methods of inspection and treatment. With the use of an unmanned aerial vehicle (UAV), a farmer has the ability to quickly and easily acquire airborne imagery of the field, which can then be used for inspection. The goal of this work is to create tools and methods for farmers to improve the health and yield of their crops from the analysis of images which can be acquired from a UAV. Various types of images can be used to investigate the health of crops. The most widely available and most inexpensive form of imagery is visual spectrum imagery. As a result, the first goal of the work was to create image processing methods to detect crop stresses using solely visual imagery. While it is unlikely that farmers will have access to near-infrared or

hyperspectral cameras, the information from these images can potentially provide a more accurate representation of crop health than the simple visual imagery. As a result, the next goal of this work was to examine the information obtained from multiple spectrums and the indices this data can produce, including known indices and the creation of new indices.

In addition to locating and treating the issues after visual damage has been done to the crops, it would be beneficial to detect some issues, such as the presence of insect pests, before the damage has been done to the crop. This also has potential to be achieved through scouting of a field with a UAV. The final goal of this work was to create methods for the detection of insects in a field from airborne imagery. Specifically, the goal was to detect brown marmorated stink bugs since these are an emerging pest in Virginia, which is the focus area of this work.

The organization of this thesis will be in the following form. Chapter 2 discusses all relevant background information required to understand the methods presented in this thesis as well as existing literature in these areas. The chapter will cover the topics of remote sensing, computer vision, and machine learning. Chapter 3 discusses my method for processing visual imagery to aid the detection of crop stress. This method includes image enhancement as well as an implementation of a machine learning algorithm to classify the problem areas in the field. Chapter 4 presents multispectral analysis for crop health monitoring, including NDVI analysis from visual and near-infrared imagery as well as hyperspectral index analysis using a hyperspectral camera. Chapter 5 covers multiple methods that have been developed to detect stink bugs on vegetation from aerial images. Two different algorithms are presented, while multiple other attempted methods are discussed. Chapter 6 will present a summary of the work in this thesis along with the conclusions which were drawn from this work. Additionally, potential areas of improvement for future work are discussed. This final chapter is then followed with the references which were used during this work.

Chapter 2 Background

This chapter discusses the background information which is necessary to understand the work of this thesis. The three main topics discussed in this section are remote sensing, computer vision, and machine learning.

2.1 Remote Sensing

Remote sensing is observing an object without physically touching the object. In the field of agriculture, remote sensing involves the ability for farmers to observe and acquire information about their fields without physically touching the fields. This generally involves images acquired from satellites and aircraft to provide a means to assess field conditions from a point of view high above the field. Many different sensors can be used for remote sensing, including ones which see in the visible wavelengths of light and others which can detect wavelengths not visible to the human eye. With recent technological advances, remote sensing has become more accessible to most agricultural producers. Images acquired through remote sensing can be used for the identification of diseases, nutrient deficiencies, insect damage, herbicide damage, water deficiencies or surpluses, and many other issues. This information allows farmers to focus treatments on only the affected areas of a field [1].

Using remotely sensed data, indices have been created for the analysis of different features of crop health. Many of these indices rely on data from wavelengths corresponding to the visible and infrared regions of the electromagnetic spectrum. Visible light ranges from wavelengths between around 400 nanometers and 700 nanometers. One portion of the infrared region of the spectrum is near-infrared (NIR) light, which ranges from wavelengths of 700 nm to 1100 nm. Chlorophyll, the pigment in plant leaves, strongly absorbs visible light for photosynthesis while strongly reflecting NIR light. These properties of the plants cause the use of an index known as the Normalized Difference Vegetation Index (NDVI), shown in Equation 2.1 [2].

$$NDVI = \frac{NIR - VIS}{NIR + VIS} \quad (2.1)$$

In this equation, *NIR* represents the spectral reflectance in the near-infrared wavelengths and *VIS* represents the spectral reflectance in the visible (red) wavelengths. Values for the NDVI range

between -1 and $+1$. Areas with high NDVI values, from 0.6 to 0.9, generally correspond to dense vegetation such as crops at their peak growth stages or forests. Sparse vegetation can result in moderate NDVI values, around 0.2 to 0.5, and areas of barren rock, sand, or snow generally result in very low NDVI values, usually 0.1 or less [3]. Many other indices can be created from the use of hyperspectral data, which involves reflectance values at specific wavelengths across a range of wavelengths. A list of crop indices was found in the dissertation work of Pavuluri [4]. From these, two indices were selected for further investigation in this thesis. The two indices were selected because these were the only two which used wavelengths for which data could be accurately acquired using our hyperspectral camera. The first of the indices was a ratio of the reflectance values at 780 nm and 740 nm in the form shown in Equation 2.2.

$$Index = \frac{R_{780}}{R_{740}} \quad (2.2)$$

An additional index taken from that work is known as the Normalized Difference Red Edge index, which uses the reflectance values at 790 nm and 720 nm. The formula for this index is shown in Equation 2.3. Many additional indices are used in remote sensing, but these were not included in the scope of this thesis.

$$Normalized\ Difference\ Red\ Edge = \frac{R_{790} - R_{720}}{R_{790} + R_{720}} \quad (2.3)$$

Current satellite sensors for remote sensing have critical limitations of the lack of imagery with optimum spatial and spectral resolutions as well as undesirable revisit time of the satellites. Manned airborne platforms for remote sensing have the issue of high operational costs. With remote sensing platforms for agriculture, high special resolution and quick turnaround times are necessary for useful results. The use of UAVs with remote sensors provides a potential solution to provide low-cost approaches for meeting the spatial, spectral, and temporal resolution requirements, which was presented in the article by Berni et al. [5]. Additionally, this article explored and validated the capabilities of thermal and narrowband multispectral remote sensing from a UAV to monitor vegetation. The idea of low-altitude remote sensing from a UAV as a potential substitute for satellite precision agriculture systems has also been explored in other works of literature [6] and the use of UAVs for remote sensing applications in agriculture has been investigated in many additional pieces of literature [7-9]. From these investigations, the use

of UAVs for remote sensing and precision agriculture is a very capable and cost-effective option when appropriate sensors are used. This is an emerging and growing technology that will likely have a large impact on the future directions of remote sensing for agriculture.

2.2 Computer Vision

Computer vision is a field which aims to make computers see. The field includes methods for acquiring, processing, analyzing, and understand images and data from the real world to produce information, such as in the form of decisions [10]. Generally, computer vision techniques are developed to replicate the abilities of human vision through the use of a computer processing images. The following will cover the computer vision concepts and techniques which have been utilized in the course of this thesis.

2.2.1 RGB Color Space

The RGB color space is an additive color space which consists of red, green, and blue. This color space is the most commonly used color space and is used by most cameras and for the related applications.

2.2.2 HSV Color Space

The HSV color space is a cylindrical-coordinate representation of points in an RGB color mode. This is one of the two most common cylindrical representations, alongside the HSL color space. The HSV name stands for hue, saturation, and value. The color space was developed for computer graphics applications and is currently used in color pickers, image editing software, image analysis as well as computer vision. The hue component is the angular measure of the color space and it represents a measure of the color. Saturation is the radial component and this represents how colorful a color is relative to its own brightness. The value component is a measure of the brightness [11].

2.2.3 Lab Color Space

The Lab color space is a color-opponent space, with the L component for lightness and a^* and b^* for the color-opponent dimensions. The a^* component represents the position of the color between red/magenta and green and the b^* component represents the position of the color

between yellow and blue. The lightness value ranges between 0 and 100, with 0 representing black and 100 representing white. For a^* and b^* , negative values indicate green and blue and positive values indicate red and yellow, respectively [12].

2.2.4 YIQ Color Space

The YIQ color space is the one that is used by the NTSC color TV system. The Y component represents the luma values, which correspond to the brightness of the image. The I and Q components represent the chrominance information. This color space representation can be used in color image processing to adjust images without altering the color balance. This is done by applying the adjustments only to the Y channel of the images [13]. The formula for converting an RGB image into the YIQ color space is shown by Equation 2.4 [14].

$$\begin{bmatrix} Y \\ I \\ Q \end{bmatrix} = \begin{bmatrix} 0.299 & 0.587 & 0.144 \\ 0.596 & -0.274 & -0.322 \\ 0.211 & -0.523 & 0.312 \end{bmatrix} \begin{bmatrix} R \\ G \\ B \end{bmatrix} \quad (2.4)$$

2.2.5 Superpixel Segmentation

Superpixels are large pixels formed from combinations of many other pixels. This allows redundancy in images to be captured and reduces the complexity of the image for later processing. One method of superpixel segmentation, which is used in this thesis, is the method of SLIC Superpixels. The algorithm for this method is called SLIC (Simple Linear Iterative Clustering) and the method is detailed in the paper [15]. This method uses a five dimensional space containing the L, a, and b values of the CIELAB color space as well as the x and y pixel coordinates and performs local clustering of the pixels based on these five dimensions. The input to this algorithm is the desired number of superpixels K , which would be approximately equal in size. For an image with a size of N pixels, each superpixel is approximately N/K pixels in size. With superpixels which are roughly equally sized, a superpixel center would be located at each grid interval $S = \sqrt{N/K}$. To begin the algorithm, K cluster centers $C_k = [l_k, a_k, b, x_k, y_k]^T$ are chosen for the superpixels with $k = [1, K]$ at regular grid intervals S . Because the approximate area of each of the superpixels is S^2 , it can be assumed that the pixels associated with this cluster center are located in a $2S \times 2S$ area around the superpixel center in the xy plane. As a result, this area becomes the search area for the pixels closest to each cluster center. For the CIELAB color space, Euclidean distances are perceptually meaningful for small distances. To prevent the

spatial pixel distances from outweighing the pixel color similarities, a distance measure D_s was used, which is defined as shown in Equation 2.5.

$$\begin{aligned}
 d_{lab} &= \sqrt{(l_k - l_i)^2 + (a_k - a_i)^2 + (b_k - b_i)^2} \\
 d_{xy} &= \sqrt{(x_k - x_i)^2 + (y_k - y_i)^2} \\
 D_s &= d_{lab} + \frac{m}{S} d_{xy}
 \end{aligned} \tag{2.5}$$

The variable m is included to control the compactness of the superpixels. Greater values of m result in a stronger emphasis on the spatial proximity and creates more compact clusters. This distance measure is used as the metric for determining how well the pixels match with the cluster centers.

The algorithm begins by sampling K regularly spaced cluster centers and then moving them to the locations in a 3 x 3 neighborhood which have the lowest gradients. This reduces the chance of choosing a noisy pixel and avoids placing them at an edge. The gradients in the image are calculated using Equation 2.6.

$$G(x, y) = \|I(x + 1, y) - I(x - 1, y)\|^2 + \|I(x, y + 1) - I(x, y - 1)\|^2 \tag{2.6}$$

In this equation, $I(x, y)$ is the vector of Lab values corresponding to the pixel at location (x, y) , which takes both color and intensity information into account. Each of the pixels in the image become associated with the nearest cluster center whose search area overlaps the pixel. Once all pixels are associated with their nearest cluster center, the average Labxy vector of all pixels in a cluster is determined to be the new cluster center. This process of associating pixels with the nearest cluster center and recalculating the cluster center continues to be repeated iteratively until the cluster centers converge within a threshold. After convergence, the connectivity of the pixel clusters is enforced. If any stray pixel labels exist, the disjoint segments will be relabeled to the labels corresponding to the largest neighboring cluster. The SLIC superpixel segmentation method has been compared to other existing methods and has been shown to outperform existing methods in nearly every respect [16].

2.2.6 Edges and Texture

Two properties of an image which can be very useful in computer vision are the edges in the image and the texture of the image. When determining edges and texture properties of an image,

the use of a filter can be advantageous. A filter is a matrix which is applied to a neighborhood of pixels around a specific pixel. Filters can be used in a variety of tasks which include smoothing, sharpening, enhancing or detecting edges, and examining image texture [17]. Two of the applications for filters which were used in the course of the work in this thesis are for edge detection and texture.

To detect edges in an image, it is necessary to find the sharp changes in the image since these correspond to edges. These changes in the image can be determined through the use of a gradient in the image. The formula in Equation 2.7 gives the gradient of an image [18]. This formula is comprised of the partial derivatives in both the x and y directions.

$$\nabla f = \begin{bmatrix} \frac{\partial f}{\partial x} & \frac{\partial f}{\partial y} \end{bmatrix} \quad (2.7)$$

When performing this gradient calculation on an image, it can be approximated through the use of a filter. One such example of a filter which can be used is the Sobel operator. Equation 2.8 shows the Sobel operators for the horizontal and vertical gradients. These operators can be used to determine the gradients in an image by convolving the operator with an image [19-21].

$$G_x = \begin{bmatrix} -1 & 0 & +1 \\ -2 & 0 & +2 \\ -1 & 0 & +1 \end{bmatrix}, \quad G_y = \begin{bmatrix} +1 & +2 & +1 \\ 0 & 0 & 0 \\ -1 & -2 & -1 \end{bmatrix} \quad (2.8)$$

To examine textures in an image, many different methods exist. The method used for the work in this thesis was the entropy of the image. Entropy is a measure of the amount of disorder in a system and a statistical measure of uncertainty. The entropy can be used to determine how ‘busy’ areas of an image are and the amount of variability in the areas. As a result, this quantity can be used to find differences in the texture of an image. The formula to calculate the entropy is shown in Equation 2.9. This equation is applied to a neighborhood, or window, around each of the pixels in an image.

$$E = - \sum_i p_i \log_2 p_i \quad (2.9)$$

In the equation, E is the entropy and p_i represents the histogram bin counts for the pixel values, which corresponds with the probabilities of those specific pixel values [22-25].

2.2.7 Binary Operations

When processing images, for some applications, a useful technique can be to create a binary image and perform operations on that binary image. A binary image is an image with only two possible values for each pixel, typically 0 and 1. The use of binary images can simplify the methods for processing images in many circumstances. One way of creating a binary image is through the use of a threshold. For image pixel values, or feature values for pixels, a binary image can be produced by comparing the pixel values to a threshold. If the pixel value fits the criteria corresponding to the threshold, the binary pixel is assigned a value of 1, while a value of 0 is assigned otherwise.

One method of processing binary images is through the use of morphological operations. These operations apply a structuring element to an image and the output pixel values are based upon comparing the input pixel values with their neighbors. A structuring element is a mask that can take on various shapes to achieve different results. The mask will have values of 1 in a desired pattern and will be applied to the input image to perform the morphological operations. The most basic operations are dilation and erosion. Dilation is a method for expanding the edges of an object in an image. For dilation, the value of the output pixel value is set to be the maximum value of the pixels in the neighborhood, as defined by the structuring element, of the input pixel. For binary images, if any pixel in the neighborhood is 1, then the output pixel value is set to 1. Erosion is a method to reduce the size of an object in an image by removing edge pixels. For erosion, the output pixel value is set to the minimum value of all pixels in the neighborhood of the input pixel. So, if any pixels in the neighborhood has a value of 0, the output pixel will be set to 0 [26]. Additional morphological operations can be achieved by the combination of the dilation and erosion operations. One such operation is opening, which is achieved by an erosion followed by a dilation. Opening can be used to remove small objects from an image and open gaps between connected objects. These are removed through the erosion process, but any regions which survive the erosion are restored to their original size by the dilation. Another operation is closing, which refers to a dilation followed by an erosion. Closing can be used to fill holes in regions of an image while keeping the same initial region size. A hole refers to pixel values of 0 within an object with pixel values of 1. The dilation operation sets the values in the hole to 1 and the following erosion is used to return the object to its initial external shape [27].

Additionally, for binary images, the region properties of the image can be useful. Regions are areas in the image which contain pixel values of 1 and are connected together by being next to the pixels. Many properties of the regions can be found and used for processing. The region properties which are used in this thesis include area, perimeter, and eccentricity. The area is the number of pixels, with a value equal to 1, contained in the region. The perimeter of a region is the total distance around the boundary of the region. This is determined by the distance between the adjoining pairs of pixels around the region. For determining the eccentricity of a binary region, a built-in Matlab method was used. For this, the eccentricity is found for an ellipse which has the same second-moments as the desired region. Once this ellipse is found, the eccentricity can be found as the ratio of the distance between the foci of the ellipse and its major axis length. This value varies between 0 and 1, with 0 representing a circle and 1 representing a line segment [28].

2.2.8 Local Illumination Adjustment

During the work of thesis, issues were found with illumination in various portions of images. These issues included shadowed as well as glared areas in an image. As a result, local illumination adjustment was required. One existing method which was explored in this thesis is Poisson image editing. For this, the following notation is introduced: “let S , a closed subset of \mathbb{R}^2 , be the image definition domain, and let Ω be a closed subset of S with boundary $\partial\Omega$. Let f^* be a known scalar function defined over S minus the interior of Ω and let f be an unknown scalar function defined over the interior of Ω . Finally, let \mathbf{v} be a vector field defined over Ω ” [29]. The guidance field for the illumination application is defined in the log-domain as shown in Equation 2.10.

$$\mathbf{v} = \alpha^\beta |\nabla f^*|^{-\beta} \nabla f^* \quad (2.10)$$

In this equation, the variable α is 0.2 multiplied by the average gradient norm of f^* over Ω and β is 0.2. For an image, a discrete Poisson solver must be used to find new, adjusted, pixel values. For this, at each pixel p in S , N_p is the set of the 4-connected neighbors for the pixel. Additionally, $\langle p, q \rangle$ is a pixel pair such that $q \in N_p$. For the discrete solver, v_{pq} is the projection of $\mathbf{v} \left(\frac{p+q}{2} \right) \cdot \vec{p}q$. The solution for the discrete Poisson solver satisfies the simultaneous linear equations shown in Equation 2.11 for all $p \in \Omega$ [29].

$$|N_p|f_p - \sum_{q \in N_p \cap \Omega} f_q = \sum_{q \in N_p \cap \partial\Omega} f_q^* + \sum_{q \in N_p} v_{pq} \quad (2.11)$$

This system of equations forms a classical, sparse, symmetric, positive-definite system and due to the arbitrary shape of the boundary, iterative solvers must be used.

2.3 Machine Learning

Machine learning was defined by Arthur Samuel in 1959 as a “Field of study that gives computers the ability to learn without being explicitly programmed” [30]. The field of machine learning explores the construction of algorithms for the use of learning from and making predictions on data. Two of the main areas of machine learning are unsupervised learning and supervised learning, which will be discussed below.

2.3.1 Unsupervised Learning

Unsupervised learning involves inferring information and making decisions about unlabeled data [31]. All information obtained from the data and decisions made about the organization must be made without any information input from a human, since no labels are given for the data. As a result of the unlabeled learning, there is no error or success information which can be used to evaluate the solution method. A common form of unsupervised learning involves clustering. Clustering is commonly used in computer vision applications for grouping together different pixels based upon how similar their features are. One such algorithm that can be used for this is the k-means clustering algorithm. The previously discussed SLIC superpixels algorithm utilized a version of k-means clustering.

2.3.2 Supervised Learning

Another form of machine learning is supervised learning, which uses labeled training data to create the function for predicting future data. In supervised learning, the features of the labeled training data are compared to the features of each new item of test data in order to predict class labels for the test data. Many different methods of supervised learning exist, which include k-nearest neighbors, support vector machines, and many others [32].

Support vector machines have become very popular for machine learning applications due to their great performance and the flexibility from the kernel-based framework. These classifiers

have the ability to achieve high classification accuracy even with only a small amount of training data. SVMs are inherently binary linear classifiers which use a set of labeled training data to build models to assign new examples into one of two classes. The kernel-based framework of an SVM also allows for efficient non-linear classification with the use of the kernel trick. Additionally, multiple SVMs can be combined in order to classify instances into more than two classes, which becomes a multiclass SVM. The algorithm for an SVM will be explained based upon the information from [33] and [34].

The most basic case of an SVM is the 2D binary linear SVM. In this case, the SVM attempts to separate the data into two classes using a hyperplane, which is reduced to a line for this case. In general, the hyperplane is described by Equation 2.12.

$$w \cdot x + b = 0 \tag{2.12}$$

In the equation, x is the input data, w is normal to the hyperplane and $\frac{b}{\|w\|}$ is the perpendicular distance from the hyperplane to the origin. A representation of a support vector machine can be seen in Figure 2.1. The data in the example in the figure is linearly separable between the two classes. The hyperplane is represented by the solid line between the two classes and the support vectors are represented by the dashed line, labeled H_1 and H_2 . The margins, or distances between the hyperplane and the support vectors, are represented in the figure by d_1 and d_2 . The overall margin of this support vector machine is the distance between the support vectors, or the sum of the individual margins.

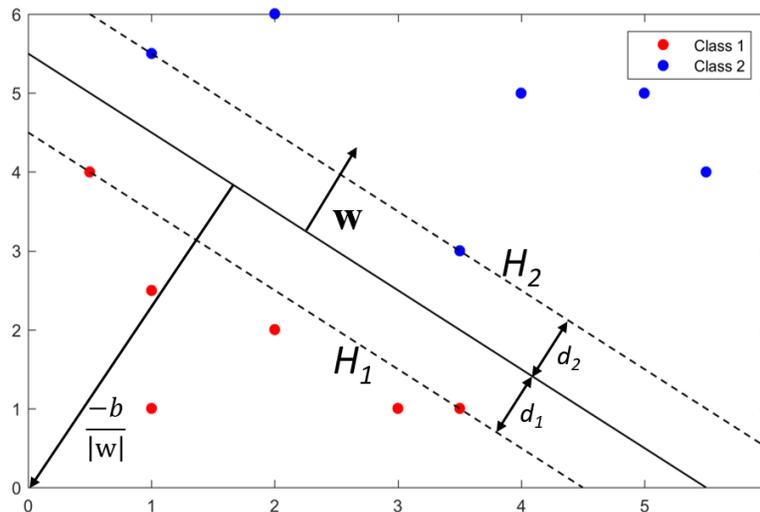


Figure 2.1: Representation of a support vector machine.

A mathematical representation of the two classes is shown by the pair of equations in Equation 2.13. As a result, implementing the SVM requires the selection of the variable w and b such that the conditions of these equations are met.

$$\begin{aligned} x_i \cdot w + b &\geq +1 \text{ for } y_i = +1 \text{ (class 1)} \\ x_i \cdot w + b &\leq -1 \text{ for } y_i = -1 \text{ (class 2)} \end{aligned} \quad (2.13)$$

When considering the points of the classes which are closest to the hyperplanes, also known as the support vectors, the plane connecting these points is represented by Equation 2.14. The equation for the line H_1 from Figure 2.1 uses $+1$ and the line for H_2 uses -1 .

$$x_i \cdot w + b = \pm 1 \quad (2.14)$$

In this model of an SVM, the distance D between any data point and the hyperplane is found through the use of Equation 2.15.

$$D = \frac{|x_i \cdot w + b|}{\|w\|} \quad (2.15)$$

Using the previous equation, the margins for each of the support vectors as well as the overall margin for the SVM can be found. The margins for each of the support vectors, d_i , can be found by substituting Equation 2.14 into Equation 2.15. The overall margin of the support vector machine can be found from combining the margins of each support vectors. The equations for these margins are shown in Equation 2.16.

$$d_i = \frac{1}{\|w\|}, \quad Margin = \frac{2}{\|w\|} \quad (2.16)$$

When determining the functions to represent an SVM, the values for w and b must be solved for. The goal when creating the SVM is to orient the hyperplane in such a way that it is located as far away from the support vectors as possible. In order to this, the margin must be maximized. Maximizing the margin creates a quadratic optimization problem and the solution for the variable w is shown in Equation 2.17.

$$w = \sum_{i=1}^L \alpha_i y_i x_i \quad (2.17)$$

For this equation, α has been introduced as a weight variable. When using the SVM in applications, the classifier function in Equation 2.18 is used. The use of the sign function for this classifier results in output label values of -1 or $+1$, which correspond to the label values for each of the classes.

$$f(x) = \text{sign}(w \cdot x + b) \quad (2.18)$$

The previous description works well for data which is linear separable, but this is not the case for many applications. In many cases, the classes cannot be separated using a simple linear classifier. An example of such data is shown in Figure 2.2. With this data, a line will not be able to separate the two classes, so a nonlinear hyperplane and support vectors would be required.

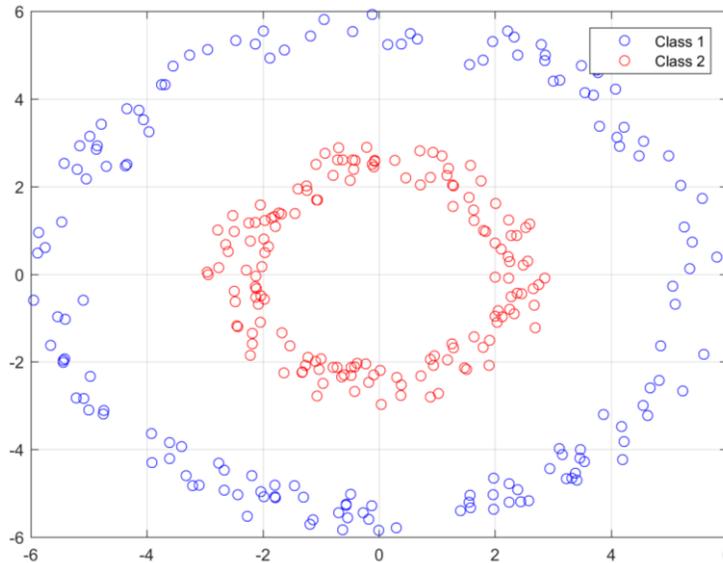


Figure 2.2: Data which is not separable with a linear hyperplane.

A technique which can be used with SVMs to handle the classification of data which is not linearly separable is the kernel trick. The feature vectors for the data can be mapped into higher dimensional space using some mapping function $x \rightarrow \phi(x)$. Applying this mapping to the linear classifier function produces the new classifier function in Equation 2.19.

$$f(x) = \text{sign}\left(\sum_{i=1}^L \alpha_i y_i x_i \cdot x + b\right) \rightarrow f(x) = \text{sign}\left(\sum_{i=1}^L \alpha_i y_i \phi(x_i) \cdot \phi(x) + b\right) \quad (2.19)$$

For the new mapping, the Kernel Function can be defined as shown in Equation 2.20.

$$K(x_i, x) = \phi(x_i) \cdot \phi(x) \quad (2.20)$$

Using this definition of the Kernel Function, the classifier function can now be written in the form of Equation 2.21.

$$f(x) = \text{sign}\left(\sum_{i=1}^L \alpha_i y_i K(x_i, x) + b\right) \quad (2.21)$$

One example of a kernel that can be used is the radial basis function (RBF) kernel, which is also known as the Gaussian kernel. The formula of this kernel is shown below in Equation 2.22. The value for this kernel can range between 0 and 1 and decreases with the distance. Additionally, there are an infinite number of dimensions in the feature space of this kernel.

$$K_{RBF}(x_i, x) = e^{-\left(\frac{\|x_i - x\|^2}{2\sigma^2}\right)} \quad (2.22)$$

Although the feature space of the kernel has an infinite number of dimensions, Figure 2.3 is included to give a visual representation of the impact of the kernel function. The figure shows the data from Figure 2.2 after the RBF kernel has been applied to the data. This data from the feature space was summed to produce the 3-dimensional plot. After the kernel, the data is clearly separable with a hyperplane, which was not the case with the original data.

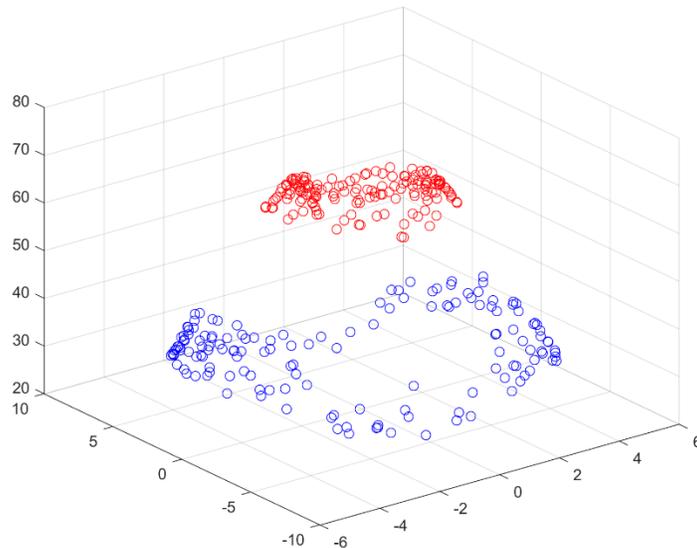


Figure 2.3: 3-dimensional representation of the impact of the RBF kernel on the data from Figure 2.2.

Although SVMs are inherently binary classifiers, their use can be extended to perform multi-class classification. Two primary methods of performing multi-class classification with SVMs are the “one vs. all” method and the “one vs. one” method. For the one vs. all method, an SVM is trained for each of the classes against all of the other data. To do this, the training data for one class is set to be the first class and all other training data, from each of the rest of the classes, is set to be the second class in this model. To perform the test stage on a test example, each SVM is applied to the test example and the class assignment is selected as the class of the SVM which returned the largest decision value. The one vs. all method requires the use of K SVMs, where K is the number of classes in the model. The second method is the one vs. one method, which requires $K(K - 1)/2$ different SVM classifiers. The implementation of this classifier requires creating an SVM for each possible pair of classes. During the test stage, each of these SVMs classifies the test example between two classes and each of these is used as a vote for which class to assign the test example. The class which receives the most votes for test example becomes the classification of that test example [35, 36].

2.3.3 Machine Learning for Crop Classification

Some examples of using machine learning for crop classification exist in the literature. Those examples used machine learning to classify the types of crops, but not the health of those crops. One method used hyperspectral data and a support vector machine for the classification. This was used to classify six different classes, which included corn, sugar beet, barley, wheat, alfalfa, and soil. Each of these classes achieved recognition rates of greater than 90 percent [37].

Another example also used hyperspectral data for separating types of crops. For this, the five crop species examined were soybean, canola, wheat, oat, and barley. This method used a discriminant function analysis approach to classify the objects based on a measure of generalized square distance. High classification accuracies were achieved throughout the growing season for this approach [38].

Chapter 3 Visual Crop Stress Detection

This chapter discusses a method for crop stress detection through the use of visible spectrum imagery. This would enable a farmer to investigate their field for health issues by flying a low-cost UAV, with a standard camera, over the crops.

3.1 Dataset

A large set of aerial crop images was provided by Tim Sexton, who works for the Department of Conservation and Recreation in Virginia. The images were taken by Tim during flights he made over fields. When farmers want their fields investigated for potential issues, Tim will be flown over the fields to take images out of a helicopter. Images of various fields from many different flights became the basis for the dataset which was used for my algorithms. The total database consists of 144 images of fields with crops in various states of health. The images encompass a wide array of crop stress issues as well as different lighting conditions. Two images from the data are shown below in Figures 3.1 and 3.2. The image in Figure 3.1 was annotated by Tim before being delivered. The image in Figure 3.2 is another corn field with some stressed areas throughout.



Figure 3.1: An example of an image from the dataset, with an annotation provided.



Figure 3.2: This is an example of an unlabeled image from the dataset showing corn with multiple stressed areas.

3.2 Image Enhancement

Many conditions can impact the quality of aerial images and the visibility of the crops in the images. These can include the lighting conditions as well as atmospheric conditions such as haze. In order to mitigate the impact of these conditions, normalize the images, and improve the contrast between differences in crops, an image enhancement method was employed to initially process the images. With the images captured from the helicopter, this enhancement is currently performed manually to allow differences in crop health to be more visually obvious. The image enhancement algorithm will automate that process.

The image enhancement algorithm has two main parts: contrast adjustment and lightness adjustment. The contrast adjustment allows variations within the crop to be seen more easily, while the lightness adjustment mitigates the atmospheric impacts and normalizes the image brightness levels.

To adjust the contrast of the images, an auto contrast adjustment algorithm was developed as an adaptation to a similar method [39]. For this, an upper and lower limit are set for the percentile of

the values that the data will be scaled to. For this implementation, values of 0.005 and 0.995 were set for the low and high limits, respectively. The R, G, and B components of the image are then sorted and the minimum and maximum RGB values are selected corresponding to the location of the high and low limits multiplied by the number of pixels. The high and low RGB pairs will then be converted to the YIQ color space for the Y, or luma, component to be used. The Y component for the high and low pairs will then be used to rescale the image as a percentage of these maximum and minimum values. The rescaling is done by Equation 3.1.

$$I_{m_{adjusted}} = \frac{I_m - Y_{min}}{Y_{max} - Y_{min}} \quad (3.1)$$

In this equation, $I_{m_{adjusted}}$ is the output image, I_m is the input image, Y_{min} is the low limit luma component and Y_{max} is the high limit luma component. The output image will have values that range from 0 to 1, so it must be multiplied by 255 to rescale the RGB values for the output.

To correct and normalize the brightness of the image, it is first converted into the Lab color space. The L, or lightness, component is extracted and used for further processing. Through analysis of manually enhanced images, the optimum median image lightness level was found to be around a value of 40. As a result of this finding, the brightness correction algorithm was designed to adjust the median lightness component of the image to be equal to a value of 40 by linearly shifting the channel values. The results of the image enhancement algorithm can be seen in the following examples. The original image in Figure 3.3 was dark and the enhancement was able to correct for that, while the image in Figure 3.4 was too bright which the enhancement was also able to appropriately correct.



Figure 3.3: An image shown before (left) and after (right) the enhancement algorithm has been applied.



Figure 3.4: An image shown before (left) and after (right) the enhancement algorithm has been applied

3.3 Machine Learning Approach

Once the images have been enhanced and normalized, the next step is to classify the areas of the images which are stressed and the areas which are healthy. In order to achieve the classification, a machine learning approach was used. Specifically, this method utilizes a support vector machine to perform the classification. For the classification, three classes are being used. The first class represents healthy crop areas. The second class represents potential problem areas in the field, which would be stressed areas to further examine in order to resolve problems. The third class contains dirt and dead areas of the fields, which would be issues that could not be resolved.

In the early stages of the development of the classification, the amount of training data was very limited, so this limited the options for the classifier selection. A k-Nearest Neighbors classifier would not have been a good option due to the lack of training data since that classifier generally requires large amounts of training data to perform well. On the other hand, a support vector machine is a classifier that has the ability to work well with very limited training data. Additionally, the Kernel-based framework of the SVM makes this classifier very powerful as well as flexible.

The first step for creating the support vector machine is to train the classifier. This requires labeling areas of the images in the dataset as one of the three classes. All images in the dataset were labeled using LabelMe [40]. Appropriate areas in the images were selected to correspond with the correct class. Figure 3.5 shows one of the training images being labeled in LabelMe.

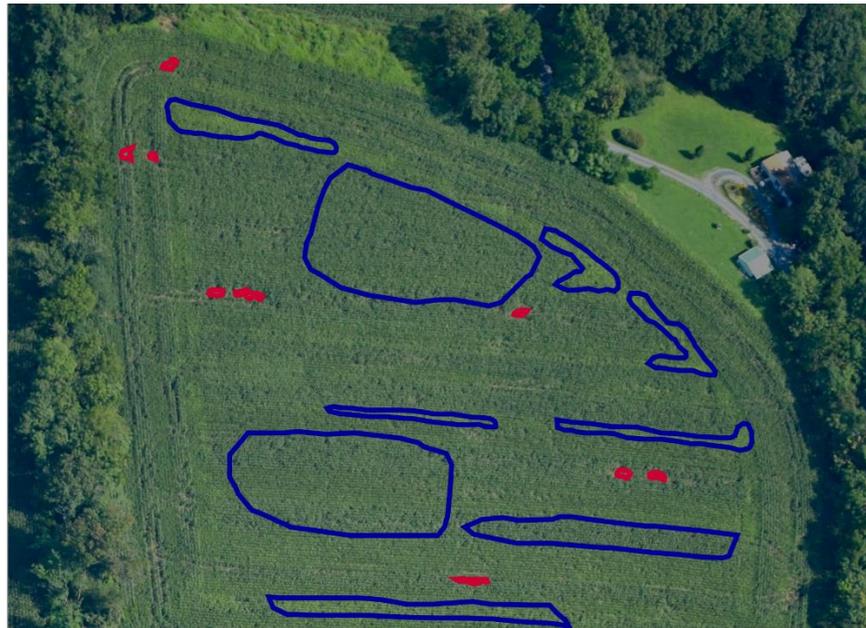


Figure 3.5: Training image labeled using LabelMe.

Once the selected areas of the image have been labeled, features can be extracted from the image in order to train the support vector machine. The features that have been selected for the support vector machine lie solely in the color aspects of the images. Multiple color spaces have been looked at, including RGB, HSV, and Lab. The decision to only use the color aspects of the image was made due to limitations in the dataset. Different features, such as texture, would also be very useful for detecting stresses and issues in the crops, however many images in the dataset did not allow for the use of texture. This was due to a lack of the required resolution to analyze the texture of the crops and to see rows of the crops.

The potential features within the color aspects of the image come from RGB, HSV, and Lab color spaces, which produces nine potential features: red, green, blue, hue, saturation, value, lightness, a^* and b^* . To select the best features to use, classifiers were trained using the different combinations of these nine features and cross validation was performed to select the best combination of features. For the cross validation, a subset of 10 percent of the training data was

held out as the test data set and the remaining 90 percent of the training data was used for training these classifiers. Using these data sets, each combination of features was tested for classification accuracy. The test began by using each feature individually, then using every combination of two, followed by three, and so on. The initial classifier used was a linear support vector machine, but other methods were tested in an attempt to improve upon the accuracy. A second method that will be discussed is a Gaussian Kernel for the support vector machine. These SVMs were implemented in Matlab through the use of the built-in function. This method uses the one vs. one approach for the multiclass SVM. The test was first run on the linear support vector machine using 10-fold cross validation and the optimal features for each step of the test are summarized in Table 3.1.

Table 3.1: Linear SVM Optimal Features

Feature	Classification Accuracy (%)
Hue	80.18
Hue & a*	82.95
Hue, a* & Red	84.72
Hue, a*, Red & Lightness	85.06
Hue, a*, Red, Lightness & Green	85.29

Once the optimal features were determined using the linear SVM, a performance comparison was run against an SVM with a Gaussian Kernel. This comparison was performed using a single subset of data as the test set for cross validation rather than the 10-fold method. This was done to compare the classification performance as well as computation time. The results are summarized in Table 3.2.

Table 3.2: Comparison Between Linear SVM and SVM with Gaussian Kernel

Features	Linear Classifier		Gaussian Kernel	
	Classification Accuracy (%)	Computation Time (s)	Classification Accuracy (%)	Computation Time (s)
Hue	80.04	21.37	83.24	28.33
Hue & a*	82.33	539.41	83.71	27.18
Hue, a* & Red	82.89	1373.66	85.20	23.36
Hue, a*, Red & Lightness	84.13	1311.34	85.27	27.16
Hue, a*, Red, Lightness & Green	84.27	1315.27	85.48	28.74

Due to the Gaussian Kernel performing much better than the Linear SVM, the original optimal feature selection method, using 10-fold cross validation, was performed using the SVM with a Gaussian Kernel. The results of the test and final feature selection are summarized in Table 3.3.

Table 3.3: Optimal Features for SVM with Gaussian Kernel

Feature	Classification Accuracy (%)
Hue	82.962
Hue & Green	84.512
Hue, Green & Saturation	85.776
Hue, Green, Saturation & a*	86.062
Hue, Green, Saturation, a* & b*	86.127
Hue, Green, Saturation, a*, b* & Blue	86.157
Hue, Green, Saturation, a*, b*, Blue & Value	86.160

The classification accuracy from Table 3.3 is plotted against the number of features used for the SVM in Figure 3.6. When the number of features used is very small, the addition of a single feature can create a large increase in the classification accuracy. As the number of features increases, the impact of adding features becomes much less significant. After a value of five features is reached, the classification accuracy completely levels off and the gains in accuracy from adding a feature are very small. Additionally, the addition of features to the algorithm increases the complexity of the algorithm, which also can increase the required processing time. Since the accuracy improvement is not significant after the addition of the fifth feature, the added complexity of additional features is not justified. As a result, the final SVM uses only five features, which are Green, Hue, Saturation, a*, and b*.

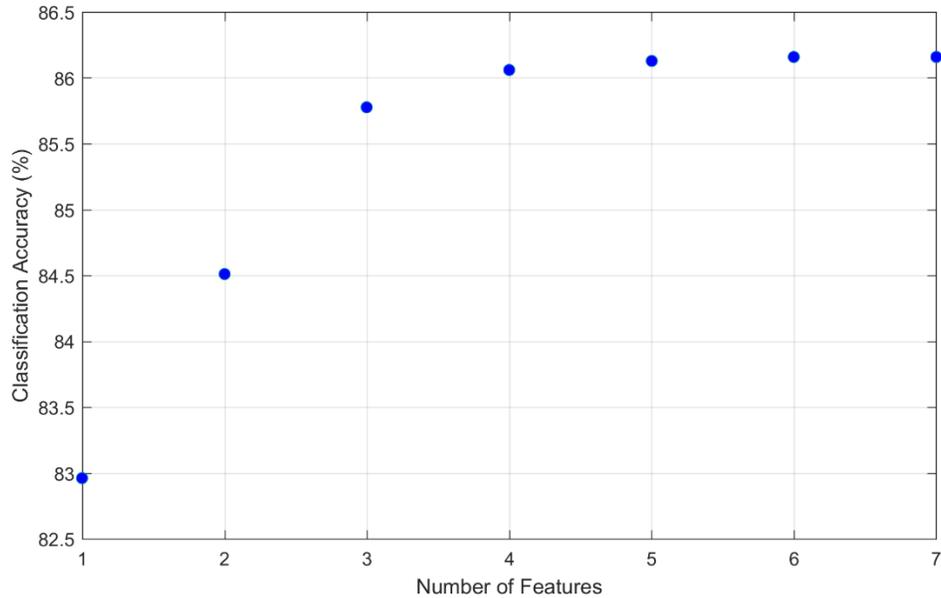


Figure 3.6: The classification accuracy of the SVM is plotted against the number of features used to train the SVM. The accuracy can be seen to level off after 5 features.

3.4 Results

Once the final SVM implementation was selected, the performance was examined with various testing data sets. To obtain a quantitative measure of the classification performance, a testing set of the labeled data was classified using the SVM. The labels from the classification were compared to the actual labels to determine the accuracy. The classification accuracy for this set of data was 86.43 percent, which is similar but slightly better than the classification accuracy during the feature selection process. For a visual understand of the accuracy of the SVM, multiple field images were used to test the SVM. Examples of this can be seen in Figures 3.7, 3.8, and 3.9. The original images are shown on the left and the classification results are shown on the right. In the classification, green corresponds with healthy crop, yellow represents potential problem areas in the field, and red shows areas classified as dirt or not being crops. The classification appears to work well for areas containing the field of crops, but does not perform as well when dealing with non-crop areas such as trees, grass, or some other objects. However, these limitations are not an issue for the farmer since these areas could be easily identified as not being located in the field. The classification performs well, but still has potential for future improvement.

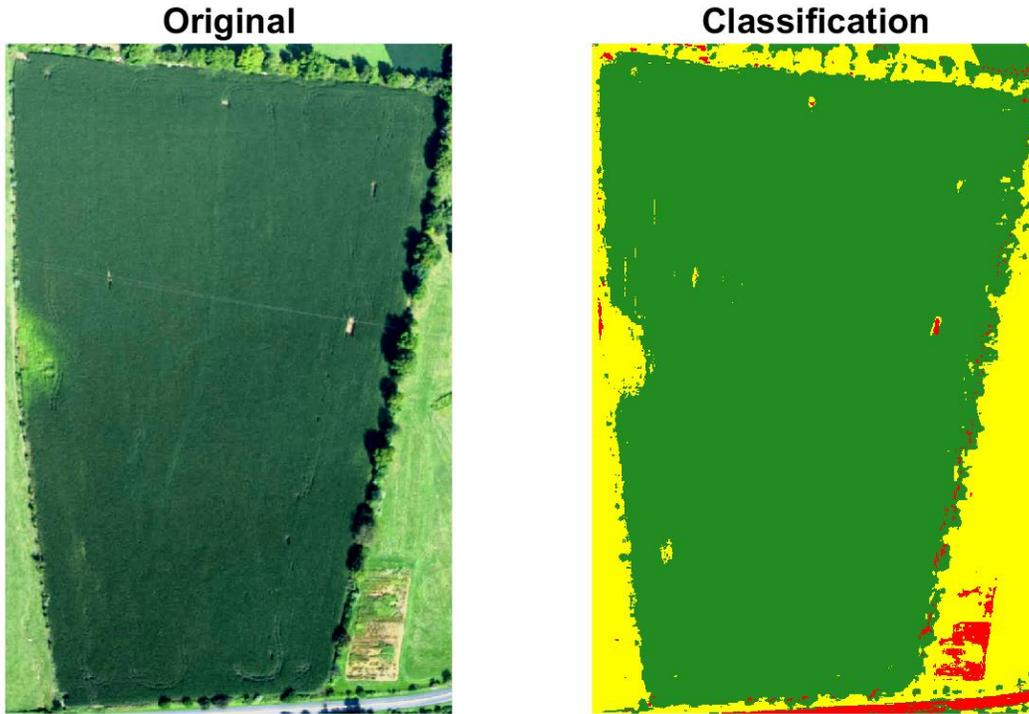


Figure 3.7: Classification results from the support vector machine. The image of the field is shown on the left and the right shows the classification results. The green corresponds to the healthy class, the yellow is the class for potential problem areas, and the red is the class for dirt.

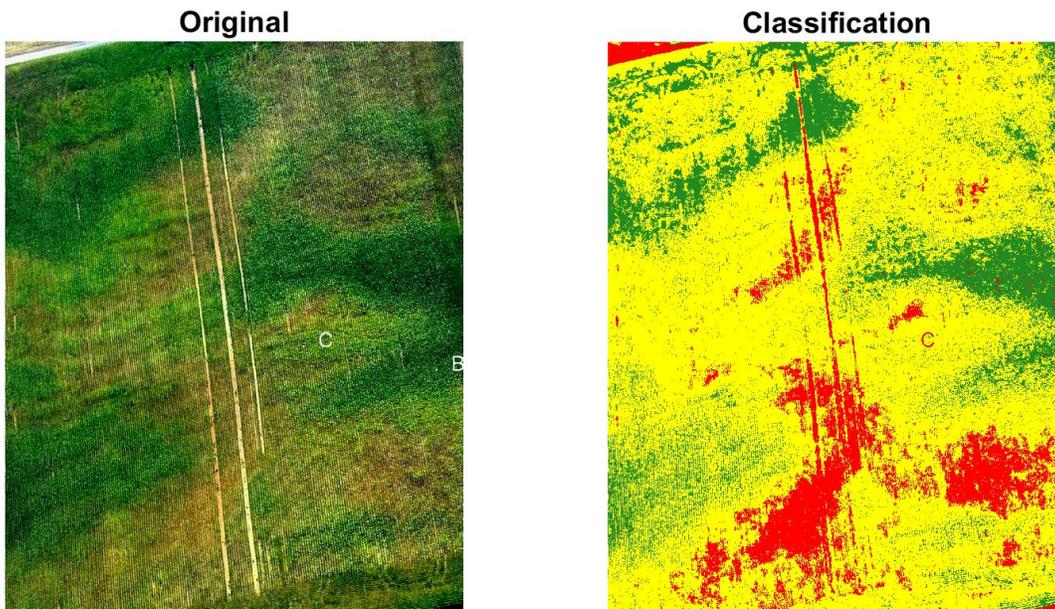


Figure 3.8: Classification results from the SVM for an image of a field. The image of the field is shown on the left and the classification is shown on the right. This field can be seen to have major issues with the crop health.

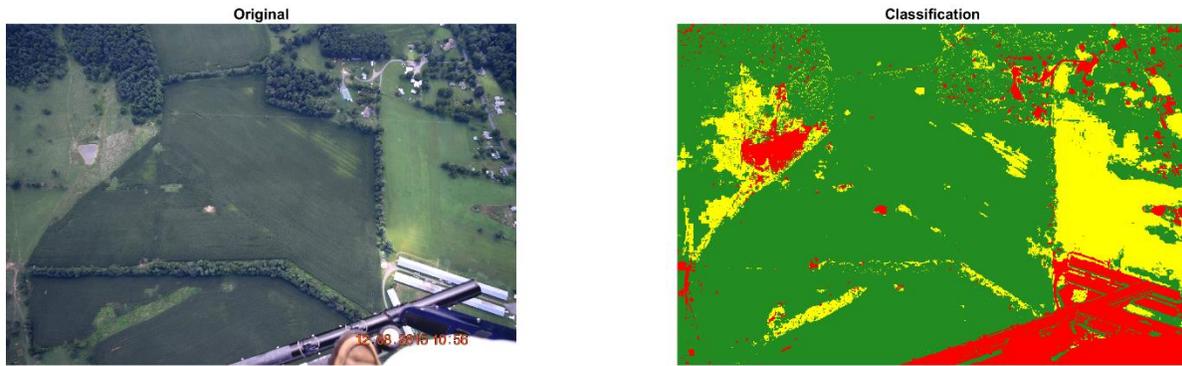


Figure 3.9: Classification results from the SVM for an image of a field. The image of the field is shown on the left and the classification is shown on the right. This image shows a broad view of field, which also contains areas not corresponding to crops.

3.5 Server

To enable the automatic processing of images taken by farmers, a server was created in the Unmanned Systems Lab. The server that was created is an FTP server with the address of <ftp://198.82.189.173>. Access to this server is restricted to people who have an account. Access can be granted for a farmer by contacting a server administrator and providing a login username and password. When a farmer requests access to the server, a new folder will be created. The farmer will only have access to this one folder and all subfolders. The subfolders within are Upload, Original, and Enhanced. The farmer using the service will upload the desired images to the Upload folder and will receive the processed results within an hour. The server is set up to check all of the Upload folders for new images and when new images are found, the images will be copied to the Original folder and processed. The processed results will be placed in the Enhanced folder and the processed images will be deleted from the Upload folder. The server processing capabilities include the algorithms discussed in this chapter. The images will first be enhanced using the previously described image enhancement method and then the enhanced image will be classified using the SVM to determine potential problem areas in the field. Both the enhanced image and the classification result will be placed into the Enhanced folder. The farmer will then be able to use these results to analyze the health of the field and determine any necessary steps for reducing issues. In the case of an error during the processing, an error log output will be created in the Enhanced folder. This log will mention possibilities for the issue. Some issues can be solved by uploading a new image, but other issues may require contacting a server administrator.

Chapter 4 Multispectral Analysis

This chapter discusses the analysis of crops through the use of multispectral data. While this technology is not as readily available for an average farmer, it has a large amount of potential in the field of remote sensing.

4.1 Hardware and Data Collection

4.1.1 Cameras

For the multispectral crop analysis, three types of cameras were used: a visible spectrum camera, a near-infrared spectrum camera, and a hyperspectral camera. Canon S100 cameras are used as both the visible spectrum and near-infrared spectrum cameras. For the near-infrared version, the filter which blocks the infrared wavelengths has been removed and replaced with a blue filter. The filter blocks the red light and captures the near-infrared spectrum data in the red channel of the recorded image. The Canon S100 cameras have a sensor resolution of 12.1 megapixels and produces images with a resolution of 12 megapixels. An image of one of these cameras is shown below in Figure 4.1 [41].



Figure 4.1: Canon S100 camera, which is used as a visual camera as well as the near-infrared camera.

The hyperspectral camera is a BaySpec OCI-UAV-2000 hyperspectral camera. The hyperspectral camera captures an image from 25 wavelength bands between 694.56 nanometers to 967.15 nanometers. The wavelengths of each of the 25 spectral bands are listed as follows: 694.556, 708.894, 732.992, 746.892, 758.992, 772.236, 784.373, 797.516, 808.470, 821.876,

839.886, 851.014, 860.955, 872.309, 881.561, 891.523, 900.802, 910.270, 925.705, 933.048, 941.019, 947.076, 956.318, 962.312, and 967.148 nanometers. The camera has a 16-degree field of view lens and is very compact, weighing only 218 grams. Images can be recorded very fast, up to 120 frames/second and are saved to a computer through a USB 3.0 connection. The exposure time can be set between 1 and 300 milliseconds and is determined based on the lighting conditions and the camera is referenced using a 95% white reference paper. This hyperspectral camera can be seen, with dimensions, in Figure 4.2 [42].



Figure 4.2: OCI-UAV-2000 hyperspectral camera that is being used.

The imaging payload combines the three cameras that have been discussed. The three cameras are mounted side by side to a piece of carbon fiber. Along with the cameras is a fit-PC to allow for interfacing with the hyperspectral camera through a USB 3.0 connection. The hyperspectral camera can be externally triggered by sending a voltage between 15 and 24 volts to the camera. The two Canon S100 cameras were also triggered by a voltage signal and the two cameras were synchronized through the use of Canon Hack Development Kit [43]. The voltage signals for triggering the cameras were controlled by an Arduino, which was set up to be either triggered by a switch or for continuous image capturing, depending on the intended mode.

4.1.2 UAV Platform

One method of acquiring aerial images with the camera payload is through the use of a UAV platform. The UAV platform used for acquiring images is the DJI S1000+ octocopter. This UAV was selected due to its ability to carry the weight of the required payload and have sufficient

flight time to cover a field of over an acre. An image of this octocopter is shown in Figure 4.3 [44] without any of the payload attached to it.



Figure 4.3: DJI S1000+ Octocopter used with the camera payload.

The entire payload included the three cameras, a fit-PC, an Arduino for triggering, as well as a Ubiquiti Bullet to allow wireless connectivity to the fit-PC. This payload was mounted to the octocopter using 3D printed components to interface with the bottom of the UAV. Figure 4.4 shows the entire UAV system with all of the hardware mounted to it. The image was taken during a test flight of the system at Kentland Farm.



Figure 4.4: Our octocopter flying at Kentland Farm with all electronics and camera payload attached.

During a test flight of the octocopter with the imaging payload at Kentland Farm, images were taken by continuously triggering the cameras for the duration of the flight. A set of simultaneously captured images is shown in Figure 4.5. The visual and near-infrared cameras have the same resolution and field of view, so the two match, however the hyperspectral camera can be seen to have a much smaller field of view than the other two.



Figure 4.5: Set of three images taken simultaneously by each of the cameras during the flight at Kentland Farm. It can be seen that the field of view for the hyperspectral camera is much smaller than for the other two cameras.

4.1.3 Imaging Pole

Due to difficulties with the ability to set up frequent flights, many images acquired for the purposes of this investigation were acquired using a 25-foot pole. The same camera payload was utilized for the images taken from the pole, however the fit-PC was not required due to the ability to use a laptop at the base of the pole, which could be connected to the hyperspectral camera using a USB 3.0 cable. The three cameras could then be triggered using a switch rather than the continuous triggering required for the flight. Figure 4.6 shows both the pole and the camera payload attached to the pole. The left image shows the pole, which shows the height relative to the corn crops that were being used for the study. The right image shows the camera payload attached to the pole and the wires can be seen running down the pole to attach with the laptop on the ground.



Figure 4.6: The height of the pole, relative to the corn, can be seen on the left and the imaging payload attached to the pole is shown on the right.

4.1.4 Corn Test Plots

For the purposes of the multispectral crop analysis, plots of corn with various sidedress nitrogen application rates were used as the test plots. The entire set of crops for the test plots contained five different nitrogen applications: 0, 40, 80, 120, and 160 lbs/acre. Each of these treatments were repeated four different times, for a total of 20 different test plots of corn. Each individual test plot consisted of four rows of corn, which were 30 inches wide and 25 feet long. When images were acquired of these plots, the cameras were aimed towards the center of the plots to reduce any border effects of the treatments. The main plots that we acquired images of for the analysis were plots 101, 102, 103, 104, and 105. The entire area of test corn test plots, with labels for the plot number and the nitrogen application can be seen in Figure 4.7. This image was taken during a flight over the field with a UAV.

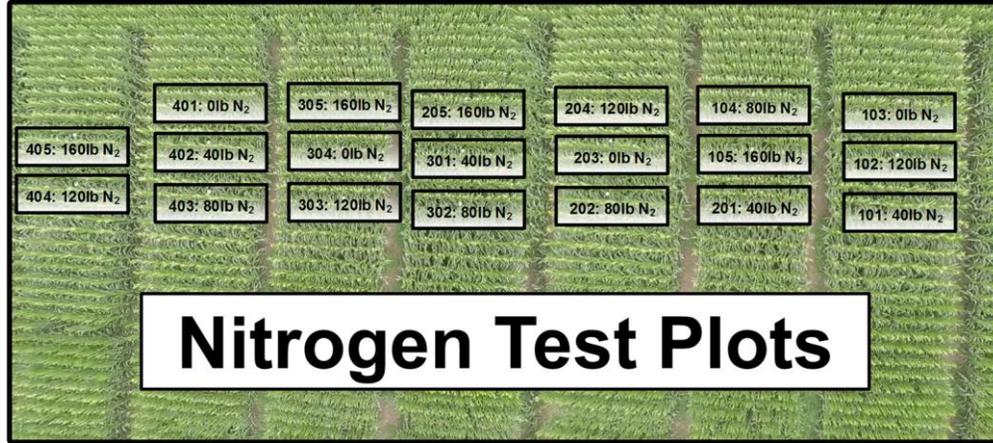


Figure 4.7: Aerial view of the nitrogen rate test plots used to acquire images of for analysis. Each plot is labeled with the plot number as well as the sidedress nitrogen application rate in lbs/acre.

4.2 NDVI Analysis

Using the data acquired, the visible spectrum images and NIR images were combined together to perform an analysis on the NDVI results.

4.2.1 Lighting Impact

From the images acquired of the test plots, the portions of the visual and near-infrared images, which corresponded with the field of the view of the hyperspectral images, were extracted. These visual and near-infrared images were used to calculate the NDVI values for the plots. Figure 4.8 shows a visual and near-infrared image of test plot 105, along with the calculated NDVI values of the plot. From this NDVI calculation, an issue was discovered with image lightness, particularly the areas of the image with shadows and glares. The NDVI calculations produced inaccurate results, sometimes with large variations in NDVI across a single leaf, when lighting changes occurred. Additionally, when a shadow occurred over an area of soil, the NDVI calculation produced a value very close to 1. The shadowed portions of the visual image produce a Red value very close to zero, which creates an NDVI value of 1, as shown in Equation 4.1.

$$NDVI = \frac{NIR - Red}{NIR + Red} = \frac{NIR - 0}{NIR + 0} = 1 \quad (4.1)$$

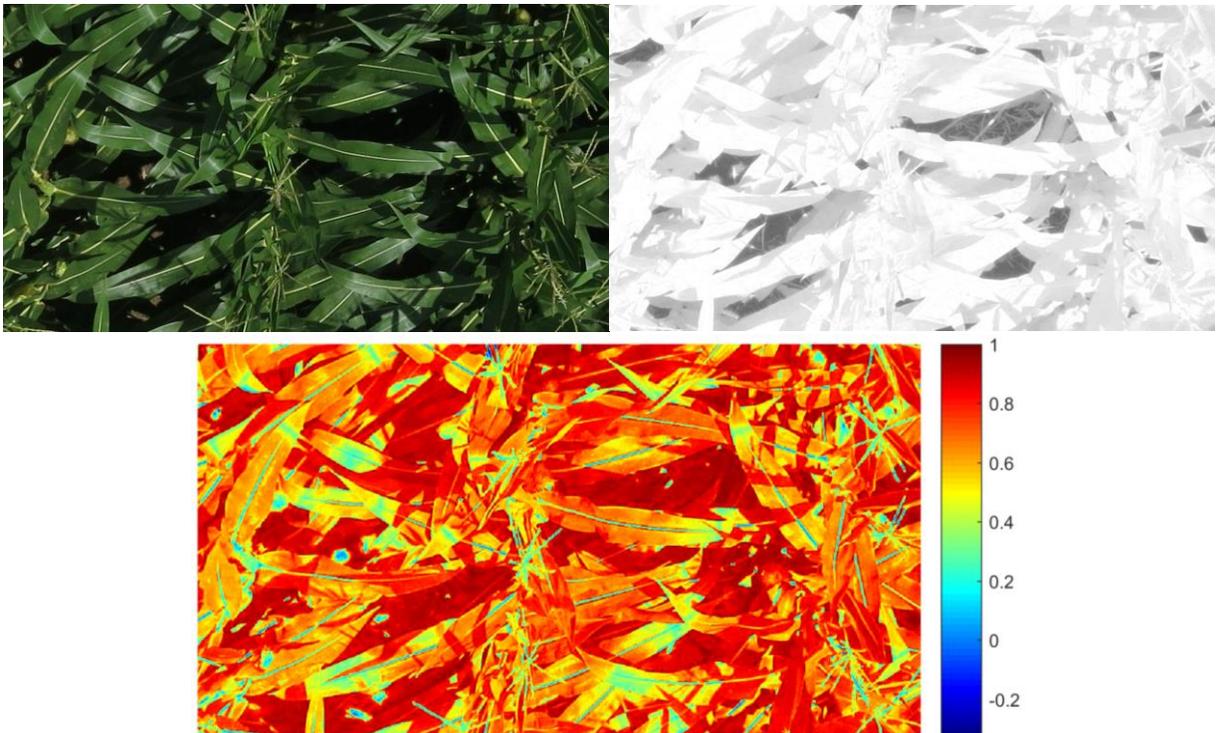


Figure 4.8: Images of corn test plot 105. The top left image is the visual image, the near-infrared image is in the top right, and the bottom image is the NDVI of the plot.

To quantify the impact of the image lighting conditions, such as shadow, a regression analysis was run using the NDVI values and the corresponding Lightness values, from the Lab plane. The plot of the NDVI and Lightness values is shown in Figure 4.9. The two variables were found to have a linear correlation with a coefficient of determination, R^2 , of 0.9488. The regression analysis confirms the lighting conditions have a strong correlation to the calculated NDVI and the trend matches with the previously mentioned errors. The lower lightness values, shadowed areas, corresponded with high values of NDVI and the high lightness values, areas with glare, corresponded with lower values of NDVI. As a result of this trend, to produce more accurate NDVI data, the images should be adjusted to reduce the impact of the lighting conditions.

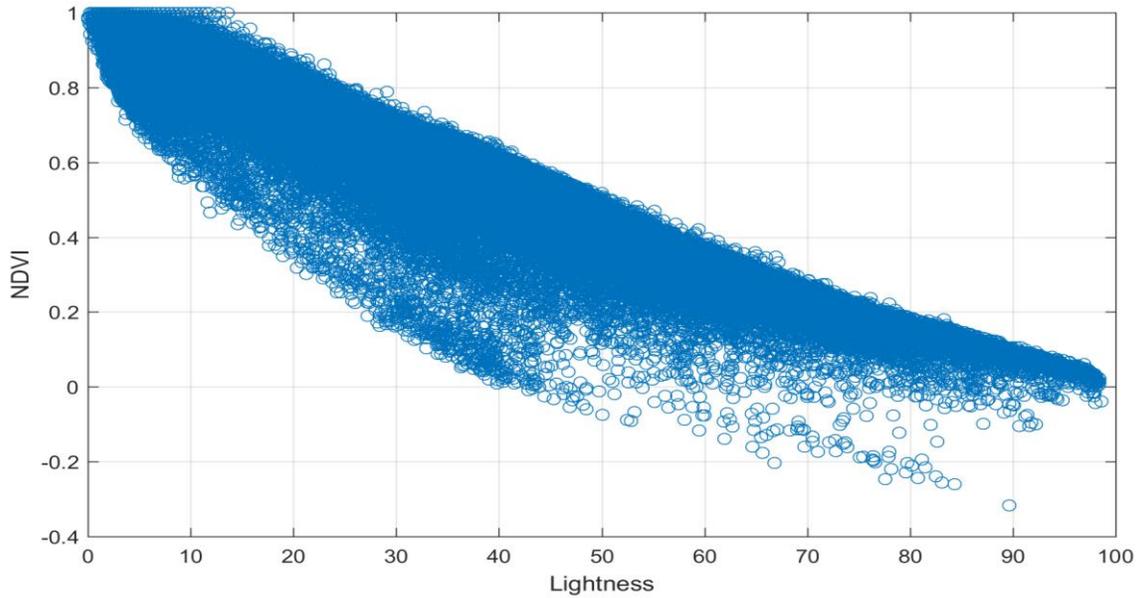


Figure 4.9: Plot of NDVI and Lightness of the images shown in Figure 4.8. A regression analysis produced a coefficient of determination of 0.9488.

4.2.2 Lightness Correction

Due to the correlation between the NDVI values and the lightness values of the images, the method for correcting the NDVI values was to normalize and adjust the lightness of the visual image. The first attempt to adjust the lightness was through the use of Poisson image editing, since this can adjust local illumination. The method for performing this was explained in Section 2.2.8. This method allows for the selection of specific areas for which the illumination adjustment is desired. This method was implemented in Matlab and tested on various images to see the performance. One result of this implementation is shown below in Figure 4.10. The image in the top left is the original image and the top right shows the NDVI calculation before the adjustment, with scaled color corresponding to the color bar on the right. The bottom left image is the image after the Poisson editing was performed and the bottom right shows the new NDVI values after the editing. This method did not perform very well in correcting the lightness for the NDVI. The adjustment changed the lightness, but the new image was slightly blurred and the extreme values, such as the shadows, were not actually removed. As a result, the NDVI calculation did not show any noticeable improvement. The Poisson image editing was also tested on a large portion of an image and was found to perform worse on a large scale. When applying to most of an image, the adjustment blurred the image into a very dark blob.

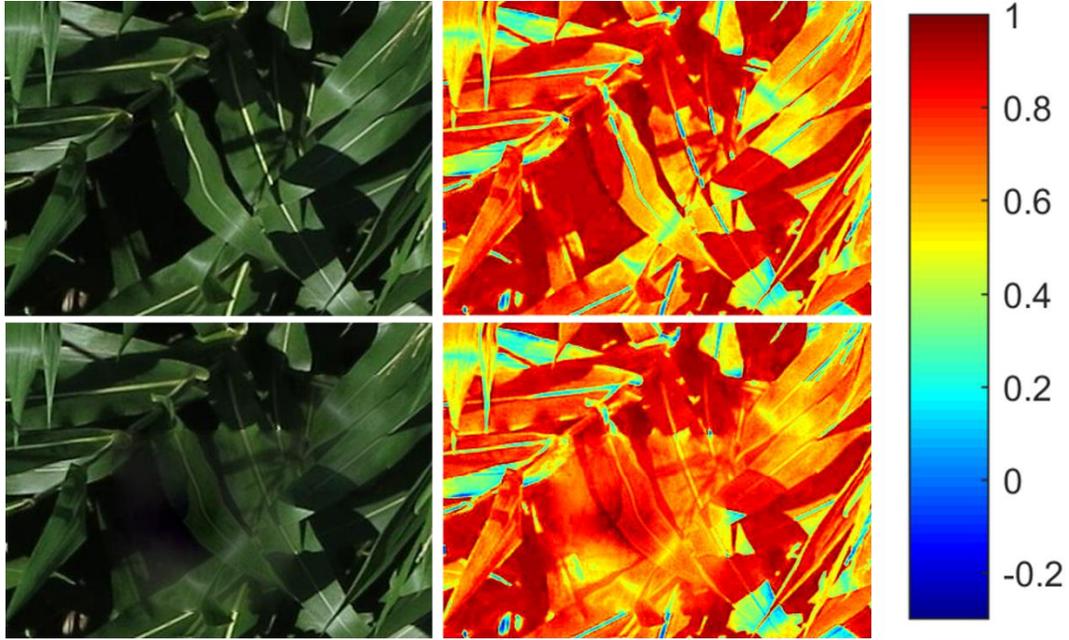


Figure 4.10: Results of the Poisson image editing method. The original image and original NDVI are shown on the top and the adjusted image and NDVI are on the bottom.

Due to the failure of the first method, the next attempt was to normalize the lightness across the image by adjusting the scale of the lightness values. Through testing all of the images, the areas which produced appropriate NDVI values generally had a lightness values between 30 and 50. As a result, a normalization method was employed to rescale the lightness values to mitigate the impact of extreme lightness values. The normalization method follows Equation 4.2.

$$\frac{L_{new} - newmin}{newmax - newmin} = \frac{L - \min(L)}{\max(L) - \min(L)} \quad (4.2)$$

In this equation, L_{new} is the new lightness for the images, $newmin$ is the selected new minimum lightness, $newmax$ is the selected new maximum lightness, and L is the current lightness of the image. The previous equation can be rearranged to solve for the new lightness values of the image, as shown in Equation 4.3.

$$L_{new} = \frac{(L - \min(L)) * (newmax - newmin)}{\max(L) - \min(L)} + newmin \quad (4.3)$$

For the purpose of these images, as a result of testing all of the images for the most appropriate values, the $newmin$ variable was set to be 30 and the $newmax$ variable was set to be 50. The

result of the lightness normalization can be seen in Figure 4.11. The visual and near-infrared images are shown on the top row, the NDVI before adjustment is on the bottom left and the bottom right image shows the calculated NDVI after lightness adjustment. The new NDVI values remove the dependence to lightness across individual leaves and also produce appropriate values for the soil portions, rather than the falsely high values previously calculated.

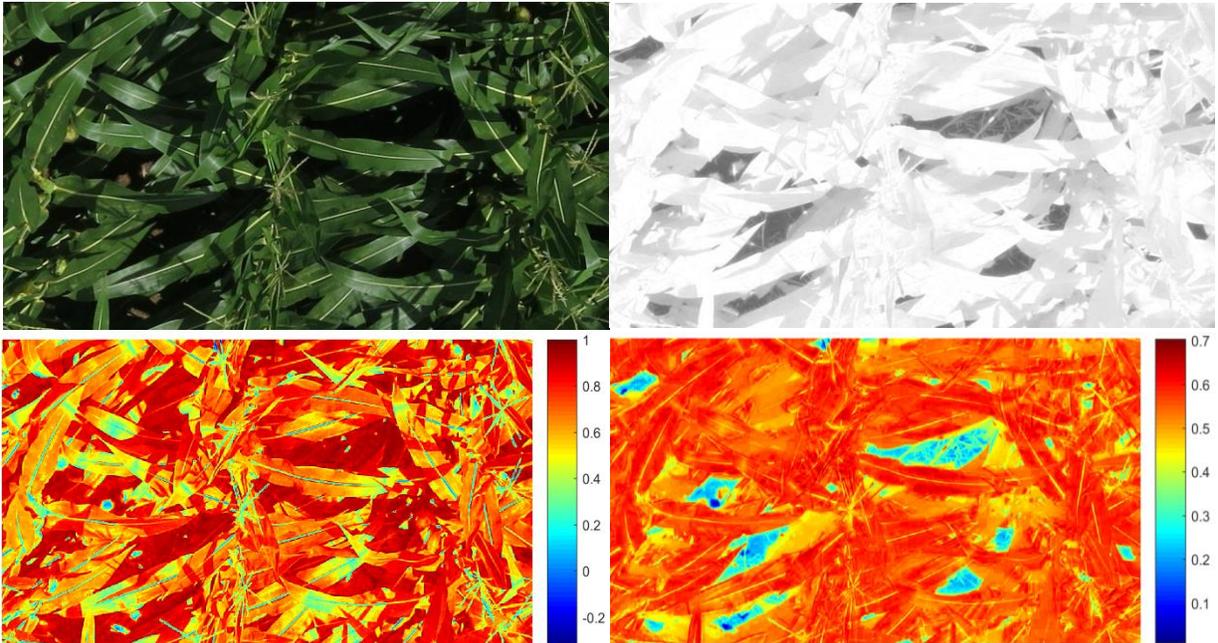


Figure 4.11: Images of corn test plot 105. The top left image is the visual image, the near-infrared image is in the top right, the bottom left image is the NDVI of the plot before the lightness adjustment, and the bottom right image is the NDVI after adjustment.

The correlation between the NDVI values and lightness values was tested again by performing a regression analysis between the NDVI calculated after lightness adjustment and the adjusted lightness values. The plot of these values can be seen in Figure 4.12 and the coefficient of determination, R^2 , was calculated to be 0.0295. The regression confirms the lightness adjustment has eliminated any significant correlation between the lightness and the NDVI values.

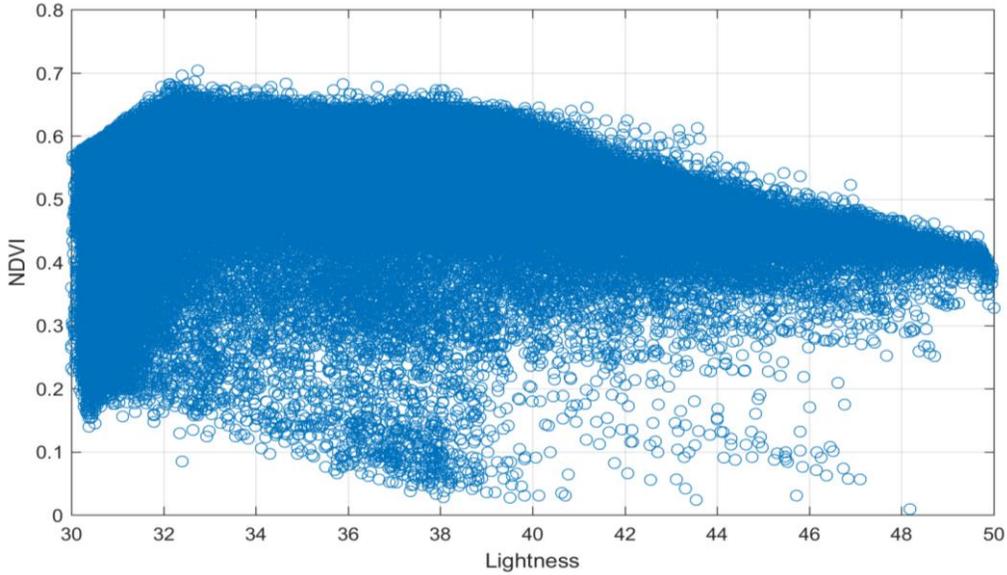


Figure 4.12: Plot of NDVI and Lightness after adjusting the images shown in Figure 4.10. A regression analysis produced a coefficient of determination of 0.0295.

4.2.3 Trend Analysis

Once the NDVI of the test plots was calculated and corrected, the data was used for analyzing any trends. The test plots that we were able to image and get usable data from were plots 101, 102, 103, 104, and 105. These five test plots covered each of the five different nitrogen sidedress application rates. After the corn plots were harvested, the yield was determined, in bushels per acre, which is plotted in Figure 4.13 for the five plots. The yield did not have a strong relationship with nitrogen application rate, with a coefficient of determination of 0.226. The highest yield came from the plot with the 80 lb/ac application, while the second highest received an application of 160 lb/ac.

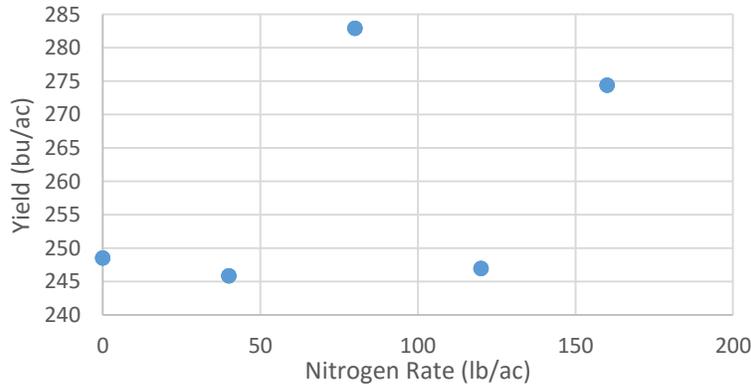


Figure 4.13: Plot of the corn crop yield, in bushels/acre, and the nitrogen application rate, in pounds/acre for the five test plots our images focused on.

The first pair of variables to analyze were the NDVI values of the plots and the nitrogen application rates of the plots. For the analysis, the average NDVI values were found for each of three separate sections of the corn along with average NDVI values of portions of soil in the images. The measured NDVI values are shown plotted against the nitrogen application rates in Figures 4.14 and 4.15. Figure 4.14 shows the results from images acquired on July 13 and Figure 4.15 corresponds to images taken on July 22, which correspond to R2 and R3 growth stages of corn, respectively. The values are shown for corn, as well as soil, before and after the lightness adjustment. No significant trend was found for these plots between the NDVI and nitrogen rate. The trend of the data is relatively flat across the nitrogen rates and none have strong coefficients of determination. The impact of the lightness correction code can be seen in the plot by the NDVI values of soil. Before the correction, the NDVI values of the soil were larger than the values for corn, which is not an accurate result. However, after the correction, the new values are much lower than the corn, which are the values that are expected for soil. Additionally, the adjusted values have a smaller variance due to a reduction of the inaccuracies caused by the lightness changes.

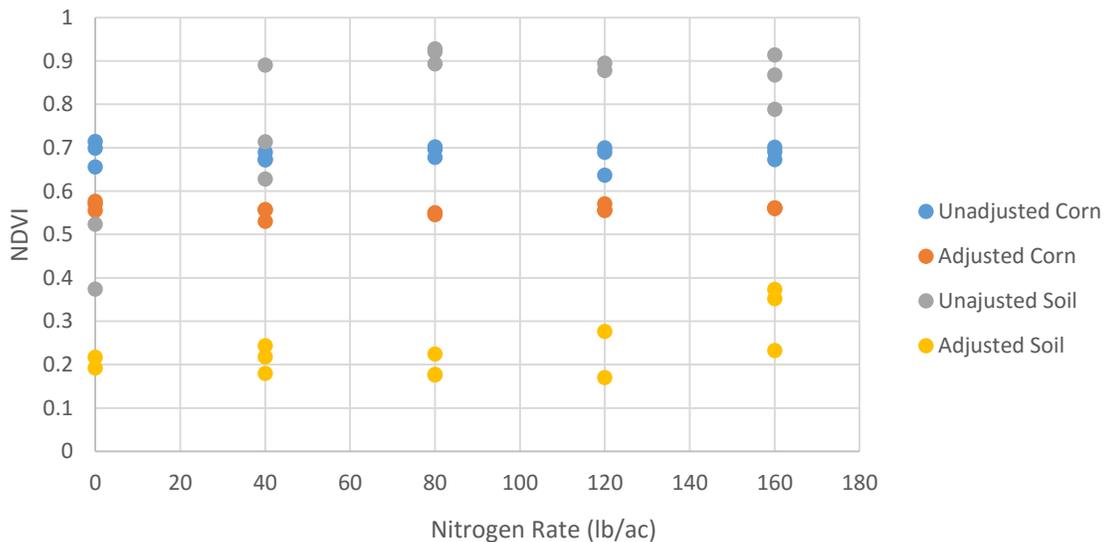


Figure 4.14: Plot of the NDVI values against the nitrogen rates of the test plots for images acquired on July 13, 2015.

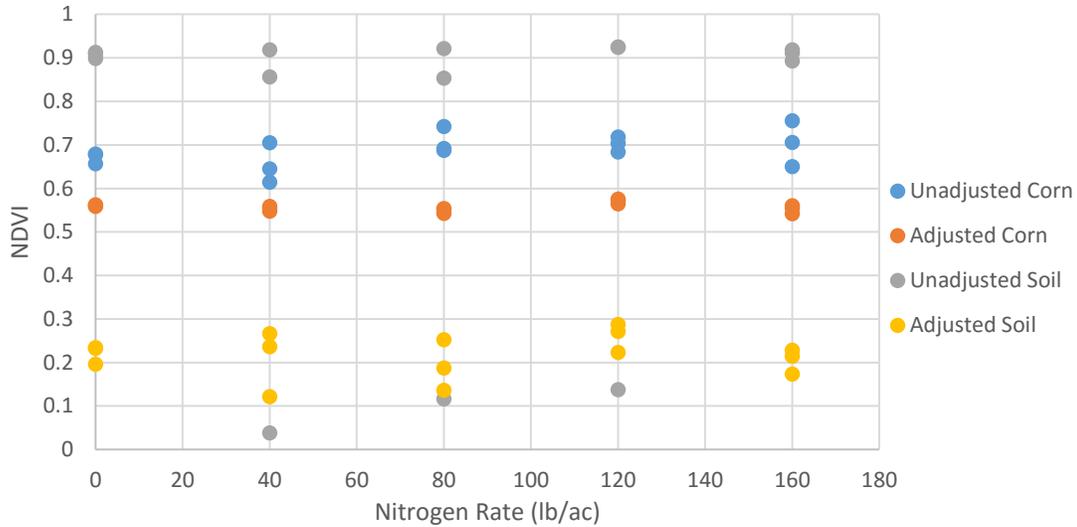


Figure 4.15: Plot of the NDVI values against the nitrogen rates of the test plots for images acquired on July 22, 2015.

The NDVI values for the corn after the lightness adjustment can be seen in Figure 4.16 for both July 13 and July 22. The NDVI values for the two different days are very similar, so the indices are consistent across the plots for different days. However, no significant trend was found between the NDVI and nitrogen application rate. Figure 4.17 shows the corn yield plotted against the NDVI values for the same two days. This also did not show any trend, while the NDVI values are all very similar across the plots. The lack of trend is likely due to a very limited response to the nitrogen at this test site. The nitrogen did not show a major effect on the yield and this is reflected by the nitrogen not affecting the NDVI values.

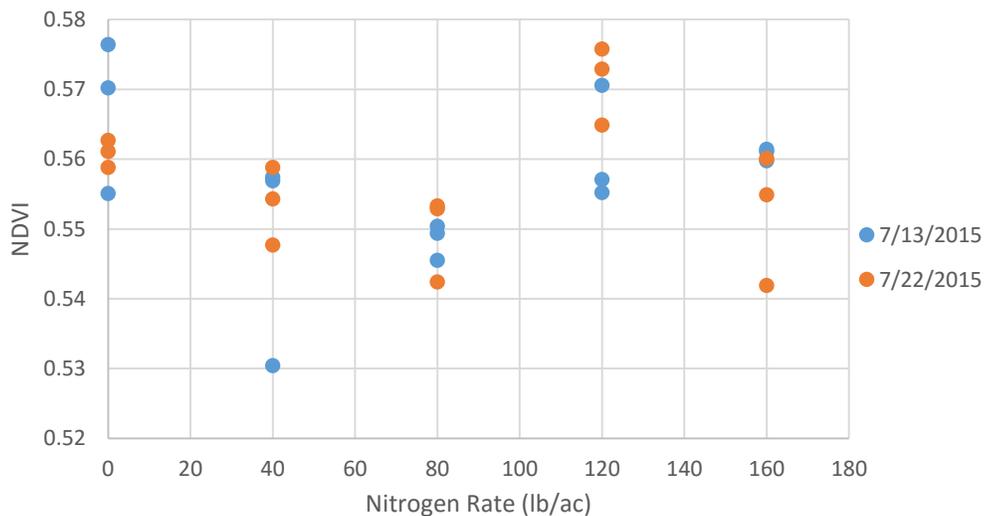


Figure 4.16: NDVI of the corn after lightness adjustment plotted against the nitrogen application for two different days.

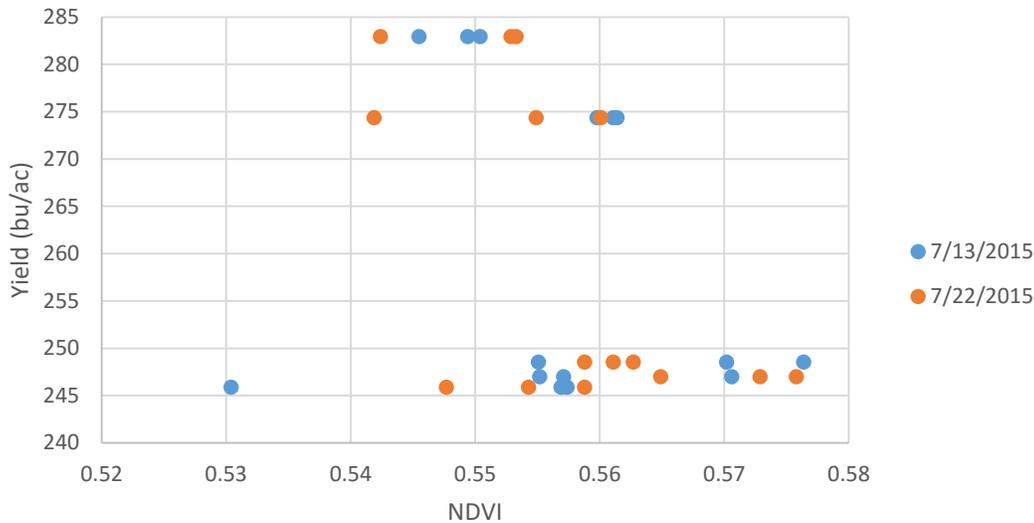


Figure 4.17: Corn yield plotted against the adjusted NDVI of the corn for two days.

4.3 Hyperspectral Data Issues

Throughout the collection and processing of the hyperspectral images and data, many different issues were experienced. These issues included referencing issues, accuracy issues, as well as issues due to local variations in lighting conditions.

4.3.1 Referencing

To acquire the hyperspectral images, the camera must be connected to a computer through the USB 3.0 connection. The camera interfaces with the computer through the BaySpec SpecGrabber program to acquire images. Before images may be capture, the camera must first be calibrated. The first step is to capture an image of a white reference. A 95 percent white reference sheet is used when performing the referencing. The camera exposure is set when acquiring the image of the white reference sheet. The camera can be auto exposed for the reference or the exposure time can be set manually. It was recommended by BaySpec that when capturing the white reference image, the max intensity of the image should be between a value of 200 and 220 for optimal results. Once the exposure time has been set, it remains the same for all of the following acquired images. A dark reference must also be captured by placing the lens cap on the camera and taking an image. Once the white and dark references have been acquired, the camera is ready to begin taking raw images. In order for accurate results, the lighting conditions

when taking raw images should be very similar to the conditions when the white references were taken. If lighting conditions change and a new exposure time is desired, a new white reference image must be captured in order to ensure accurate results. The referencing and exposure time are crucial to ensuring accurate results and any errors during this process can lead to completely useless data. Additionally, adjusted or resetting the exposure time during an experiment often results in changes in the measurements. Figure 4.18 shows a raw image acquired without proper referencing and the result is an overexposed image, which will lead to inaccurate data.



Figure 4.18: Raw image acquired of bushes when the referencing was not performed properly. This image is overexposed and leads to bad data when processed.

Once the raw images are acquired, they are processed in the BaySpec CubeCreator program to produce the cube of hyperspectral images for the different wavelengths. Once the cube is created, the data can be analyzed in CubeCreator or loaded into Matlab for further processing. Figure 4.19 shows the processed results of the raw image shown in Figure 4.18. The left image shows the first band of hyperspectral data and the image is mostly white due to the overexposure. The plot on the right shows the reflectance for the selected areas in the image on the left. The blue line corresponds to the blue box and the red line corresponds to the red box. The reflectance values range between 340 percent and 475 percent, so the data from these images is useless due to the overexposure from improper referencing.

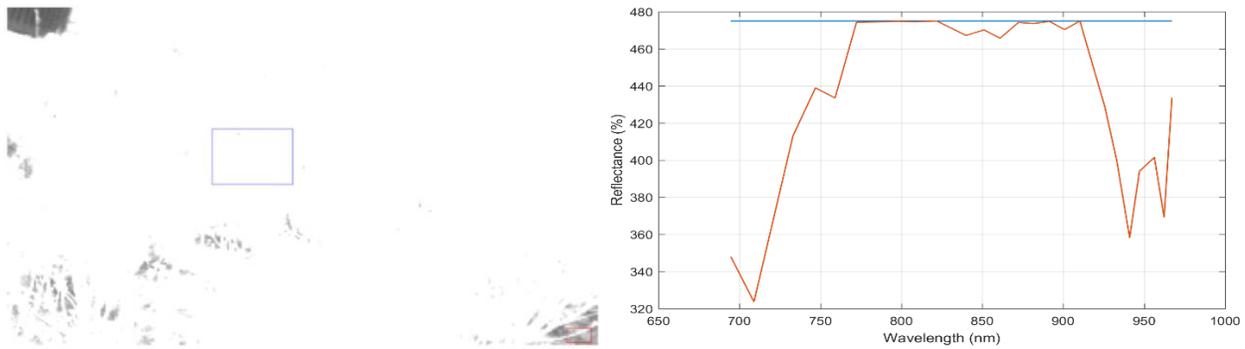


Figure 4.19: The first band of the hyperspectral cube is shown in the left image. The overexposure resulted in a mostly white image. The plot of the reflectance is shown on the right for the two selected portions in the left image.

4.3.2 Hyperspectral Camera Accuracy

Once the issues in the reference process were resolved and images could be properly acquired, the accuracy of the camera was analyzed. To look at the accuracy of the camera, it was compared against a higher quality and more expensive hyperspectral imager, which is known to be accurate. This device was used as the baseline comparison for investigating the accuracy of our hyperspectral camera. The baseline device had a field of view of 8 degrees, while our camera has a field of view of 16 degrees. To account for this, only the center portion of the image was used, corresponding the 8-degree field of view of the other device. For the comparison test, three different objects were used: concrete, a patch of grass, and a healthy leaf from a tree. For each case, the two cameras were referenced at the same time and in the same lighting conditions. Once the referencing was complete, the images of the objects were taken consecutively, while lighting conditions were consistent. The baseline comparison device recorded the average reflectance for wavelengths between 350 nm and 2500 nm, by increments of one nanometer. The results of this comparison are shown in Figures 4.20, 4.21, and 4.22. Figure 4.20 shows the comparison for the results with concrete and the two devices achieved similar results. Both curves have a very flat shape across the wavelength spectrum, however there is a small bias error. The comparison of the devices imaging grass is shown in Figure 4.21. The middle bands of the data produce similar shapes, but there is still a bias error in the data, with our hyperspectral camera recording slightly smaller reflectance values. The biggest difference occurs at the high and low ends of the spectrum, with large error between the two devices. Similar results can be seen in Figure 4.22, which shows the results from the leaf comparison. Again, the reflectance

curves have similar shapes in the middle wavelengths, but have a bias error, with our camera recording smaller values. The error becomes very large at the high and low wavelengths, so our camera does not appear to be very accurate at those ends of the spectrum. For wavelengths below 720 nm and above 940 nm, our camera appears to produce extremely inaccurate reflectance data for vegetation, so as a result, the data from bands on those outside edges cannot be trusted to be accurate.

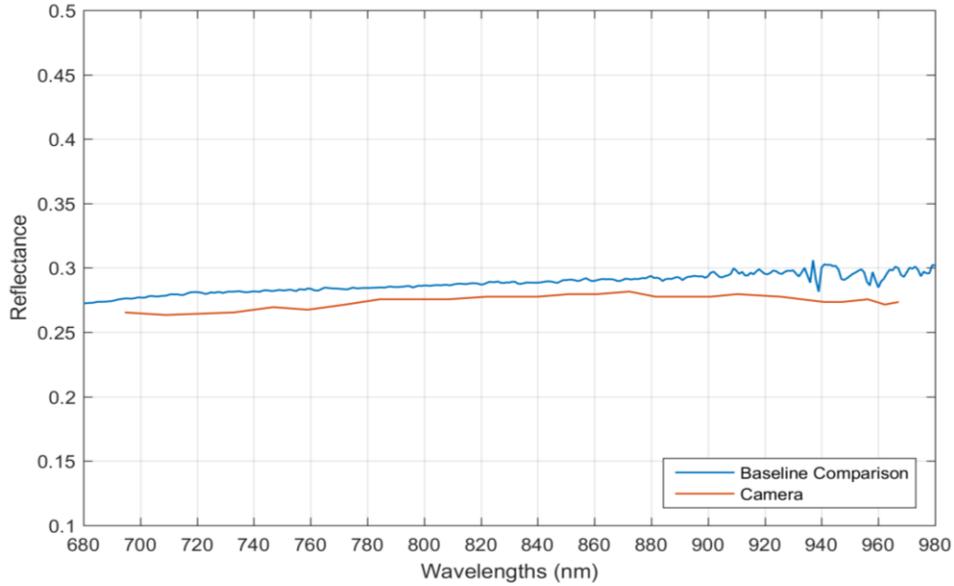


Figure 4.20: Reflectance curves of concrete for each of the two hyperspectral devices.

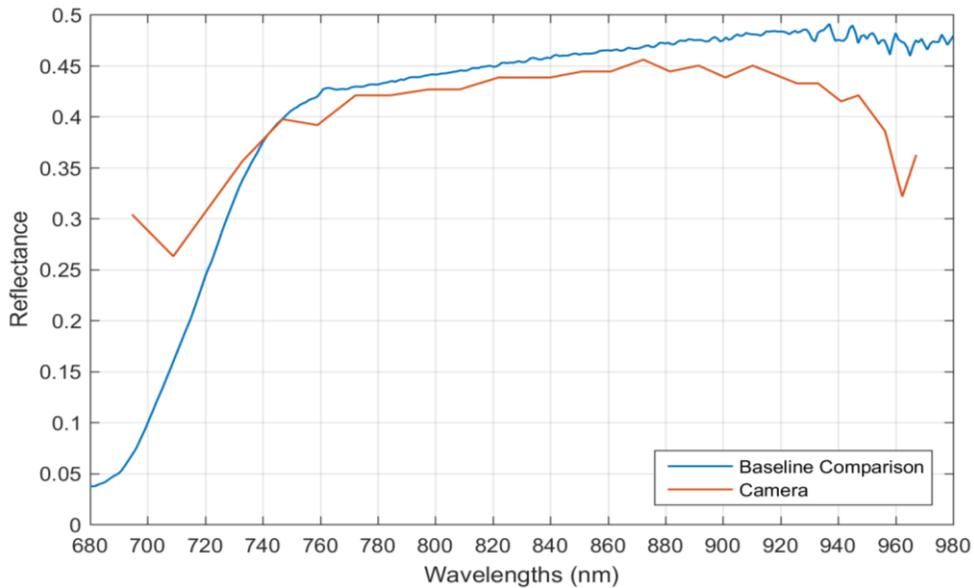


Figure 4.21: Reflectance curves of grass for each of the two hyperspectral devices.

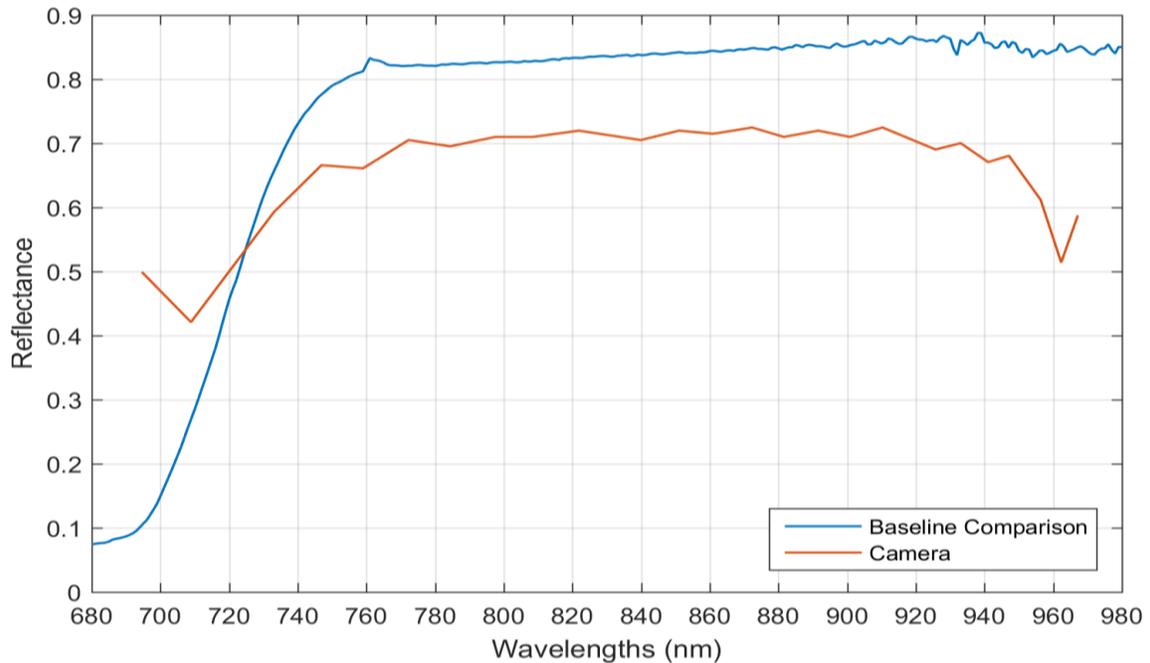


Figure 4.22: Reflectance curves of a leaf for each of the two hyperspectral devices.

A final comparison of this data can be seen in Figure 4.23, which shows reflectance curves of the grass, concrete, and leaf for each of the two cameras. Despite the inaccuracies of our camera from the baseline hyperspectral device, each similarly show distinct differences between the grass, concrete, and leaf. The leaf had the highest reflectance over most of the spectrum, with the grass being second, and the concrete having the lowest reflectance. The high and low ends of the grass and leaf curves from our camera can be seen to display similar shapes, so the inaccurate results are likely due to issues with our camera. As a result, the wavelengths below 720 nm and above 940 nm will be ignored in future analysis due to the inaccuracies.

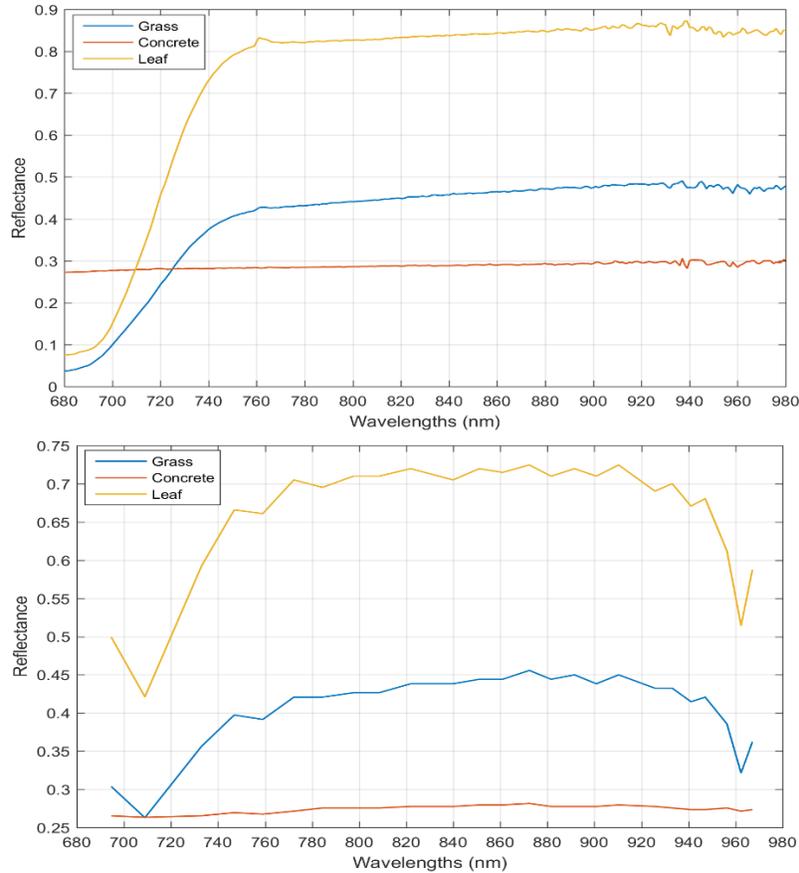


Figure 4.23: Reflectance curves of the three objects for each of the two devices. The top plot shows the curves produced from the baseline camera and the bottom plot shows the curves produced from our camera.

4.3.3 Shadows and Light Variations

After the referencing issues were resolved, the next issues discovered with the data are shadows in the images. Due to the conditions and heights at which the images were taken, the hyperspectral data was impacted by variations in lighting conditions, particularly shadows. The portions of the hyperspectral images that were impacted by the shadows showed a reflectance curve similar to that of a stressed crop, even when the leaf was healthy. An example of this can be seen in Figure 4.24. The visual image is shown along with one band of the hyperspectral image. The blue and red boxes selected on the hyperspectral image correspond to the blue and red reflectance curves on the plot. These portions were selected on the same leaf, with one in a shadowed portion and the other without any shadows. The portions are also nearby on the same leaf, so the reflectance characteristics of the two portions should be very similar in the same

lighting conditions. The reflectance curve plot shows that the shadowed portion of the leaf resulted in much lower reflectance values, which would be similar to an unhealthy or stressed leaf. The large differences in reflectance values due to variations in lighting conditions can be problematic to ensuring the accuracy of the data.

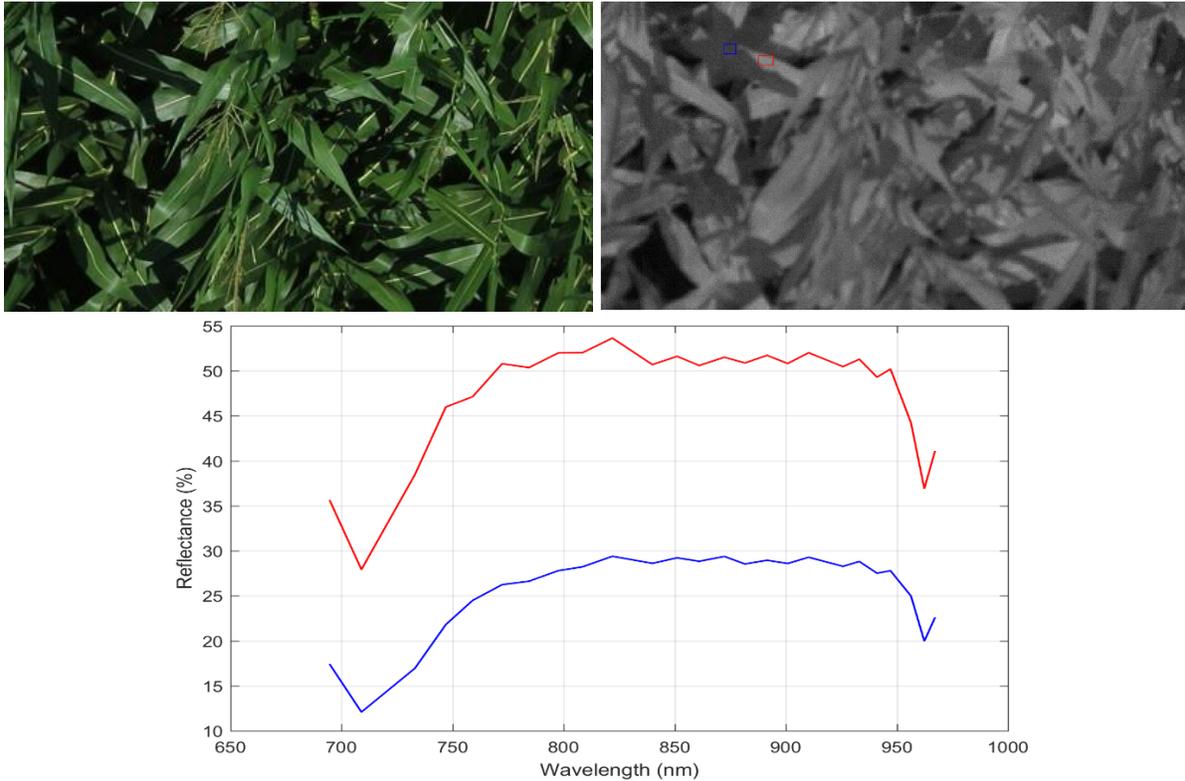


Figure 4.24: Test plot image impacted by shadows. The visual image (top left) and corresponding hyperspectral image (top right) are shown. Red and blue boxes on the hyperspectral image correspond to the Reflectance curves on the plot.

4.4 Hyperspectral Shadow Correction

In order to ensure that the data acquired in the hyperspectral images is more accurate and usable, it will be important to reduce the impact of the shadows on the data. A method for correcting the shadows in the images was created through a combination of the visual and hyperspectral images. To determine the relationship between the local lighting conditions of the image and the hyperspectral reflectance, combinations of matched images were used. Using pairs of matched visual and hyperspectral images, areas with varying lighting conditions were selected. The conditions varied from shadowed to glared areas and included lighting conditions in between as

well. For the areas selected, the Lightness from the Lab plane of the visual image was determined as well as the hyperspectral reflectance. Using these pairs of values, the data was plotted to determine a relationship. The plot is shown below in Figure 4.25 and contains values for 20 of the 25 different spectral bands. The extreme high and low bands were removed since these bands were not very accurate. The reflectance and lightness values for each of the bands showed similar trends, but were biased from each other vertically. To get a true grasp for how well the data fits, an average of the bands was taken before fitting a line to the data.

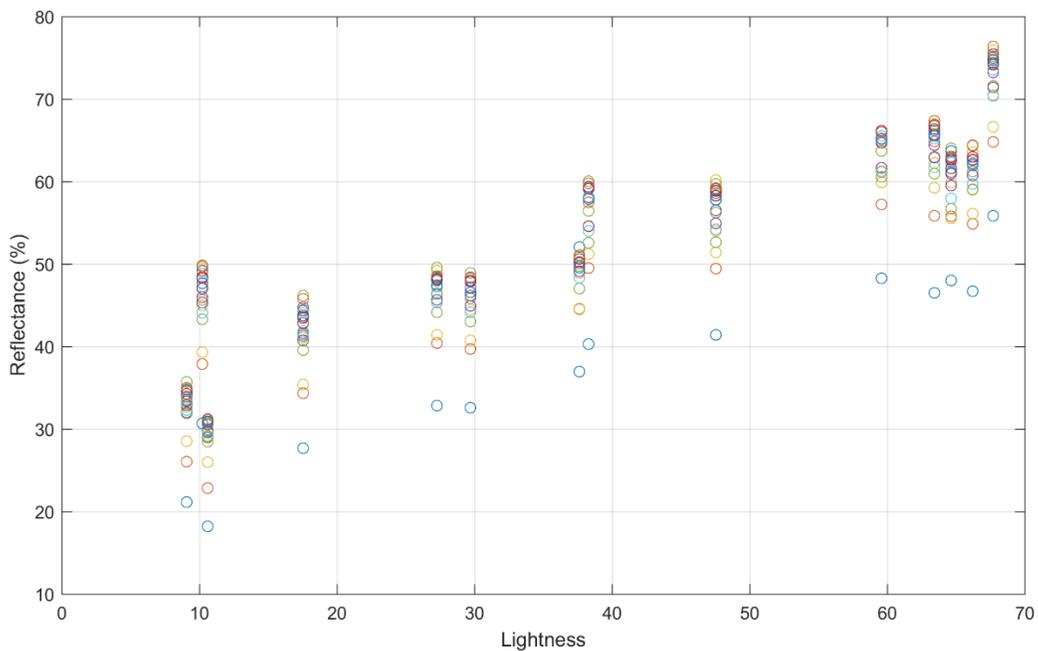


Figure 4.25: Plot of the hyperspectral data points for 20 bands at various lighting conditions.

The averaged data is shown in figure 4.26 with a best fit regression line shown for the data. Higher order polynomial fits were also looked at, but the addition of a second order term was not statistically significant. This additional term produced a p-value of 0.713, while a statistically significant term would have a p-value less than 0.05. As a result, a linear fit was chosen for the data.

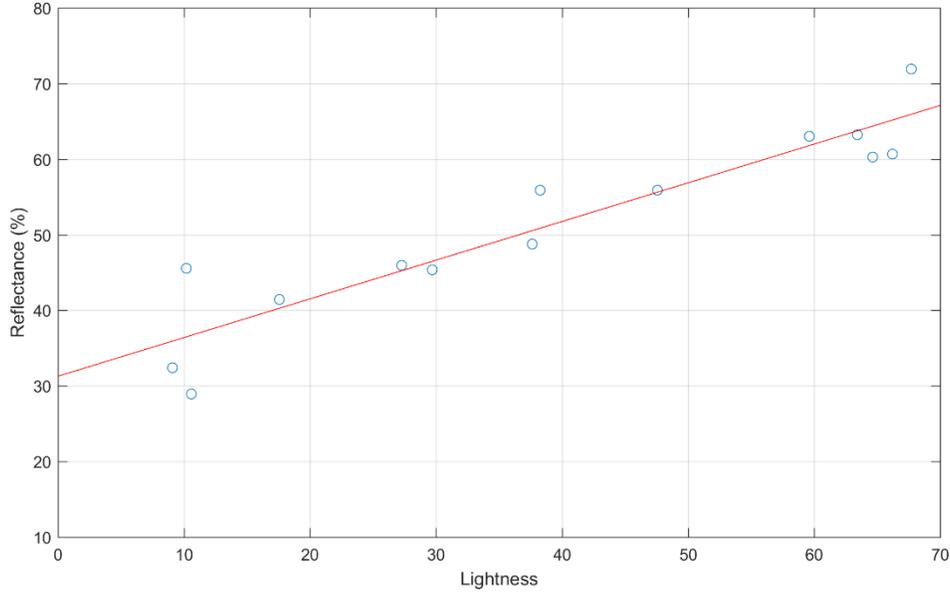


Figure 4.26: Plot of the average of the hyperspectral data points with various lighting conditions. A trend line was fit to the data with an R^2 value of 0.868.

The trend line of best fit for the Lightness and Reflectance data was found to be Equation 4.4.

$$R = 0.51179 * L + 31.284 \quad (4.4)$$

In this equation, R is the reflectance, in percent, and L is the Lightness value of the Lab plane of the visual image. This line fit the data with a coefficient of determination of 0.868. After an equation was determined for the relationship between the two variables, it could be implemented as a method for correcting the lighting conditions in the hyperspectral images. The constant term at the end of the equation can vary depending upon the magnitude of the reflectance values, so this is set as a constant variable, c. The optimal image Lightness for the correction was determined through investigating the dataset of images. Through manual tuning, a value of 45 for the Lightness was found to be the optimal condition which contained hyperspectral reflectance values corresponding to the healthy corn as well as the absence of both shadows and glare in the visual image. The Lightness value of 45 was targeted when correcting the lighting conditions. When plugging the Lightness value of 45 into Equation 4.4 to determine the final adjusted reflectance value, the result is Equation 4.5.

$$R_{adjusted} = 0.51179 * (45) + c \quad (4.5)$$

From this equation, $R_{adjusted}$ is the new reflectance value after being adjusted for the shadows and c is the constant term. To adjust an image, the current Lightness and reflectance values must also

be taken into account, so plugging these into Equation 4.4 creates the result shown below in Equation 4.6.

$$R = 0.51179 * (L) + c \quad (4.6)$$

R represents the reflectance of the current hyperspectral image and L is the current lightness of the visual image at the corresponding location. Rather than solving for the constant term, the equations can be combined together by subtracting Equation 4.6 from Equation 4.5. Once the two equations are subtracted, the constant value cancels out and after rearranging variables, the final equation becomes Equation 4.7.

$$R_{adjusted} = R + 0.51179 * (45 - L) \quad (4.7)$$

These variables are the same as previously defined. Equation 4.7 was then used as the method for removing the impact of the shadows from the hyperspectral reflectance values. The results of the method can be seen in Figures 4.27 and 4.28. The results show the method reduces the impact of the shadows and normalizes the reflectance across leaves and the image. The adjustment performs well overall, however some areas in the images are not perfectly aligned between the visual and hyperspectral and this can lead to undesired results in some small sections.

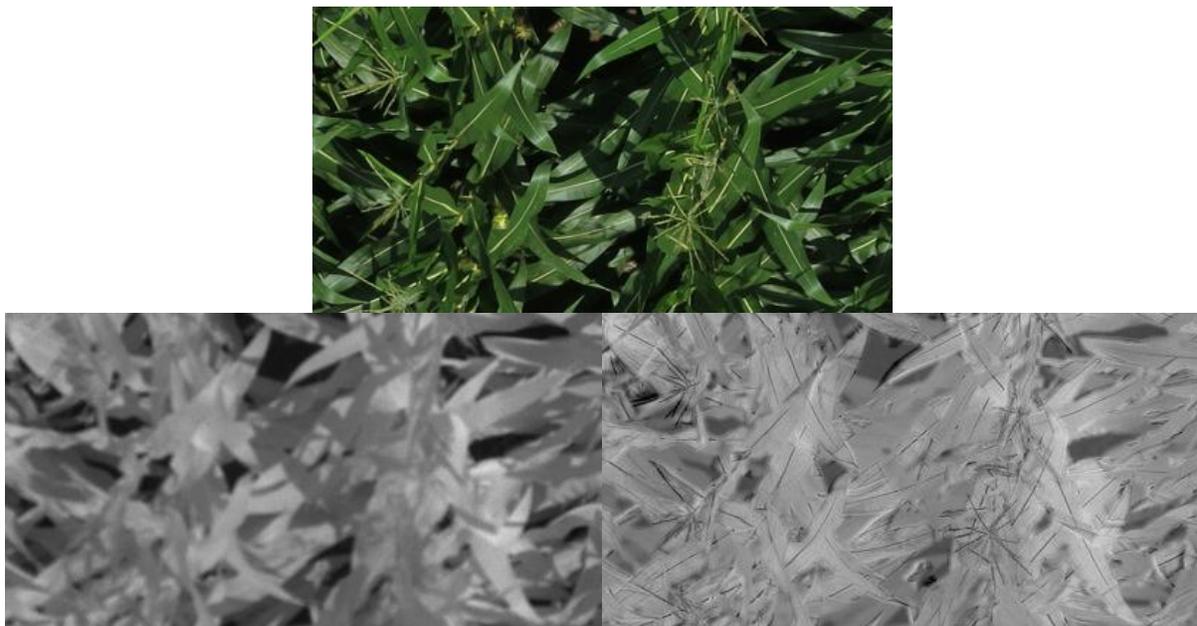


Figure 4.27: The top image is the visual image which corresponds to the hyperspectral image for test plot 101. A single band of the hyperspectral image is shown in the bottom left. The bottom right image is the result of the reflectance adjustment code on the hyperspectral image.

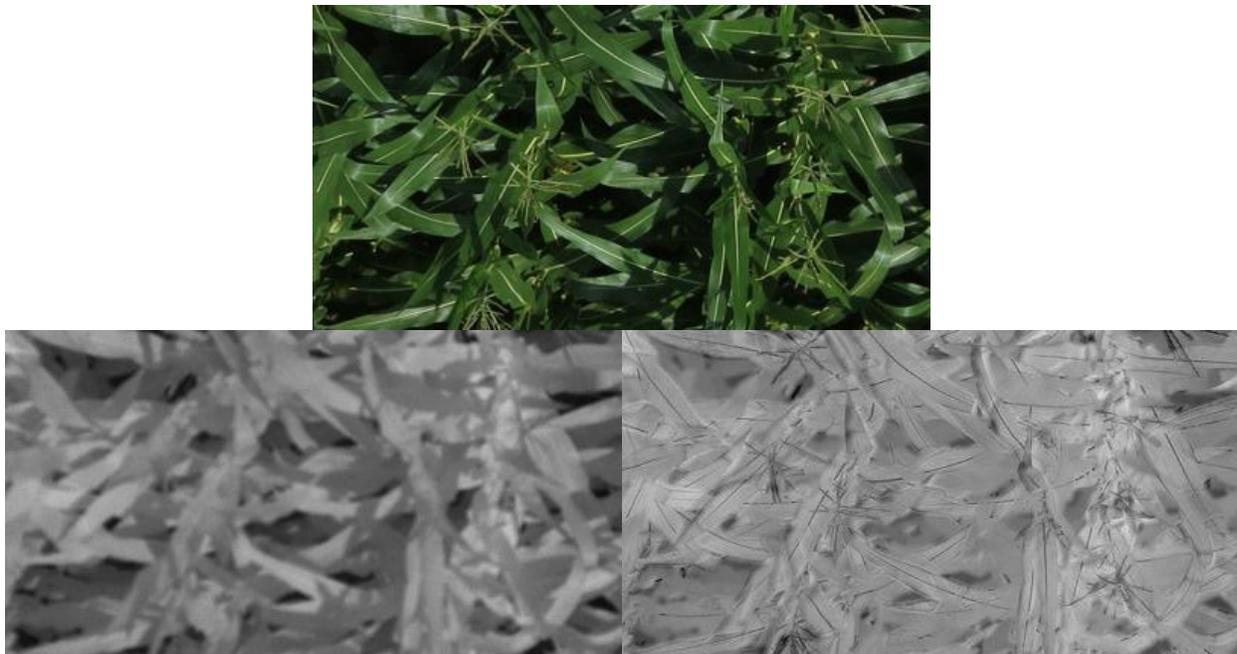


Figure 4.28: The top image is the visual image which corresponds to the hyperspectral image for test plot 102. A single band of the hyperspectral image is shown in the bottom left. The bottom right image is the result of the reflectance adjustment code on the hyperspectral image.

The impact of the reflectance adjustment can be seen across the entire spectrum of the camera in Figure 4.29. The Soil and Healthy Corn curves correspond to reflectance curves of shadowed areas before the adjustment and the Adjusted curves show the reflectance as a result of the adjustment method. The reflectance of the shadowed areas increases from the adjustment. Figure 4.30 shows how well this method normalizes the reflectance in an image. The reflectance is shown for a shadowed area of the corn as well as an area, on the same leaf, with a glare, both before and after the adjustment. The adjusted results end up with reasonably close reflectance curves over the middle portion of the spectrum, which is the desired result. The reflectance curve is also shown for a shadowed portion of soil both before and after adjustment, which also increases the reflectance to values still reasonable for soil.

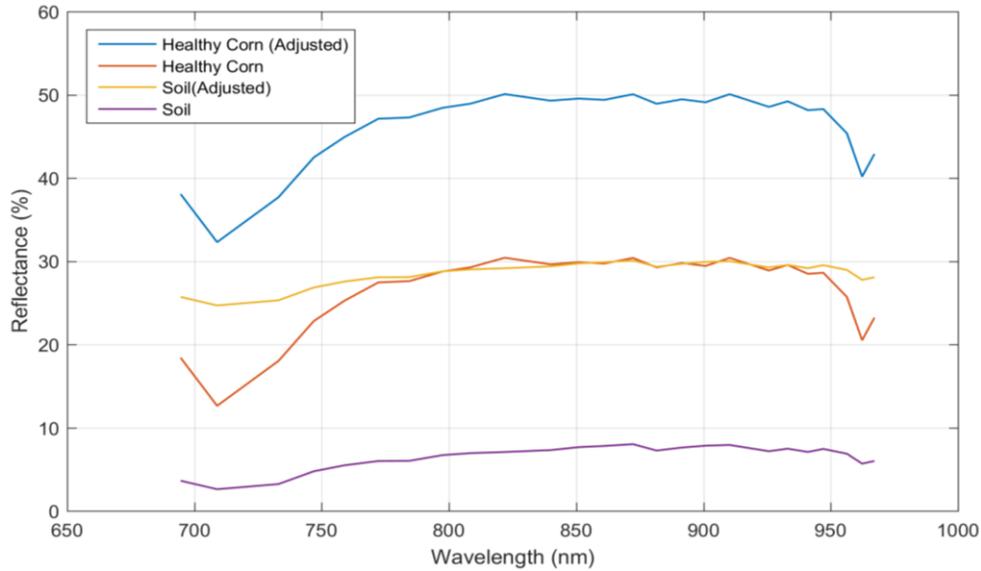


Figure 4.29: Reflectance curves for shadowed areas of corn before and after adjustment.

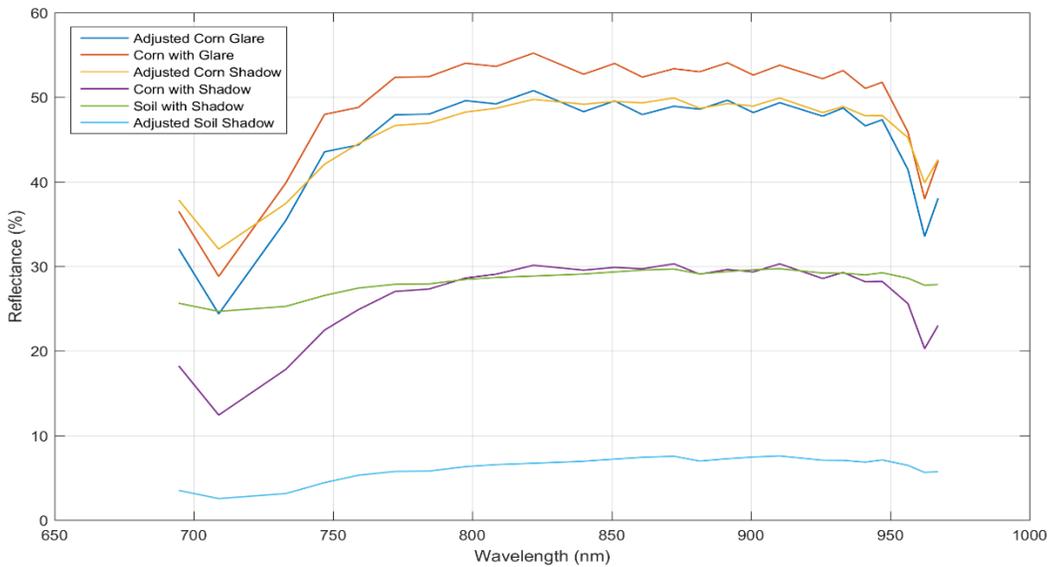


Figure 4.30: Reflectance curves for corn and soil in various stages of lighting before and after adjustment.

4.5 Hyperspectral Indices

The hyperspectral reflectance data for each of the plots can be used to investigate crop indices to help indicate the vigor of the crops. A wide variety of indices have been discovered through various research, however only a few known indices focus on the bands within the range of our camera. Two known indices include R_{780}/R_{740} as well as the normalized difference red edge. The

first index is simply the ratio between the reflectance at 780 nm and the reflectance at 740 nm and the second index, normalized difference red edge, is determined from Equation 4.8.

$$\text{Normalized Difference Red Edge} = \frac{R_{790} - R_{720}}{R_{790} + R_{720}} \quad (4.8)$$

In this equation, R_{790} and R_{720} are the reflectance values at 790 and 720 nm, respectively. The investigation of our test plots began with these two indices to analyze any trends for these given test conditions. These bands are within the range of our hyperspectral camera, however these exact values are not among the 25 bands collected by the camera. As a result, it was necessary to interpolate the reflectance results in order to estimate the reflectance values at those wavelengths. Through discussions with BaySpec, the company who produces our camera, we were informed that the reflectance values at wavelengths between the measured bands of the camera were linearly related and could be estimated by using linear interpolation between the measured wavelengths. The linear interpolation was performed between values measured by the camera in order to produce the desired reflectance values at 790, 780, 740, and 720 nm, which could be used in the previously mentioned indices. To calculate the indices, only portions of the hyperspectral image were used, due to images containing both corn leaves and background soil through the image. Three separate portions of leaves were selected from the image and then these selected areas were averaged to determine the final index value of the corn in the image. This process was performed in order to isolate the corn crop values from the background soil contained in the image and the averaging of multiple sections produces a more representative value of the entire image, rather than isolating one localized section.

Once the indices were determined, the results were analyzed to determine any possible trends of the data. Figure 4.31 shows the R_{780}/R_{740} index values for the test plots on July 13, which corresponds to the R2 growth stage. The plot on the top shows the index plotted against the yield of the corn plots and the bottom plot shows the index against the nitrogen application rate. The relationship between this index and the corn yield exhibits a positive trend with a correlation coefficient of 0.6554. The index and nitrogen application rate show a slightly positive trend, but only a correlation coefficient of 0.4534. A small relationship was found between this index and the nitrogen application rate, but a stronger one was found between the index and crop yield. While the relationship does exist, it is still not shown to be very significant.

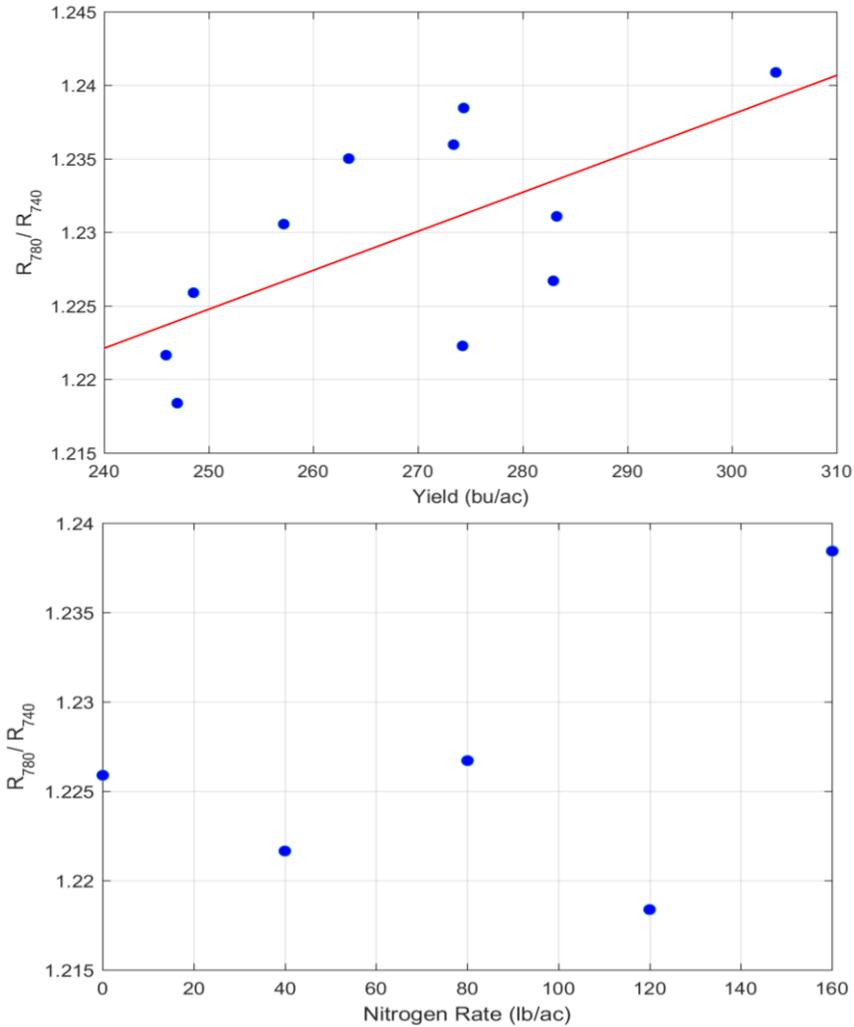


Figure 4.31: Plots of the R_{780}/R_{740} index. The top plot shows the index against the crop yield and the bottom plot shows the index against the nitrogen application rate.

The next index to be analyzed was the Normalized Difference Red Edge index and the results can be seen in Figure 4.32. The left plot shows the index against the yield and the right plot shows the index against the nitrogen application rate. Both of these exhibited a slightly negative trend but neither trend was significant with correlation coefficients of -0.1085 for the yield and -0.1315 for the nitrogen rate.

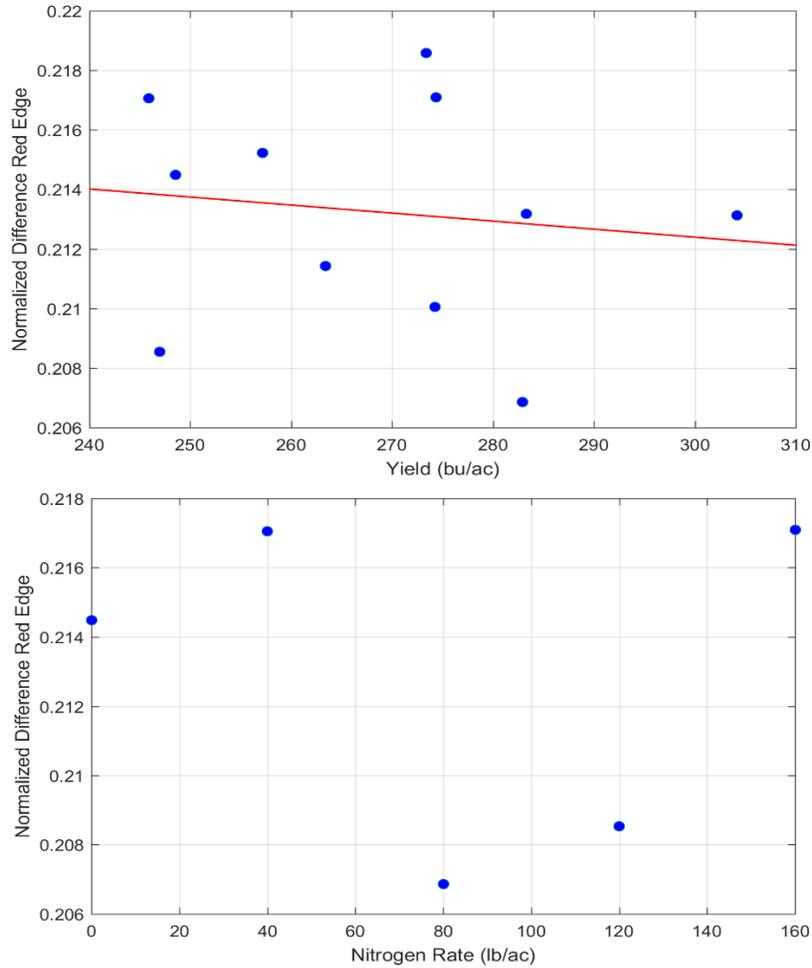


Figure 4.32: Plots of the Normalized Difference Red Edge index. The top plot shows the index against the crop yield and the bottom plot shows the index against the nitrogen application rate.

Due to the lack of strong trends with the previous indices, the creation of new indices was desired for these crop conditions. To create a new index, the hyperspectral data was interpolated to determine the reflectance for each wavelength that is divisible by 10, such as 720 nm, 730 nm, etc. Only wavelengths at which the camera was accurate were used for this investigation, so that included only the wavelengths between 720 nm and 940 nm. The two forms of indices that were looked at were ratios of the reflectance at two distinct wavelengths and normalized differences of the reflectance between two wavelengths. All of the possible combinations of wavelengths were used for this data and the correlation was calculated between the new index and the yield of the crop to find the best indices. An additional criteria used to assess the validity of the indices was the value of the index for areas of background soil to ensure that the index values of the soil match up with the trend observed for the crop.

After testing all combinations of wavelengths, the pair of wavelengths which produced the highest correlation between the index and the crop yield were 760 and 740 nm. Two different indices using these two bands were looked at. The first one is the ratio of the reflectance at each of these wavelengths, which is shown in Figure 4.33. The top plot shows the index value plotted against the yield of the corn in the plot. A least squares regression line was fit through the points, which can be seen as the red line in this plot. The correlation coefficient between this index and the yield was found to be 0.8404, which shows a much stronger correlation than the previous indices. The bottom plot in the figure shows the same index values along with additional points showing the index values for soil. These were plotted at a yield of zero to show the separation between the index magnitudes and the index trend can be seen as an extension of the trend within the corn.

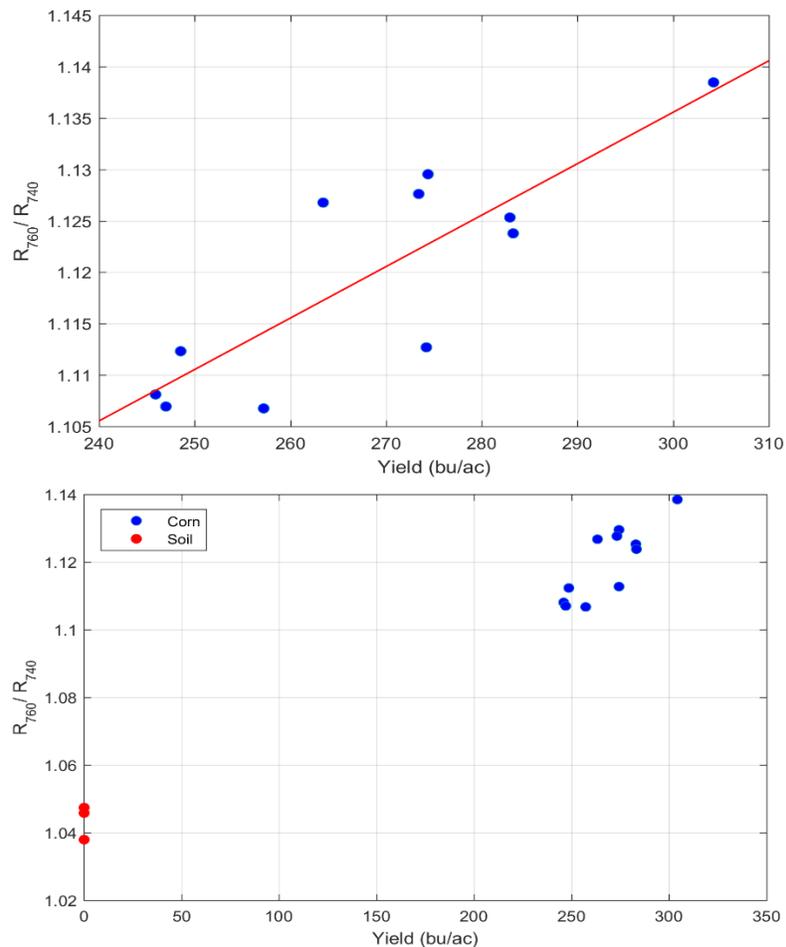


Figure 4.33: Plots of the relationship between the corn yield and the new index created from the ratio between the reflectance at 760 nm and 740 nm.

The same pair of wavelengths, 740 nm and 760 nm, was used to create a second index in the form of a normalized difference equation. The index produced is shown in Equation 4.9.

$$Index = \frac{R_{760} - R_{740}}{R_{760} + R_{740}} \quad (4.9)$$

This index produced similar results to the previous one, which used the same wavelengths. The index was found to have a correlation coefficient with the corn yield of 0.84. Plots of this index against the corn yield are shown in Figure 4.34. The top plot shows the index against the yield for the plots of corn with a least squares regression line fit to the data. The bottom plot shows the index values for the corn along with index values for soil. The difference between the largest and smallest index values was much smaller for this index than the previous index. The difference for the corn was 0.015, compared to 0.035 and the difference between the corn and soil was 0.046 compared to 0.1.

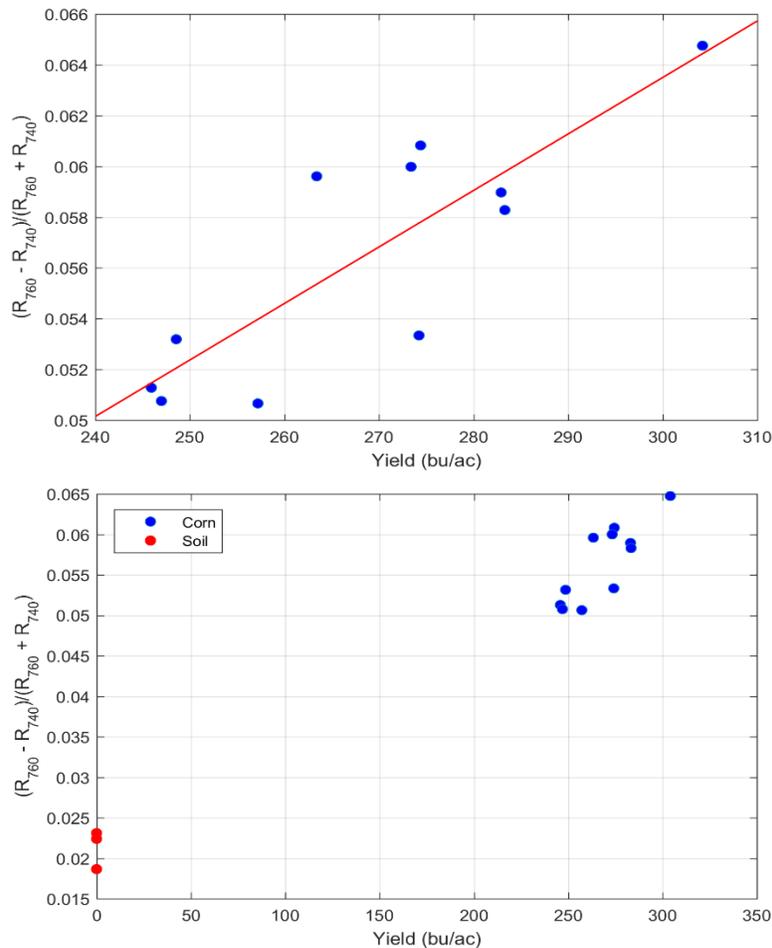


Figure 4.34: Plots of the relationship between the corn yield and the new index created from the normalized difference between the reflectance at 760 nm and 740 nm.

The pair of wavelengths which produced the second highest correlation between the index and the corn yield was 860 nm and 930 nm. The first combination of these wavelengths was a simple ratio of the reflectance, shown by Equation 4.10.

$$Index = \frac{R_{860}}{R_{930}} \quad (4.10)$$

This index produced a correlation coefficient of 0.8224 between the index and the corn yield. Figure 4.35 shows the relationship between this index and the yield of the corn. The top plot shows the index of the corn and corn yield plotted for the test plots along with a least squares line fit to the trend of the data. The bottom plot shows the indices plotted in the top plot along with indices of various portions of soil in the image. A large separation can be seen between the index values of corn and soil. The difference between index values for the corn is .011 and the difference between the corn and soil is 0.06.

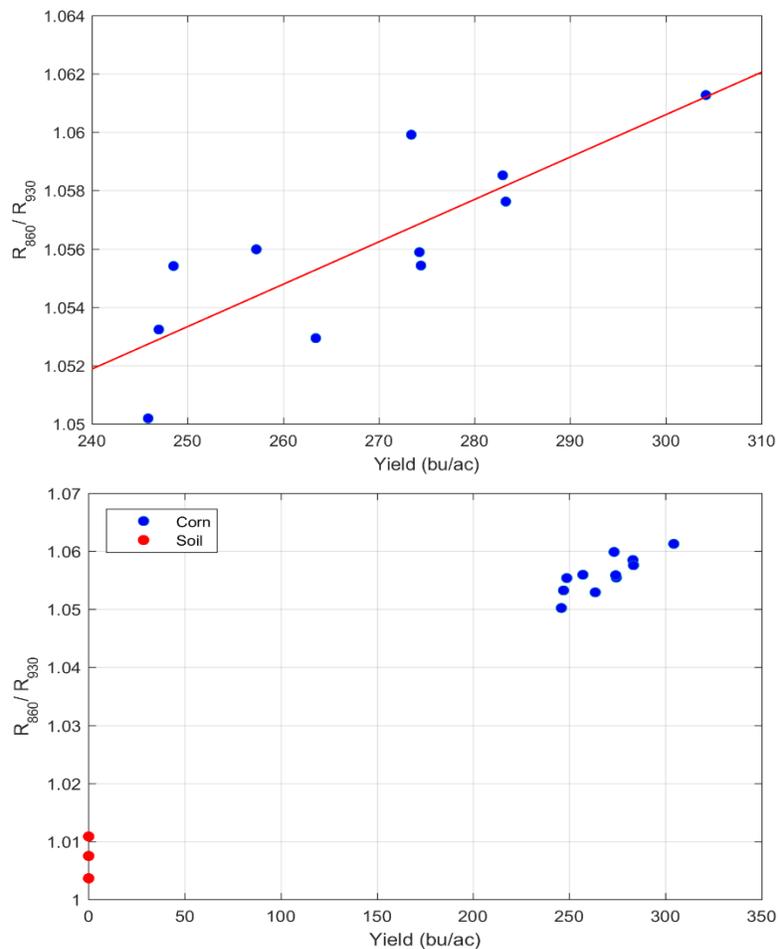


Figure 4.35: Plots of the relationship between the corn yield and the new index created from the ratio between the reflectance at 860 nm and 930 nm.

This same pair of wavelengths, 860 and 930 nm, was combined using a normalized difference equation to be used as another index. This index was calculated using Equation 4.11.

$$Index = \frac{R_{860} - R_{930}}{R_{860} + R_{930}} \quad (4.11)$$

The results from this index are shown in Figure 4.36 and this index produced a correlation coefficient of 0.8221 with the corn yield. The top plot shows the index values and yield for the corn in the test plots along with a least squares regression line and the bottom plot adds index values of soil. The separation between the corn values was very small, only 0.005, while the difference between the soil and corn index values was a much larger value of 0.027.

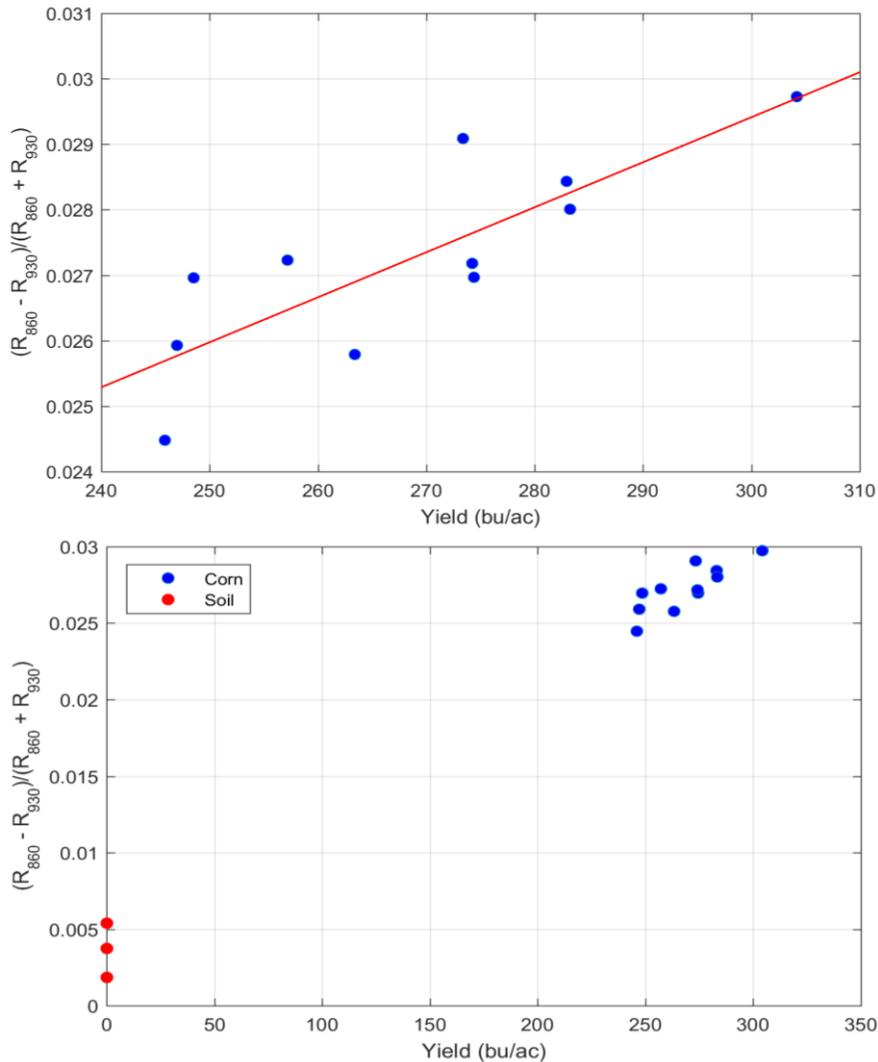


Figure 4.36: Plots of the relationship between the corn yield and the new index created from the normalized difference between the reflectance at 860 nm and 930 nm.

The third highest correlation between the index and the yield resulted from the use of 810 and 930 nm wavelengths. The ratio index created from these two indices is the ratio shown in Equation 4.12.

$$Index = \frac{R_{810}}{R_{930}} \quad (4.12)$$

This index produced a correlation coefficient of 0.8101 with the corn yield. The relationship between this index and the corn yield can be seen in Figure 4.37. The top plot shows the index values for the corn portions of the test plots and shows a least squares regression line for the data. The bottom plot shows the points from the top plot along with index values for portions of the test plot containing soil. The separation between the maximum and minimum values for corn was very small, 0.015, while the separation between the corn and soil was slightly larger, 0.085.

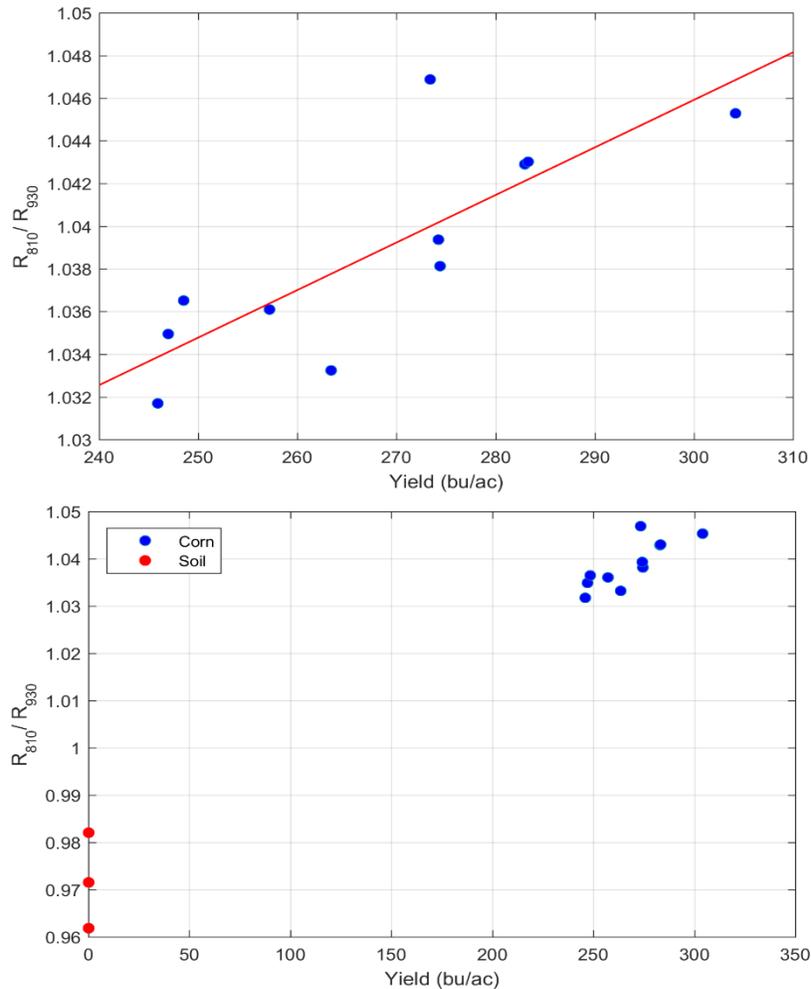


Figure 4.37: Plots of the relationship between the corn yield and the new index created from the ratio between the reflectance at 810 nm and 930 nm.

The pair of wavelengths from the previous index, 810 and 930 nm, was used to create a different equation in a normalized difference form. This index was calculated using Equation 4.13.

$$Index = \frac{R_{810} - R_{930}}{R_{810} + R_{930}} \quad (4.13)$$

The correlation coefficient between this index and the corn yield was found to be 0.8103. Figure 4.38 shows plots of the relationships between this index and the yield of the corn. The top plot displays the index for sections of corn in the image along with the yield for the corresponding plots. A least squares regression line was fit to the data and is also displayed. The bottom plot includes index values which correspond to areas of soil. These index values are very small and the difference maximum difference between the recorded corn index values was only 0.007. The separation between the corn and soil was much larger, with a value of 0.042. Moreover, the index values for the soil were all negative, compared the positive values for the corn.

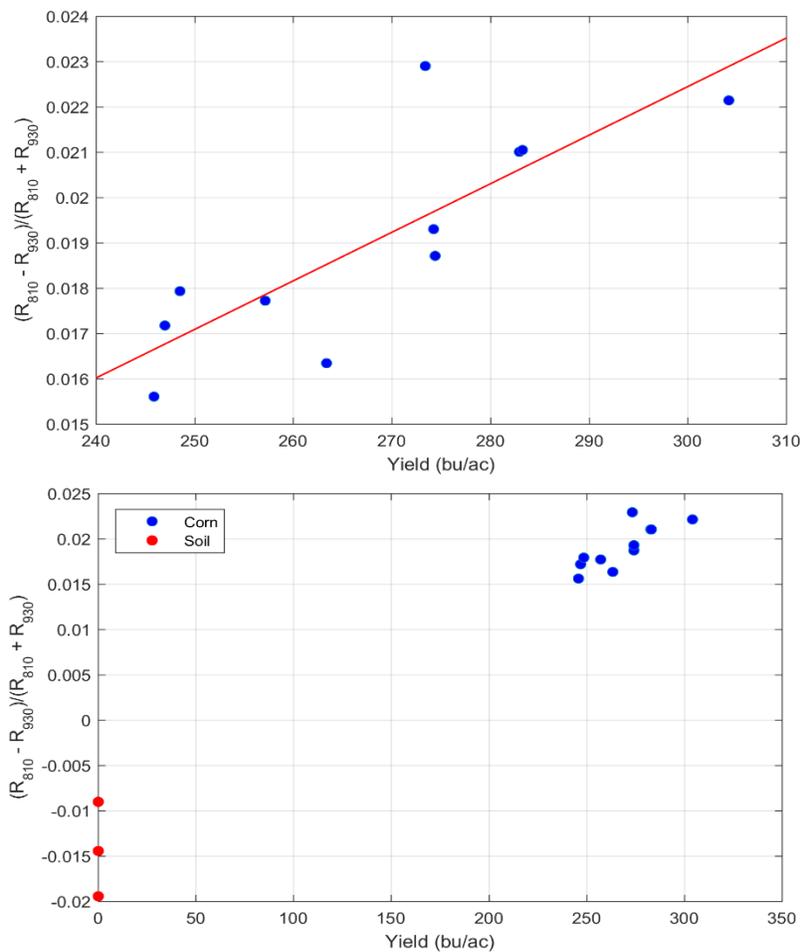


Figure 4.38: Plots of the relationship between the corn yield and the new index created from the normalized difference between the reflectance at 810 nm and 930 nm.

The fourth index was created using the wavelengths of 730 nm and 760 nm. The index using these wavelengths was in the ratio form represented by Equation 4.14.

$$Index = \frac{R_{760}}{R_{730}} \quad (4.14)$$

This index produced a correlation coefficient of 0.7580 between the index value and the yield of the corn. This relationship can be seen in the top plot of Figure 4.39, which shows the index values plotted against the corn yield, along with a least squares regression line. The bottom plot in the figure includes index values for soil. A large separation can be seen between the index values for corn and soil. The maximum difference between index values of corn was 0.037, compared to the difference of 0.19 between the index values for corn and soil.

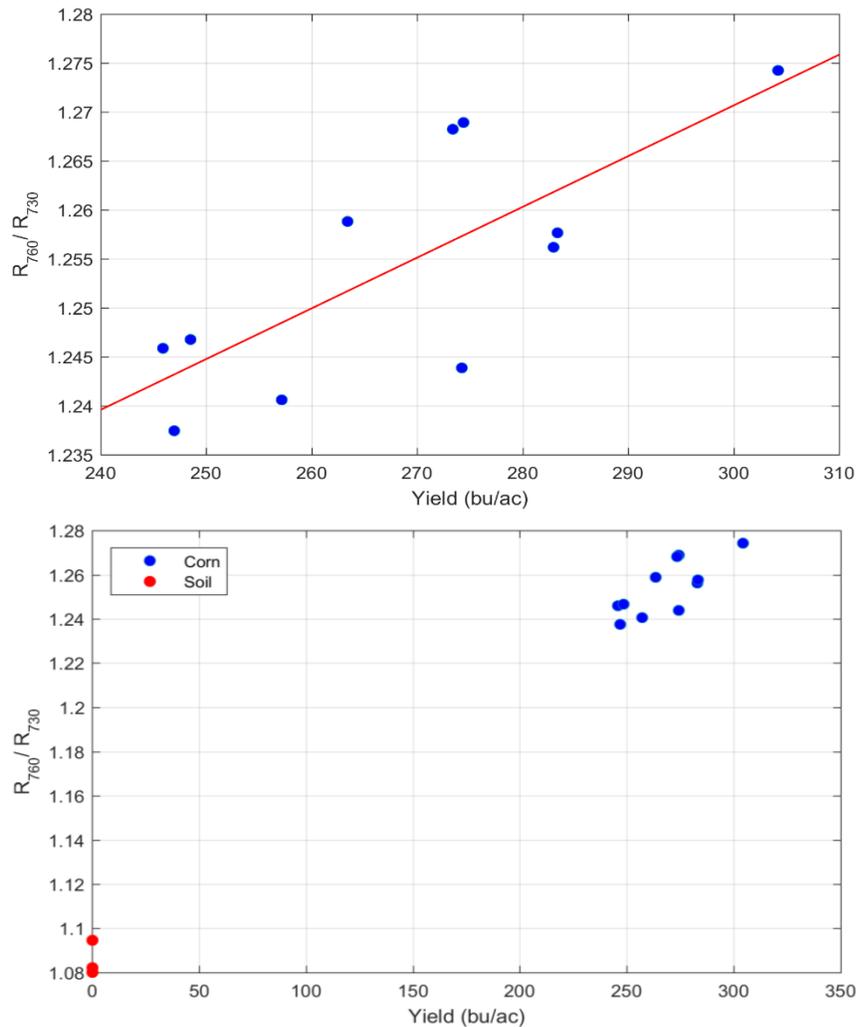


Figure 4.39: Plots of the relationship between the corn yield and the new index created from the ratio between the reflectance at 760 nm and 730 nm.

A second index was created in the form of a normalized difference equation using the previous pair of wavelengths, 730 nm and 760 nm. This index is shown in Equation 4.15.

$$Index = \frac{R_{760} - R_{730}}{R_{760} + R_{730}} \quad (4.15)$$

This index resulted in a correlation of 0.7577 between the index values and the corn yield. This index can be seen below in Figure 4.40. The top plot shows the index values for the test plots against the corn yield of the plot along with a least squares regression line. The bottom plot displays the indices for corn along with index values for soil. The difference between the index values for corn was 0.014 and the separation between the index values for corn and soil was 0.081.

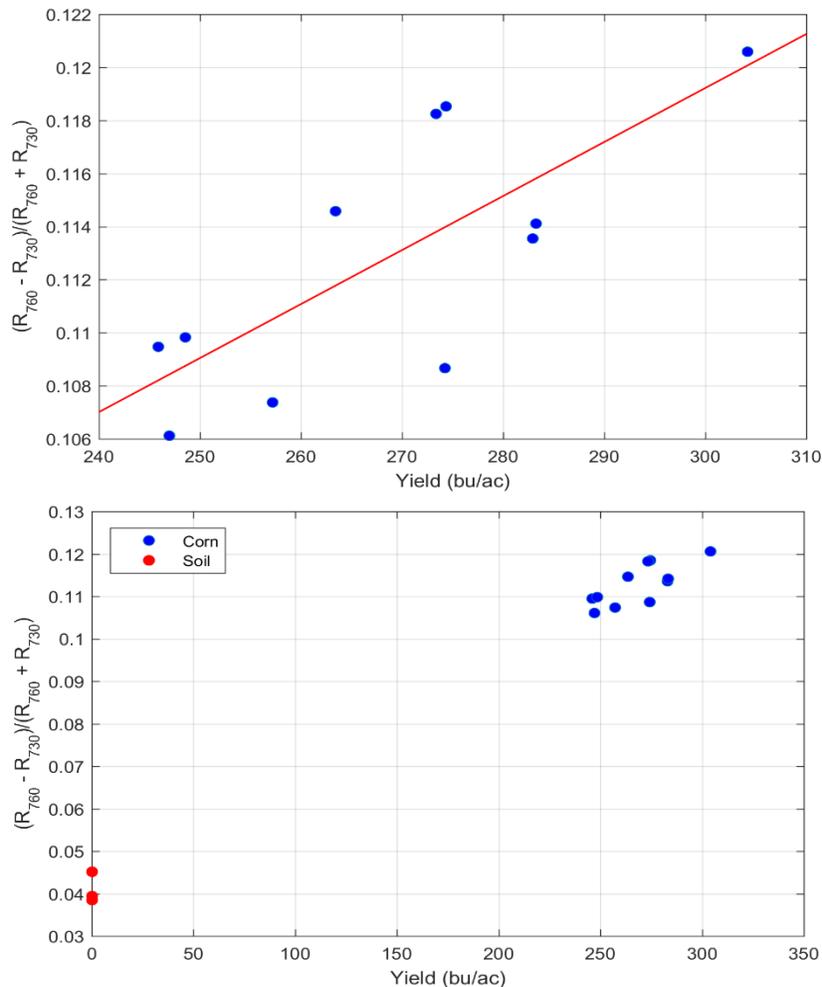


Figure 4.40: Plots of the relationship between the corn yield and the new index created from the normalized difference between the reflectance at 760 nm and 730 nm.

The results of the investigation of indices are summarized in Table 4.1. The correlation between each of these indices and the corn yield were larger than the correlation for the existing indices that were looked at. For each of the four combinations of wavelength pairs, the correlation coefficient for the ratio index and the normalized difference index were almost identical. Additionally, in all cases, the index values for areas of soil followed along with the trend of the crop, so these extend well to determining differences between areas of soil and areas containing the corn crops.

Table 4.1: Indices and Correlation to Corn Yield

Wavelengths (nm)	Index 1	Correlation Coefficient	Index 2	Correlation Coefficient
740 and 760	$\frac{R_{760}}{R_{740}}$	0.8404	$\frac{R_{760} - R_{740}}{R_{760} + R_{740}}$	0.8400
860 and 930	$\frac{R_{860}}{R_{930}}$	0.8224	$\frac{R_{860} - R_{930}}{R_{860} + R_{930}}$	0.8221
810 and 930	$\frac{R_{810}}{R_{930}}$	0.8101	$\frac{R_{810} - R_{930}}{R_{810} + R_{930}}$	0.8103
730 and 760	$\frac{R_{760}}{R_{730}}$	0.7580	$\frac{R_{760} - R_{730}}{R_{760} + R_{730}}$	0.7577

Among these top four indices, two wavelengths were present in two separate indices. These two wavelengths were 760 nm and 930 nm. However, the use of these wavelengths together did not produce a strong correlation for the index value. Some of these indices may be picking up on the water absorption of the test plots since there are generally strong water vapor absorption lines around 930, 820, and 740 nm. For each of the pairs of wavelengths, the two different forms of the indices produced very similar correlations with the yield. While the magnitudes of the indices were much different, the associated trends between the indices and yield were almost identical.

Chapter 5 Stink Bug Detection

This chapter discusses methods for enabling the detection of stink bugs on crops through the use of airborne imagery. The presence of brown marmorated stink bugs can have extremely detrimental effects on crops. As a result, it is very important for a farmer to be aware of the presence of these insect pests in order to eliminate the issue and save crops from harm. Scouting a field for can be very difficult, so the goal is to enable the detection of these stink bugs through the use of images acquired from a UAV. The following will elaborate on the image dataset used for the creation of algorithms and various stages of algorithm development. The created algorithms are discussed and their performance is analyzed.

5.1 Image Collection

The image dataset for the stink bug detection was collected from two main sources. The first is a set of images provided by a contact who works at DuPont and the second set of images are taken by me using dead stink bugs and placing them on various forms of vegetation. Two images from this dataset are shown in Figure 5.1, with the left image being one provided from our contact at DuPont and the image on the right being taken at our lab by placing dead stink bugs on canola.



Figure 5.1: Two example images from the dataset. The left image was provided by a contact at DuPont and the image on the right was taken using canola at our lab.

5.2 Edge and Contour Methods

Stink bugs have a very distinct shape to their bodies, so this can be utilized to facilitate their detection. The first segmentation method that was looked at was a contour based segmentation

method. By extracting the contours in the image, the outline of the shape of the insect can be distinctly seen. Once this contour has been extracted, the shape can be matched with a template of the appropriate shape for the stink bug to determine if one is in the image. To begin examining the feasibility of this approach for the detection of stink bugs, the contour method was first applied to an image of a single stink bug with a background of a white piece of paper. An output of contours from a stink bug image are shown below in Figure 5.2. In this case, the contour of the stink bug can be clearly seen.

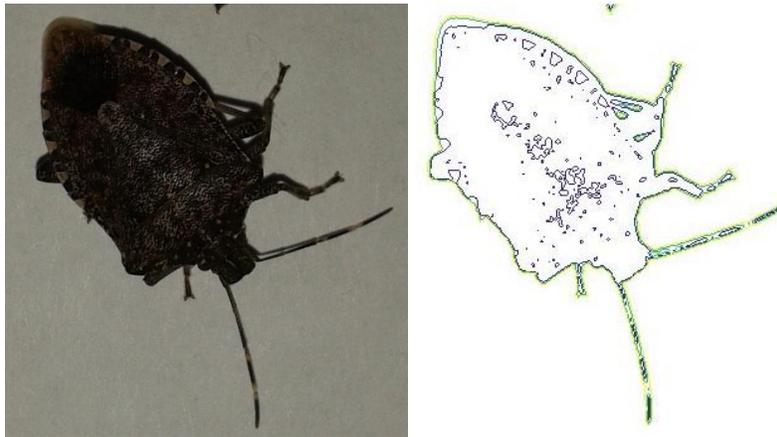


Figure 5.2: Contour extracted for a high resolution image of a stink bug on a plain white background.

In addition to this, simple edge detection methods were tested to determine the effectiveness of these methods. Edge detection methods can also extract the outline of the stink bug, which can then be used in the detection process. An example of an edge detection output is shown in Figure 5.3. The image of the stink bug on a white background from the previous figure was used to determine the effectiveness of the edge detection. For the edge detection in the image, the Sobel operator was used.

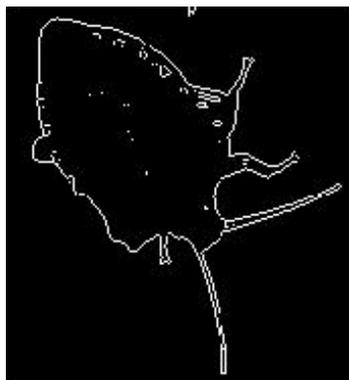


Figure 5.3: Edge segmentation results from the stink bug in Figure 5.2.

The biggest issue for the segmentation and detection of the stink bugs was found to be the resolution of the image. Very high resolution images are required in order for accurate edges and contours to be found. The previous images had a resolution of 34000 pixels in the direct vicinity of the stink bug and produced very good results. Figure 5.4 shows the same segmentation performed using only 2700 pixels. The shape can still be seen, but this is near the lowest resolution at which the shape can still be accurately found. The stink bugs tend to be very small, generally less than an inch, so images taken close to the stink bug or with very high resolution are required to achieve enough pixels over the insect.

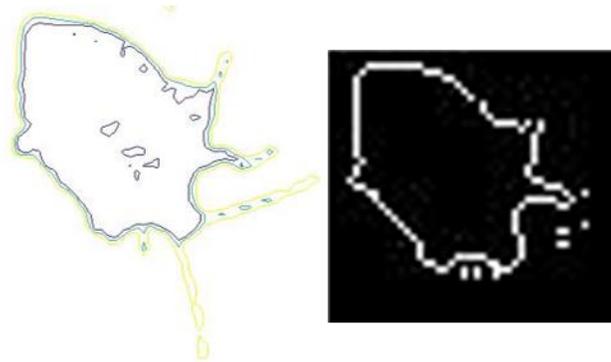


Figure 5.4: Contour and edges extracted from a resized version of the stink bug in Figure 5.2. The new image had a resolution of 2700 pixels.

The main issue with these two detection methods is when the stink bug is placed in a background containing many edges. In a real-world setting, such as with the insect on a leaf, many other edges and contours result in increased difficulty for finding the shape of the stink bug. In these cases, the previous, simple methods no longer work effectively. This can be seen in Figure 5.5.

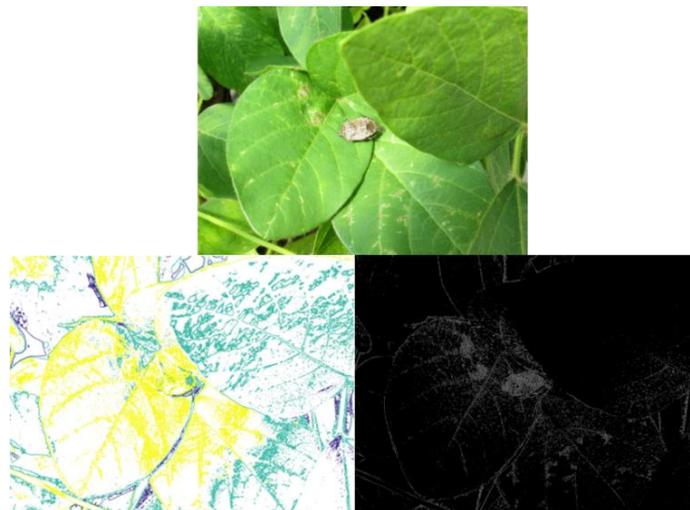


Figure 5.5: Contours and edges for a stink bug on leaves of a plant.

Due to the issues with the previous methods, more sophisticated methods were required. As a result, a more sophisticated edge detection method was looked at. This was a structured edge detection method [45, 46]. This was performed on some images of stink bugs and the results are shown in Figures 5.6 and 5.7. This extracted the edges, which would allow the shape of the stink bugs to be matched. Again, high resolution is required to achieve the accuracy needed.



Figure 5.6: Structured edge detection performed on an image with a stink bug on a plant.



Figure 5.7: Structured edge detection performed on an image with a stink bug on a plant.

5.3 Superpixel Segmentation

In an attempt to reduce the resolution required for the stink bug detection, a superpixel segmentation method was investigated. To segment the image into superpixels, the SLIC Superpixels method was used. The SLIC Superpixels method was implemented using an existing version, which is publicly available [47]. Adult brown marmorated stink bugs have consistent

sizes, so this size can be used for the target superpixel size. With knowledge of the height at which an image was taken and the resolution of the image, a number of superpixels can be selected based upon the expected insect size. After an appropriate number of superpixels is selected, the image can be segmented into the superpixels using the SLIC superpixels method. When an appropriate amount of superpixels was selected, the segmentation effectively grouped the individual stink bugs in the image into individual superpixels. Examples of this can be seen in Figure 5.8. The image on the top left shows an image segmented into 300 superpixels and the top right image is one that has been segmented into 600 superpixels. The images on the bottom show zoomed-in versions of the images above them, which focus on the superpixels containing the stink bugs.

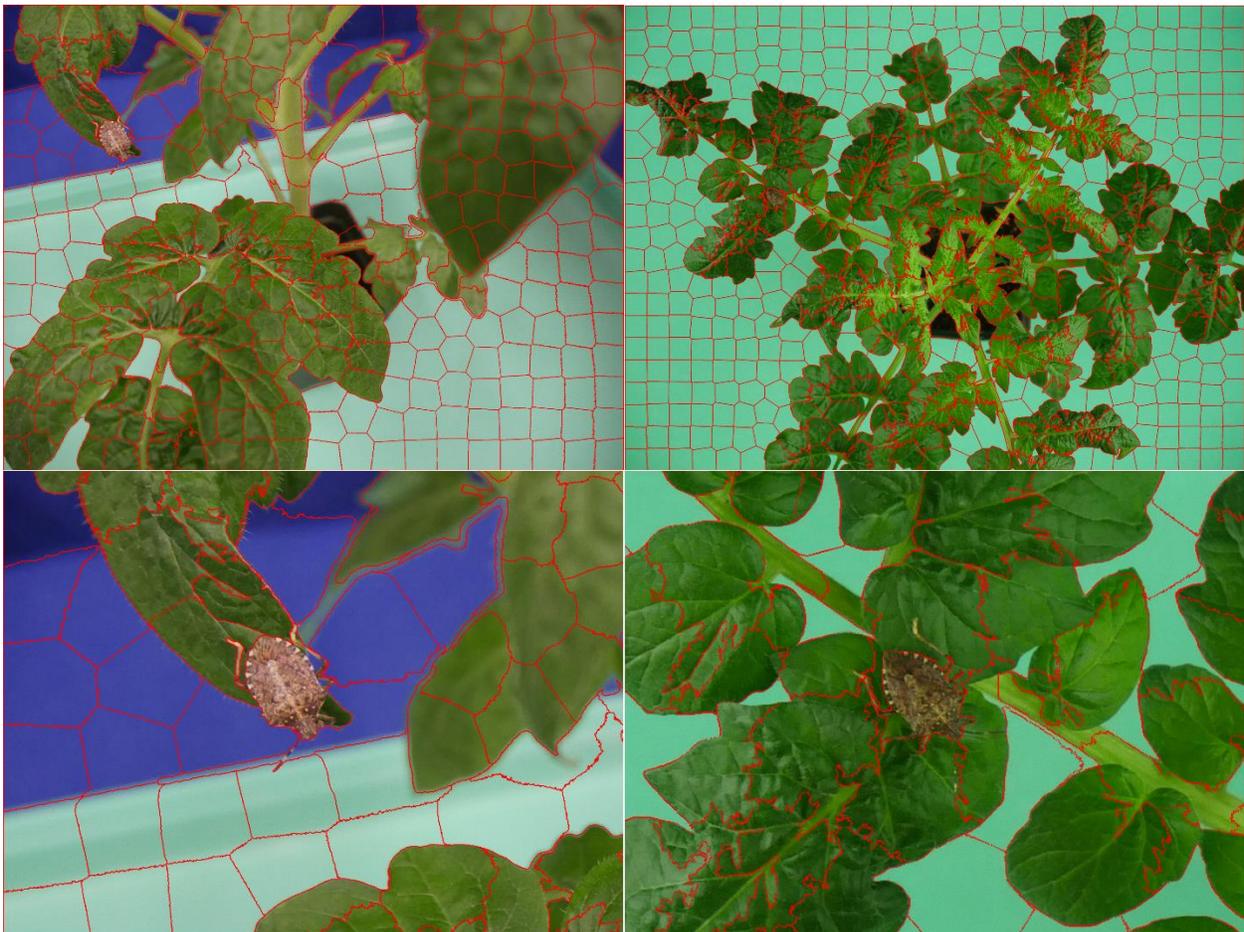


Figure 5.8: Superpixel segmentation results for two images. The red lines indicate superpixel borders. The bottom images zoom in on the stink bugs from the top images.

Once an image has been segmented into superpixels, the next step is to determine which of the superpixels contain a stink bug. The superpixels can be filtered based upon whether they meet

specific criteria, which will eliminate the superpixels which do not contain a stink bug. The first methods of filtering were based upon the color contents of the superpixels. Specifically, the hue, red, and a^* color channels of the image were used for the filtering process. These channels were selected to be used for the filtering because they were found to have large distinctions between the pixel values of the stink bugs and the values for the surrounding leaves. These distinctions are demonstrated by the images in Figure 5.9, which shows an image of a stink bug on a plant in the different color spaces. The image on the top left shows the RGB image of the stink bug on the plant. The top right image shows a color-scaled representation of the red plane image values. The bottom left image shows a scaled color representation of the hue values of the image and the bottom right image shows a scaled color representation of the a^* plane. These planes, especially the hue and a^* planes show very distinct differences between the stink bugs and the surrounding leaves.

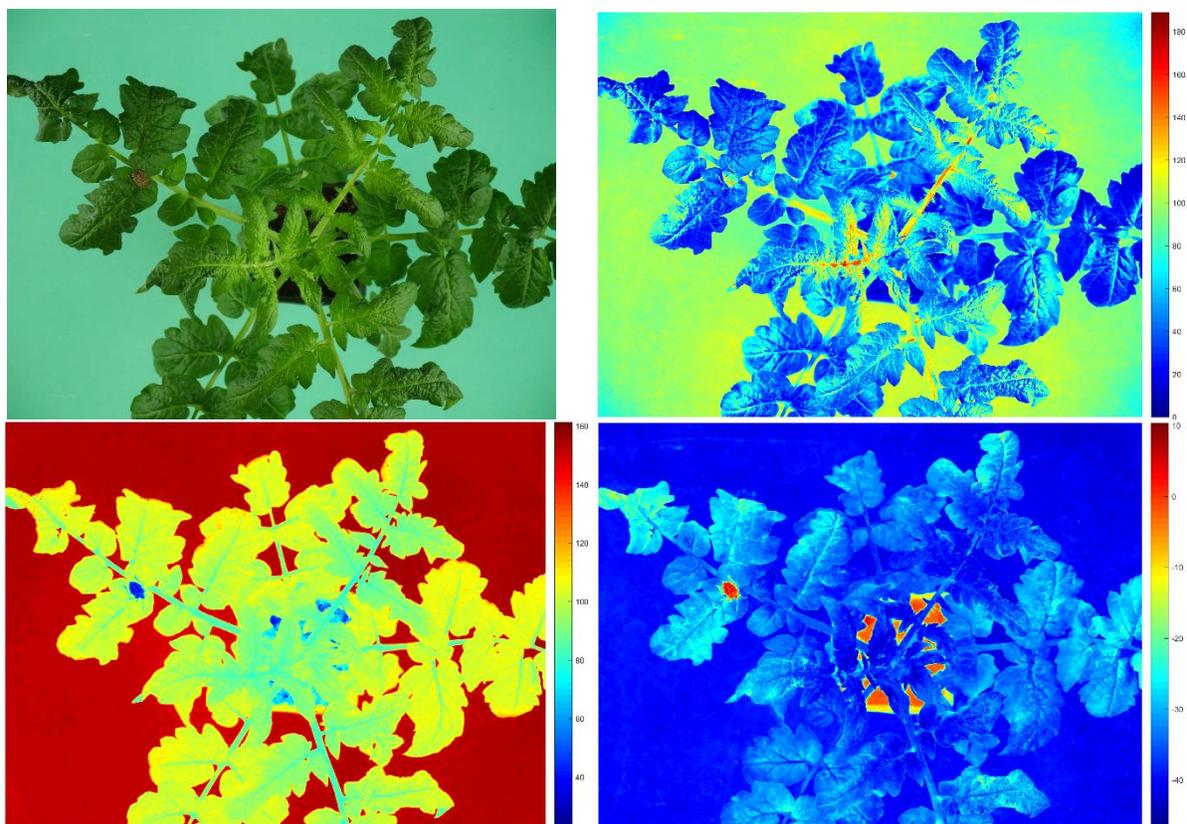


Figure 5.9: RGB image of a stink bug on a plant along with scaled representations of the red, hue, and a^* planes. The top right image shows the red values, the bottom left image shows the hue, and the bottom right images shows the a^* values.

The first step of the filtering process was to check the hue, a^* , and red values of each of the superpixels. To determine a single pixel value for each of the superpixels, a method invariant to outliers should be used. As a result, the mean of the pixels would not be a good choice. The median is a better method than the mean, however even better options exist. The chosen approach is to look at the histogram of the pixel values in the superpixel and use the largest peak of the histogram as the representative value. By using this method, a value representing a portion of the stink bug will be selected as long as the stink bug takes up a majority of the superpixel. A representation of this method is shown in Figure 5.10. The superpixel containing the stink bug is shown in the left image, with all other superpixels being set to black. The histogram on the right shows the hue values of all of the pixels which comprise the stink bug superpixel. The largest peak can be seen to be at a low hue value, around 20, which corresponds to the stink bug. Smaller peaks can be seen at larger hue values, corresponding to the green leaf portions and the blue portion of the superpixel. The histogram peak method is invariant to the higher peaks and, as a result, finds a value which solely represents the actual stink bug.

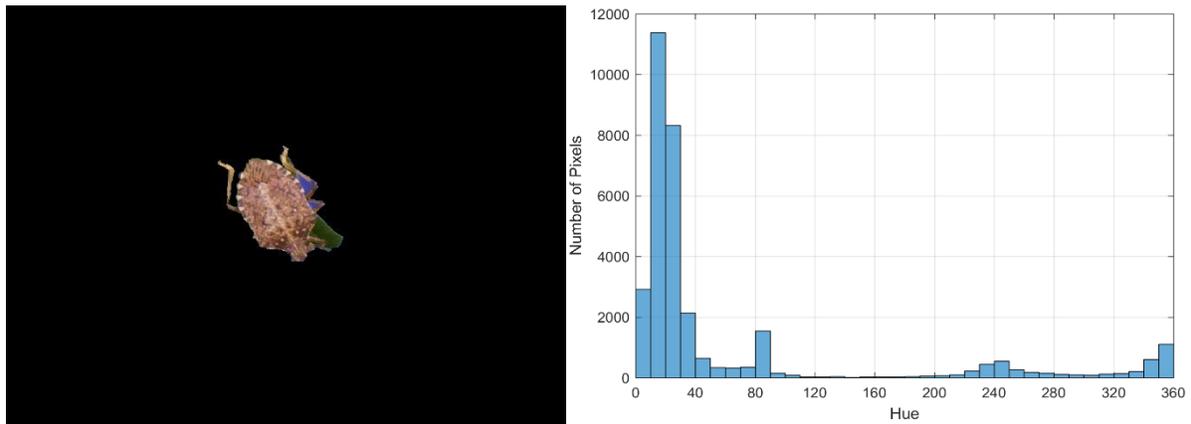


Figure 5.10: Superpixel of a stink bug and the corresponding histogram of hue values. The largest peak will be selected to represent the superpixel.

Once a representative value for each of the superpixels was determined, it could be compared to threshold values in order to filter out many of the superpixels. The threshold values were selected through investigating the values of many different superpixels containing stink bugs. For the hue, the selected threshold was between 10 and 56. The selected threshold for the red image plane was the range between 50 and 180. For the a^* color plane, the selected threshold was between -15 and 20. Every superpixel which meets all three threshold criteria remains as a potential

candidate for a stink bug while all other superpixels are classified as not being a stink bug and are eliminated.

After the superpixels have been filtered through the color thresholds, the superpixels get filtered by their shape and size. One of the shape characteristics examined is the eccentricity of the superpixel. Eccentricity is a measure of the amount that a conic section deviates from being circular. A circle has an eccentricity of 0 and an ellipse has an eccentricity greater than 0 but less than 1 [48]. The shapes of stink bugs, and as a result, the corresponding superpixels containing stink bugs, were found to generally be an elliptical shape with an eccentricity of less than 0.83. This eccentricity threshold was used for the superpixels to filter out inappropriately shaped pixels. Additionally, the size of the superpixels was taken into account by examining that number of pixels in the superpixels. Using the number of superpixels, the target superpixel size can be calculated from the image resolution. The target size can be compared to the actual size of the superpixel to check if the superpixel corresponds to a size which is appropriate for a stink bug. During the filtering process, superpixels which are much larger, determined to be greater than 1.75 times the size of the target size, are classified as not being a stink bug. The final method of filtering is by examining another characteristic of the stink bug shape. This method aims at removing very oddly shaped superpixels. This is done by comparing the size of the bounding box of the superpixel to the filled area of the superpixel. This method is effective for eliminated superpixels with very odd shapes, which likely correspond to a pixel without a stink bug. An example of such a superpixel can be seen in Figure 5.11. The very thin and long portions of the superpixel create a wide bounding box with only a small percentage of the bounding box containing the superpixel. The threshold of the ratio of the number of pixels in the bounding box to the number of pixels in the superpixel was set to a value of 4. All superpixels which have a ratio less than 4 remain as potential stink bug candidates, while all other superpixels will be classified as not containing a stink bug.

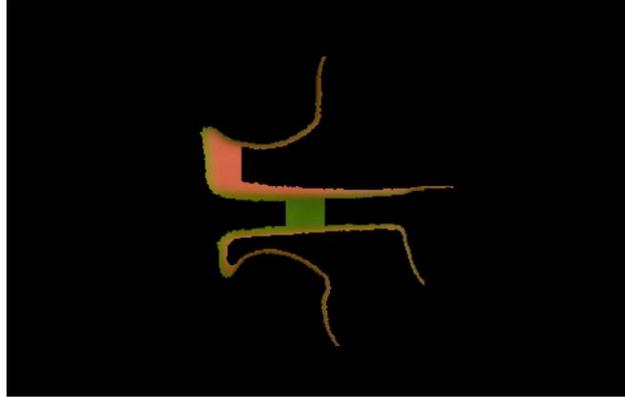


Figure 5.11: Superpixel with a very odd shape, which will be filtered out with the bounding box method.

The superpixel segmentation method worked very effectively at detecting stink bugs, even when the stink bug has a low resolution. An example of this can be seen in Figure 5.12 with a stink bug detected with only 666 pixels in the superpixel. The left image shows the original image and the image on the right shows the detected stink bug. Despite the low resolution on the stink bug, the detection method was able to accurately detect the presence of the stink bug in the image.



Figure 5.12: Stink bug detection result using the superpixel method. The detected stink bug is shown on the right.

In addition to low resolution cases, this method also detected stink bugs which were positioned at angles as well as ones which were partially obscured by leaves. These detection cases can be seen in Figure 5.13. The top images show the detection of a stink bug, which is partially covered by a leaf and the bottom pair of images shows a detected stink bug facing perpendicular to the camera. Despite these conditions, the superpixels allowed the pixels representing the stink bug to be grouped together and the color and shape characteristics can be matched up with values corresponding to a stink bug in order to allow detection.



Figure 5.13: Superpixel stink bug detection results. The top shows a stink bug detected when partially obscured by a leaf and the bottom shows a stink bug detected when perpendicular to the camera.

5.4 Multispectral

An additional investigation was made into the usefulness of multispectral imagery for the stink bug detection process. This investigation was performed through the use of the three cameras discussed earlier, which were visual, near-infrared, and hyperspectral cameras. The visual and near-infrared cameras could be combined to investigate the NDVI of the images and the hyperspectral imagery can be used to see the spectral signatures of the stink bugs. Investigating the NDVI produced results showing significant differences between the values for vegetation and for the stink bug. However, the NDVI values for the stink bug and the soil were very similar. These differences between vegetation and stink bugs can be found through the use of simpler methods with only visual images, so the use of NDVI was not pursued further.

The hyperspectral images of the stink bugs did not produce any significantly distinct spectral characteristics. Across the wavelength range of our hyperspectral camera, the reflectance values for the stink bug produced a generally flat spectral reflectance signature. The overall shape of the reflectance curve across the spectrum of the camera was very similar to the shape of the reflectance curve of soil. The only difference between these curves were that the stink bug produced slightly higher reflectance values than the soil, which can be seen in Figure 5.14.

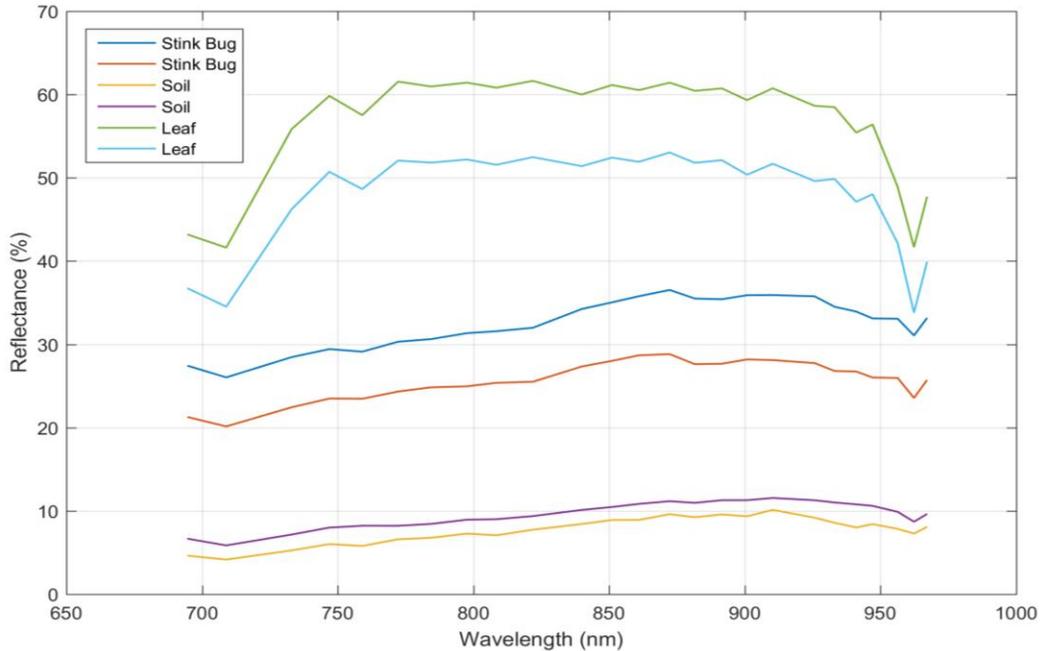


Figure 5.14: Reflectance curves for selected areas of vegetation (leaf), soil, and stink bugs.

5.5 Additional Method

While the superpixel segmentation method worked effectively for detecting the stink bugs, the algorithm is very slow. As a result of the slow execution speed, a faster alternative was desired and this led to the creation of an additional method for detecting the stink bugs. The new method uses color, texture, size, and shape to locate the stink bugs in the image. The first filtering method used on the image is based on the color of the image. The color filter is based upon the hue of the pixels in the image. The hue values corresponding to green are eliminated to filter out the plants from the image. Additionally, the hue values corresponding with shades of blue are eliminated since these also do not correspond with the stink bugs. The filtering eliminates pixels in the image with hue values between 71 and 250 and keeps all other pixels. An example image from this hue filtering process is shown below in Figure 5.15. This image contains two stink bugs, which are located near the vertical center of the image. The hue filtering removes much of the vegetation in the image due to the green color.



Figure 5.15: Result of filtering an image by hue. The hue filtering removes most of the vegetation in the image while the stink bugs remain.

The next filtering step uses the texture of the image by finding the entropy. This entropy calculation was performed on the image after the hue filtering was performed and used a nine by nine window for the calculation. For the entropy, the stink bugs resulted in high values due to the texture which is present on the back of the stink bugs, while areas of soil resulted in much lower entropy values. The entropy values for the filtered image from Figure 5.15 can be seen in the image on the left in Figure 5.16. The stink bugs can be seen in the scaled color image to be red, which corresponds to high entropy values. From many different images, a threshold value of 5 was selected for the filtering. Unlike the previous filtering step, this was not performed on a pixel by pixel basis, but rather by groups. The binary mask of the image created from the result of the hue filtering was used to collect all groups of remaining pixels, which have binary values of 1. The median value of the entropy of all of the pixels in each of these groups was calculated and this was then compared to the threshold. All of the regions with sufficient entropy values were kept as potential candidates for being stink bugs. The image result after this processing is shown in the image on the right in Figure 5.16. At this point, the amount of possible stink bugs in the image has been reduced significantly and the stink bugs can be seen to remain in this example image.

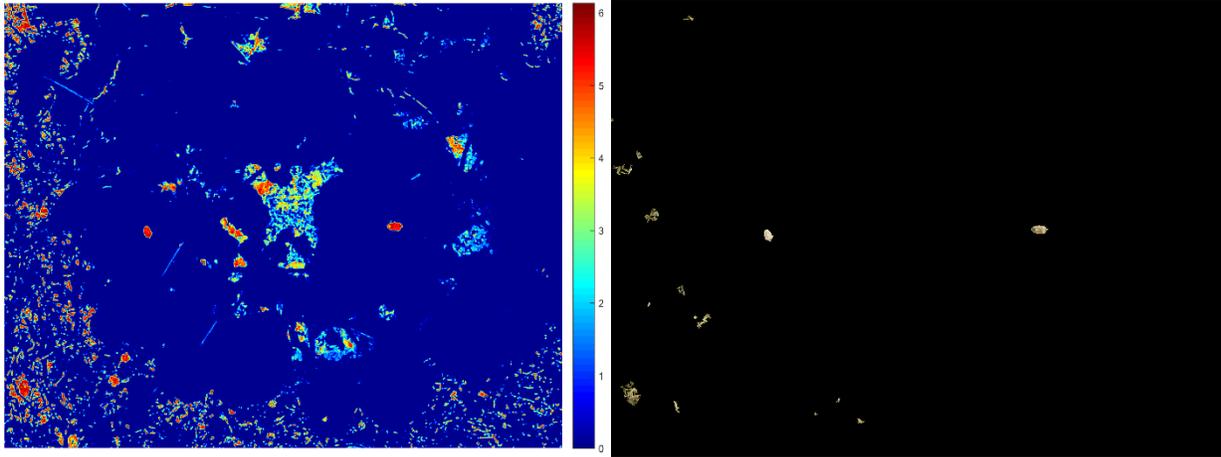


Figure 5.16: Entropy filtering of the resulting image from Figure 5.15. The image on the left shows the scaled color representation of the entropy values and the image on the right shows the image result after the entropy filtering process.

The next step in the filtering process is to look at the shape and size characteristics of the binary blobs which remain in the image. A binary closing operation is first performed on the image mask to remove the small holes in the blobs in the image, then shape and size characteristics are examined. The image is filtered based upon the ratio of the area to the perimeter of each of the binary regions. All regions which are greater than a threshold remain in the binary mask as potential stink bugs. This step can remove very oddly shaped regions, with large perimeters and small areas, as well as many small regions in the image.

The size of the regions in the image are examined to determine if they match well with the size of the stink bug. This portion of the algorithm requires the knowledge of the height, relative to the crop, at which the image was taken. The size of stink bugs tends to be relatively uniform, with the length of an adult being around 17 mm or 0.67 inches [49]. As a result, regions in the image which are larger than the size corresponding to an average stink bug can be removed. To determine this size, images were acquired of a large stink bug at various heights and these heights were recorded along with the amount of pixels contained in the stink bug and the image resolution. Using this information, the ratio between the number of pixels in the stink bug and the number of pixels of the image was plotted against the height at which the image was taken to determine the trend of the data. The plot of this data is shown below in Figure 5.17. A curve was fit to this data and the best curve was the power function in Equation 5.1.

$$Size = 0.1071H^{-1.878} \quad (5.1)$$

The size in this equation is the ratio of the number of pixels for the stink bug to the number of pixels in the image and the variable H represents the height at which the image was taken, in inches, relative to the crop or search area for the stink bug. This curve has an R^2 value of 0.9989 and it is displayed as the dotted line on the figure. The size function is used as the maximum size of a stink bug, so all regions in the image larger than this determined ratio can be eliminated as potential stink bug candidates.

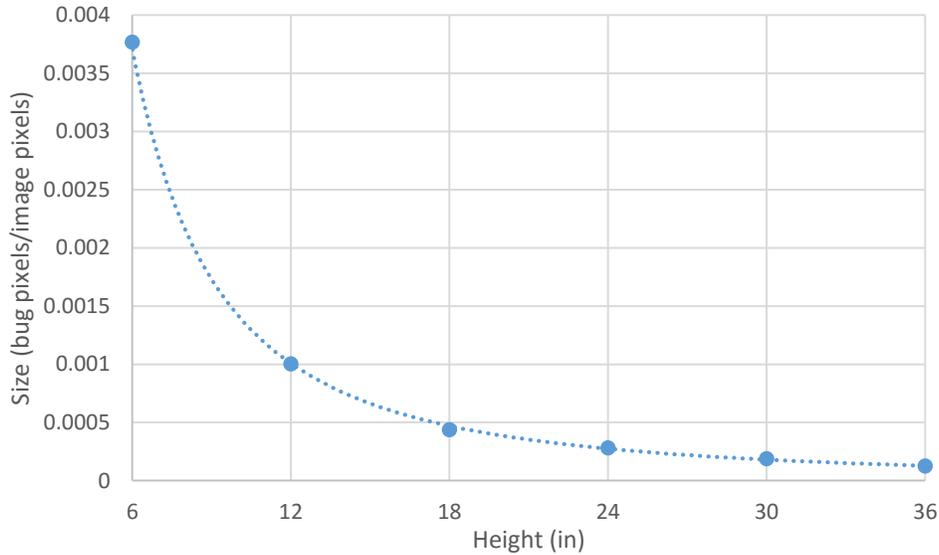


Figure 5.17: Plot of the stink bug size against the height at which the image was acquired. A curve was fit to the trend of the data.

The final portion of the filtering process examines shape characteristics, particularly the circularity of the regions. The shapes of stink bugs are very similar and have an oval shape to them. To examine the shape, the eccentricity as well as circularity of the regions are found. The eccentricity of a shape is a measure of how much it deviates from the shape of a circle. A value of 0 corresponds to a circle and an ellipse will have a value between 0 and 1. Through testing with the shapes of many stink bugs, a range of values was selected as ones that correspond to most stink bugs. This range of values is between 0.68 and 0.92, so all regions with values in this range remain. The final property is a measure of circularity based on the isoperimetric quotient [50]. The formula used for this is shown in Equation 5.2, with P representing the perimeter of the region and A representing the area of the region.

$$C = \frac{P^2}{4\pi A} \quad (5.2)$$

This measure of circularity produces a value of 1 for a circle and increases to infinity. A threshold value of 2.5 was selected, so all regions with a circularity less than this threshold could be a stink bug.

The final result for the image in Figure 5.15 is shown in Figure 5.18. The top image shows the whole image with only the two stink bugs remaining and all other portions turned to black. The bottom two images show zoomed in versions of the two stink bugs which were detected by this algorithm.



Figure 5.18: Final detection result of the image from Figure 5.15. The top image shows the detected results and the bottom two images show zoomed-in version of the detected stink bugs.

5.6 Results

Two potential methods for detection of the stink bugs have been proposed. Each of these methods successfully detected stink bugs in various settings, but each have some disadvantages.

For the method using the superpixel segmentation, in an image set containing 33 stink bugs, 31 of those were successfully detected, which is a recognition rate of 93.9%. In these cases, there were also two false positives detected. The detection was able to work in cases with very low stink bug pixel resolution, as low as 666 pixels for a bug. The biggest problem with this method is the processing time. The processing required around 30 seconds for images very close to the stink bug, which require only 300 superpixels. In images much higher up, for instance one requiring 4000 superpixels, the processing required as much as 3 minutes to complete. However, this algorithm was not optimized and could likely be speed up substantially through the use of another coding language, such as C instead of Matlab.

The second method performed well on a small test set of images and did the processing much faster than the first method. In a set of 11 images which contained 21 stink bugs, the algorithm correctly detected 20, 95.2%, of these stink bugs, but did also have one image with 2 false positives. The algorithm processed a 16-megapixel image in approximately 17 seconds. Most of the processing time is spent on the entropy filtering portion. This algorithm is written in Matlab and the entropy filtering portion utilizes a built-in function that runs using a MEX file. The MEX file is written in C, so it would likely be difficult to achieve large improvements in processing speed by writing the code in a different language, such as C. This method has some potential failure cases when the images do not achieve required resolution. The entropy method requires high resolution on the back of the stink bug to detect the texture on the back of the stink bug. The one failed detection in the dataset was a stink bug that was not well focused in the image, so texture could not be detected. The test of the code worked up to a height of 36 inches and the smallest detected stink bug had a resolution of 1137 pixels across the stink bug. Other potential issues with this algorithm contain cases where stink bugs are obscured by the leaves or sideways at odd angles. If a stink bug is oriented in a way that the texture of it cannot be picked up by the camera, it will likely not be detected and if the stink bug is partially or completely obscured by leaves, it will also not be detected.

Chapter 6 Summary & Conclusions

The current methods for monitoring crop health and detecting issues in crop fields are lacking in many areas, such as the cost and turnaround times. Satellite or airplane remote sensing can be very expensive and very slow to detect the issues, which can also limit the treatment possibilities. A potential improvement on these current methods is through the use of UAVs for remote sensing. This thesis has presented methods for UAV remote sensing using visible spectrum and multispectral images. A method has been proposed to use an SVM for the detection of crop issues from the use of airborne images of corn fields. The created SVM achieved a detection accuracy of 86.4% over a test set of images. Additionally, multispectral analysis was performed to determine indices appropriate for determining the health of corn. Along the way of this process, a method for correcting hyperspectral images for shadows and other lighting issues was created. Several indices were created and the best performance was achieved using 760 and 740 nm wavelengths, which produced a correlation coefficient of 0.84 with the corn yield. In addition to this, stink bug detection algorithms were also created to detect stink bugs on crops using aerial visible spectrum images. Two methods for this were created, which include a superpixel segmentation method and a color and texture filtering method. The superpixel method achieved a recognition rate of 93.9%, but had high processing times in some cases. The second method achieved a recognition rate of 95.2%, while improving upon the speed of the first method.

For each of the algorithms which have been presented, there are potential areas of improvement and future work. For the crop health detection using only visible spectrum imagery, which was the work in Chapter 3, the biggest area for future work is the addition of features outside of color, such as texture. Currently, this work is limited by the training data lacking the resolution necessary for texture features. Training data would need to be acquired by flying UAVs over fields to acquire high resolution imagery of the fields. In addition to this, the current training data lacks ground truth data for specific issues. The current data is labeled for areas which are known to have crop health issues, but the ground truth data does not exist to identify the specific issues. If this data can also be acquired, the machine learning algorithm could be extended to detect specific health issues, such as water stress, nutrient deficiency, or specific abiotic or biotic stress impacts, which could include stink bugs and nematodes. Another possibility to examine for the

machine learning approach is the use of superpixels for the classification in order to reduce the required processing as well as reduce the impacts of noisy pixels. Additional improvements to the image enhancement method could be made by using a localized adjustment approach to reduce the impacts of localized shadows in the images.

Future work for the multispectral crop analysis would include another repeated test of the index study to validate the determined indices. Additionally, it would be useful to examine index values throughout the crop growth stages to see how the indices vary from early stage growth to late stage crops. This additional data would allow for further validation and the potential to improve upon the indices. Additional improvements in the system could be obtained through the use of a light sensor to determine the lighting conditions during the image acquisition. This study did not include any way to measure the lighting changes, outside of watching the sky to notice when clouds obscure the sun. The addition of lighting condition values would allow for more robust results, which can be more invariant to changes in lighting conditions.

For the stink bug detection, additional testing is required to determine the performance of these algorithms in a real crop field. The images used to test the algorithms were acquired from a stationary person taking an image of a stink bug on a plant, rather than from a UAV flying over a field. To better evaluate the performance of these algorithms, images should be acquired by flying a UAV over a field of crops to obtain realistic testing images for the algorithms. Additionally, my implementations of these algorithms have not been optimized for speed, so reducing the processing time will be an important step for implementing these in practice.

From the work in this thesis, the most significant contribution, with the most potential for future application, is the work from Chapters 3 and 5. The work from each of these sections has significant potential to be improved upon and implemented in practice by farmers. The combination of the methods presented in these two chapters could allow for the detection and prevention of many issues in a field. The work presented here creates a foundation for a method to allow a farmer to use a UAV to automatically scout a field for the presence of bugs, specifically stink bugs from this work, as well as issues with the field. The detection of these issues would allow the farmer to treat the appropriate areas of the field to reduce the impact of these issues and improve upon the yield of the crop. Additional development of these methods would be required to achieve that goal, but this potential exists.

The work presented in this thesis can hopefully be built upon for the benefit of farmers to analyze the conditions of their fields and improve the health and yield of their crops with proper treatments. The use of UAVs in the area of remote sensing has a large amount of potential to overtake other methods for remote sensing in agriculture, which can allow for more widespread use of remote sensing by average farmers.

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